Niche modelling
Biological control and climate change

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Research Article
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**Assessing the current and future biological control potential of
Trichogramma ostrinae on its hosts *Ostrinia furnacalis* and *Ostrinia
nubilalis***

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Abstract

BACKGROUND: Understanding interactions between biocontrol agents and their pest hosts under climate change should assist implementation of biocontrol strategies, by identifying appropriate biocontrol agents for release or determining the optimal timing of releases. Species Distribution Models (SDMs) are applied to evaluate the distributions of *Trichogramma ostrinae* and its native host, *Ostrinia furnacalis*, in southeastern Asia, and a non-native host, *Ostrinia nubilalis*, in a novel range, North America, using MAXENT and CLIMEX modelling approaches.

RESULTS: Both models led to similar predictions about the expected distribution of the two species in Asia, and emphasized likely mismatches between host and natural enemy. *Trichogramma ostrinae* was predicted to occur in the summer corn region of China, with distribution limits linked to its sensitivity to cold, seasonality of radiation and precipitation. Modelled *Ostrinia nubilalis* distribution overlaps with the main corn production areas of northeastern United States and Canada; temporary/seasonal suitable habitat was predicted across southeastern states as well. Climate change scenarios are predicted to favor *T. ostrinae* over its hosts in northeastern China and North America.

CONCLUSIONS: The modelling approaches used here proved useful for assessing environmental factors linked to an egg parasitoid and its lepidopteran hosts and identifying areas potentially suitable for inundative releases.

Keywords: biocontrol, CLIMEX, species distribution model, egg parasitoids, environmental niche model, MAXENT

1 INTRODUCTION

Weather extremes driven by climate change are set to challenge crop production, with the potential to destabilize agricultural ecosystems and threaten local to global food security.¹ To maintain sustainable pest control in crop production systems, an understanding of the extent to which climate change will affect both pests and their natural enemies (including predators and parasitoids) is needed. Climate change is expected to drive population asynchrony or coextinction of hosts, parasites and mutualists,² highlighting that increased research attention should be focused on these important ecological relationships. There is a range of approaches for examining effects of climate change on these interactions,³ particularly for changes in the distributions and abundances of host and parasitoid.⁴

To make predictions of species distributions, different types of niche models are increasingly used both in biodiversity conservation and in applied ecology.^{5,6,7} The choice of which type of model to use is typically determined by the type data that is available for the species.⁸ A popular form of model is the environmental niche model

(ENM) such as MAXENT, that aims to approximate the realized niche of species by characterising species-environment relationships using species occurrence data and environmental covariates. The realized niche inherently includes not only abiotic constraints but also other implicit factors, such as biotic interactions including competition and host-availability, as well as the ability of species to disperse through the environment.⁷ Correlative models are thus useful for inferring species-environment relationships and examining response of the species across its range to specific predictors to better understand distribution limits.^{9,10} While ENMs provide important tools to infer species-environment relationships and limits to distributions, there are issues of transferability when extrapolating these models in time and space. Species distributions have also commonly been analyzed with CLIMEX, a semi-mechanistic niche modelling tool, adopting a combination of life history parameters with abundance and distribution records. It is broadly used to investigate the interactions between biological invasions, potential response and efficiencies to climate change.^{4,10,11} By constructing niche models grounded on species ecophysiological traits, CLIMEX avoids some issues of extrapolation or transferability across regions and future climate change scenarios.¹⁰ CLIMEX

determines an ecoclimatic index reflecting suitability of a location through a combination of growth and stress parameters.¹²

One area where CLIMEX models are useful is for predicting the distribution of classical biocontrol agents. CLIMEX has been used to identify American regions most climatically similar to native localities (southern Africa) of biocontrol agents, and thus efficiently narrowed the search range for natural enemies of two alien pest plants of significant invasion concern worldwide, *Chromolaena odorata* and *Acacia nilotica* spp.^{13,14} Further analyses of this nature should provide valuable information for classical biocontrol strategies, by identifying regions where natural enemies may become mismatched with their hosts, and by identifying periods of time when inundative releases with parasitoids and predators are likely to be effective.¹⁵

An important group of natural enemies are the *Trichogramma* (Hymenoptera: Trichogrammatidae) wasps, small egg parasitoids that help control a range of lepidopteran pests. *Trichogramma* spp. are often cultured and released in inundative biocontrol programs to successfully aid agricultural production.¹⁶ In comparison to

other insect guilds, parasitoids are **more** likely to be sensitive to climate change¹⁷ and other environmental disturbances because of their position in the trophic web, and because of host specialization (e.g. host generalists responded faster with larger expansion/host shift ranges within habitats).^{18,19} To this end, parasitoids will not only be affected by the direct effects of climate change, but also by indirect effects mediated through changes in host population dynamics.^{17,20}

The Asian corn borer (ACB), *Ostrinia furnacalis* Guenée (1854), is a serious pest of corn and causes more than 9 million tons of yield loss per year.²¹ The geographic range of ACB extends from eastern and southeastern Asia, through to western Pacific islands.²² The endemic egg parasitoid, *Trichogramma ostrinae* Pang & Chen (1974), is the predominant species parasitizing ACB across the native range, which consists of most corn cropping regions of southeastern Asia.^{21,23} In China, *Trichogramma* biocontrol for ACB has been conducted since the early 1950s; today nearly four million hectares of cornfields are treated with *T. ostrinae* and other *Trichogramma* species across southeastern Asia annually.²¹ Surveys of both *T. ostrinae* and ACB in the native range provide an opportunity to model the current distribution and

investigate potential distributions of the host and this parasitoid under climate change scenarios across southeastern Asia.

Because of its success in controlling ACB in China, *T. ostriniae* has been introduced to control the closely related European corn borer (ECB) *Ostrinia nubilalis* (Hübner 1917) in northeastern USA (New York, Massachusetts, Delaware and other locations) and Canada (southern Quebec) since the early 1990s.^{24,25,26} The ECB is endemic to Europe and western Asia and was introduced to northwestern America in the early 19th century. Early inoculative releases of *T. ostriniae* showed promising results, with up to 80% of parasitism achieved in cornfields in this region.²⁷ Inoculative releases are currently available as an option to help control European corn borer in sweet corn in New York, Kentucky and other States in northeastern US.²⁸ Given the status of these important lepidopteran pests and the active *Trichogramma* biocontrol programs in place, these data provide a unique opportunity to test projections of niche models within and between continents, and assess the future effectiveness of biocontrol under climate change scenarios.

In this current study we employ niche modelling to determine climatic factors associated with two lepidopteran host species and the biocontrol parasitoid, and examine potential changes in their distributions due to climate change. The specific aims of the current study are: (1) to analyze and compare species-environment relationships and niche overlap of *T. ostriniae* and its host moth ACB across their native distribution in Asia; (2) to better understand the climate suitability for *T. ostriniae* in North America for control of ECB; and (3) to examine how host-parasitoid relationships of ACB in Asia and ECB in North America may be altered under climate change. Determining how broad-scale climate patterns may change host-parasitoid relationships should help plan future biological control programs.

2 MATERIALS AND METHODS

As we had data from different knowledge domains, including field observations of distribution and phenology, physiological response information, and models of diapause, we constructed different models where appropriate.

2.1 Distribution data

Occurrence data were obtained from published references (see Appendix Table S1) ($n = 111$ for *Trichogramma ostrinae*, $n = 149$ for *Ostrinia furnacalis* (ACB)), field surveys around southwestern Taiwan in 2014 and 2015 ($n = 9$ for *T. ostrinae*, $n = 28$ for ACB, Wu *et al.*²³ and Hsieh, 2016, pers. comm.) and the Global Biodiversity Information Facility (GBIF) database ($n = 404$ unique points for *O. nubilalis* (ECB)) (Fig. 1). *Trichogramma ostrinae* is not currently used in Europe as a biological control agent for ECB. The North American distribution of *T. ostrinae* listed (Table S1) was not used in model training (see below) as these data were obtained from successful inundative releases in biological control programs, and it is not clear if permanent populations have established at these release points.²⁹ Instead, these points were used to test whether the predictions from the models matched those where inundative releases have taken place.²⁹

2.2 Environmental data

We obtained environmental data from CliMond,³⁰ which provides data formats

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suitable for both correlative ENMs and CLIMEX models. For ENM model construction, CliMond provides 35 bioclimatic variables describing means, seasonality and trends for temperature, precipitation, solar radiation and soil moisture. Data for CLIMEX uses the same observations, but uses daily minimum and maximum temperatures, monthly precipitation and relative humidity. We used a grid cell resolution of 10' for both approaches. The model was run for the equator.

2.3 ENM construction

For the construction of correlative distribution models for *Trichogramma ostrinae* and its host *Ostrinia furnacalis* (Fig. 2), we ran MAXENT (Version 3.3.3i)³¹ which uses the principle of maximum entropy to relate distribution records of a species to environmental variables for estimating the potential distribution of the targeted organism.^{31,32} MAXENT uses information on conditions in the region of interest as a basis for comparison with conditions at known presence sites, so the regions ("background") need to be defined before model construction. We first set the background for the native range to occupied bioclimatic regions from Köppen-Geiger

classifications within (or intersecting) the borders of China, Japan, North Korea, South Korea and Taiwan, because all our data points were contributed by references published from these areas (see Figure 1). Köppen-Geiger classifications followed the rules defined in Kriticos *et al.*³⁰ as applied to the 10 arc-minute resolution CLIMAT dataset (sourced from the WORLDCLIM and Climate Research Unit (CRU) datasets, <http://worldclim.org>, Version 1.4, release 2013³³; <https://crudata.uea.ac.uk/cru/data/hrg/>, Version CL2.0³⁴). This provides a broad geographic area to account for data availability and occurrence record biases, but omits large geographic areas that are not relevant to modelling species distributions.³⁵

To determine key variables for model inclusion, we applied Environmental Niche Factor Analysis (ENFA)³⁶ for the two species datasets independently across their respective backgrounds using the ‘adehabitat’ package³⁷ in R (version 3.1.1).³⁸ Variables were ranked on marginality, which describes the difference between the total range of environmental variables and the range occupied by the species, within the accessible area.³⁶ The top 10 variables for both species were then checked for spatial autocorrelation. We generated 10000 random points across each background,

extracted variable information for each of the environmental predictors, and conducted pairwise Pearson correlation tests across all variables. For variables that were highly correlated ($r > 0.75$), a variable from the pair was removed based on a lower marginality score.³⁹ By removing variables in this manner, we could reduce issues that may be caused by multicollinearity, and its bias on models projected to novel regions.⁴⁰ To enforce smooth response curves, we only enabled hinge features (see Elith *et al.*⁹; Hill & Terblanche⁴¹), all other parameters were left at default. Models were then run using 10 cross-validation replicates, taking the average of all replicates for spatial predictions. The respective predictor variable sets were then examined using the jackknife feature of MAXENT together with percentage contribution to the model.

To test for any changes to model performance from possible sampling error, we also created a buffer zone of 20 km around each location assigned from literature records and randomly jittered locations (see Hill *et al.*⁴²). The resulting mapped predictions were compared through niche overlap analysis (Schoener's D statistics) using the 'dismo' package⁴³ in R. We then performed a one-tailed t test to determine whether

the actual data set produced models different to those with random error incorporated (Appendix S1).

2.4 Environmental Space

In addition to modelling the distributions of all species, we examined overlap in multivariate environmental conditions across the different geographic regions. We extracted predictor information for a common predictor variable set that included all important variables across the distribution of all species. We then sampled 10000 random points across the Asia and North America backgrounds and extracted the same climatic information at these points. We performed principal components analysis (PCA) for the total combined data, and plotted the first two-axes to visually examine overlap between the different species, and the background datasets (Appendix S2)⁴¹.

2.5 CLIMEX modelling

The CLIMEX model processes a series of weekly growth and stress indices that are combined to derive an Ecoclimatic Index (EI). The EI indicates regions that are

unsuitable for the species to persist, compared to those that provide an ideal environment for the species. The EI scale ranges 0–100, with 0 representing an unfavourable environment, values over 20 considered to be ideal conditions, between 10 and 20 considered suitable, and between 0.01 and 10 considered marginal,⁴¹ though interpretations of EI are species-specific to some extent.¹⁰ For ACB and *T. ostriniae* CLIMEX models, initial parameter values were provided by a template species (Colorado beetle *Leptinotarsa decemlineata* (Say)), as it is also an important agricultural pest inhabiting similar latitudes, and its default simulation fitted well with known distributions. For ECB, initial parameter values were acquired from previous modelling attempts⁴⁵ and refined using its native European distribution. To fit the CLIMEX model for each of *T. ostriniae*, *O. furnacalis* and *O. nubilalis*, the species' known native distributions together with reported range descriptions were used to iteratively adjust the model until the CLIMEX simulated geographical distribution coincided with its known distribution.

Both species here undergo facultative diapause as later stage larvae to escape suboptimal cold conditions, especially at higher latitudes. Diapause in CLIMEX is

incorporated through a relationship between photoperiod and temperature. The diapause index is an important component in determining the yearly Growth Index, and can be adjusted according to observations from trap captures and field observations for the species. Below we describe where we made parameter changes, and the rationale for each, for the three species under investigation.

2.6 Parameters for *O. furnacalis* (ACB)

2.6.1 Temperature index

Lower developmental threshold **ranges** were 6.8 to 10.4°C for *O. furnacalis*.²² In the current study, the minimum temperature for development (DV0) was set at 6.8°C. The lower and upper optimum temperatures (DV1, **DV2**) were set at 20 **and 28°C, respectively**. The upper temperature threshold was close to 33-35°C, so the upper threshold temperature (DV3) was set at 33°C.

2.6.2 Moisture index

Moisture index values limited the establishment of both species in western China and

across the southeastern mountain areas. The lower soil moisture limit for development (SM0) was set to 0.15 to indicate the lowest moisture threshold points of the egg parasitoid and its lepidopteran host. This limit normally corresponds to approximately 10% of soil moisture level. The lower and upper limits for optimal growth (SM1, SM2 and SM3) were set to levels considered biologically reasonable for current distributions, or 0.35, 0.80 and 1.5, respectively.

2.6.3 Cold stress

For ACB cold stress, temperature threshold (TTCS) and cold stress accumulation rate (THCS) were set to 0.5°C and -0.0001 week⁻¹. EI in northeastern China would be too low if its TTCS parameter had higher values. Furthermore, higher TTCS values would not be consistent with the known distribution of ACB in northeastern China, where it is widespread and causes high damage levels. THCS were adjusted iteratively to fit the distribution boundaries for ACB.

2.6.4 Heat stress

Heat stress was increased by setting the heat stress temperature (TTHS) to 33°C and heat stress accumulation (THHS) to 0.002 week⁻¹, to prevent southern parts of Indochina and other tropical areas becoming suitable.

2.6.5 Diapause index

In its subtropical and temperate geographical range, ACB enters a facultative cold diapause in the late larval stage from late summer to early spring.²² Diapause has been studied in different parts of the species range, including China and Japan. In general, declining photoperiod experienced by the larval stage determines the proportion of the population entering cold diapause, with warm temperatures and photoperiod in spring determining the timing of termination.^{22,46} In China, Guo *et al.*⁴⁷ noted that the critical day lengths for diapause induction of ACB among four geographic populations declined from the north towards the south and the sensitivity to photoperiod also declined from north to south populations. At the same time, the critical day length would be shortened when the rearing temperature increased from 20 to 27°C. We set the diapause induction daylength to 14.88 hours, the induction temperature to 17°C and the termination temperature to 6°C. These values were obtained from Lu and

Zhou⁴⁸ and then iteratively fitting the growth index around population observations from Yang *et al.*⁴⁶.

2.7 Modelling for *O. nubilalis* (ECB) in Europe and North America

2.7.1 Temperature index

The mean lower developmental threshold was 10°C in previous modelling attempts.^{45,49} In the current study, the minimum temperature for development (DV0) was set at 5.5°C to capture the northern distribution points in Scandinavia better; the lower and upper optimum temperatures (DV1, DV2) were set at 28 and 38°C, respectively; in line with parameter estimation from Maiorano *et al.*⁴⁹, we set the upper limit to 41°C.

2.7.2 Moisture index

Moisture index values limited the establishment of *O. nubilalis* in central America and across the mountain areas. The lower soil moisture limit for development (SM0) was set to 0.06 to indicate the lowest moisture threshold points. The lower and upper

limits for optimal growth (SM1, SM2 and SM3) were set to levels considered biologically reasonable for current distributions, which were 0.15, 1.2 and 2, respectively.

2.7.3 *Cold, dry and heat stress*

As the temperature index for *O. nubilalis* alone was able to define the northern (colder) range, adjustments of cold stress parameters were not expected to affect the model outcome and were omitted. Dry stress relates to the moisture index and was set to 0.05 for dry stress threshold (SMDS); -0.04 week^{-1} for dry stress rate (HDS). Heat stress was lowered, by increasing heat stress temperature (TTHS) to 30.5°C and heat stress accumulation (THHS) to 0.01 week^{-1} to prevent southern parts of USA and Mexico becoming suitable. Parameters used in CLIMEX models are listed in Table 1.

2.7.4 *Diapause index*

The diapause induction daylength and temperature were set at 14.5 hour and 12°C in previous modelling attempts.^{45,49} In the current study, the diapause termination temperature was lowered to 10°C , capturing emergence time of ECB adult for

Pontevedra in north-western Spain better.⁵⁰

The CLIMEX model for ECB was compared to previous model results from Trnka *et al.*⁵¹ and Svobodová *et al.*⁵². Because CLIMEX also models the weekly suitability of climate for population growth, it is possible to verify phenology as well as different geographical establishments.³⁰ The modelled growth index phenology (GI_w) was compared to field moth capture numbers from two locations in Spain (Pontevedra & Barrantes) for *O. nubilalis* validation in Europe.⁵⁰ For North America, we tested our model performance with published data points.^{26,53,54}

2.8 Parameters for *T. ostriniae*

2.8.1 Temperature index

Lower developmental thresholds were around 11.7 to 11.8°C for *T. ostriniae*.⁵⁵ In the current study, the minimum temperature for development (DV0) was set at 11.7°C. The lower and upper optimum temperatures (DV1) were set at 22 and 29°C, respectively.²⁹. The upper temperature threshold for both species was close to 33-35°C, so the upper threshold temperature (DV3) was set at 35°C.

2.8.2 Cold stress

For *T. ostriniae* cold stress temperature threshold (TTCS) and cold stress accumulation rate (THCS) were set to 1°C and -0.0001 week⁻¹. THCS thus were adjusted iteratively to fit the distribution boundaries for *T. ostriniae*.

2.9 Host-parasitoid dynamics under climate change in Asia and North America

To explore possible host-parasitoid dynamics in distribution under climate change for ACB and *T. ostriniae* in Asia, and for ECB and *T. ostriniae* in North America, we used the CSIRO Mk. 3.0 models (available from CliMond)³⁰ for the year 2080 under the A1B SRES for future climate change, one of the higher rates of warming in the Fourth Assessment Report (AR4).⁵⁶ We further investigated the impact of the predicted changes by plotting the difference in the GI for both corn borers (ACB/ECB) and the GI for *T. ostriniae* as a proportion of the GI for ACB/ECB in Asia and North America, and presented proportions as a measure of the likely effectiveness of *T. ostriniae* as an inundative biological control agent among those two continents.

3 RESULTS

3.1 Distribution fit and model projections for *O. furnacalis* and *T. ostriniae* in Asia

The predicted native range of ACB overlaps with the main corn production areas that are within northeastern (Heilongjiang, Jilin and Shenyang provinces), eastern (Beijing, Hebei, Henan, Shandong, Shanxi, Jiangsu and Anhui provinces), and southeastern (Guangdong, Guangxi and Yunnan including Hainan provinces) China, Japan and southwestern coastal regions of Taiwan (Figs 2a & 2c). The predicted range of *T. ostriniae* (Figs 2b & 2d) is similar to its host *O. furnacalis*, except for its more constrained northern limits in China and Japan, and there are no presence data for Hainan Island.

Variable importance was similar and a final suite of five variables was common to both species after our selection procedure (Table 2). MAXENT performed well in terms of AUC scores, estimated on held-out cross-validation data within the training areas (Table 2). Varying locations up to 20 km did not significantly change model

predictions (Appendix S1). ‘Mean temperature of wettest quarter (°C)’ (bio08) was the most influential variable for the two species; precipitation and radiation seasonality (bio15 & bio23) were also important, each contributed more than 10% (Table 2). CLIMEX predicted suitable areas of *O. furnacalis* in Jiangsu and Yunnan (Figs. 2a & 2c), whereas MAXENT predicts the constrained distribution of *T. ostriniae* well in Jiangsu, Gansu, Shaanxi and Hainan provinces, although its performance tended to be over-conservative in Yunnan province (Figs. 2b & 2d).

Figure 3 shows the response curves for the variables used in our MAXENT models. Each curve represents the response to a specific variable with all others set constant, and should therefore be interpreted cautiously, although comparisons between species and models do allow for testable biogeographical insights.⁴² The responses indicate that the distributions of both species are correlated with mean temperature of wettest quarter in summer rainfall corn production regions (bio08) in the range 10-29°C (Fig. 3), with both species preferring areas with higher summer precipitation and precipitation seasonality (bio18 & bio15) within areas that possess moderate isothermality (bio03). Overall, the shape of response curves is similar between ACB

and *T. ostriniae*, although the starting points for precipitation and radiation seasonality were higher for ACB. The response curve displayed an opposite trend to radiation seasonality in ACB. This difference suggests that *T. ostriniae* is more sensitive to climatic seasonality. Permutation importance of precipitation and radiation seasonality in both species models was 18.13 and 23.33 respectively for *T. ostriniae*, compared to 14.08 and 10.03 respectively for ACB (Table 2).

3.2 Distribution fit and model projections for *O. nubilalis* and *T. ostriniae* in Europe and North America

The locations for inundative release for *T. ostriniae* are within central and northeastern locations of USA and Canada (Kentucky, Virginia, Massachusetts, New York states and Quebec, Fig. 1). Each of the final variable sets used in the MAXENT, PCA analysis and MESS map for North American projections (see Appendix S2 for detail) indicated accessible climate space available to *T. ostriniae* distributed in native Asia is not accessible in novel North America regions. Dissimilar climate space between those two continents might have constrained the MAXENT projection for *T.*

ostrinae, thus, only the CLIMEX model was adopted for model projection across continents.

The modelled native distribution of *O. nubilalis* distributes across whole Europe areas including England and the southern margin of the Scandinavian peninsula (Fig. 4). The distribution is consistent with model results from Trnka *et al.*⁵¹ and Svobodová *et al.*⁵² in Czech Republic and most European areas, taking into account the use of different climate backgrounds here, and the expected minor idiosyncrasies between datasets. Another test of the CLIMEX model is to generate weekly changes in GI_w over an average year for a location and compare this “modelled suitability” with observed phenology based on trapping of adult moths (Fig. 4). GI_w would be expected to be an indicator of population growth rate for *O. nubilalis*. For both sites, trapped moths occur at times which are climatically suitable for growth, with a seven-week delay for adults observed in Barrantes (northwestern Spain). Moth numbers will of course reflect additional landscape level effects of crop availability and “management”, such as extensive and intensive spraying of insecticides.^{57,58}

The modelled distribution of *O. nubilalis* in North America overlaps with the main corn production areas and published distribution points for *O. nubilalis*^{26,53,54} that are within northeastern United States (Iowa, Illinois, Indiana, Ohio and nearby areas) and Canada (Quebec) (Fig. 5a). Moreover, there is contiguous suitable habitat across southeastern states for the modelled distribution of *T. ostriniae* (Fig. 5b).

3.3 Host-parasitoid dynamics under climate change in Asia and North America

Using the difference in GI between ACB/ECB and *T. ostriniae* allows us to comment on the effectiveness of *T. ostriniae* as a biological control agent under current and future condition. Current climates in Asia are predicted to favor *T. ostriniae* over ACB across China (Figure 6a, purple shading), ACB only has higher GI in the Sichuan area (Figures 6a, yellow shading). By 2080, it is predicted to favor *T. ostriniae* over ACB in northeastern China as the difference between GI for ACB and GI for *T. ostriniae* increases in these regions. On the other hand, GI of ACB is predicted to decline through the Sichuan area (Figure 6b, previously yellow shading (Figure 6a) changes to light purple shading). The predicted higher temperatures by 2080 is likely to favor *T. ostriniae* in southern China, where GI for the parasitoid is predicted to be greater

than GI for ACB (Figure 3c, light purple shading (Figure 6b) changes to dark purple shading).

For North America projections, the climate change scenario is predicted to favor *T. ostriniae* over ECB in the area with least difference presented around most of the US and Canada except some southern area (Figure 6c). By 2080, there is a northward shifting tendency for these areas (Figure 6d). The predicted higher temperatures by 2080 could favor *T. ostriniae* from regions in southern Canada (Figure 6d, dark purple shading) as the difference between GI for ECB and GI for *T. ostriniae* decreases in these regions. At the same time, higher temperature is predicted to favor ECB in most southern US, where their GI difference is predicted to be amplified (Figure 6d, light purple shading (Figure 6c) changes to yellow and light-yellow shading).

4 DISCUSSION AND CONCLUSIONS

Understanding the extent to which host-parasitoid relationships may be altered by climate change has important implications for future biological control release programs. Identifying where putative spatial mismatch between species may occur is

particularly important. By employing modelling methods appropriate to the data available on the Asian and European corn borers (ACB and ECB, respectively), and the *Trichogramma* parasitoid (*T. ostriniae*), we identified broad scale patterns that provide insights into the importance of precipitation and radiation seasonality that might determine the distribution of a pestiferous host (ACB) and its parasitoid (*T. ostriniae*). Our analyses also enabled identification of suitable areas for inundative release of *T. ostriniae*, which convey areas of likely successful parasitism.

As for most insects, temperature is the key factor that determines the distribution of parasitoids and their hosts across multiple scales.^{17,59} Differences in thermal requirements appear to be responsible for large discrepancies between host and parasitoid distributions. In northern areas of Asia where our models typically predicted more suitable climate space for ACB over *T. ostriniae* under current conditions, two factors might contribute. First, the lower development temperature threshold of *T. ostriniae* is higher than that of its host (6.8 to 10.4°C for ACB; 11.7 to 11.8°C for *T. ostriniae*).^{22,55} Second, ACB demonstrates broader thermal requirements for development than *T. ostriniae*, which may reflect geographical variation in

physiological thermal limits; Xie *et al.*⁶⁰ previously noted variation in the supercooling point (SCP) and survival rate after low temperature treatments in this species, with mean SCP significantly declining from -22°C in a tropical population (Haikou, N 19.98°) to -28.5°C for a univoltine low temperature population (Gongzhuling, N 43.55°).

For southern areas of Asia, *T. ostrinae* appears to be more sensitive to the lower seasonality of radiation and precipitation (Fig. 3) than its host. The different responses of *T. ostrinae* and ACB to seasonality of both precipitation and radiation reflect findings from a meta-analysis for *Trichogramma* parasitism,⁶¹ which found that parasitism was higher in regions with less temperature and precipitation seasonality.

Trichogramma displays a more restricted climate tolerance than its hosts as the modelling approach is based on occurrence data, and the occurrence and abundance of *Trichogramma* is dependent on the abundance of its hosts. Occurrence data for the hosts are relatively better than for *Trichogramma* parasitoids. At the edge of the geographical range of the hosts, we expect low host population densities and thus low parasitoid densities, making parasitoids particularly hard to detect. As a result, the

abiotic tolerance of the parasitoid is estimated as being lower. However, this may also reflect the poor mobility of the parasitoid compared to its host (suggesting that the parasitoid cannot easily track changes in favorable conditions, and, being restricted to the static egg stage of its ACB host, is more exposed to extreme temperature and precipitation), or the dependence of *T. ostriniae* on passive transport (mainly wind dispersion), compared to its larger bodied lepidopteran hosts.^{62,63}

The Ecoclimatic index of *T. ostriniae* and its host ECB in North America were associated with different temperature, moisture, dry and heat stress parameters. In low latitudes of North America, we found that there is more suitable climate space for *T. ostriniae* than ECB (Fig. 5), which is different from predictions in Asia, despite the upper development temperature threshold being higher for the host (41°C for ECB; 35°C for *T. ostriniae*).^{49,55} In this case we suspect that much lower heat stress temperature (30.5°C for ECB; 35°C for *T. ostriniae*) and higher heat stress accumulation for ECB (0.01 week⁻¹ for ECB; 0.02 week⁻¹ for *T. ostriniae*) reflect constrained growth and dispersion in southern areas of North America.

Constructing niche models at a global scale inherently assumes that all populations respond to predictor variables, or ecophysiological indices, in much the same way. This is highly unlikely, as differences in temperature-related life history parameters for *T. ostriniae* are likely to exist. For example, individuals from North American populations grow faster than Chinese populations (9 days at 24°C for America; 10 to 11 days at 25°C for China).²⁹ There may also be genetic differentiation among North America populations of ECB related to voltinism.⁵³ Such differences could influence the successful establishment of *T. ostriniae* populations in new areas. For species that encompass large geographic ranges, it can be challenging to define diapause as a single function for all populations. For ACB and ECB there are different strains and varying responses to light and temperature according to latitude.^{47,60,64} Whilst we have fitted single diapause indices to these species, it should be noted that regional assessments of distributions and field efficacy will need to identify specific temperature limits and diapause cues for the local populations. Additionally, identifying different populations and strains through genetic methods could allow for partitioning correlative models like MAXENT to investigate species-environment relationships at finer scales.⁶⁵

The underprojection of the *T. ostriniae* Asia-trained MAXENT model into North America highlights some potential limits of extrapolating correlative models.⁸ Dissimilar climate space between continents may have constrained the modelling projections presented here (Appendix S2). However, the CLIMEX models **were able to project from the native occurrence points without these issues, reinforcing both the importance of comparing different modelling approaches, and using physiologically based approaches (i.e. CLIMEX) when projecting to novel areas of climates as well**^{11,39} Distribution data for both *T. ostriniae* and *O. furnacalis* were limited due to recent survey efforts being restricted regionally and a lack of precise locality data associated with most published records.^{66,67} Our modelling approaches consider these two species in isolation, and inherently overlook competitive interactions with other *Trichogramma* species present across the distributions. Meta-analyses have suggested for competing natural enemy species that antagonism may result in effects on the target being less than additive, and this deleterious outcome is most obvious when the same life stage of a host is attacked.¹⁵ Therefore, any inference regarding areas suitable for *T. ostriniae* ought to be made with caution and requires empirical studies

that include molecular identification of potential competing *Trichogramma* species that might have been incorrectly identified.^{23,68}

In terms of biological control, there are perhaps implications for areas with GI differences that favour the parasitoid over the host (Fig. 6a & 6c). Such areas may represent increased efficacy of the parasitoid wasp in those parts of Asia and North America and indicate areas with a higher probable success rate of inundative releases. In areas where *T. ostrinae* is in marginal climate space, the efficiency of biocontrol might be improved through increasing the number of wasps used in inundative releases.²¹ For example, Gagnon *et al.*²⁶ recently reported that the use of *T. ostrinae* around Quebec (marginally suitable in our models) could effectively decrease the pressure exerted by ECB and its damage on sweet corn ears. Biocontrol efficiency might also be improved by timing releases to avoid extreme radiation/rainfall events based on meteorological forecasts. **Better regional temporal forecasts should emerge by considering host-parasitoid relationships through dynamic population models that consider the interaction between trophic levels.**

Under the future climate change scenarios investigated here, both ACB and *T. ostriniae* are expected to have broad geographic distribution in southeastern Asia into the next few decades. While this suggests a reasonable level of biological control potential for the future, there are also some likely challenges. The high degree of variation in life history traits within ACB populations across Asia,^{21,22} and the occurrence of multiple generations per year, could provide opportunities for rapid adaptation to novel environments.^{11,60} Areas representing a relative advantage for ECB through the central United States (Fig. 6d) contain low similarity to Asian climate space of *T. ostriniae* (Appendix S2), which might limit its control efficacy across central North America in the future. To sum up, the efficacy of *T. ostriniae* biocontrol across areas of northern America and Canada is likely to be effectively improved under warming temperature around summer.^{26,61} Control efforts through central North America area, on the other hand, may be improved by increasing the numbers and release frequency of *T. ostriniae* and optimizing corn integrated control strategies based on further phenological dynamics studies.

Overall the modelling approaches used here proved useful for examining broad-scale

environmental limits to an egg parasitoid and its lepidopteran hosts. Global investigations provide important hypotheses on key traits that limit distributions and define synchrony in host-parasitoid relationships. Such hypotheses can help to guide future experimental work to understand how climate change, such as climate seasonality/extremity, may affect these species at finer-scales. We also identified areas potentially suitable for inundative releases, however more regional-scale assessments will need to determine which strains of the hosts are present to determine how important life-history traits such as diapause are represented in local populations. Finally, further phenological and diapause studies for *T. ostrinae* and its hosts are required to optimize corn integrated control strategies.

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References

1. Lesk C, Rowhani P and Ramankutty N, Influence of extreme weather disasters on global crop production. *Nature* **529**: 84-87 (2016).
2. Dunn RR, Harris, NC, Colwell, RK, Koh, LP and Sodhi NS, The sixth mass coextinction: are most endangered species parasites and mutualists? *P R Soc B* **276**(1670): 3037-3045 (2009).
3. Thomson LJ, Macfadyen S and Hoffmann AA, Predicting the effects of climate change on natural enemies of agricultural pests. *Biol Control* **52**(2): 296-306 (2010).
4. Furlong MJ and Zalucki MP, Climate change and biological control: the consequences of increasing temperatures on host-parasitoid interactions. *Curr Opin Insect Sci* **20**: 39-44 (2017).
5. Guisan A and Thuiller W, Predicting species distribution: offering more than simple habitat models. *Ecol lett* **8**(9): 993-1009 (2005).
6. Franklin J, *Mapping Species Distributions: Spatial Inference and Prediction*. Cambridge University Press, Cambridge, UK (2009).
7. Franklin J, Species distribution models in conservation biogeography:

- developments and challenges. *Divers Distrib* **19**(10): 1217-1223 (2013).
8. Hill MP and Thomson LJ, Species distribution modelling in predicting response to climate change, in *Climate Change and Insect Pests*, ed. by Björkman C and Niemelä P, CABI. Oxfordshire, pp. 16-37 (2015).
9. Elith J, Kearney M and Phillips S, The art of modelling range-shifting species. *Methods Ecol Evol* **1**(4): 330-342 (2010).
10. Hill MP, Bertelsmeier C, Clusella-Trullas S, Garnas J, Robertson MP and Terblanche JS, Predicted decrease in global climate suitability masks regional complexity of invasive fruit fly species response to climate change. *Biol Invasion* **18**(4): 1105-1119 (2016).
11. Lozier JD and Mills NJ, Predicting the potential invasive range of light brown apple moth (*Epiphyas postvittana*) using biologically informed and correlative species distribution models. *Biol Invasions* **13**: 2409-2421 (2011).
12. Sutherst RW, Maywald GF and Kriticos D, *CLIMEX Version 3: User's Guide*. Hearne Scientific Software Pty Ltd, Melbourne (2007).
13. Robertson MP, Kriticos DJ and Zachariades C, Climate matching techniques to narrow the search for biological control agents. *Biol Control* **46**(3): 442-452

(2008).

14. Senaratne KADW, Palmer WA and Sutherst RW, Use of CLIMEX modelling to identify prospective areas for exploration to find new biological control agents for prickly acacia. *Aust J Entomol* **45**(4): 298-302 (2006).
15. Hoddle MS, Warner K, Steggall J and Jetter KM, Classical biological control of invasive legacy crop pests: New technologies offer opportunities to revisit old pest problems in perennial tree crops. *Insects* **6**(1): 13-37 (2015).
16. Smith SM, Biological control with *Trichogramma*: Advances, successes, and potential of their use. *Annu Rev Entomol* **41**: 375-406 (1996).
17. Hance T, van Baaren J, Vornon P and Boivin G, Impact of extreme temperatures on parasitoids in a climate change perspective. *Annu Rev Entomol* **52**: 107–126 (2007).
18. Le Roux RC and McGeoch MA, Rapid range expansion and community reorganization in response to warming. *Glob Change Biol* **14**(12): 2950-2962 (2008).
19. Walther G, Community and ecosystem responses to recent climate change. *Philos T R Soc Lon B* **1549**(365): 2019-2024 (2010).

20. Stireman JO III, Dyer LA, Janzen DH, Singer MS, Lill JT, Marquis RJ, Ricklefs RE, Gentry GL, Hallwachs W, Coley PD, Barone JA, Greeney, HF, Connahs H, Barbosa P, Morais HC and Diniz IR, Climatic unpredictability and parasitism of caterpillars: Implications of global warming. *P Natl Acad Sci USA* **48**(102): 17384-17387 (2005).
21. Wang ZY, He KL, Zhang F, Lu X and Babendreier D, Mass rearing and release of *Trichogramma* for biological control of insect pests of corn in China. *Biol Control* **68**: 136-144 (2014).
22. Nafus DM and Schreiner IH, Review of the biology and control of the Asian corn borer, *Ostrinia furnacalis* (Lep: Pyralidae). *Trop Pest Manage* **37**(1): 41-56 (1991).
23. Wu LH, Hoffmann AA and Thomson LJ, *Trichogramma* parasitoids for control of Lepidopteran borers in Taiwan: species, life-history traits and *Wolbachia* infections. *J Appl Entomol* **140**(5): 353-363 (2016).
24. Wang B, Ferro DN and Hosmer DW, Importance of plant size, distribution of egg masses, and weather conditions on egg parasitisms of the European corn borer,

Ostrinia nubilalis by *Trichogramma ostrinae* in sweet corn. *Entomol Exp Appl* **83**(3): 337-345 (1997).

25. Etilé E, Boisclair J, Cormier D, Todorova S and Lucas É, Evaluation of two biological control strategies against the European corn borer (*Ostrinia nubilalis*) in southern Quebec fresh sweet corn, in *4ème Conférence Internationale sur les Méthodes Alternatives en Protection des Cultures. Evolution des cadres réglementaires européen et français*, Nouveaux moyens et stratégies Innovantes, Nouveau Siècle, Lille, France, pp. 614-620 (2011).
26. Gagnon AE, Audette C, Duval B and Boisclair J, Can the use of *Trichogramma ostrinae* (Hymenoptera : Trichogrammatidae) to control *Ostrinia nubilalis* (Lepidoptera : Crambidae) be economically sustainable for processing sweet Corn ? *J Econ Entomol* **110**(1): 59-66 (2016).
27. Seaman A, Hoffmann H, Gardner J and Chenus S, Pilot testing of *Trichogramma ostrinae* releases in fresh market sweet corn for control of European corn borer, in *New York State Vegetable Project Reports Relating to IPM*, NY IPM Publication 121, pp. 149-154. (1996).
28. Hoffmann MP, Pitcher SA, Cheever SA, Gardner J, Losey JE, Kuhar TP, Laub CA

- and Youngman RR, Efficacy of inoculative releases of *Trichogramma ostriniae* (Hymenoptera: Trichogrammatidae) against European corn borer *Ostrinia nubilalis* (Lepidoptera: Crambidae) in Weld corn. *Biol Control* **36**(3): 345-349 (2006).
29. Wang B, Ferro DN, Wu J and Wang S, Temperature-dependent development and oviposition behavior of *Trichogramma ostriniae* (Hymenoptera: Trichogrammatidae), a potential biological control agent for the European corn borer (Lepidoptera: Crambidae). *Environ Entomol* **33**(4): 787-793 (2004).
30. Kriticos DJ, Webber BL, Leriche A, Ota N, Macadam I, Bathols J and Scott JK, CliMond: global high-resolution historical and future scenario climate surfaces for bioclimatic modelling. *Methods Ecol Evol* **3**(1): 53-64 (2012).
31. Phillips SJ, Anderson RP and Schapire RE, Maximum entropy modeling of species geographic distributions. *Ecol Model* **190**(3-4): 231-259 (2006).
32. Elith J, Phillips SJ, Hastie T, Dudík M, Chee Y and Yates CJ, A statistical explanation of MAXENT for ecologists. *Divers Distrib* **17**(1): 43-57 (2011).
33. Hijmans RJ, Cameron S, Parra JL, Jones PG and Jarvis A, Very high resolution

- interpolated climate surfaces for global land areas. *Int J Climatol* **25**(15): 1965-1978 (2005).
34. New M, Lister D, Hulme M and Makin I, A high-resolution data set of surface climate over global land areas. *Clim Res* **21**(1): 1-25 (2002).
35. Barve N, Barve V, Jiménez-Valverde A, Lira-Noriega A, Maher SP, Peterson AT, Soberón J and Villalobos F, The crucial role of the accessible area in ecological niche modeling and species distribution modeling. *Ecol Model* **222**(11): 1810-1819 (2011).
36. Hirzel AH, Hausser J, Chessel D and Perrin N, Ecological-niche factor analysis: how to compute habitat-suitability maps without absence data? *Ecology* **83**(7): 2027-2036 (2002).
37. Calenge C, The package ‘adhabitat’ for the R software: a tool for the analysis of space and habitat use by animals. *Ecol Model* **197**(3-4): 516-519 (2006).
38. R Core Team, R: A language and environment for statistical computing. R foundation for statistical computing, Vienna, Austria. ISBN 3-900051-07-0 (2014).
39. Hill MP, Axford J and Hoffmann AA, Predicting the spread of *Aedes albopictus* in

Australia under current and future climates: Multiple approaches and datasets to incorporate potential evolutionary divergence. *Austral Ecol* **39**(4): 469-478 (2014).

40. Rödder D and Lötters S, Niche shift versus niche conservatism? Climatic characteristics of the native and invasive ranges of the Mediterranean house gecko (*Kemidactylus turcicus*). *Global Ecol Biogeogr* **18**(6): 674-687 (2009).
41. Hill MP and Terblanche JS, Niche overlap of congeneric invaders supports a single-species hypothesis and provides insight into future invasion risk: implications for global management of the *Bactrocera dorsalis* complex. *PLoS One* **9**(2): e90121(2014).
42. Hill MP, Hoffmann AA, Macfadyen S, Umina PA and Elith J, Understanding niche shift: using current and historical data to model the invasive redlegged earth mite, *Halotydeus destructor*. *Divers Distrib* **18**(2): 191-203 (2012).
43. Hijmans RJ, Phillips S, Leathwick J and Elith J, Package 'dismo'. Available online at: <http://cran.r-project.org/web/packages/dismo/index.html> (2011).
44. Sutherst RW and Maywald GF, A climate-model of the red imported fire ant, *Solenopsis invicta* Buren (Hymenoptera: Formicidae): implications for invasion

of new regions, particularly Oceania. *Environ Entomol* **34**(2): 317-335 (2005).

45. Kocmánková E, Trnka M, Eitzinger J, Formayer H, Dubrovský M, Semerádová D, Žalud Z, Juroch J and Možný M, Estimating the impact of climate change on the occurrence of selected pests in the Central European region. *Clim Res* **44**(1): 95-105 (2010).
46. Yang HZ, Tu XY, Xia QW, He HM, Chen C and Xue FS, Photoperiodism of diapause induction and diapause termination in *Ostrinia furnacalis*. *Entomol Exp Appl* **153**(1): 34-46 (2014).
47. Guo JQ, Zhang HG, Wang ZY and He KL, Effects of photoperiod and temperature on diapause induction in *Ostrinia furnacalis* (Lepidoptera: Crambidae). *Acta Entomol Sinica* **56**(9): 996-1003 (2013).
48. Lu X and Zhou DR, On the photoperiodic reactions in different voltinism ecotypes of asian corn borer. *J Plant Prot* **27**: 12-16 (2000).
49. Maiorano A, Bregaglio S, Donatellia M, Fumagalli D and Zucchini A, Comparison of modelling approaches to simulate the phenology of the European corn borer under future climate scenarios. *Ecol Model* **245**(24): 65-74 (2012).

50. Cordero C, Malvar RA, Butrón A, Revilla P, Velasco P and Ordás A, Population dynamics and life-cycle of corn borers in south Atlantic European coast. *Maydica* **43**: 5-12 (1998).
51. Trnka M, Muška F, Semerádová D, Dubrovský M, Kocmánková E and Žalud Z, European Corn Borer life stage model: Regional estimates of pest development and spatial distribution under present and future climate. *Ecol Model* **207**(2-4): 61-84 (2007).
52. Svobodová E, Trnka M, Dubrovský M, Semerádová D, Eitzinger J, Žalud Z and Štěpánek P, Pests occurrence model in current climate-validation study for European domain. *Acta U Agr Silvi Mendelianae Brunensis* **1**: 205–214 (2013).
53. Coates BS, Sumerford DV and Hellmich RL, Geographic and voltinism differentiation among North American *Ostrinia nubilalis* (European corn borer) mitochondrial cytochrome c oxidase haplotypes. *J Insect Sci* **4**(35): 1-9 (2004).
54. Merrill SC, Walter SM, Peairs FB and Schleip EM, The distribution of European Corn Borer (Lepidoptera : Crambidae) moths in pivot-irrigated corn. *J Econ Entomol* **106**(5): 2084-2092 (2013).
55. Hirashima Y, Miura K, Miura T and Shiro K, Effect of temperature on the

development of the egg-parasitoids *Trichogramma chilonis* and *Trichogramma ostrinae*. *Sci B Fac Agr Kyushu* **44**: 71-76 (1990).

56. Intergovernmental Panel on Climate Change, in *Managing the Risks of Extreme Events and Disaster to Advance Climate Change Adaptation*, ed. by Field CB, Barros V, Stocker TF, Dahe Q, Dokken DJ, Ebi KL, Mastrandrea MD, Mach KJ, Plattner GK, Allen SK, Tignor M and Midgley PM, Cambridge University Press, Cambridge, pp. 1-19 (2013).
57. Zalucki MP, Adamson D and Furlong MJ, The future of IPM: whither or wither? *Aust J Entomol* **48**(2): 85–96 (2009).
58. Kriticos DJ, Ota N, Hutchison WD, Beddow J, Walsh T, Tay WT, Borchert DM, Paula-Moreas SV, Czapak C and Zalucki MP, The potential distribution of invading *Helicoverpa armigera* in North America: is it just a matter of time? *PLoS ONE* **10**: e0119618 (2015).
59. Jeffs T and Lewis OT, Effects of climate warming on host-parasitoid interactions. *Ecol Entomol* **38**(3): 209-218 (2013).

60. Xie HC, Li DS, Zhang HG, Mason CE, Wang ZY, Lu X, Cai WZ and He KL, Seasonal and geographical variation in diapause and cold hardiness of the Asian corn borer, *Ostrinia furnacalis*. *Insect Sci* **22**(4): 578-586 (2015).
61. Wu LH, Hoffmann AA and Thomson LJ, Potential impact of climate change on parasitism efficiency of egg parasitoids: A meta-analysis of *Trichogramma* under variable climate conditions. *Agr Ecosyst Environ* **231**: 143-155 (2016).
62. Romeis J, Babendreier D, Wäckers L and Shanower TS, Habitat and plant specificity of *Trichogramma* egg parasitoids-underlying mechanisms and implications. *Basic Appl Ecol* **6**(3): 215-236 (2005).
63. Mills N, Egg parasitoids in biological control and integrated pest management, in *Egg Parasitoids in Biological Control and Integrated Pest Management*, ed. by Consoli FL, Parra JRP and Zucchi RA, Springer, New York, pp. 389-411 (2010).
64. Onstad DW and Brewer EF, Modeling induction of diapause in North American *Ostrinia nubilalis* (Lepidoptera: Pyralidae) populations. *Popul Ecol* **25**(5): 1140-1146 (1996).
65. Lozier JD and Mills NJ, Ecological niche models and coalescent analysis of gene

flow support recent allopatric isolation of parasitoid wasp populations in the Mediterranean. *PLoS ONE* **4**: e5901 (2009).

66. Graham CH, Ferrier S, Huettman F, Moritz C and Peterson AT, New developments in museum-based informatics and applications in biodiversity analysis. *Trends Ecol Evol* **19**(9): 497-503 (2004).
67. Pearson RG, Raxworthy CJ, Nakamura M and Peterson AT, Predicting species distributions from small numbers of occurrence records: a test case using cryptic geckos in Madagascar. *J Biogeogr* **34**(1): 102-117 (2007).
68. Wu LH, Hoffmann AA and Thomson LJ, Taiwanese *Trichogramma* of Asian corn borer: morphology, ITS-2 rDNA characterization, and natural *Wolbachia* infection. *J Insect Sci* **16**(1): 1-7 (2016b).

Tables and Figures

Table 1 CLIMEX parameter values for the final model for *Ostrinia furnacalis* and *Trichogramma ostriniae* and *Ostrinia nubilalis* determined by iteratively changing parameter values until the performance provided best fit to the known native distributions (see Fig. 1)

Index	Parameter	Value ^a		
		<i>O. furnacalis</i>	<i>T. ostriniae</i>	<i>O. nubilalis</i>
Temperature	DV0 = Lower threshold	6.8°C	11.7°C	5.5°C
	DV1 = Lower optimum temperature	20°C	22°C	28°C
	DV2 = Upper optimum temperature	28°C	29°C	38°C
	DV3 = Upper threshold	33°C	35°C	41°C
Moisture	SM0 = Lower soil moisture threshold	0.15	0.15	0.06
	SM1 = Lower optimum soil moisture	0.35	0.35	0.15
	SM2 = Upper optimum soil moisture	0.80	0.80	1.20
	SM3 = Upper soil moisture threshold	1.50	1.50	2.00
Cold stress	TTCS = Cold stress temperature threshold	0.5°C	1°C	-
	THCS = Cold stress accumulation rate	-0.0001 week ⁻¹	-0.0001 week ⁻¹	-
Heat stress	TTHS = Heat stress temperature threshold	33°C	35°C	30.5°C
	THHS = Heat stress accumulation rate	0.002 week ⁻¹	0.02 week ⁻¹	0.01 week ⁻¹
Dry stress	SMDS = Dry stress threshold	0.1	0.1	0.05
	HDS = Dry stress rate	-0.02 week ⁻¹	-0.02 week ⁻¹	-0.04 week ⁻¹
Wet stress	SMWS = Wet stress threshold	1.5	1.5	1.5
	HWS = Wet stress rate	0.0002 week ⁻¹	0.0002 week ⁻¹	0.0002 week ⁻¹
Diapause	Diapause induction daylength	14.88 hour	-	14.5 hour
	Diapause induction temperature	17°C	-	12°C

^aValues without units are dimensionless proportions. Cold stress parameters for

O. nubilalis were left blank as these would not affect model outcome.

Table 2 Sample size and percentage based on mean area under the curve of receiver-operator characteristic (AUC) test gain across 10 cross-validation replicates for MAXENT models; final variable[†] contributions and importance assessed through MAXENT selected for *Trichogramma ostrinae* and *Ostrinia furnacalis*

Species	<i>n</i>	AUC	Variable contribution and importance									
			Bio03		Bio08		Bio15		Bio 18		Bio 23	
			PC	PI	PC	PI	PC	PI	PC	PI	PC	PI
<i>O. furnacalis</i>	183	0.778	9.64	8.22	62.11	61.88	12.77	14.08	5.43	5.80	10.06	10.03
<i>T. ostrinae</i>	129	0.854	5.10	5.27	58.07	52.47	19.29	18.13	1.05	0.80	16.48	23.33

[†]Bio03 = Isothermality (mean of monthly (max temp – min temp)/ temperature annual range) (*100); Bio08 = Mean temperature of wettest quarter (°C); Bio15 = Precipitation seasonality (coefficient of variation); Bio18 = Precipitation of warmest quarter; Bio23 = Radiation seasonality (coefficient of variation). PC, percent contribution; PI, permutation importance.

Figure 1 Distributions of Asian corn borer, *Ostrinia furnacalis* (purple circles), European corn borer, *Ostrinia nubilalis* (blue circles) and *Trichogramma ostrinae* (yellow circles). Sources are published references (see Table S1) and field surveys in 2014 and 2015 in China, Japan, Korea and Taiwan. Locations of *T. ostrinae* inundative releases undertaken in North America from published references (see Table S1). Colored areas represent backgrounds used in Environmental Niche Model (ENMs) construction based on Köppen-Geiger classifications (Green = Asia model; Pink = America model).

Figure 2 Distribution fitting models between *Ostrinia furnacalis* (ACB) and *Trichogramma ostrinae* (To) in southeastern Asia using two different niche modelling approaches. (a) *O. furnacalis* logistic MAXENT fitting model: MAXENT logistic output as a continuous suitability (0-1) layer, and darker shading indicates higher climate space suitability. (b) *T. ostrinae* logistic MAXENT fitting model. (c) *O. furnacalis*, (d) *T. ostrinae* CLIMEX fitting model: EI output as a continuous suitability (0-65) layer, and darker shading indicates higher climate space suitability. Provinces are indicated on the map by abbreviations: Ah, Anhui; Bj, Beijing; Fj, Fujian; Gd, Guangdong; Gs, Gansu; Gx, Guangxi; Gz, Guizhou; Hb, Hebei; Hlj, Heilongjiang; Hn, Henan; Hnl, Hainan Island; Hub, Hubei; Hun, Hunan; IM, Inner Mongolia; Jl, Jilin; Js, Jiangsu; Jx, Jiangxi; Ln, Liaoning; Sax, Shaanxi; Sc, Sichuan; Sd, Shandong; Sx, Shanxi; Yn, Yunnan; Zj, Zhejiang.

Figure 3 Variable response curves for three dominant predictor variables across models of *Ostrinia furnacalis* and *Trichogramma ostrinae* built in MAXENT. The line on the graphs is the mean of 10 cross-validation replicates, and the shaded area is the standard deviation across the replicates. Red: *O. furnacalis* (ACB); blue: *T. ostrinae* (To).

Figure 4 CLIMEX output for European corn borer *Ostrinia nubilalis* in Europe built on CliMond data sets. Shading reflecting Ecoclimatic Index (EI), and various measures of population abundance for *O. nubilalis* (Major Y-axis) and Weekly Growth Index (minor Y-axis) values over a year derived from current model (time in weeks, 1-52) match with each other in summer for Pontevedra (blue point) in North western Spain (modified from Cordero *et al.* 1998).

Figure 5 Distribution fitting models between *Ostrinia nubilalis* and *Trichogramma ostrinae* in north America built on CliMond data sets. (a) *O. nubilalis*, (b) *T. ostrinae* CLIMEX fitting EI model: output as a continuous suitability (0-80) layer, and darker shading indicates higher climate space suitability. Blue points represent distribution points of *O. nubilalis*, yellow points represent distribution points of *T. ostrinae*.

Figure 6 Effects of predicted climate change on the relationships between ACB/ECB and *Trichogramma ostriniae* in southeastern Asia/north America. Relationship between the current differences between ACB, ECB and *T. ostriniae* growth indices (GIs); and the predicted differences in these indices by 2080. (a) Differences between ACB GI and *T. ostriniae* GI expressed as a proportion of ACB current GI. (b) Predicted differences between ACB GI and *T. ostriniae* GI expressed as a proportion of ACB GI for 2080. (c) Differences between ECB GI and *T. ostriniae* GI expressed as a proportion of ECB current GI. (d) Predicted differences between ECB GI and *T. ostriniae* GI expressed as a proportion of ECB GI for 2080. In figures dark purple shading shows locations where GI differences of *T. ostriniae* are greatest, lighter purple shows locations where differences are less pronounced; yellow shading shows locations where ACB/ECB GI is greater than *T. ostriniae* and white shading shows locations where ACB/ECB GI is equal to *T. ostriniae* GI.

Assessing the current and future biological control potential of *Trichogramma ostrinae* on its hosts *Ostrinia furnacalis* and *Ostrinia nubilalis*

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Our study is proved useful for assessing environmental factors linked to *Trichogramma* and its lepidopteran hosts and identifying areas suitable for inundative releases under climate change scenarios.

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