Spiro-epoxyglycosides as Activity-Based Probes for Glycoside Hydrolase Family 99 Endomannosidase/Endomannanase


Abstract: N-Glycans direct protein function, stability, folding and targeting, and influence immunogenicity. While most glycosidases that process N-glycans cleave a single sugar residue at a time, enzymes from glycoside hydrolase family 99 are endo-acting enzymes that cleave within complex N-glycans. Eukaryotic Golgi endo-1,2-α-mannosidase cleaves glucose-substituted mannose within immature glucosylated high-mannose N-glycans in the secretory pathway. Certain bacteria within the human gut microbiota produce endo-1,2-α-mannanase, which cleaves related structures within fungal mannan, as part of nutrient acquisition. An unconventional mechanism of catalysis was proposed for enzymes of this family, hinted at by crystal structures of imino/azasugars complexed within the active site. Based on this mechanism, we developed the synthesis of two glycosides bearing a spiro-epoxide at C-2 as electrophilic trap, to covalently bind a mechanistically important, conserved GH99 catalytic residue. The spiro-epoxyglycosides are equipped with a fluorescent tag, and following incubation with recombinant enzyme, allow concentration, time and pH dependent visualization of the bound enzyme using gel electrophoresis.

Introduction

N-Linked glycans are complex oligosaccharides linked to asparagine (Asn) residues in eukaryotic proteins.[1] They play important roles in protein function, stability, folding and targeting and are essential for a range of cellular functions.[2] Erroneous N-glycan composition is associated with various diseases including viral infections, Alzheimer’s disease and metastatic cancer.[3–5] Assembly of the N-glycan commences in the endoplasmic reticulum (ER) where the 14-mer polysaccharide Glc3Man9GlcNAc2-diphosphodolichol is coupled to the Asn residue of the target protein by the enzyme oligosaccharidyl transferase. The glycan undergoes stepwise ”trimming” of the non-reducing end glucose residues by α-glucosidase I and II, after which α-mannosidase I truncates the resulting oligomannoside.[6] The resulting Man3GlcNAc2 structure is redecorated to yield complex N-glycans. Because α-glucosidases I and II play important roles in the early stages of glycan maturation, these enzymes were investigated as therapeutic targets to control diseases involving incorrect N-glycosylation.[7–10] However, inhibition of these enzymes did not block N-glycosylation; mouse lymphoma cells inhibited with the α-glucosidase inhibitor castanospermine as well as mutant cell lines lacking α-glucosidase II retained up to 80% of normal N-glycan maturation.[11–13] Spiro and co-workers identified endo-1,2-α-mannosidase,[14, 15] (later classified as a member of glucoside hydrolase family 99 (GH99); see http://cazypedia.org),[16] residing in the Golgi apparatus, which circumvents inhibition of ER α-glucosidase I and II. The enzyme cleaves glucose-substituted mannose from the A-branch of ER escaped immature N-glycoproteins bearing Glc3-Man9GlcNAc2, releasing Glc3-Man. The resulting Man3GlcNAc2 glycoprotein subsequently re-enters the normal processing route in the Golgi apparatus.

Bacterial GH99 orthologs including Bacteroides thetaiotaomicron (Bt) and Bacteroides xylanisolvens (Bx) enzymes possess endo-1,2-α-mannosidase activity, but are more appropriately described as endo-1,2-α-mannanases, as they act on yeast...
mannan and exhibit a tenfold preference for mannan-based substrates versus the equivalent glucose-substituted mannans. Several imino/azasugar inhibitors for GH99 endomannosidase have been developed, including α-glucopyranosyl-1,3-isofagomine (GlcIFG, 1, Figure 1) and α-mannopyranosyl-1,3-isofagomine (ManIFG, 2). Due to a preference for a manno- pyranosyl residue in subsite -2 GH99 endo-1,2-α-mannanases show a greater affinity for 2 than for 1. Recently, mannoeuromycin (ManNOE, 3), which features a 2-hydroxyl allowing interaction with the proposed general base/acid residue, has been reported as the most potent endo-1,2-α-mannanase inhibitor for bacterial GH99 enzymes with \( K_D \) values in the low nanomolar range. Additionally, fluorescent and fluorogenic substrates have been developed for monitoring endo-1,2-α-mannosidase/mannanase activity.

Family GH99 endo-1,2-α-mannosidases/mannanases cleave their substrate glycosides with retention of anomeric stereochemistry; however, instead of the classical Koshland double-displacement mechanism for retaining enzymes, an unusual neighboring group participation hydrolytic mechanism was proposed in which a glutamate residue (Glu333 in BxGH99) acts as a general base assisting OH-2 to displace the aglycon via a 1,2-anhydro sugar that is subsequently hydrolyzed by water (Figure 1B). In order to study enzyme function in biological settings, screen for inhibitors, as well as to further illuminate the catalytic reaction mechanism, the development of a mechanism-based irreversible inhibitor would be of interest. Here, the synthesis is described of two putative covalent inhibitors 4 and 5, designed to, respectively inhibit eukaryotic GH99 endo-1,2-α-mannosidases and bacterial endo-1,2-α-mannanases and which vary in the nature of the pyranoside at the non-reducing end (Figure 1A, right). Both compounds contain a spiro-epoxide at position C-2 to serve as an electrophile to trap the general base residue. Inspection of the crystal structures of BxGH99 suggests that the general base will be situated close to the methylene group of the spiro-epoxide, where it may open the ring via nucleophilic attack resulting in a covalent intermediate (Figure 1C). The compounds are also equipped with a reporter tag, allowing active enzyme labeling by activity-based protein profiling (ABPP) protocols, the efficiency of which is reported as well.

**Results**

Acceptor 7 was synthesized by 4,6-silylidene protection of compound 6, followed by formation of the 2,3-orthobenzoate and final treatment with acid (Scheme 1). Glucopyranoside donor 9 was synthesized from thiophenyl β-glucopyranoside 8. While 4,6-silyliden protection proceeded smoothly, elevated temperatures were required to install the TBS-

**Figure 1.** (A) Known GH99 endo-1,2-α-mannosidase inhibitors (1–3) and fluorescent spiro-epoxyglycosides 4 and 5 subject of this study. \( K_D \) values are for *B. thetaiotaomicron* endo-1,2-α-mannosidase (BtGH99). (B) The proposed catalytic mechanism for GH99 enzyme (amino acid numbering for *B. xylanisolvens* endo-1,2-α-mannosidase (BxGH99)). (C) Anticipated covalent inhibition mechanism of GH99 enzymes.
groups onto both secondary hydroxyl groups, presumably due to steric hindrance. Using a similar approach, thiophenyl \(\alpha\)-mannopyranoside \(10^{[26]}\) was converted to protected thiomanoside donor \(11\).

Glycosylation of acceptor \(7\) by \(9\) or \(11\) was achieved in an N-iodosuccinimide (NIS)/trimethylsilyl triflate (TMSOTf) mediated coupling at low temperature, affording \(12\) or \(12\) respectively. Both glycosylations proceeded in excellent yield and stereoselectivity. Pedersen, Boll and co-workers\(^{[29]}\) recently reported that silylideneprotected mannosyl donors can be used for stereoselective \(\beta\)-mannosylation. The contrasting selectivity obtained here is likely the result of the steric buttressing effect of the large silyl ether protecting groups at the C-2- and C-3-hydroxyls of \(7\), consistent with the steric effects that large protecting groups and functionalities have in glycosylations of otherwise \(\beta\)-selective benzylidene-protected mannosyl donors.\(^{[30]}\) Thus, the \(\beta\)-face of mannosyl donor \(11\) is shielded from attack by the incoming nucleophile. The high reactivity. The “arming” silyl protecting groups allow this donor to readily form an oxocarbenium ion, which will likely take up a \(\alpha\)-\(\text{H}_{2}\)like conformation, which is preferentially attacked from the \(\alpha\)-face to provide the 1,2-\(\text{cis}\)-linked product.\(^{[31,32]}\) Next, the benzyl groups were deprotected under Zemplén conditions affording compounds \(13\) and \(13\). The alcohol was then oxidized with Dess–Martin periodinane (DMP) to ketones \(14\) and \(14\), which appeared to be in equilibrium with the corresponding hydrates.

Transformation of ketones \(14\) into their corresponding epoxides was explored next (Table 1). Reaction of \(14\) with diazomethane as methylenating agent\(^{[33]}\) resulted in the formation of the equatorial \(15\) and axial \(16\) methylenes in a 1:1 ratio and in good yields (entry 1). Their absolute configuration was determined by 1D-NOE difference experiments (see Supplementary Information). Reaction of \(14\) with diazomethane also resulted in a mixture of \(15\) and \(16\), in a 3:1 ratio, in favor of the equatorial methylene group in almost quantitative yield (entry 2). We anticipated that a Corey–Chaykovsky epoxidation\(^{[34]}\) using stabilized dimethylsulfoxonium methyldie would favor the formation of the equatorial methylenes \(15\) and \(15\). Indeed, also in these cases both isomers were obtained, however the formation of axial methylenes was still favored in both cases (entries 3 and 4). Finally, using the more reactive dimethylsulfoxonium methyldie, only the kinetically favored axial methylenes \(16\) and \(16\) were formed, albeit in moderate yields (entries 5 and 6). With epoxides \(16\) and \(16\) in hand, global deprotection was accomplished by reaction with tetrabutylammonium fluoride (TBAF). Finally, a fluorescent Cy5 tag was installed at the azide handle using copper(I) catalyzed click chemistry, which after HPLC purification afforded spiro epoxyglycosides \(4\) and \(5\).

The ability of \(4\) and \(5\) to label recombinant \(Bt\)- and \(Bx\)GH99 endo-1,2-\(\alpha\)-mannanase was evaluated (Figure 2A). The compounds label both enzymes in a concentration-dependent manner, at concentrations as low as 100 nM. Previous studies indicated a preference for a mannosyl residue at the \(-2\) subsite of both enzymes.\(^{[18]}\) However, no difference in potency of labeling was observed. Studies on the effect of the pH dependence on labeling revealed that both spiro epoxyglycosides label the enzymes maximally at pH 6–8, corresponding to

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**Table 1.** Transformation of ketones \(14\) and \(14\) into their corresponding spiro epoxides.

<table>
<thead>
<tr>
<th>Entry</th>
<th>s.m.</th>
<th>Conditions</th>
<th>15:16 Yield [%]</th>
</tr>
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<tbody>
<tr>
<td>1</td>
<td>14g</td>
<td>CH(_2)N(_2), EtOH, 0°C</td>
<td>1:1 78</td>
</tr>
<tr>
<td>2</td>
<td>14m</td>
<td>CH(_2)N(_2), EtOH, 0°C</td>
<td>3:1 97</td>
</tr>
<tr>
<td>3</td>
<td>14g</td>
<td>SOMe(_2), nBuLi, THF, 0°C</td>
<td>1:5 83</td>
</tr>
<tr>
<td>4</td>
<td>14m</td>
<td>SOMe(_2), nBuLi, THF, 0°C</td>
<td>1:2 85</td>
</tr>
<tr>
<td>5</td>
<td>14g</td>
<td>SMe(_2), NaH, DMSO, THF, 0°C</td>
<td>0:1 50</td>
</tr>
<tr>
<td>6</td>
<td>14m</td>
<td>SMe(_2), NaH, DMSO, THF, 0°C</td>
<td>0:1 53</td>
</tr>
</tbody>
</table>

[a] Combined yield after column chromatography. s.m. = starting material.
the pH optimum of GH99 enzymatic activity (Figure 2B).[23] Notably, more than one band is evident, suggesting enzyme degradation under reducing SDS-PAGE conditions or alternatively that multiple labelling events may be occurring. Next, labeling of wild-type (WT) BxGH99 was compared to analogous active-site mutants (Figure 2C). While WT enzyme is labeled by spiro-epoxyglycosides 4 and 5 within 5 minutes, the general base mutant E333Q and the catalytic acid mutant E336Q were not labeled in the same time period with these compounds, suggesting that labeling is indeed activity-based, and is consistent with reaction occurring in a mechanism-based manner. However, incubation for longer times resulted in labeling of the mutant enzymes, indicating that either the spiro-epoxide is susceptible to ring opening by the mutant catalytic residues, or that other residues may also be involved in covalent labeling. Denaturation of BtGH99 and BxGH99 completely abrogated labeling by spiro-epoxyglycosides 4 and 5, indicating that labeling requires the natively folded enzyme (Figure 2D).

To further evaluate whether covalent inhibition of BtGH99 and BxGH99 is activity-based, the processing of human a-galactosidase A (GLA) by these enzymes was investigated (Figure 3A). GLA contains three N-glycosylation sites, of which two are decorated with oligo-mannose structures, and one contains complex oligosaccharides low in mannose content.[35, 36] We have previously demonstrated that fluorescent a-galacto-cyclophellitol aziridines such as TB340 covalently label GLA in activity-based manner.[37] Here, GLA was pre-labeled with TB340 to enable fluorescent detection on gel. Without additives, GLA gives a distinct major band at ≈50 kDa (Figure 3B, lane 1). Incubation of GLA with BtGH99 results in demannosylation of the two high-mannose N-glycans of GLA, resulting in a shift of the GLA band into lower bands at ≈42 kDa (lane 2). This shift in molecular weight is similar to the shift observed when GLA is incubated with Endo-H (lane 4), which causes demannosylation of high-mannose N-glycans by cleaving within the chitobiosyl core leaving a residual GlcNAc on Asn. Treatment of GLA with PNGase-F

![Figure 2. Fluorescent labeling of GH99 endomannanases. (A) Detection limit of Bt and Bx GH99 endomannanases (left and right, respectively), labeled with various concentrations of fluorescent spiro-epoxyglycosides 4 or 5. (B) Effect of pH on labeling of Bt and Bx GH99 enzymes with 4 or 5. (C) Labeling of wild-type and mutant BxGH99 with 4 or 5 (left or right, respectively) for 5, 15 or 30 minutes. (D) Effect of denaturation with 1% (w/v) SDS and boiling on labeling of Bt and Bx GH99 enzymes (left and right, respectively) with 4 or 5. The marker is annotated with an asterisk (*).](image)

![Figure 3. (A) Schematic representation of processing of human a-galactosidase GLA by GH99 endomannosidase. GLA is pre-labeled by fluorescent TB340, and contains high-mannose N-glycans which can be truncated by endomannosidase, resulting in a decrease in GLA molecular weight. Activity-based labeling of endomannosidase by spiro-epoxyglycosides 4 or 5 (prior to incubation with GLA) blocks its activity, and is therefore unable to process GLA. (B) BtGH99 wild-type demannosylates GLA, causing a shift in molecular weight for the protein bands. Pre-labeling BtGH99 wild-type with 4 or 5 abrogates GLA demannosylation. Endo-H cleaves high-mannose structures, PNGase-F cleaves full N-linked glycan (leaving Asp-GlcNAc). (C) BxGH99 wild-type demannosylates GLA, while BxGH99 pre-labeled with 4 or 5 is unable to do so. BxGH99 active-site mutants E333Q and E336Q are unable to process GLA. (D) Fluorescent labeling of BtGH99 (top) and BxGH99 (bottom) by 4 or 5 competed by different concentrations of 17, 18, ManIFG (2) and yeast mannan. The marker is annotated with an asterisk (*).](image)
(lane 5), which cleaves most N-glycans leaving Asn, results in a band of a lower molecular weight, most likely as a result of complete deglycosylation of all three N-glycans. When βTG9H9 was pre-incubated with 4 or 5, demannosylation of TB340-labeled GLA was mostly inhibited (lane 3), indicating that binding of 4 and 5 occurs in the βTG9H9 active site. An identical experiment was conducted with βxGH99 wildtype and the E333Q and E336Q mutant enzymes (Figure 3C). Similar to βTG9H9, wildtype βxGH99 is able to process the N-glycans decorating the surface of the enzyme, giving rise to a shift in molecular weight (lane 2), similar to processing by Endo-H (lane 8). Pre-incubation of WT βxGH99 by 4 or 5, prior to consecutive incubation with TB340-labeled GLA resulted in an observed absence of glycan processing (lane 3), indicating that binding of 4 and 5 abrogates enzymatic activity. Interestingly, while mutants E333Q and E336Q are labeled by spiro-epoxyglycosides 4 and 5 after prolonged reaction times, they are evidently unable to process TB340-labeled GLA (lanes 4–7).

Finally, the inhibitory potencies of 2, 17, 18 and yeast mannan from S. cerevisiae (an α-1,6-linked mannose backbone branched with α-1,2 and α-1,3 mannoses)[39] towards βTG9H9 were investigated using spiro-epoxyglycoside 5 as fluorescent read-out (Figure 3D). The enzyme was first pre-incubated with the competitor for 30 min at 37 °C, followed by labeling with 1 μM 5. Compounds 17 and 18 both show a concentration-dependent competition of fluorescent labeling in the range of 10–1,000 μM, although full competition with labeling could not be achieved under these conditions. Similarly, the azasugar ManII FG (2) gave concentration-dependent competition but again full competition was not achieved. However, pre-incubation by yeast mannan achieved full competition with labeling, suggesting that processing of spiro-epoxyglycoside 5 by βTG9H9 endomannanase is specific and activity based. A similar competition experiment was performed for βxGH99, and it was shown that while pre-incubation with 17 did not fully abrogate labeling, pre-incubation with 1000 μM of 18 provided full competition, possibly hinting at a slight preference for a mannosyl residue in subsite –2. Additionally, yeast mannan showed concentration dependent (albeit incomplete) competition, and azasugar 2 fully competed with labeling at 1,000 μM, suggesting that processing of spiro-epoxyglycoside 5 by βxGH99 is specific and activity-based.

Discussion

Epoxide-based probes have been investigated as mechanism-based inhibitors of a range of glycosidases. Early work led to the development of epoxalkyl glycosides,[39] which were initially proposed as reagents that could specifically label the nucleophile of retaining glycosidases, however X-ray crystallography later revealed labelling of both acid/base and nucleophile residues.[40] In one classic study, conduritol C epoxide, which was originally believed to label the nucleophile of E. coli LacZ β-galactosidase, was subsequently shown to covalently label the acid/base catalyst.[41] Work from our laboratory has investigated related pyranose-mimicking cyclophellitol epoxides and aziridines and shown that these typically exhibit excellent selectivity for labelling the nucleophile of assorted α- and β-glycosidases.[42] We have shown that introduction of a reporter tag (e.g. biotin or a fluorescent dye) onto these small molecule inhibitors affords chemical probes that enable quantification of activity,[43, 44] and have distinct advantages over techniques such as transcription analysis and antibody-based detection. We report here the first activity-based probes for detection of GH99 enzymes, which were designed based on the proposed mechanism of this enzyme. In this proposed mechanism a 1,2-anhydro-epoxide intermediate is formed by general base assisted deprotonation of O2 by a carboxylate residue.[23] Our design strategy includes a reactive C2 spiro-epoxide that can potentially covalently label the general base (acting as a nucleophile), and includes a fluorescent label for visualization. Gel-based analysis of labelled bacterial GH99 endo-1,2-α-mannanases demonstrated concentration dependent labelling which occurs in a pH dependent manner consistent with the pH optimum of enzyme activity. Labelling could be competed by various substrates and inhibitors, providing evidence that it is active-site directed. While mutation of the key general base and general acid residues inactivated the enzyme towards processing of natural substrate N-glycans in GLA, the mutants could be labelled with the spiro-epoxyglycosides, albeit with reduced potency. Collectively, our data suggests that these spiro-epoxides do result in labelling at the active site, presumably through the catalytic general base. However, the high reactivity of the primary epoxide means that labelling is most likely not exclusive at a single residue. While endo-1,2-α-mannanase has a preference for mannosyl residues at the −2 binding subsite, there was minimal differences in the efficiency of labelling for spiro-epoxides bearing either a mannosyl or glucosyl residue. We believe these compounds represent an important first step in devising probes that take advantage of the unique mechanism proposed for this family. Future studies will seek to better understand the mode of labelling by identifying the covalently labelled residue(s) by X-ray crystallography or MS based techniques. By analogy to previously described irreversible cyclophellitol activity-based probes,[43] we propose these fluorescent spiro-epoxyglycosides could ultimately lead to chemical tools for functional investigation of GH99 endo-1,2-α-mannosidase/mannanases, both as isolated species and in tissue extracts.

Experimental Section

Chemicals were purchased from Acros, Sigma Aldrich, Biosolve, VWR, Fluka, Merck and Fisher Scientific and used as received unless stated otherwise. Tetrahydrofuran (THF), N,N-dimethylformamide (DMF) and toluene were stored over molecular sieves before use. All reactions were performed under an argon atmosphere unless stated otherwise. TLC analysis was performed using Screening Device b.v. silica gel sheets (Silica gel 60 F254) with the indicator ninhydrin. CHROMATOGRAPHY was performed using Screening Device b.v. silica gel (particle size of 40–63 μm, pore diameter of 60 Å) with the indicated eluents. For reversed-phase HPLC purifications an Agilent Tech-

nologies 1200 series instrument equipped with a LTQ Orbitrap mass spectrometer (Thermo Finnigan), equipped with an electrospray ion source in positive mode (source voltage 3.5 kV, sheath gas flow 10 mL min⁻¹, capillary temperature 250 °C) with resolution R = 60000 at m/z 400 (mass range m/z 150–2000) and diocetyl phthalate (m/z 391.28428) as lock mass. The high-resolution mass spectrometer was calibrated prior to measurements with a calibration mixture (Thermo Finnigan). ManiFiG was prepared as previously reported. Recombinant expression of B. thetaiotaomicron (8) and B. xylanisolvens (8x) GPH9 was achieved as previously described. Recombinant α-galactosidase (GLA) was purchased from Genzyme (Cambridge, MA, USA). The α-galactosidase ABP TBS40 was synthesized as described earlier. Yeast mannann from S. cerevisiae was purchased from Sigma.

Synthesis and characterization

**(4aR,6S,7S,8aR,9S)-6-(3-Azidoxypropoxy)-2,2-di-tert-butylyl-8-hydroxyhexahydropyrano[3,2-d][1,3,2]dioxasilin-7-yl benzoate (7):** Compound 6**19** (1.00 g, 3.80 mmol) was co-evaporated with dry toluene and dissolved in dry DMF (38 mL). The resulting solution was cooled to −50 °C and SiBu₃(OTf) (1.11 mL, 3.42 mmol, 0.9 EQ) and 2,6-lutidine (0.44 mL, 3.80 mmol) were added. The reaction was stirred for 30 minutes and subsequently quenched with brine (400 mL). The aqueous layer was extracted with Et₂O (4×100 mL). The combined organic layers were washed with 1M aqueous HCl (2×100 mL), H₂O (100 mL) and brine and dried over Na₂SO₄. The solvents were removed under reduced pressure and the crude product was purified by gradient column chromatography (EtOAc/pentane, 1:4 to 1:2). The 4,6-silylde product was obtained as white solid (970 mg, 70%). **1H NMR** (400 MHz, CDCl₃): δ = 4.81 (d, J = 1.4 Hz, 1H), 4.11 (dd, J = 10.0, 5.0 Hz, 1H), 4.07–4.00 (m, 2H), 3.96 (t, J = 10.2 Hz, 1H), 3.86–3.76 (m, 2H), 3.69 (td, J = 9.1, 2.5 Hz, 1H), 3.96 (t, J = 9.1 Hz, 1H), 3.90 (t, J = 10.2 Hz, 1H), 3.68 (t, J = 9.0 Hz, 1H), 3.60 (t, J = 8.7 Hz, 1H), 3.51–3.37 (m, 2H), 2.92 (s, 1H), 2.77 (s, 1H), 1.04 ppm (s, 9H), 0.98 ppm (s, 9H).

**13C NMR** (101 MHz, CDCl₃): δ = 132.9, 131.7, 129.1, 128.3, 86.6, 77.8, 76.4, 74.5, 71.8, 66.1, 27.4, 27.0, 22.7, 19.9 ppm. IR (neat): α = 3241, 2932, 1659, 1477 ppm. HRMS (ESI) m/z [M + Na]⁺: calcd for C₂₆H₂₂O₃SNa 343.16319, found 343.16315.

**4,6-silylde product (1.0 g, 2.42 mmol) was co-evaporated with toluene (3 x), dissolved in dry pyridine (5 mL) and cooled to 0 °C. DMAP (30 mg, 0.24 mmol) and TBSOTf (5 mL) were added and the mixture was heated to 60 °C and stirred overnight. The mixture was carefully diluted with water (25 mL) and extracted with DCM (3 x 50 mL). The combined organic layers were washed with aq. 1M HCl (3 x 25 mL) and brine, dried over Na₂SO₄, filtered and concentrated. The crude product was purified by gradient column chromatography (pentane/EtOAc, 400:1 to 200:1), affording the title product as a white solid (850 mg, 93%). Analytical data were in accordance with those reported in literature.**

**(4aR,6R,7S,8aR,9R)-2,2-Di-tert-butylyl-7,8-bis(tert-butylimethylsilyloxy)-6-(phenylthio)hexahydropyrano[3,2-d][1,3,2]dioxasilin (9):** Compound 8**20** (2.6 g, 9.5 mmol) was dissolved in dry DMF (100 mL) under Ar-atmosphere. The mixture was cooled to −50 °C and 2,6-lutidine (3.3 mL, 28.5 mmol) and SiBu₃(OTf) (3.4 mL, 10.5 mmol) was added. After 2,6-lutidine (3.3 mL, 28.5 mmol) was added and the solution was allowed to reach room temperature, the mixture was subsequently quenched with H₂O (100 mL). The water layer was extracted with EtOAc (3 x 100 mL). The organic layers were combined and washed with H₂O (2 x 200 mL) and brine (200 mL) and dried over MgSO₄. The solvents were removed under reduced pressure and the crude product was purified by gradient column chromatography (EtOAc/pentane, 1:4 to 1:2). The 4,6-silylde product was obtained as a white solid (3.58 g, 91%). **1H NMR** (400 MHz, CDCl₃): δ = 7.57–7.46 (m, 2H), 7.38–7.28 (m, 3H), 4.60 (d, J = 9.7 Hz, 1H), 4.21 (dd, J = 10.2, 5.1 Hz, 1H), 3.90 (t, J = 10.2 Hz, 1H), 3.68 (t, J = 9.0 Hz, 1H), 3.60 (t, J = 8.7 Hz, 1H), 3.51–3.37 ppm (2H, 2H), 2.92 (s, 1H), 2.77 (s, 1H), 1.04 ppm (s, 9H), 0.98 ppm (s, 9H).

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\[ m/z: [M+H]^+ \text{ calc for } C_{23}H_{36}O_{5}SiS_{5}Na 641.35420, \text{ found } 641.36460.\]

\[(4aR,6S,7S,8R,8aR)-2,2-di-tert-butyl-8-\]

\[\text{[(4aR,6R,7R,8S,8aR)-2,2-di-tert-butyl-7,8-bis[(tert-butylidimethyl-}

\[\text{silyloxy)hexahydropyran(3,3-d,1,3),2]dioxsilin-6-yl]oxy}hexa-

\[\text{hydropyran(3,3-d,1,3),2]dioxsilin-7-yl benzoate (12g): Compound 12g}

\[\text{m/z: [M+Na]^+ calc for } C_{24}H_{37}O_{5}NaSiS_{5}Na 956.53099, \text{ found } 956.53097.\]

\[\text{for C}_{29}\text{H}_{59}\text{O}_{10}\text{SiS}_{19} \text{Na} \text{ calc for } 1219.4874, \text{ found } 1218.4905.\]

\[\text{for C}_{30}\text{H}_{61}O_{11}\text{SiS}_{20} \text{Na} \text{ calc for } 1333.5346, \text{ found } 1333.5367.\]

\[\text{for C}_{31}\text{H}_{63}O_{12}\text{SiS}_{21} \text{Na} \text{ calc for } 1447.5722, \text{ found } 1447.5745.\]

\[\text{for C}_{32}\text{H}_{65}O_{13}\text{SiS}_{22} \text{Na} \text{ calc for } 1561.6098, \text{ found } 1562.6110.\]

\[\text{for C}_{33}\text{H}_{67}O_{14}\text{SiS}_{23} \text{Na} \text{ calc for } 1679.6474, \text{ found } 1679.6497.\]

\[\text{for C}_{34}\text{H}_{69}O_{15}\text{SiS}_{24} \text{Na} \text{ calc for } 1797.6850, \text{ found } 1797.6873.\]

\[\text{for C}_{35}\text{H}_{71}O_{16}\text{SiS}_{25} \text{Na} \text{ calc for } 1915.7226, \text{ found } 1915.7249.\]

\[\text{for C}_{36}\text{H}_{73}O_{17}\text{SiS}_{26} \text{Na} \text{ calc for } 2033.7592, \text{ found } 2033.7615.\]

\[\text{for C}_{37}\text{H}_{75}O_{18}\text{SiS}_{27} \text{Na} \text{ calc for } 2151.7958, \text{ found } 2151.7981.\]

\[\text{for C}_{38}\text{H}_{77}O_{19}\text{SiS}_{28} \text{Na} \text{ calc for } 2269.8324, \text{ found } 2269.8347.\]

\[\text{for C}_{39}\text{H}_{79}O_{20}\text{SiS}_{29} \text{Na} \text{ calc for } 2387.8690, \text{ found } 2387.8713.\]

\[\text{for C}_{40}\text{H}_{81}O_{21}\text{SiS}_{30} \text{Na} \text{ calc for } 2505.8956, \text{ found } 2505.8979.\]

\[\text{for C}_{41}\text{H}_{83}O_{22}\text{SiS}_{31} \text{Na} \text{ calc for } 2623.9222, \text{ found } 2623.9245.\]

\[\text{for C}_{42}\text{H}_{85}O_{23}\text{SiS}_{32} \text{Na} \text{ calc for } 2741.9488, \text{ found } 2741.9511.\]

\[\text{for C}_{43}\text{H}_{87}O_{24}\text{SiS}_{33} \text{Na} \text{ calc for } 2859.9754, \text{ found } 2860.9778.\]

\[\text{for C}_{44}\text{H}_{89}O_{25}\text{SiS}_{34} \text{Na} \text{ calc for } 2977.9920, \text{ found } 2977.9943.\]

\[\text{for C}_{45}\text{H}_{91}O_{26}\text{SiS}_{35} \text{Na} \text{ calc for } 3095.0086, \text{ found } 3095.0109.\]

\[\text{for C}_{46}\text{H}_{93}O_{27}\text{SiS}_{36} \text{Na} \text{ calc for } 3213.0252, \text{ found } 3213.0275.\]

\[\text{for C}_{47}\text{H}_{95}O_{28}\text{SiS}_{37} \text{Na} \text{ calc for } 3331.0418, \text{ found } 3331.0441.\]

\[\text{for C}_{48}\text{H}_{97}O_{29}\text{SiS}_{38} \text{Na} \text{ calc for } 3449.0584, \text{ found } 3449.0607.\]

\[\text{for C}_{49}\text{H}_{99}O_{30}\text{SiS}_{39} \text{Na} \text{ calc for } 3567.0740, \text{ found } 3567.0763.\]

\[\text{for C}_{50}\text{H}_{101}O_{31}\text{SiS}_{40} \text{Na} \text{ calc for } 3685.0906, \text{ found } 3685.0929.\]
(25a,4a,R,6,5,8,5,8a,R)-6′-([3-Azidopyrrolo]2,2′-di-tert-butyl-butylicate-7-bis([tert-butyl(dimethyl)silyl]oxy)hexahydropropano[3,2-d][1,3,2]dioxasilin-6-yloxy)tetrahydro-6-H-spironorxane-2,7-pyran-3,2-d[1,3,2]dioxasiliniline (15g) and (2R,4a,R,6,5,8,5,8a,R)-6′-([3-Azidopyrrolo]2,2′-di-tetrahydro-6,6-bis([tert-butyl(dimethyl)silyl)oxy]hexahydropropano[3,2-d][1,3,2]dioxasilin-6-yloxy)tetrahydro-6-H-spironorxane-2,7-pyran-3,2-d[1,3,2]dioxasiliniline (16g): Method A: *dimethyl sulfoxide methylene: A 1 M solution of dimethyl-sodium was prepared from sodium hydride (60 wt %, 200 mg, 5 mmol) in dry DMSO (2.5 mL) and heating this mixture to 70 °C for 1 h. The olive-green solution was diluted to room temperature and diluted with dry THF (2.5 mL). A fraction (0.12 mL, 0.19 mmol) of this mixture was added to a dried flask containing a mixture of tetrabutylammonium iodide (26.3 mg, 0.129 mmol) in dry DMSO (0.43 mL) and dry THF (0.4 mL) was added drop wise and the mixture was stirred for 5 minutes. Then, compound 14g (100 mg, 0.11 mmol, co-evaporated with toluene (3×) beforehand) in dry THF (0.64 mL) was added and the mixture was stirred for 30 minutes. The mixture was diluted with water (20 mL) and extracted with EtO/0 phosphate (2:1, 4× 15 mL). The combined organic layers were washed with water (20 mL), dried over Na2SO4, filtrated and concentrated. The crude product was purified by gradient column chromatography (pentane/ETOA, 60:1 to 50:1) to afford solely product 15g (51 mg, 50%) as an oil.

Method B: *dimethyl sulfoxonim methylene: Trimethylsulfoxonium iodide (37.8 mg, 0.172 mmol) was suspended in dry THF (2 mL) and cooled to 0 °C. n-Butyllithium (2 mL in pentane, 80 μL, 0.16 mmol) was added and the mixture was heated to 60 °C. Compound 14g (100 mg, 0.11 mmol) was co-evaporated with toluene (3×), dissolved in dry THF (1 mL) and added drop wise to the ylide solution. After 10 minutes, the mixture was cooled to room temperature and quenched with MeOH (0.5 mL). The mixture was evaporated and the crude product was purified by gradient column chromatography (pentane/ETOC, 60:1) to give a mixture of compounds 15g and 16g (90 mg, ratio 15g:16g 1:5, total yield 88%) as a colorless oil.

Method C: *diazomethane: To a glass tube were added acq. KOH (40 %, 5 mL) and Et2O (20 mL) and this mixture was cooled to 0 °C. Then, 1-methyl-3-nitro-1-nitrosoguanidine (2.9 g, 10 mmol) was added in portions with swirling. A fraction (2 mL) of the bright yellow ether layer was added drop-wise to a solution of compound 14g (100 mg, 0.11 mmol) in EtOH (3 mL) at 0 °C. After stirring for 10 minutes, acetic acid (glacial) was added drop wise until the yellow mixture turned colorless. The mixture was concentrated and co-evaporated with toluene (3×). The crude products were purified by column chromatography (pentane/acetone, 150:1, affording compound 15g (10 mg, ratio 15g:16g 1:5, total yield 78%).

Data for compound 15g (equatorial methylene): 1H NMR (400 MHz, CDCl3): δ = 5.33 (d, J = 2.9 Hz, 1H), 4.27–4.15 (m, 3H), 4.18 (s, 1H), 4.10–4.01 (m, 2H), 4.00–3.95 (t, J = 10.1 Hz, 1H), 3.92–3.76 (m, 3H), 3.75–3.59 (m, 4H), 3.50–3.35 (m, 3H), 3.15 (d, J = 5.1 Hz, 1H), 2.61 (d, J = 5.1 Hz, 1H), 1.86 (quintet, J = 6.3 Hz, 2H), 1.04 (s, 9H), 1.04 (s, 9H), 0.98 (s, 18H), 0.91 (s, 9H), 0.90 (s, 9H), 0.13 (s, 3H), 0.10 (s, 3H), 0.09 (3H), 0.08 ppm (3H). 13C NMR (101 MHz, CDCl3): δ = 102.5, 97.4, 79.3, 78.5, 76.0, 74.8, 67.8, 67.7, 67.1, 66.3, 64.2, 59.4, 48.3, 46.4, 29.2, 29.1, 27.6, 27.2, 27.1, 26.3, 22.9, 22.7, 20.1, 18.4, 18.3, 18.3, –3.6, –3.6, –4.0 ppm. IR (neat): ν = 2930, 2858, 2099, 1472, 1252, 1091, 1043 cm⁻¹. [M+H]⁺ cated for C31H38N5O12Si, found 496.54904.

Data for compound 16g (axial methylene): 1H NMR (400 MHz, CDCl3): δ = 5.34 (d, J = 3.0 Hz, 1H), 4.40–4.22 (m, 2H), 4.09 (dd, J = 10.4, 5.1 Hz, 2H), 4.04–3.93 (m, 3H), 3.87–3.76 (m, 4H), 3.66 (t, J = 8.7 Hz, 1H), 3.56–3.39 (m, 4H), 3.23 (d, J = 5.6 Hz, 1H), 2.63 (d, J = 5.6 Hz, 1H), 1.89 (q, J = 5.7 Hz, 2H), 1.07 (s, 18H), 1.05 (s, 9H), 1.02 (s, 9H), 0.96 (s, 9H), 0.95 (s, 9H), 0.16 (s, 3H), 0.13 (s, 3H), 0.11 (s, 3H), 0.10 ppm (3H). 13C NMR (101 MHz, CDCl3): δ = 101.2, 96.9, 79.7, 78.2, 74.7, 73.8, 67.7, 67.4, 66.9, 66.8, 66.1, 64.4, 58.3, 48.1, 48.0, 29.0, 27.6 (3x), 27.3 (3x), 27.2 (20x) (3x), 26.5 (3x), 26.4 (3x), 22.8, 22.5, 20.1, 20.0, 18.2, 18.1, –3.08, –3.38 (2x), –4.03 ppm. IR (neat): ν = 2934, 2858, 2320, 2094, 1095, 1043, 827, 773 cm⁻¹. [M+H]⁺ cated for C31H38N5O12Si, found 496.54953.
in THF, 2.3 mL, 5 \times 10^{-5} \text{M}, J_8 = J_9 \text{ for } 30 \text{ min at } 37 ^\circ \text{C, to afford the title product (20 mg, 74 %).}

- B. xylanisolvens dissolved in McIlvaine buffer, pH 7.0 (citric acid-NaHPO4) for 1h at 37°C. This biomass was prepared from yeast mannan (1 mmol) as described for the preparation of 4 to afford the product (2.9 mg, 34 %) as a blue solid. 

- \( \text{H NMR (500 MHz, MeOD): } \delta = 8.24 \text{ (t, } J = 13.0 \text{ Hz, 2H), 7.90 \text{ (s, 1H), 7.49 \text{ (d, } J = 7.4 \text{ Hz, 2H), 7.44-7.39 \text{ (m, 2H), 7.28 (dt, } J = 16.4, 7.6 \text{ Hz, 4H), 6.62 \text{ (t, } J = 12.4 \text{ Hz, 1H), 6.28 \text{ (d, } J = 13.7 \text{ Hz, 2H), 5.19 \text{ (d, } J = 1.3 \text{ Hz, 1H), 4.85 \text{ (s, 1H), 4.35 (t, } J = 6.5 \text{ Hz, 2H), 3.10 \text{ (d, } J = 4.5 \text{ Hz, 1H), 2.81 \text{ (d, } J = 4.5 \text{ Hz, 1H), 1.95-1.78 \text{ ppm (m, 2H).}} \]

- \( \text{H NMR (101 MHz, D}_2\text{O): } \delta = 100.0, 99.2, 73.0, 72.5, 71.2, 71.5, 70.91, 70.4, 65.3, 60.3, 60.2, 58.7, 48.2, 48.1, 27.8 \text{ ppm. IR (neat): } \tilde{\nu} = 3369, 2927, 2108, 1521, 1026 \text{ cm}^{-1} \).

- The pH optimum was analyzed using 4 ng GH99 wild-type, E333Q and E336Q enzymes was assessed by incubating 400 ng for 5, 15 or 30 min with 1 \mu M 4 or 5 dissolved in Mcllvaine buffer, pH 7.0. The pH optimum was analyzed using 4 ng enzyme incubated with 1 \mu M 4 or 5 dissolved in Mcllvaine buffer, pH 3–8, for 30 min at 37°C. Time-dependent labeling of BxGH99 wild-type, E333Q and E336Q enzymes was assessed by incubating 400 ng for 5, 15 or 30 min with 1 \mu M 4 or 5 dissolved in Mcllvaine buffer, pH 7.0. The effect of denaturation was assessed on 4 ng wild-type BxGH99 and BxGH99 by boiling for 4 min at 100°C prior to incubating with 1 \mu M 4 and 5 for 30 min at 37°C. Competitive ABPP assays utilized 4 ng BxGH99 and BxGH99 enzyme that was pre-incubated with 10–1000 \mu M 17, 18 or MannF5G, or 0.3–30 \mu M 1–3 yeast mannan (S. cerevisiae), at pH 7.0 for 30 min at 37°C, followed by labeling with 1 \mu M 4 and 5 for 30 min at 37°C.

### Functional GLA assay

Recombinant \( \alpha \)-galactosidase GLA was diluted 1:2 in 50 mM Mcllvaine buffer, pH 4.6, and pre-labeled with 2 \mu M TB340 for 1 h at 37°C. Subsequently, the mixture was diluted to 1:500 in 150 mM Mcllvaine buffer, pH 7.0. In parallel, 400 ng BxGH99 wild-type, E333Q and E336Q were incubated in the presence or absence of 10 \mu M 4 or 5, dissolved in 150 mM Mcllvaine buffer, pH 7.0 for 1 h at 37°C. Subsequently, the BxGH99 mixture (10 \mu L) was incubated...
with 10 μL TB340-labeled GLA for 8 h at 37 °C. Hereafter, samples were denatured, separated on SDS-PAGE gel and visualized by fluorescence scanning, as described above (vide supra). As control, 10 μL TB340-labeled GLA was treated by either Endo-H or PNGase-F, following the manufacturer’s instructions (New England Biolabs).

Acknowledgements

We thank the Netherlands Organization for Scientific Research (NWO-CW ChemTherm grant to JMFGA and HS0), and the European Research Council (ERC-2011-AdG-290836 “Chembiosphering” to HS0, and ERC-2012-AdG-32294 “Glycopoise” to GJD). GJD thanks the Royal Society for the Ken Murray Research Professorship.

Conflict of interest

The authors declare no conflict of interest.

Keywords: activity-based probes · endomannosidase · GH99 · glycosidase · inhibitors

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Title:
Spiro-epoxyglycosides as Activity-Based Probes for Glycoside Hydrolase Family 99 Endomannosidase/Endomannanase

Date:
2018-07-11

Citation:

Persistent Link:
http://hdl.handle.net/11343/255266

File Description:
Published version

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