

Probabilistic evaluation of low-quality DNA profiles

K. Ryan, D. Gareth Williams^a, David J. Balding^{b,c}

a. DGW Software Consultants LTD

b. UCL Genetics Institute, University College, London WC1E 6BT

c. Centre for Systems Genomics, University of Melbourne, VIC 3010, Australia

ABSTRACT

Many DNA profiles recovered from crime scene samples are of a quality that does not allow them to be searched against, nor entered into, databases. We propose a method for the comparison of profiles arising from two DNA samples, one or both of which can have multiple donors and be affected by low DNA template or degraded DNA. We compute likelihood ratios to evaluate the hypothesis that the two samples have a common DNA donor, and hypotheses specifying the relatedness of two donors. Our method uses a probability distribution for the genotype of the donor of interest in each sample. This distribution can be obtained from a statistical model, or we can exploit the ability of trained human experts to assess genotype probabilities, thus extracting much information that would be discarded by standard interpretation rules. Our method is compatible with established methods in simple settings, but is more widely applicable and can make better use of information than many current methods for the analysis of mixed-source, low-template DNA profiles.

1 Introduction

Crime scene samples often contain DNA from multiple individuals, some or all of whom contribute small amounts of DNA that may have suffered degradation following environmental exposure. An electropherogram (epg) arising from such a sample can show stochastic effects that make it difficult to identify all of the alleles in the underlying genotypes [1,2]. Uncertain alleles are often encoded as “unknown”. For example the UK National DNA Database uses the coding F which can match any allele in a subsequent database search [3]. But if there is only limited uncertainty about the genotypes at several loci, designating them all as unknown is wasteful of potentially valuable information. We propose instead to encode the epg information as probability distributions over genotypes. We then compute a likelihood ratio (LR) assessing the evidence for a donor of interest in each of two samples to have a specified genetic relationship, including the important special case that they are the same person.

We expect that our proposed method will generate substantial benefits from its use as an intelligence tool. Given one or more complex profiles, the method permits speedy and accurate inferences to guide an investigation, for example in seeking a contributor to multiple, noisy crime scene profiles (CSP). However, our approach is also suitable for conveying weight of evidence to a finder of fact within a judicial system. We believe that the approach has great potential in improved database searches: partial profiles can be better represented in the database and therefore be more likely to generate matches, and they can be successfully searched against the database. Rigid rules for the “quality” of a profile to be included in the database, or for searches of it, can be greatly relaxed. With probabilistic encoding of epg information, poorer quality samples will automatically generate more diffuse probability distributions and hence weaker LRs, without the need for rules based on thresholds.

In standard settings there is a reference profile that specifies a unique genotype, while the information from the crime scene epg may be represented as a list of alleles with peak heights. Here, we allow each of the two profiles being compared to be represented as a probability distribution. In the special case of a reference profile, one genotype is assigned probability one.

We briefly discuss below automated epg encoding using a statistical model, but our approach can also be used in conjunction with an expert forensic scientist who encodes the epg information as a probability distribution over a matrix of possible genotypes. This encoding implies an element of subjectivity, but inter-expert comparisons and blind proficiency testing can satisfy a court's requirement for objectivity. Our approach is designed to exploit the ability of experts to make use of complex information quickly and efficiently, without the need for inflexible rules or sophisticated software. Experts can make better use of the epg than is the case in much current practice because probabilities allow gradations of judgement that better reflect the available information than do interpretation rules based on thresholds.

2 Encoding an epg in a genotype probability matrix

Although the interpretation of an epg can largely be automated, an expert forensic scientist is usually called on to confirm the automatic interpretation, and sometimes over-rule decisions relating to apparent artefacts such as split peaks and stutter. We distinguish this process of “interpretation” from that of “evaluation” in which numerical summaries of evidential weight are computed. For a single-source sample of good quality DNA, the result of an interpretation is the genotype of the donor at the tested loci. However, for many CSPs the genotype of an unknown donor of interest cannot be inferred with certainty from the epg. Reasons for this can include low DNA template and/or degraded DNA leading to the possibility of dropout of some alleles, DNA from more than one individual (mixed profile), and uncertainty about possible artefacts.

The need for objectivity in legal settings has led to the adoption of interpretation rules based on thresholds that have been shown in laboratory trials to have good properties over many real or simulated examples [4, 5]. For example, peaks below a threshold (typically 25 to 50 relative fluorescence units, RFU) are dismissed as noise, while single peaks above another threshold (typically 200 to 300 RFU) are interpreted as homozygotes (thus ruling out dropout of another allele from the donor at that locus) [3]. Similarly, a peak is interpreted as stutter or allelic according to whether or not its height is below a threshold fraction of the peak height at the “parent” allele (at one repeat unit larger than the peak under consideration).

Most importantly, many laboratories have rules for deciding when the genotype of the major donor to a mixture can be confidently identified, referred to as deconvolution of a mixed profile. Because of masking, it is almost never possible to determine the genotype of any donor other than a clear major. Typically, deconvolution is not permissible if the two greatest DNA donors gave similar amounts of DNA to the sample, but as the discrepancy in the amounts of DNA increases, it becomes increasingly possible to make a confident inference of the genotype of the major donor. Choosing criteria for when the major genotype can be inferred with sufficient confidence is problematic, and the implications can be considerable, because the evidential weight attached to a single genotype

deconvolved from a mixture can be many orders of magnitude greater than for the mixed profile.

All of these interpretation rules suffer from a “cliff edge” effect [6]. For example, a peak in a stutter position can be classified as certainly stutter, yet if it were slightly higher it would be regarded as certainly allelic. The setting of thresholds is usually intended to be “conservative”, such that in laboratory trials an allelic designation is rarely made when not correct. However, a reduction in one kind of error for a binary classification inevitably implies an increase in the other error, so that true alleles are wrongly designated as stutter. Moreover, this “conservative” policy does not necessarily favour defendants: broadly speaking, fewer alleles called from a crime scene epg means less information to identify donors, which in general helps defendants, but calling a peak as stutter rather than allelic can disfavour a defendant whose profile does not include that allele.

Experts are capable of making fine judgements about the plausibilities of different possible states of nature (e.g. stutter/allele). We believe that in general they do this well, better than decision processes based on thresholds, and that their ability can be assessed by inter-expert comparisons in blind trials. We propose to exploit the ability of a trained human expert to accurately process complex information, by allowing them to specify a probability distribution for the genotype of a donor of interest to a complex DNA profile. Typically, this donor will be the source of the largest amount of DNA, excluding known donors. For example, if the genotype of the major donor is clear at most loci, but there is some ambiguity at one or two loci, this information can be fully captured by specifying a unique genotype (probability one) at the relevant loci, but encoding the ambiguous loci as probability distributions concentrated on the genotypes consistent with the epg. If the donor of interest is a minor donor, while the major donor genotype is known, then the assignment of probabilities to genotypes should allow for any uncertainty due to masking of alleles by the major profile. We can also specify a joint genotype distribution for more than one unknown contributor, from which marginal genotype distributions can be obtained.

The genotype of an individual at a short tandem repeat (STR) locus consists of an unordered pair from a set of possible alleles. We encode an uncertain genotype as a symmetric matrix, with non-negative entries that sum to one. The number of rows and columns is the number of alleles recognised at the locus. The diagonal entries are the probabilities of the homozygote genotypes, and the (i, j) th and (j, i) th entries are each half the probability of an ij heterozygote. The full DNA profile of an individual is then represented as a genotype probability matrix (GPM) for each locus. GPMs can be used to encode epg information. Alternatively GPMs can encode information about an individual's genotype using genotypes of their relatives, in which case background allele probabilities are used for alleles inherited from ungenotyped relatives.

3 Calculating single-locus LR's using GPMs

Consider two GPMs, G^1 and G^2 , and suppose that we seek to compare the propositions:

η_1 : G^1 and G^2 represent the genotype of the same individual.

η_0 : G^1 and G^2 represent the genotypes of two unrelated individuals (who can have the same genotype).

In a standard setting, one GPM is from a CSP and may reflect uncertainty as discussed above, while the other encodes a reference profile that is usually assumed to be measured without error. However, our framework treats G^1 and G^2 in the same way, and hence allows for uncertainty in both profiles. An uncertain reference profile can arise if the individual is not available to give a good-quality DNA sample, and it has been obtained from a personal item such as a comb or toothbrush, or when it has been inferred from the genotypes of one or more relatives. In other situations there may be no reference profile, and the task is to assess whether the same individual has contributed to different CSPs.

We will initially assume that, under η_0 , the two individuals are unrelated and have no coancestry (that is $F_{ST} = 0$, see below). The case that the two individuals have a specified relationship, or are unrelated but share coancestry for example due to population structure, is discussed below. Genotypes are regarded a priori as random draws from a GPM representing the population genotype distribution, which we denote B (for background). It is standard practice in forensic DNA analysis to assume independence of each individual's two alleles at a locus, which is called Hardy-Weinberg Equilibrium (HWE) [7]. In that case B can be expressed in the form $B = \mathbf{b}^T \mathbf{b}$ where \mathbf{b} denotes a row vector of allele probabilities. Here, T denotes transpose, so \mathbf{b}^T is a $k \times 1$ column vector and B is $k \times k$. In practice, the elements of \mathbf{b} are relative frequencies obtained from a population database. There may be evidence suggesting a specific ethnic background of a donor of interest and hence an appropriate choice of B , or the weight of evidence may be assessed for multiple B matrices reflecting different possible ethnic backgrounds.

The LR conveying the weight of DNA evidence in support of η_1 relative to η_0 is the ratio of the probability of the evidence if η_1 is true, to its probability if η_0 holds [7].

Theorem 1

$$LR(\eta_1, \eta_0) = \sum_{ij} \frac{G_{ij}^1 \cdot G_{ij}^2}{B_{ij}} \quad (1)$$

where the subscript ij denotes the ij th entry of a matrix. The proof is in Appendix A1.

Each element of the numerator of (1) is the product of the probabilities that an ij genotype underlies each of samples 1 and 2. The denominator, B_{ij} is the probability for an unknown, unprofiled individual to have genotype ij . In (1), the ratio of these terms is summed over all genotypes at the locus.

Here, we assume that only one population is relevant, so that the B_{ij} apply to both contributors under η_0 . Equation (1) holds approximately if the two individuals come from different populations each with their own background genotype frequency matrix: in this case the B that represents the alternative contributor under η_0 should be used. In Section (6) we generalise (1) to allow for the two individuals under η_0 to come from the same subpopulation of the population from which the B_{ij} are obtained (coancestry correction).

In the special case that both G^1 and G^2 assign probability 1 to one genotype (spread over two entries of 0.5 for a heterozygote), (1) simplifies to $1/(2 B_{ij})$ (using $B_{ij} = B_{ji}$), if genotype ij is specified by both G^1 and G^2 ($1/B_{ii}$ for a homozygote), and zero otherwise. This corresponds to the familiar result that the LR is the reciprocal of the population genotype probability, which is the special case of the match probability when $F_{ST} = 0$.

4 LRs under specified relationships

A GPM for any individual can be used to generate GPMs for their parents and children, and subsequently other relatives. To see this, note that a GPM specifies an allele probability vector via its row or column sums (because GPMs are symmetric, these are the same). If \mathbf{p}_1 and \mathbf{p}_2 denote allele probability vectors for two parents, under HWE the GPM for their child can be written

$$\mathbf{G} = (\mathbf{p}_1^T \mathbf{p}_2 + \mathbf{p}_2^T \mathbf{p}_1) / 2. \quad (2)$$

We ignore the possibility of mutation here, and we continue to assume $F_{ST} = 0$ (no coancestry). If there is no genotype information available for one parent, then \mathbf{b} , the population allele probability vector, can be used instead:

$$\mathbf{G} = (\mathbf{p}^T \mathbf{b} + \mathbf{b}^T \mathbf{p}) / 2. \quad (3)$$

The GPM of a parent given a child's allele probability vector \mathbf{c} is

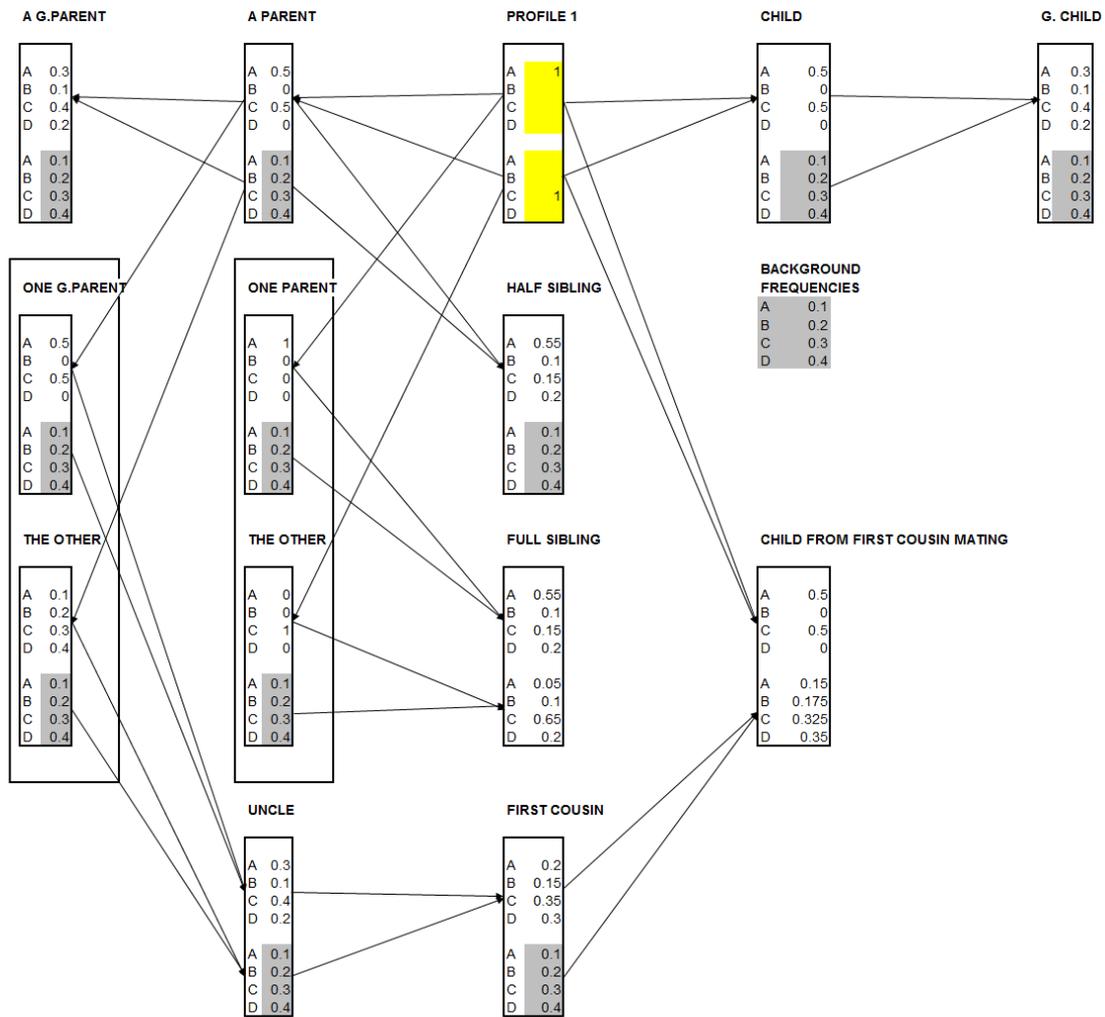
$$\mathbf{G} = (\mathbf{c}^T \mathbf{b} + \mathbf{b}^T \mathbf{c}) / 2 \quad (4)$$

which is equivalent to (3), reflecting the symmetry of the parent-child relationship (and any unilineal relationship) when no other genotyped relatives are available. Given a GPM for an individual, the GPM for any unilineal relative can be obtained by applying the above two steps for every parent-child link separating the relatives (1. generate an allele frequency vector from the GPM; 2. use (3) or (4) to generate a new GPM). Explicit formulae for some common relationships are developed in Appendix A2.

Specifying the joint probability distributions for two parents is more complex, because their genotypes are dependent given their child's genotype. For simplicity, we focus only on a pair of relatives, one of whom is genotyped. Relationships among multiple individuals pose no problem in principle: Bayesian networks provide a good framework for propagating GPMs through arbitrary networks of related individuals, and GPMs can be assigned to any specified members of the network [8].

Figure 1 illustrates the derivation of GPMs via relatedness in some important special cases. Assuming HWE, these GPMs can always be expressed in terms of two allele probability vectors. This simpler representation is not always available for GPMs derived directly from an epg. In Figure 1 the genotype for Profile 1 is known with certainty, but we can also combine uncertainty from a noisy epg with uncertainty due to relatedness.

Figure 1: GPMs for relatives of a profiled individual.



Under HWE, each GPM can be specified by two parental allele probability vectors, \mathbf{p}_1 and \mathbf{p}_2 , which are given here for each individual at a four-allele locus. The GPM of a reference individual is specified (here Profile 1 has genotype AC, indicated by the \mathbf{p}_1 and \mathbf{p}_2 assignments highlighted in yellow). The population allele probability vector \mathbf{b} is shaded in grey at each instance. For example, CHILD has one allele from Profile 1, which is equally likely to be A or C, and one allele chosen according to \mathbf{b} . The diagram shows both the marginal GPM for a parental genotype (“A PARENT”), and also the joint GPMs of both parents (“ONE PARENT” and “THE OTHER”, both included in a box). A similar situation applies to the grandparents through a specified parent.

If G denotes a GPM for an individual, we will write $R(G)$ to denote the GPM derived from G for a specified relative, using one or more instances of (3) or (4). Note that $R(B) = B$, since the probability distribution for an individual of unknown genotype is the same as for any of their relatives. In place of hypothesis η_1 , we now contrast η_0 with

η_R : the individual underlying G^1 is an R -relative of the individual underlying G^2 .

Theorem 2

$$LR(\eta_R, \eta_0) = \sum_{ij} \frac{G_{ij}^1 \cdot R(G_{ij}^2)}{B_{ij}} \quad (5)$$

See Appendix A3 for proof. Equation (5) simplifies to standard LR formulae for relatives (see e.g. [9]) in the special case that the GPMs G^1 and G^2 assign probability 1 to a single genotype at each marker.

5 Mutation models.

In Figure 1, and in deriving (5), we assumed no mutation at the locus under consideration in the genetic lineage linking the two relatives. That assumption was for convenience only and is not required.

Mutations occur at forensic STR loci roughly once every 500 generations in males, and once every 2000 generations in females [10]. About 97% of mutations change the repeat count by one. Thus the stepwise mutation model, in which all mutations are between alleles that differ by one repeat unit, is incorrect but can provide a good approximation. The further simplifying assumption of symmetry (steps up as likely as steps down) may be adequate over small numbers of generations, but steps up are more common [10]. We introduce a mutation matrix M , whose ij th entry is the probability that a child receives a j allele given that the parental allele was an i . Thus $1-M_{ii}$ is the total mutation rate for an i allele.

To compute the GPM of an individual, given the GPM of their relative and allowing for mutation in parent-child transmissions, we proceed as described in Section 4 and illustrated in Figure 1 except that a parental allele probability vector \mathbf{p} should be replaced with $\mathbf{p}M$ when computing the child's GPM. The population allele probability vector \mathbf{b} is assumed to be the same across generations. This implies $\mathbf{b}M = \mathbf{b}$, which holds at least approximately for realistic M . Theorem 2 continues to hold if R is replaced with R_M , denoting the transformation of a GPM due to relatedness allowing for mutation.

6 Population structure and coancestry

Even if two individuals have no close relatedness, they may share common ancestors a handful of generations in the past, so that their genotypes are positively correlated and the independence assumption underpinning Theorems 1 and 2 is not valid. The simplest scenario in which coancestry is important is the case that, under η_0 , the individuals whose genotypes underlie G^1 and G^2 both come from the same subpopulation of the population from which \mathbf{b} has been obtained. A dependence between the genotypes arises because the genotype of either one of them is informative about the local allele probabilities which alters the probability distribution for the other individual. A mathematical model has been developed for such dependence, sometimes called the Balding-Nichols conditional model [7]. It is the basis of equation 4.10 in the NRCII report [11]. It assumes in effect that the local allele probability vector has a Dirichlet distribution with mean \mathbf{b} and variance determined by the population genetics parameter F_{ST} (sometimes called θ). Theorems 1 and 2 assume $F_{ST} = 0$, corresponding to zero variance.

The details of the Balding-Nichols model are not important here. All we need is notation

for conditional genotype probabilities. B still specifies the marginal GPM of an unknown individual, but the GPM for a second unknown individual requires a GPM conditional on the genotype, say ij , of the first individual; we will denote this conditional GPM $B_{|ij}$. Thus $B_{pq|ij}$ is the pq entry of $B_{|ij}$, the probability that an unknown individual has genotype pq given the observation of an unrelated individual in the same subpopulation with genotype ij . See Appendix A3 for derivation of the LR which we now state, corresponding to Theorem 2 which allows for relatedness and mutation but now further extended to allow for coancestry between the two individuals under η_0 :

$$LR(\eta_R, \eta_0) = \frac{\sum_{ij} \frac{G_{ij}^1 \cdot R_M(G_{ij}^2)}{B_{ij}}}{\sum_{ij} G_{ij}^1 \cdot H_{ij}^2} ; H_{ij}^2 = \sum_{pq} \frac{G_{pq}^2 \cdot B_{pq|ij}}{B_{pq}} \quad (6)$$

The denominator of (6) we call the ‘‘GPM subpopulation correction factor’’. It reduces to 1 when $F_{ST} = 0$, in which case $B_{|ij} = B$.

Equation (6) is our most general expression for the LR.

7 Encoding an epg as a GPM

7.1 Using a statistical model

A number of statistical treatments of low-template DNA (LTDNA) profiles have been published in recent years[12, 13]. A 2010 review paper [14] distinguishes the *biological* or *consensus* model and the *statistical* model and goes on to assert:

‘‘It may seem unusual to state that one can and should interpret LT-DNA profiles without ever trying to infer what genotype(s) the epg(s) represent. However this is exactly what we advocate ...’’.

This viewpoint has prevailed, and recently new statistical models and software implementing them have been proposed [2]. LRs are calculated by summing over the possible genotypes of unknown contributors, using explicit models of processes such as dropout, dropin, stutter and degradation [15,16]. Some degree of expert judgement is still required in processing the epg into input data for the software, for example in encoding apparent artefacts.

In contrast, our approach uses the epg to infer contributor genotype(s) as a first step, but since the recent statistical models each imply a probability distribution for these genotypes there is no conflict. Indeed the statistical models can be used to derive contributor GPMs from an epg. However, here we emphasise that expert judgement can also form a valid basis for deriving GPMs: we return to discussing the advantages of our approach in the Conclusion below.

In the special case that G^1 specifies a unique genotype (a reference profile), encoding a full GPM G^2 is not required since only the entries corresponding to the genotype specified by G^1 are used in (6). However, the full GPM can be useful if there are multiple suspected contributors, such as when searching a database of possible contributors. Moreover when both genotypes being compared are uncertain, then our approach which uses two GPMs appears to be the only one available for computing LRs.

7.2 Expert assessment

When assigning genotype probabilities, the expert makes assessments of the number of significant contributors and the relative amounts of DNA from each of them, based on the observed peaks, particularly their heights. Note that the expert can choose to specify a GPM only for the major contributor, or for the two most important contributors, without drawing inferences about the genotypes of lesser contributors. There is no requirement to assess the total number of contributors of DNA to the sample: this number is of little importance when some individuals contribute negligible amounts of DNA. The GPM assessments should take into account the genotypes of known contributors, and allow for stochastic phenomena such as variability in peak heights, as well as possible drop out and drop in. When there is uncertainty about the genotype of a contributor, for example if there is a single epg peak that may represent a homozygote genotype or a heterozygote with one allele dropped out, then the expert may also take into account a vector \mathbf{b} of background allele probabilities in assigning probabilities to dropout alleles.

The following examples illustrate, for a range of common situations, how an expert may take these factors into account. An encoding scheme and shorthand notation can facilitate this task, see Appendix A4 for further discussion.

7.2.1 One contributor of interest

If the contributor of interest is the major contributor of DNA to the sample, and allelic peaks are well above noise levels, encoding is reasonably straightforward. At loci where there are one or two peaks much higher than the others, the expert can assign high probability to a single genotype, whereas at other loci there may be more ambiguity about the major peaks, requiring probability judgements based on peak heights and possibly also \mathbf{b} .

Example 1: Single contributor, allowing for dropin and dropout

Consider two peaks with approximate height ratio of $A:B = 2:1$ at a locus with 3 alleles; A, B, C. In Table 1 we show three schemes for the expert to encode the epg information, all assuming that there is at least one A allele present, while the peak at B may be allelic or a dropin. Row (i) corresponds to coding the genotype as AF (see discussion above): it is assumed that the epg conveys no information about the second allele, which may have dropped out, and so the allele probabilities are given by \mathbf{b} . This corresponds to some current practice but it ignores relevant information.

		Genotypes		
		AA	AB	AC
options	i	b_A	b_B	b_C
	ii	Γ	$1-\Gamma$	0
	iii	$\Gamma(1-\Delta)+\Delta b_A$	$(1-\Gamma)(1-\Delta)+\Delta b_B$	Δb_C

Table 1: Genotype probability allocations for options (i) – (iii) discussed in the text.

In row (ii), the expert decides that no dropout has occurred and so the second allele is A with probability Γ , in which case the B peak is regarded as a dropin, otherwise (with probability $1-\Gamma$) the genotype of the contributor of interest is AB. Row (iii) represents an

interpolation between row (i) (case $\Delta = 1$) and row (ii) (case $\Delta = 0$). The value of Δ reflects the probability of dropout: $\Delta = 0$ implies zero dropout and so the AC genotype is impossible, whereas $\Delta = 1$ implies a high probability of dropout. A GPM resulting from a particular instance of the row (iii) coding is shown in Table 2.

	A	B	C
A	0.250	0.200	0.175
B	0.200	0	0
C	0.175	0	0

Table 2: EPG encoding from row (iii) of Table 1 with $b_A = 0.1, b_B = 0.2, b_C = 0.7, \Gamma = 0.4, \Delta = 0.5$

7.2.2 Two contributors of interest

It is difficult in general to specify a joint genotype distribution for multiple contributors, because for example the genotype probabilities for a second contributor will depend on genotype assignments for the first contributor. However, it is feasible in some settings.

If the differences in peak heights between the two contributors are large, it may be possible to infer the genotype of the major with certainty, in which case the task reduces to encoding the genotype of a minor with known major (see Example 2 below).

Our approach can also be used when the two main contributors can to some extent be distinguished through peak heights, but there remains some uncertainty about the genotype of the major at some loci, and considerable uncertainty about the minor at most loci. We have outlined above how to specify a GPM for the major contributor. A GPM for a minor can be similarly derived conditional on each possible genotype for the major. A marginal GPM for the minor is then obtained by summing over the major contributor genotypes, weighted by their probabilities. Then the GPMs for major and minor contributors can both be used to compute LRs as outlined above, for example to search against databases.

A joint genotype distribution can be represented, if required, as a 4-dimensional matrix of allele probabilities, or more conveniently as a 2-dimensional matrix of genotype probabilities as in Table 3.

Example 2 : Two distinguishable contributors.

Consider three peaks with heights A:B:C = 5:5:2. Suppose that the expert decides that the major is AB, and the minor includes at least one C allele, while the other is equally likely to be any of A, B or C, so that the joint genotype probabilities are as shown in Table 3. The corresponding GPMs for major and minor contributors are shown in Table 4 and Table 5.

		Major contributor					Minor Marginal
		AA	AB	AC	BB	BC	
Minor contributor	AA						
	AB						
	AC		0.33				0.33
	BB						
	BC		0.33				0.33
	CC		0.33				0.33
Major Marginal			1				

Table 3: Encoding of Example 2

	A	B	C
A	0	0.5	0
B	0.5	0	0
C	0	0	0

Table 4: GPM corresponding to Major in Example 2.

	A	B	C
A	0	0	0.167
B	0	0	0.167
C	0.167	0.167	0.333

Table 5: GPM corresponding to Minor in Example 2.

Example 3 : Two distinguishable contributors, neither certain

Consider four peaks with heights A:B:C:D = 5:3:3:2. The expert may decide that the four alleles are A, B, C and D; that the A and D peaks are major and minor alleles respectively, while B and C are each equally likely to be major and minor alleles¹. These assignments are shown in Table 6, while the corresponding marginal GPMs are shown in Table 7 and Table 8. Note that the joint genotype probabilities do not correspond to the product of the marginal probabilities. For example, there is uncertainty about the origin of the peaks at both B and C, but resolving one of them determines the other.

¹ In Example 3 since four alleles have been identified dropout may be ignored, and since the same four alleles are identified in either scenario, differences in their relative background frequencies do not affect the assignment.

		Major contributor				Minor Marginal
		AA	AB	AC	AD	
Minor contributor	AD					
	BD			0.5		0.5
	CD		0.5			0.5
	DD					
Major Marginal			0.5	0.5		

Table 6: Encoding of Example 3

	A	B	C	D
A	0	0.25	0.25	0
B	0.25	0	0	0
C	0.25	0	0	0
D	0	0	0	0

Table 7: GPM corresponding to Major in Example 3.

	A	B	C	D
A	0	0	0	0
B	0	0	0.25	0.25
C	0	0.25	0	0
D	0	0.25	0	0

Table 8: GPM corresponding to Minor in Example 3.

Example 4: Two indistinguishable contributors

Consider three peaks with heights A:B:C = 5:4:3. Suppose that the expert decides there is insufficient information to distinguish between contributors, and that alleles A, B and C are all present, while the fourth allele is likely to be another A but may be another B.

Using her expert judgement (taking into account the peak heights and background frequencies), suppose she assigns probabilities as to which alleles are present as follows:

$$P(AABC) = 0.9$$

$$P(ABBC) = 0.1$$

Writing p and q for two constants in the ratio 9:1, Table 9 assigns probability p to each genotype pair containing alleles AABC, and q to each pair containing ABBC. The row sums (= column sums) specify the marginal genotype distribution, and equating the overall sum to one specifies p and q (here $p = 9/120$, $q = 1/120$).

		First contributor					Marginal
		AA	AB	AC	BB	BC	
Second contributor	AA					2p	2p
	AB			4p		4q	4p+4q
	AC		4p		2q		4p+2q
	BB			2q			2q
	BC	2p	4q				2p+4q
Marginal		2p	4p+4q	4p+2q	2q	2p+4q	

Table 9: Joint genotype probabilities for two indistinguishable contributors in Example 4.

	A	B	C
A	18q = 0.15	20q = 0.17	19q = 0.16
B	20q = 0.17	2q = 0.02	11q = 0.09
C	19q = 0.16	11q = 0.09	0

Table 10: Marginal GPM corresponding to Table 1 ($q = 1/120$).

The GPM in Table 10 represents the probability distribution for either contributor to the mixture: since the expert does not attempt to deconvolve the mixture, the GPMs are the same for each contributor. If searched against a database of GPMs, a non-zero LR would be returned for any GPM that assigns a non-zero probability to any of the genotypes AA, AB, AC, BB or BC. The LR computed using GPMs for two non-deconvolved mixtures measures the evidence for the proposition that the mixtures have a contributor in common, without specifying the genotype of the common contributor.

For example, the LR for the proposition that the major contributor in example 3 is one of the contributors to example 4 is obtained by using the GPMs of Table 7 and Table 10 in Theorem 1, which gives $0.083/b_A b_B + 0.079/b_A b_C$ which equals 16.2 in the special case that $b_A = b_B = b_C = 0.1$.

8 Conclusion

We propose computing LRs to evaluate poor-quality DNA evidence (due to low-template, degradation, and/or multiple donors) using a probability distribution for the genotype(s) of donor(s). While the donor genotypes are strictly nuisance variables and it is possible to compute LRs without inferring them, we see several advantages to explicitly using genotype probability distributions for unknown or contested contributors. Firstly, this is an intuitive quantity for which an expert can reasonably assign probabilities, thus taking

advantage of their expert judgment. Secondly, summarising the information in a complex epg in terms of one or more genotype distributions allows rapid searching of even complex profiles against large databases, as well as the comparison of two complex profiles, perhaps from different crime scenes, to assess hypotheses about common contributors. A final advantage is that for an individual with a known genotype, probability distributions for the genotypes of their relatives are readily computed, allowing us to assess hypotheses about relatedness of contributors to different profiles even when one or both profiles are complex. We believe that our approach can assist both in rapid identification of intelligence leads and in evaluating evidence within a judicial process.

Although the use of “subjective” encoding based upon expert opinion may seem problematic, there is now a substantial literature on the elicitation of expert opinion in a subjective Bayesian methodology [17, 18]. Moreover the performance of experts can be tested in blind trials and through assessments of between-expert agreement. The expert can make conservative assessments, for example by assigning more weight to background frequencies for noisy profiles.

Initially the benefits of the method will be in its use as an intelligence tool; providing additional leads to an investigation. New and historical epgs that could not be used conventionally can now be searched and stored to the advantage of ongoing and “cold case” investigations. We envisage crime datasets made up of probabilistic epg encodings that would be a subset of, or complementary to, conventional national crime databases. Any possible matches suggested could then be subject to conventional examination. The use of the method within judicial systems would, we hope, follow. There are clear advantages in being able to give evidential strengths rather than a narrative along the lines of “could not be excluded as a donor to...”.

References

[1] Butler J, *Advanced Topics in Forensic DNA Typing: Interpretation*. Academic Press, ISBN 9780124052130, 2014.

[2] Steele CD, Balding DJ, Statistical evaluation of forensic DNA profile evidence. *Annu Rev Stat Appl*, 1: 361–384, 2014.

[3] Gill P, Guinness J, Iveson S, *The interpretation of DNA evidence (including low-template DNA)*, July 2012.
<https://www.gov.uk/government/publications/the-interpretation-of-dna-evidence>

[4] Scientific Working Group on DNA Analysis Methods. *SWGDM Interpretation Guidelines for Autosomal STR Typing*, 2010.
<https://www.fbi.gov/about-us/lab/biometric-analysis/codis/swgdam.pdf>

[5] Puch-Solis R, Kirkham AJ, Gill P, Read J, Watson S, Drew D. Practical determination of the low template DNA threshold. *Forensic Sci Int Genet* 2011 5(5):422-7. doi: 10.1016/j.fsigen.2010.09.001.

[6] Robertson B, Vignaux G, *Interpreting evidence: evaluating forensic science in the*

courtroom, Wiley 1995

[7] Balding D, Steele C, Weight of evidence for forensic DNA Profiles 2nd ed, Wiley (2015).

[8] Taroni F, Biedermann A, Bozza S, Garbolino P, Aitken C, Bayesian Networks for Probabilistic Inference and Decision Analysis in Forensic Science, Wiley, Chichester, UK (2014).

[9] Fung WK, Hu YQ. Statistical DNA Forensics: Theory, Methods, Computation. Wiley, Chichester, UK (2008).

[10] Lu D, Liu Q, Wu W, Zhao H, Mutation analysis of 24 short tandem repeats in Chinese Han population, International Journal of Legal Medicine, 126(2): 331-335 (2012).

[11] National Research Council, The Evaluation of Forensic DNA Evidence, National Academy Press, Washington DC (1996).

[12] Bright J-A, Evett IW, Taylor D, Curran JM, Buckleton J. A series of recommended tests when validating probabilistic DNA profile interpretation software. Forensic Sci Int Genet 5(14):125-31 (2015).

[13] Cowell RG, Graverson T, Lauritzen SL, Mortera J, Analysis of forensic DNA mixtures with artefacts, Appl. Statist. 64(1): 1–32, (2015)

[14] Gill P, Buckleton J, A universal strategy to interpret DNA profiles that does not require a definition of low-copy- number, Forensic Science International: Genetics 4: 221-227 (2010).

[15] Gill P, Whitaker J, Flaxman C, Brown N, Buckleton J, An investigation of the rigor of interpretation rules for STRs derived from less than 100 pg of DNA, Forensic Science International 112: 17-40 (2000).

[16] Buckleton J, Triggs C, Walsh S, Forensic DNA Evidence Interpretation, CRC Press, Boca Raton, Florida (2005).

[17] O'Hagan A, Buck C, Daneshkhah A, Eiser J, Garthwaite P, Jenkinson D, Oakley J, Rakow T, Uncertain Judgements: Eliciting Experts' Probabilities, Wiley, Chichester, UK (2006).

[18] Cooke R, Experts in Uncertainty: Opinion and Subjective Probability in Science, Oxford University Press, New York (1991)

A1 Theorem 1

Given eggs E^1 and E^2 and hypotheses

η_1 : G^1 and G^2 represent the genotype of the same individual

η_0 : G^1 and G^2 represent the genotypes of two unrelated individuals

the likelihood ratio (LR) is

$$LR(\eta_1, \eta_0) = \frac{P(E^1, E^2 | \eta_1)}{P(E^1, E^2 | \eta_0)} \quad (\text{A1-0})$$

We write S^1 and S^2 for the genotypes of the contributors to E^1 and E^2 . Under η_1 we have $S^1 = S^2$ and the numerator is obtained by summing over the two alleles of the genotype.

Under η_0 , we marginalize over the four alleles $A_i A_j A_p A_q$ where $S^1 = A_i A_j$: $i \leq j$ and $S^2 = A_p A_q$: $p \leq q$. Then

$$LR = \frac{\sum_{ij} P(E^1, E^2 | S^1 = S^2 = A_i A_j, \eta_1) \cdot P(S^1 = S^2 = A_i A_j | \eta_1)}{\sum_{pqij} P(E^1, E^2 | S^1 = A_i A_j, S^2 = A_p A_q, \eta_0) \cdot P(S^1 = A_i A_j, S^2 = A_p A_q | \eta_0)} \quad (\text{A1-1})$$

We now make the following assumptions

1. In the numerator, E^1 and E^2 are assumed independent given the underlying genotype $A_i A_j$ so that $P(E^1, E^2 | S^1 = S^2 = A_i A_j, \eta_1)$ becomes $P(E^1 | S^1 = A_i A_j) \cdot P(E^2 | S^2 = A_i A_j)$.
2. $P(S^1 = S^2 = A_i A_j | \eta_1)$ is the background genotype probability B_{ij} .
3. In the denominator $P(E^1, E^2 | S^1, S^2, \eta_0) = P(E^1 | S^1 = A_i A_j) \cdot P(E^2 | S^2 = A_p A_q)$ again assuming E^1 and E^2 are independent given the genotypes.
4. $P(S^1 = A_i A_j, S^2 = A_p A_q | \eta_0) = P(S^1 = A_i A_j | \eta_0) \cdot P(S^2 = A_p A_q | S^1 = A_i A_j, \eta_0) = B_{ij} \cdot B_{pq|ij}$ using the notation for \mathbf{B}_{ij} introduced in Section 6.

Then

$$LR = \frac{\sum_{ij} P(E^1 | S^1 = A_i A_j) \cdot P(E^2 | S^2 = A_i A_j) \cdot B_{ij}}{\sum_{ij} \left[P(E^1 | S^1 = A_i A_j) \cdot B_{ij} \cdot \sum_{pq} P(E^2 | S^2 = A_p A_q) \cdot B_{pq|ij} \right]} \quad (\text{A1-2})$$

This equation contains terms of the form $P(E | S = A_i A_j)$. This is appropriate when considering explicit models of the generation of eggs from profiles. We now use Bayes' theorem to write the LR in terms of the form $G_{ij} = P(S = A_i A_j | E)$, which correspond to entries of GPM matrices. By Bayes' theorem:

$P(E | S = A_i A_j) = G_{ij} P(E) / B_{ij}$ where $P(E)$ is the *prior probability* of the egg E .
Substituting this expression into (A2-2) we get

$$LR = \frac{\sum_{ij} \frac{G_{ij}^1 P(E^1)}{B_{ij}} \cdot \frac{G_{ij}^2 P(E^2)}{B_{ij}} \cdot B_{ij}}{\sum_{ij} \left[\frac{G_{ij}^1 P(E^1)}{B_{ij}} \cdot B_{ij} \cdot \sum_{pq} \frac{G_{pq}^2 P(E^2)}{B_{pq}} \cdot B_{pq|ij} \right]}$$

And simplifying

$$LR = \frac{\sum_{ij} \frac{G_{ij}^1 \cdot G_{ij}^2}{B_{ij}}}{\sum_{ij} \left[G_{ij}^1 \cdot \sum_{pq} \frac{G_{pq}^2 \cdot B_{pq|ij}}{B_{pq}} \right]} \quad (A1-3)$$

If S^1 and S^2 are independent then $B_{pq|ij} = B_{pq}$ and the denominator simplifies to unity (Since $\sum_{ij} G_{ij}^1 = \sum_{ij} G_{ij}^2 = 1$), which establishes Theorem 1.

In the general case $B_{pq|ij} \neq B_{pq}$ (A1-3) should be used.

A2 Relatedness

In this section we derive formulae equivalents to Figure 1 section 4.

We define $Rel_R(A_p A_q)_{ij}$ to be the probability that an R-relative of an individual with genotype $A_p A_q$ has genotype $A_i A_j$.

The function $Rel_R(A_p A_q)_{ij}$ is calculated on the following assumptions:

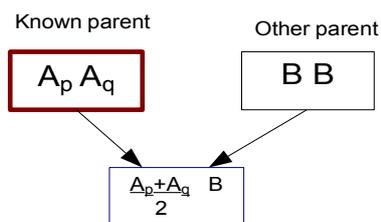
- (a) Mendelian segregation with independent assortment.
- (b) Genes contributed by unknown individuals are drawn from the background allele probability vector \mathbf{b} .

A2.1 Degree 1

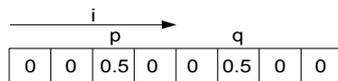
We consider first the parent-child relationship under the assumption of HW Equilibrium. We will show that the GPMs for “Child of X” and “Parent of X” are the same.

A2.1.1 Child

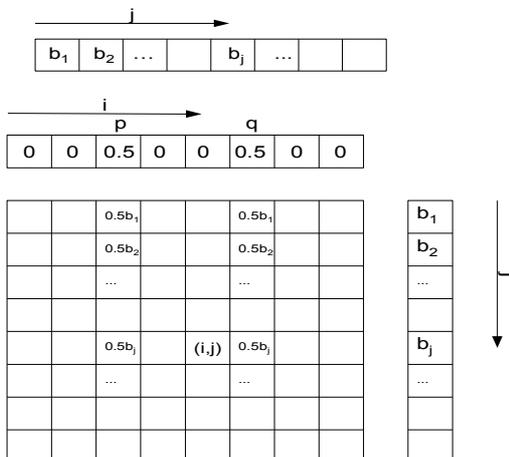
We seek an expression for $Rel_{Child}(A_p A_q)_{ij}$ (i.e. component (i, j) of the matrix representing $Rel_{Child}(A_p A_q)$) or in words: “The probability that the child of an individual with genotype $A_p A_q$ is $A_i A_j$ ”



The child of an individual with genotype $A_p A_q$ will inherit A_p or A_q , each with probability 0.5. The inherited allele may be written as the vector² $(\delta_{ip} + \delta_{iq})/2$



The other parent contributes an allele distributed according to the background probabilities \mathbf{b} . The matrix $Rel_{Child}(A_p A_q)$ is then a symmetric product of these allele vectors:

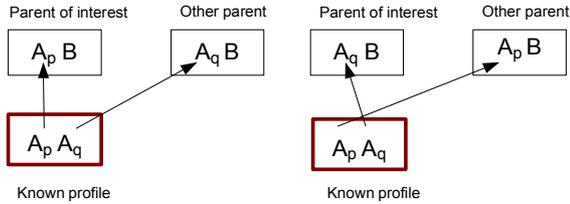


² Where δ_{ij} is the Kronecker delta symbol, defined to be equal to 1 if $i = j$ and 0 otherwise.

and it can be seen that the general term ((i,j) in the diagram) is

$$Rel_{Child}(A_p A_q)_{ij} = \frac{(\delta_{ip} + \delta_{iq})}{2} b_j \quad (A2-1)$$

A2.1.2 Parent



In the case of a parent there are two cases to consider: The parent contributed either A_p or A_q , with the other allele drawn from \mathbf{b} . Each of these cases may be constructed as above, and then averaged. (In the diagrams, arrows represent deductions, rather than gene flow).

$$Rel_{Parent}(A_p A_q)_{ij} = \frac{1}{2}(\delta_{ip} b_j + \delta_{iq} b_j) = \frac{(\delta_{ip} + \delta_{iq})}{2} b_j \quad (A2-2)$$

It can be seen the expressions for a parent and a child are identical, and we term both these relationships *Degree 1 or D1()*.

A2.2 Degree n

We term an n th generation descendant or ancestor (i.e. (n-2) greats grandchild or grandparent) a *Degree n* relative $Dn()$. Applying the construction for degree 1 repeatedly yields

$$Rel_{Dn}(A_p A_q)_{ij} = \frac{(\delta_{ip} + \delta_{iq} + (2^n - 2) b_i)}{2^n} b_j \quad (A2-3)$$

As special cases, we have the formulae for the D2 and D3 cases:

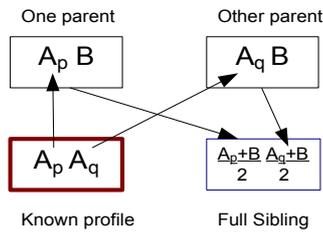
$$Rel_{D2}(A_p A_q)_{ij} = \frac{(\delta_{ip} + \delta_{iq} + 2 b_i)}{4} b_j \quad (A2-4)$$

$$Rel_{D3}(A_p A_q)_{ij} = \frac{(\delta_{ip} + \delta_{iq} + 6 b_i)}{8} b_j \quad (A2-5)$$

A2.3 Sibling

The full sibling relationship is the simplest example of a *collateral bilineal* relationship (the individuals have common ancestors via both parents), which may be constructed as follows:

$$Rel_{Sib}(A_p A_q)_{ij} = \frac{1}{2}(b_i + \delta_{pi}) \cdot \frac{1}{2}(b_j + \delta_{qj}) \quad (A2-6)$$



NB in this case adding the alternative term obtained by interchanging p and q is equivalent to adding the symmetric term obtained by interchanging i and j , so it is omitted. This is not always true for bilinear relationships.

In the general case where the profile is represented by the GPM \mathbf{G}

$$Rel_{Sib}(\mathbf{G})_{ij} = \sum_{pq} G_{pq} \cdot \frac{1}{2}(b_i + \delta_{pi}) \cdot \frac{1}{2}(b_j + \delta_{qj})$$

which may be simplified to

$$Rel_{Sib}(\mathbf{G})_{ij} = \frac{1}{4}(b_i b_j + b_i \sum_p G_{pj} + b_j \sum_q G_{iq} + G_{ij}) \quad (A2-7)$$

The LR for $A_p A_q$ and $A_r A_s$ being siblings (versus unrelated) is from Theorem 2)

which simplifies to

$$LR(A_p A_q, Rel_{Sib}(A_r A_s)) = \frac{1}{8} \left[\left(1 + \frac{\delta_{pr}}{b_p}\right) \left(1 + \frac{\delta_{qs}}{b_q}\right) + \left(1 + \frac{\delta_{qr}}{b_q}\right) \left(1 + \frac{\delta_{ps}}{b_p}\right) \right] \quad (A2-8)$$

This formula reproduces all seven cases given in Table (A2-1) below, in agreement with published formulae, e.g. [13] at p137, Table 4.10.

P1	P2	LR (Full Sibling/Unrelated)
xx	xx	$(1+b_x)^2 / 4 b_x^2$
xx	xy	$(1+b_x) / 4 b_x$
xx	yy	1/4
xx	yz	1/4
xy	xy	$(1+b_x+b_y+2b_x b_y) / (8 b_x b_y)$
xy	xz	$(1+2b_x) / 8 b_x$
xy	zw	1/4

Table (A2-1): Sibling Likelihood Ratio

A2.4 Tables

Relationship	Components of $Rel(A_p A_q)_{ij}$	Vector notation: GPM of relative, given GPM X with marginal x , background $B = b^T b$
D1	$(\delta_{ip} + \delta_{iq}) b_j / 2$	$(x^T b + b^T x) / 2$
Dn	$(\delta_{ip} + \delta_{iq} + (2^n - 2) b_i) b_j / 2^n$	$(x^T b + b^T x + (2^n - 2) B) / 2^n$
Full Sibling	$(b_i + \delta_{pi}) \cdot (b_j + \delta_{qj}) / 4$	$(X + x^T b + b^T x + B) / 4$

Table (A2-2): Relationship formulae

D1	Parent, Child
D2	Grandparent, Grandchild, Uncle, Nephew, Half-sibling
D3	Great-grandparent, Great-grandchild, Great-aunt, Great-niece, First Cousin

Table (A2-3): Examples of relationships

A3 Theorem 2

Given an individual with GPM \mathbf{G} , the GPM \mathbf{G}^R of a relative of that individual may be expressed as a function of \mathbf{G} : $\mathbf{G}^R = R(\mathbf{G})$ where function $R()$ depends on the particular relationship, e.g. we may have $\mathbf{G}^R = Child(\mathbf{G})$, $\mathbf{G}^R = Sib(\mathbf{G})$ etc.

We now show how the likelihood ratio may be expressed in terms of $R()$.

We consider two GPMs G^1 and G^2 representing eggs E^1 and E^2 respectively. We consider the LR for the following hypotheses:

η_R : the individual underlying G^1 is an R -relative of the individual underlying G^2 .

η_0 : G^1 and G^2 come from unrelated individuals

Again we write S^1 and S^2 for the (unknown) genotypes of the contributors to E^1 and E^2 . And $Rel_R(A_p A_q)_{ij}$ is defined as in Appendix A2. Then under η_R the GPM for the relative S^1 is given by

$$R(\mathbf{G}^2)_{ij} = \sum_{pq} G_{pq}^2 Rel_R(A_p A_q)_{ij} \quad (\text{A3-1})$$

Proceeding as in A2 we define the likelihood ratio for a familial match

$$LR = \frac{P(E^1, E^2 \mid \eta_R)}{P(E^1, E^2 \mid \eta_0)} \quad (\text{A3-2})$$

First, marginalize both numerator and denominator over the exclusive and exhaustive set of alleles $A_p A_q A_i A_j$ where $S^1 = A_i A_j$: $i \leq j$ and $S^2 = A_p A_q$: $p \leq q$:

$$LR = \frac{\sum_{pqij} P(E^1, E^2 \mid S^1 = A_i A_j, S^2 = A_p A_q, \eta_R) \cdot P(S^1 = A_i A_j, S^2 = A_p A_q \mid \eta_R)}{\sum_{pqij} P(E^1, E^2 \mid S^1 = A_i A_j, S^2 = A_p A_q, \eta_0) \cdot P(S^1 = A_i A_j, S^2 = A_p A_q \mid \eta_0)}$$

We proceed as in Appendix A1 with regard to the denominator.

In the numerator we have $P(E^1, E^2 \mid S^1, S^2, \eta_R) = P(E^1 \mid S^1, S^2, \eta_R) \cdot P(E^2 \mid E^1, S^1, S^2, \eta_R)$. Since under η_R the sources of S^1 and S^2 are assumed to be related, it is *not* true that E^1 is independent of S^2 , nor that E^2 is independent of S^1 and E^1 . However it is clear that if S^1 were known, knowledge of S^2 or E^2 would not add any new information about E^1 . Therefore $P(E^1 \mid S^1, S^2, \eta_R) = P(E^1 \mid S^1, \eta_R)$ and similarly $P(E^2 \mid E^1, S^1, S^2, \eta_R) = P(E^2 \mid S^2, \eta_R)$. Then

$$LR = \frac{\sum_{pqij} P(E^1 \mid S^1 = A_i A_j) \cdot P(E^2 \mid S^2 = A_p A_q) \cdot P(S^1 = A_i A_j \mid S^2 = A_p A_q, \eta_R) \cdot P(S^2 = A_p A_q \mid \eta_R)}{\sum_{pqij} P(E^1 \mid S^1 = A_i A_j) \cdot P(E^2 \mid S^2 = A_p A_q) \cdot P(S^2 = A_p A_q \mid S^1 = A_i A_j, \eta_0) \cdot P(S^1 = A_i A_j \mid \eta_0)}$$

Noting that $P(S^1 = A_i A_j \mid S^2 = A_p A_q, \eta_R) = Rel_R(A_p A_q)_{ij}$, and substituting in the background probabilities:

$$LR = \frac{\sum_{pqij} P(E^1 | S^1 = A_i A_j) \cdot P(E^2 | S^2 = A_p A_q) \cdot Rel_R(A_p A_q)_{ij} \cdot B_{pq}}{\sum_{pqij} P(E^1 | S^1 = A_i A_j) \cdot P(E^2 | S^2 = A_p A_q) \cdot B_{pq|ij} \cdot B_{ij}}$$

Again by Bayes' theorem we have $P(E^1 | S^1 = A_i A_j) = G_{ij}^1 P(E)/B_{ij}$, and $P(E^2 | S^2 = A_p A_q) = G_{pq}^2 P(E)/B_{pq}$ and simplifying with the help of (A3-1) we have

$$LR = \frac{\sum_{ij} \left[\frac{G_{ij}^1 \cdot R(\mathbf{G}^2)_{ij}}{B_{ij}} \right]}{\sum_{ij} \left[G_{ij}^1 \cdot \sum_{pq} \frac{G_{pq}^2 \cdot B_{pq|ij}}{B_{pq}} \right]} \quad (\text{A3-3})$$

Under the same assumption as Appendix A1 (i.e. HWE or an unconditional population model) we have

$$LR = \sum_{ij} \frac{G_{ij}^1 \cdot R(\mathbf{G}^2)_{ij}}{B_{ij}}$$

which establishes theorem 2. In the general case $B_{pq|ij} \neq B_{pq}$ (A3-3) should be used.

A3.1 Example

As an example in the calculation of a LR for a familial relationship, consider a locus with alleles with STR repeat numbers (8, 9, 10, 11) with background frequencies (0.1, 0.2, 0.3, 0.4) respectively. Alice has profile (8,F) and Bob has profile (8, 9). This corresponds to the following matrices (written for convenience in upper-triangular form).

$$\text{Alice} = \begin{bmatrix} 0.1 & 0.2 & 0.3 & 0.4 \\ & 0 & 0 & 0 \\ & & 0 & 0 \\ & & & 0 \end{bmatrix}; \quad \text{Bob} = \begin{bmatrix} 0 & 1 & 0 & 0 \\ & 0 & 0 & 0 \\ & & 0 & 0 \\ & & & 0 \end{bmatrix}; \quad \text{Background} = \begin{bmatrix} 0.01 & 0.04 & 0.06 & 0.08 \\ & 0.04 & 0.12 & 0.16 \\ & & 0.09 & 0.24 \\ & & & 0.16 \end{bmatrix};$$

Suppose we wish to find the LR for the relationship "Alice is a sibling of Bob" (vs "Alice and Bob are unrelated") We first calculate the matrix for "Sibling of Bob" from Equation A2-6

$$Sib(\mathbf{Bob})_{ij} = \sum_{pq} Bob_{pq} \frac{1}{2}(b_i + \delta_{pi}) \cdot \frac{1}{2}(b_j + \delta_{qj})$$

Since the only element of $Bob_{pq} \neq 0$ is $Bob_{10} = 1$ we find

$$Sib(\mathbf{Bob})_{ij} = \frac{1}{4}(b_i + \delta_{0i}) \cdot (b_j + \delta_{1j}) \text{ and therefore}$$

$$Sib(\mathbf{Bob})_{00} = \frac{1}{4}(b_0 + \delta_{00}) \cdot (b_0 + \delta_{10}) = \frac{1}{4}(0.1 + 1) \cdot (0.1 + 0) = 0.0275$$

$$Sib(\mathbf{Bob})_{01} = \frac{1}{4}(b_0 + \delta_{00}) \cdot (b_1 + \delta_{11}) = \frac{1}{4}(0.1 + 1) \cdot (0.2 + 1) = 0.3300 \text{ etc,}$$

giving the components of the matrix shown below.

$$Sib(\mathbf{Bob}) = \begin{bmatrix} 0.0275 & 0.3300 & 0.0825 & 0.1100 \\ 0.0050 & 0.0600 & 0.0150 & 0.0200 \\ 0.0075 & 0.0900 & 0.0225 & 0.0300 \\ 0.0100 & 0.1200 & 0.0300 & 0.0400 \end{bmatrix} \text{ or } \begin{bmatrix} 0.0275 & 0.3350 & 0.0900 & 0.1200 \\ & 0.0600 & 0.1050 & 0.1400 \\ & & 0.0225 & 0.0600 \\ & & & 0.0400 \end{bmatrix}$$

The LR for a sibling relationship between Alice and Bob is obtained by matching "Alice" vs "Sibling of Bob" using Equation (5):

$$\Sigma \begin{bmatrix} 0.1 & 0.2 & 0.3 & 0.4 \\ & 0 & 0 & 0 \\ & & 0 & 0 \\ & & & 0 \end{bmatrix} \times \begin{bmatrix} 0.0275 & 0.3350 & 0.0900 & 0.1200 \\ & 0.0600 & 0.1050 & 0.1400 \\ & & 0.0225 & 0.0600 \\ & & & 0.0400 \end{bmatrix}$$

$$\begin{bmatrix} 0.01 & 0.04 & 0.06 & 0.08 \\ & 0.04 & 0.12 & 0.16 \\ & & 0.09 & 0.24 \\ & & & 0.16 \end{bmatrix}$$

The result of which is

$$\frac{0.1 \times 0.0275}{0.01} + \frac{0.2 \times 0.3350}{0.04} + \frac{0.3 \times 0.09}{0.06} + \frac{0.4 \times 0.12}{0.08} = 0.275 + 1.675 + 0.45 + 0.6 = 3$$

A4 Encoding as vectors in GPMDNA

The first two authors have developed a computer program GPMDNA to calculate LR_s using the method described in this paper. GPMDNA allows the expert to encode an epg as a single GPM for a major contributor, or as two marginal GPMs for a major and a minor as discussed in the main text.

An expert using GPMDNA may also encode an N -contributor GPM as $2N$ allele vectors, each normalised to 1 and representing the probability distribution of one allele. The allele vectors are used by GPMDNA to produce marginal GPMs for contributors to the mixture. Encoding GPMs as products of allele vectors was used in Section 4 for propagating an individual's GPM to his/her relatives. Using allele vectors for interpreting an epg is not completely general, because it implies that allele assignments are independent, but we have had considerable experience of this approach and found it be very flexible, permitting good approximations for a wide range of judgements that an expert might wish make.

We introduce a shorthand notation to specify allele vectors. Any probability not explicitly assigned by the expert is assumed by GPMDNA to be a multiple of background probabilities (denoted '@B'), chosen so that each allele vector is normalised to 1. The B values can be supplied by the software so the expert does not need to know them at the time of encoding. We now illustrate the allele vector encoding and notation using the examples in the main text, assuming a locus with STR alleles 11, 12, ... and associated background probabilities b_{11}, b_{12}, \dots .

Example 1

Having decided there is a single contributor the expert designates the two allele vectors \mathbf{u} and \mathbf{v} , and the program generates the components as shown in Table A4-1.

		STR allele			
Case	Allele vector	11	12	13	GPMDNA designation
(i)	\mathbf{u}	1	0	0	11
	\mathbf{v}	b_{11}	b_{12}	b_{13}	F
(ii)	\mathbf{u}	1	0	0	11
	\mathbf{v}	0.4	0.6	0	11@0.4/12@0.6
(iii)	\mathbf{u}	1	0	0	11
	\mathbf{v}	$0.2+0.5b_{11}$	$0.3+0.5b_{12}$	$0+0.5b_{13}$	11@0.2/12@0.3

Table A4-1: Allele vector encoding of Example 1. The pair of rows labelled (i), (ii) or (iii) correspond to the row of Table 1 with the same label. The columns show the allele probability assignments explicitly and in GPMDNA notation. In each case, vector \mathbf{u} indicates that allele 11 is certainly present, while \mathbf{v} specifies probabilities for the other allele. In (i) 'F' indicates that background probabilities are assigned to all alleles. In (ii) probabilities of 0.4 and 0.6 are assigned to 11 and 12, corresponding to $\Gamma=0.4$ in Example

1. In (iii) the allele probabilities shown have sum < 1; the residual probability is assumed to follow the background probabilities, corresponding to $\Gamma=0.4, \Delta=0.5$ in Table 1..

Example 2

Having decided there are two contributors the expert must designate four alleles : **u**, **v**, **w**, **z**. In this case the expert attempts to deconvolve the major and minor, and assigns **u**, **v** to the major and **w**, **z** to the minor.

		STR allele				
Contributor	Allele vector	11	12	13	14	GPMDNA designation
Major	u	1	0	0	0	11
	v	0	1	0	0	12
Minor	w	0	0	1	0	13
	z (i)	0.33	0.33	0.33	0	11/12/13@0.33
	z (ii)	$\frac{b_{11}}{b_{11}+b_{12}+b_{13}}$	$\frac{b_{12}}{b_{11}+b_{12}+b_{13}}$	$\frac{b_{13}}{b_{11}+b_{12}+b_{13}}$	0	11/12/13@B
	z (iii)	$\frac{b_{11}}{b_{11}+b_{12}}$	$\frac{b_{12}}{b_{11}+b_{12}}$	0	0	11/12@B

Table A4-2: GPMDNA encoding of Example 2 is shown in the rows down to and including **z(i)**. Also shown are two alternative codings of vector **z** that take the background probabilities into account for the fourth allele, either allowing for it to be any of alleles 11, 12 and 13 (vector **z(ii)**, coding “11/12/13@B”) or only one of 11 or 12 (vector **z(iii)** coding “11/12@B”).

The 4-allele GPM (shown in Table 3) may be constructed automatically as follows. The GPM for the major is formed from the allele vectors **u** and **v** as in Equation (2), and we will write this as

$$[\mathbf{uv}] = (\mathbf{u}^T \mathbf{v} + \mathbf{v}^T \mathbf{u})/2$$

Similarly the minor is $[\mathbf{wz}]$. We will always permute the vectors for each major or minor component of a profile because we can not distinguish the maternal and paternal alleles. When there is uncertainty about which contributor an allele belongs to, we must perform additional permutations (see examples 3, 4). In this case no more permutations are needed and the 4-allele GPM is just the outer product $A = [\mathbf{uv}] [\mathbf{wz}]$. In components $A_{ijkl} = (u_i v_j + v_i u_j)(w_k z_l + z_k w_l)/4$. The marginals for the major and minor (Tables 4 and 5) can be got from A , or directly as $[\mathbf{uv}]$ and $[\mathbf{wz}]$ respectively.

Example 3

In example 3 the GPMDNA designation is straightforward, shown in Table A4-3.

		STR repeat number				
Contributor	Allele vector	11	12	13	14	GPMDNA designation
Major	u	1	0	0	0	11
Major/ Minor	v	0	1	0	0	12
	w	0	0	1	0	13
Minor	z	0	0	0	1	14

TableA4-3: GPMDNA encoding of Example 3

Since the expert has flagged alleles **u** and **v** as belonging to either the major or the minor we must form all possible permutations to get the 4-allele GPM :

$([\mathbf{uv}] \quad [\mathbf{wz}] + [\mathbf{uw}] \quad [\mathbf{vz}])/2$ Alternatively we can calculate the 2-allele marginal GPMs directly: $([\mathbf{uv}] + [\mathbf{uw}])/2$ and $([\mathbf{wz}] + [\mathbf{vz}])/2$ for the major and minor respectively. These GPMs are shown in Tables 6, 7 and 8.

Example 4

In example 4 there is no attempt to assign alleles to major or minor, so we have:

		STR repeat number			
Allele vector	11	12	13	GPMDNA designation	
u	1	0	0	11	
v	0	1	0	12	
w	0	0	1	13	
z	0.9	0.1	0	11@0.9/12@0.1	

Table A4-4: GPMDNA encoding of Example 4.

Again we can form the 4-allele GMP of Table 9 by permutation if we need it:

$([\mathbf{uv}] \quad [\mathbf{wz}] + [\mathbf{uw}] \quad [\mathbf{vz}] + [\mathbf{uz}] \quad [\mathbf{vw}] + [\mathbf{vw}] \quad [\mathbf{uz}] + [\mathbf{vz}] \quad [\mathbf{uw}] + [\mathbf{wz}] \quad [\mathbf{uv}])/6$ while forming the permuted product $([\mathbf{uv}] + [\mathbf{uw}] + [\mathbf{uz}] + [\mathbf{vw}] + [\mathbf{vz}] + [\mathbf{wz}])/6$ yields the marginal of Table 10 directly.

GPMDNA Software

The software may be downloaded from: <https://github.com/GPMSoftware/GPM.git>



Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Ryan, K;Williams, DG;Balding, DJ

Title:

Probabilistic evaluation of low-quality DNA profiles

Citation:

Ryan, K., Williams, D. G. & Balding, D. J. Probabilistic evaluation of low-quality DNA profiles

Persistent Link:

<http://hdl.handle.net/11343/118558>