

Video Article

A Step Beyond BRET: Fluorescence by Unbound Excitation from Luminescence (FUEL)

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Abstract

Fluorescence by Unbound Excitation from Luminescence (FUEL) is a radiative excitation-emission process that produces increased signal and contrast enhancement *in vitro* and *in vivo*. FUEL shares many of the same underlying principles as Bioluminescence Resonance Energy Transfer (BRET), yet greatly differs in the acceptable working distances between the luminescent source and the fluorescent entity. While BRET is effectively limited to a maximum of 2 times the Förster radius, commonly less than 14 nm, FUEL can occur at distances up to μm or even cm in the absence of an optical absorber. Here we expand upon the foundation and applicability of FUEL by reviewing the relevant principles behind the phenomenon and demonstrate its compatibility with a wide variety of fluorophores and fluorescent nanoparticles. Further, the utility of antibody-targeted FUEL is explored. The examples shown here provide evidence that FUEL can be utilized for applications where BRET is not possible, filling the spatial void that exists between BRET and traditional whole animal imaging.

Video Link

The video component of this article can be found at <http://www.jove.com/video/51549/>

Introduction

The genetic modification of organisms, such as viruses^{1,2}, bacteria³, or small mammals⁴ to either induce or constitutively express bioluminescence, has been highly successful and widely demonstrated⁵⁻⁷. Bioluminescence, an *in vivo* chemiluminescent reaction involving naturally occurring reagents, has the advantage of producing light without the need for an external light source. As such, bioluminescent imaging does not suffer from the common drawbacks of auto- and non-specific signal found from fluorescence imaging⁸. Consequently, bioluminescence has a significant signal-to-noise ratio since any detected signal originates solely from the intended source. While many models have exploited the lux operon from *Photobacterium luminescens* (emission maximum centered between 480 and 490 nm) for *in vitro* and *in vivo* applications⁹, its use in small mammals has been problematic due to the very nature of the imaging conditions; the pervading existence of optical absorbers, such as hemoglobin, and scattering agents, such as tissue and bone, strongly affect blue to yellow wavelengths³. The expression of an engineered firefly luciferase (emission maximum at 617nm) has been recently developed and incorporated, providing a tool that greatly overcomes optical absorption¹⁰, but is still subject to scattering effects.

In response, there have been multiple attempts to red-shift the emitted signal into the desired optical window of 650-900 nm, a region of minimized absorption and scatter, using bioluminescence resonance energy transfer (BRET)¹¹⁻¹³. As a tool to enhance signal detection, BRET, which uses a bioluminescent source as the donor and an added fluorophore as the acceptor, has found limited success. As a seminal example of this phenomenon, "self-illuminating quantum dots" (SIQDs)¹⁴ consist of modified *Renilla reniformis* luciferases bound to the external polymer-lysine layer of commercially available quantum dots (QDs). Upon substrate addition, the resulting bioluminescent reaction induces fluorescence emission from the QDs, generating a significant production of red photons. However, these SIQDs have limited applicability to *in vivo* visualization of physiologically relevant events. This limited applicability is likely due to the difficulty of linking the dual probe to the organ, cell or gene of interest, since the SIQDs cannot be genetically encoded and therefore would require a secondary modification of the polymer shell. To improve their applicability, alternative SIQDs, where the luciferases are bound directly to the luminescent core, have recently been employed¹⁵. Building off of the SIQD concept, a more applicable BRET system was achieved by attaching *Cypridina* luciferase to an indocyanine

dye¹⁶, which was capable of specifically targeting tumors in mice while producing a substantial red shift from 460 nm to 675 nm. To undergo non-radiative energy transfer, BRET follows the same primary constraints as its fluorescent counterpart: there must be a strong spectral overlap between the donor emission and the acceptor excitation spectra and the working distance between the two moieties must be on the order of the Förster radius (5–14 nm depending on the donor-acceptor pair, with an effective maximum distance of twice the Förster radius¹⁷). This distance dependence greatly limits the types of events that can be observed using BRET as a means to enhance detection.

Recently a new approach was identified and demonstrated under both *in vitro* and *in vivo* conditions. Building off the foundation of BRET, Fluorescence by Unbound Excitation from Luminescence (FUEL)^{18,19} also requires a strong spectral overlap between the luminescent and fluorescent components. However, unlike BRET, FUEL is a completely radiative process whereby the emitted photon from the luminescent source is absorbed by an optically accessible fluorophore, which subsequently emits a red-shifted photon according to the fluorophore quantum yield. Akin to BRET, this approach can also be used to overcome the constraints of imaging in the presence of optical absorbers. The resulting red shift provides an overall increase and specificity in the detected signal due to a decrease in attenuation and a reduction of optical scattering effects. FUEL has been reported to occur between bioluminescent *Escherichia coli* expressing the *lux* operon and QDs^{18,19}. While experimentally similar to the SIQDs, a fundamental difference exists: in FUEL, it is not necessary for the luminescent source to be physically bound to the fluorophore, which allows for genetic encoding of the luminescent probe. Due to the successful detection of FUEL between luminescent bacteria and QDs, it is possible that this technique could be applied to both superficial (skin) and deep tissue (lung, liver) infections such as *Staphylococcus aureus* and *Klebsiella pneumoniae*.

Since the report of its experimental significance, FUEL has evolved to include a robust mathematical model²⁰ that can be used to predict acceptable luminescent and fluorescent pairs, and its applications have expanded to include use in identification of photophysical characteristics such as quantum yield. We describe below some of the basic techniques of FUEL. First, we show evidence for this phenomenon over both short (μm) and long (cm) working distances, which fundamentally distinguishes FUEL from BRET. Second, we expand upon the possible FUEL pairs by examining a wide variety of fluorophores and fluorescent nanoparticles. Third, FUEL applications are investigated by comparing targeted and non-targeted FUEL pairs.

Protocol

1. Reagents

1. Purchase or develop luminescent bacteria and appropriate culture media such as modified *Escherichia coli* expressing the *luxABCDE*²¹, *Vibrio fischeri*, *Photobacterium* sp. *Klebsiella pneumoniae*²².
2. Prepare physiological saline (0.9% NaCl) and culture media solutions according to standard recipes. *E. coli* and *K. pneumoniae* were grown in Luria Bertani (LB) at 37 °C, and *V. fischeri* and *Photobacterium* sp in LB supplemented with 0.5 M NaCl at 22 °C.
3. Purchase or acquire fluorescent probes such as Q-tracker 705 (40 nm diameter, referred to as QD705), Q-tracker 800 (40 nm diameter, referred to as QD800), fluorescent (40 nm diameter) and non-fluorescent polystyrene microspheres (48 nm diameter), and conventional fluorescent dyes.
4. Prepare the necessary reagents for bacterial labeling.
5. Ensure access to a whole animal bioluminescence imager, such as an IVIS Spectrum, that is capable of detecting signal under a wide range of emission wavelengths and exposure times. A plate reader capable of bioluminescence measurements will suffice for most of the experiments described here.

2. Basic Recapitulation of FUEL

1. Starting from individual colonies on standard culture plates, start overnight cultures of luminescent bacteria. Here, *V. fischeri* were used.
2. The day of experimentation, initiate fresh subcultures and allow them to progress until an OD₆₀₀ of 1–1.5 is achieved. In order to produce comparable results it is best to use bacteria in similar growth states in which they produce intense signals. This can be assured by keeping the OD₆₀₀ constant.
3. Combine aliquots of 100 μl from each with either 5 μl of QD705 or physiological saline (PS), and then add to 895 μl of PS into standard spectroscopic cuvettes. Place the filled cuvettes into the IVIS Spectrum and take the measurements under the appropriate filter sets. Here, the 710–730 nm emission filter was used.

3. FUEL Over Varying Distances

1. Fill two reduced volume (1 ml) plastic photometric cuvettes with either 50 μl of QD705 or 97.2 μl of non-fluorescent 48 nm polystyrene microspheres in a total volume of 1 ml PS. This ensures similar solid surface area per total volume between the two entities.
2. Prepare a light source cuvette by encasing a third cuvette with standard black tape or some other opaque material capable of blocking light. Carefully prepare two identical optical windows on opposite sides of the cuvette.
3. Place the previously filled cuvettes directly onto either side of the light source cuvette. Add a 1 ml aliquot of *V. fischeri* or other culture (*i.e.* luminescent *E. coli* or *Photobacterium* sp) from a fresh subculture into the light source cuvette and cover it with an opaque material, such as black paper to reduce any light contamination.
4. Visualize the three cuvettes under the appropriate emission filters such as the Total Light and 710–730 nm emission filters, with exposure times of 10, 30, and 30 sec, respectively.
5. Reposition both external cuvettes at equivalent further distances from the central cuvette, and then visualize again. Repeat until a final face-to-face (central to reduced volume cuvette) distance of 3cm is achieved. At each step, acquire a fluorescence image (450–480 nm excitation, 710–730 nm emission) to validate the QD705 location.

4. Investigating Potential FUEL Pairs

1. Fill two reduced volume cuvettes with 1 ml solutions containing either a fluorophore or a fluorescent nanoparticle of interest in one, and PS or PS with non-fluorescent nanoparticles as the negative control in the other. In the work demonstrated here, a variety of potential commercially available FUEL fluorophores and fluorescent nanoparticles were investigated as outlined in **Table 1**.
2. Add a 1 ml aliquot of fresh *V. fischeri* or other luminescent bacteria to a blacked-out third cuvette containing two physically equal optical windows situated on opposite sides. Cover the cuvette with an opaque substance such as a piece of black paper.
3. At a short but equal distance place the two reduced volume cuvettes on either side of the blacked-out cuvette and visualize under the appropriate filters. Here, a distance of 0.7 cm was used. Repeat with the other fluorophores.

5. Targeted FUEL

1. Prepare biotinylated antibodies specific to the luminescent bacteria. For example, here primary antibodies specific to *K. pneumoniae* (α -Kp) were biotinylated using a solution containing EZ-Link Sulfo-NHS-LC-Biotin.
2. Remove excess reagent from the antibodies using desalting columns under centrifugation (1,500 x g) for 2 min.
3. Use a standard Bradford assay to determine the protein concentration and determine the degree of antibody biotinylation by HABA assay.
4. Obtain the α -Kp-biot biotinylated antibody (Ab) batches by mixing 100 μ l (10 mM, dissolved in 1x PBS) with 40 μ l α -Kp antibody, resulting in a final degree of substitution of 16 biotin residues per Ab and a final protein concentration of 10 mg ml⁻¹.
5. Using multiple overnight replicate cultures, target the washed bacteria with the aforementioned antibodies following the appropriate protocol.
 1. Specifically, wash the *K. pneumoniae* in 1x PBS, and resuspend in 1x PBS to an OD of 4 (ca. 4 x 10⁸ CFU ml⁻¹ OD⁻¹).
 2. For each replicate, incubate 100 μ l PBS, α -Kp antibodies (10 μ l α -Kp-biotB or 10 μ l PBS for the control) and 100 μ l cells for 90 min at 30 °C. Wash the cells three times in 1x PBS, and resuspend in 196 μ l PBS.
6. Label the cells with the QD705 streptavidin conjugate and divide them into two equivalent volumes.
7. Wash one solution three times and then resuspend it into the appropriate volume.
8. Distribute 100 μ l of the washed and unwashed solutions into individual wells of a black 96-well plate. Measure the resulting luminescence under the Total Light, 490-510 nm, and 710-730 nm filters. Fluorophore concentration can be determined using 450-480 nm excitation and 710-730 nm emission.

Representative Results

Resonance Energy Transfer (RET) is a non-radiative interaction between a luminescent donor and a fluorescent acceptor, whereby the energy from the excited donor is capable of inducing a fluorescence response from the acceptor through a strong dipole-dipole interaction¹³. RET, which has been described using fluorescent²³, chemiluminescent²⁴, and bioluminescent¹³ donors, principally requires: 1. Strong spectral overlap between the donor emission and acceptor excitation spectra; 2. Appropriate rotational alignment between the two entities; and 3. A working distance no greater than 0.5- to 2-times the Förster radius, R_0 , between the donor and acceptor¹⁷. Contrasting with RET, FUEL occurs when a luminescent source, such as a bioluminescent bacteria, emits a photon that is absorbed and re-emitted by a second entity, such as a fluorophore or a fluorescent nanoparticle, red-shifting the emission spectrum of the original luminescent source. Thus, FUEL follows a standard excitation-emission process akin to standard epifluorescent conditions, yet without the use of a focused excitation. Evidence of this can be readily observed by the simple mixing of luminescent bacteria and highly-fluorescent quantum dots. In the presence of the *Vibrio* sp, a significant increase in red signal is observed when QD705 were also in solution compared to a non-fluorescent polystyrene microsphere control (**Figure 1A**). The components necessary to create bioluminescence, essentially the aldehyde substrate, luciferase, and ATP, are all produced and contained within the cytoplasm. Furthermore, the enzymatic luminophore production occurs within the bacterial cytoplasm (**Figure 1B**). As has been reported elsewhere, the distance between the inner and outer membrane of bacteria is typically greater than 30 nm²⁵⁻²⁷, a distance that does not allow for significant RET to occur. As well, the fluorescent nanoparticles do not cross into the bacterial cytoplasm since there is no endocytosis or other means of uptake. Together, these constraints indicate that FUEL is the dominant excitation-emission phenomenon that occurs when luminescent bacteria are mixed with fluorescent nanoparticles.

As was previously shown, FUEL exists beyond the range of RET. To investigate the FUEL dependence on distance, standard reduced volume spectrophotometric cuvettes can be used, with one containing a fluorophore solution, a second containing an appropriate control that can control for scatter, and the third containing an aliquot of fresh luminescent solution. It is essential that the central cuvette be enveloped using black tape or another opaque material, save for at least two identical optical windows located on opposite faces, to reduce any potential light contamination from the luminescent source. Further, the two remaining cuvettes need to be placed equidistantly onto either side of the central cuvette. Finally, a fresh luminescent solution, using bacteria or chemiluminescence, should be used with each experiment to ensure maximum light production. Upon appropriate placement, acquire the luminescence signal under the desired filters. The Total Light and 710-730 nm filters were used in this case, though only the data from the latter is shown. After each acquisition, increase incrementally the face-to-face distance from 1.0 cm to 3 cm (**Figure 2**). Finally, normalize all the data to the brightest point. In the examples used here, this occurred at the shortest distance ($D = 1$ cm) between the luminescent source and the cuvette containing the QD705. Using this approach, viable FUEL signal can be observed up to the final acquisition suggesting that, in the absence of any optical absorber, FUEL can occur at distances beyond any possible resonance energy transfer and can only be explained to be a purely radiative effect. Fitting the data reveals a decrease in signal as a function of distance D following a D^{-1} to D^{-2} dependence. Depending upon the geometric configuration, the former corresponds to the capacitor distance dependence and the latter to the inverse square law for point sources. This result is therefore consistent with our overall acquisition setup, given the size of the cuvette aperture and the distance between the luminescent source and the fluorophore.

FUEL not only applies to quantum dots, but also can be observed using a wide range of fluorophores ranging from the Alexa series to fluorescent microspheres (**Table 2**). In order to be comparable to the control, equal concentrations of the various fluorophores should be used and the total surface area of the nanoparticles held constant (**Table 1**). By comparing the fluorophores and nanoparticles to their appropriate controls (PS or PS with non-fluorescent polystyrene microspheres), a significant increase in signal is observed at the emission maximum of

each fluorescent entity. The largest relative increase in signal was found to occur where the fluorescent emission maximum was farthest from the luminescent emission maximum. This is most likely due to the increased specificity of the fluorescent signal compared to the broad emission spectrum of the luminescent source. On the contrary, the least reliable FUEL signal was found when the two emission maxima were not well separated. Interestingly, a discernable FUEL signal was found with the Yellow microspheres even though the spectral difference is minimal, due most likely to the substantial quantity of the fluorophore present per bead (350 fluorescein equivalents). The results shown here indicate that under appropriate conditions FUEL can be achieved with a variety of fluorophores, which enables the tailoring of probes chosen for more relevant applications under both *in vitro* and *in vivo* conditions. The significance of the observed FUEL signal was determined using a standard two-tailed Student's t-test. As such, only the Alexa555 was found to be incompatible with the luminescent source used.

In order to explore the effects of targeting on FUEL, biotinylated antibodies were used to target luminescent bacteria and streptavidin-linked QDs. It is important that the bacteria or luminescence source provide a layer thick enough to minimize the possibility of RET between the luminophore and the corresponding fluorophore. After incubation and washing to remove unbound antibodies, the biotin-labeled bacteria are then exposed to either streptavidin-conjugated QD705 or non-functionalized QD705 as the control, the solutions divided and one set exposed to further washes in order to remove any non-adhered QD705. Here, for all four conditions, the resulting luminescence was observed under the Total Light, 490-510 nm, and 710-730 nm filters, though only the latter two filters are used for the data analysis. The presence of the QD705 was compared using 450-480 nm excitation and 710-730 nm emission filters. Upon investigation we have found little to no difference in red-shifted signal when the solutions were left in an unwashed state (Figure 3). The resulting fluorescence signal under this condition is also found to be quite similar, suggesting that an equal number of QD705 was present under both the targeted and non-targeted state. However, washing the samples to remove any unbound QD705 provides a nearly two-fold increase in relative red signal for the targeted bacteria compared to their non-targeted control. Investigation by fluorescence can be used to verify the presence or absence of the QD705. Comparatively, the targeted washed condition resulted in a fluorescence intensity that was almost three times less than the unwashed condition, yet the resulting red-shift decreased by only 30%, suggesting that under purely bound conditions the targeting of the bacteria will result in an increase in red-shifted emission. This strongly implicates the utility of targeted FUEL for future applications.

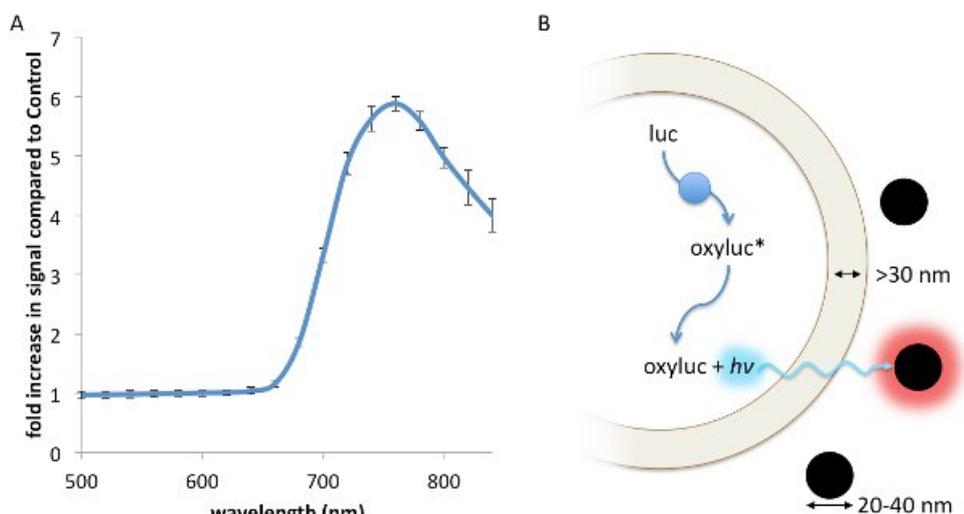


Figure 1. A FUEL interaction between luminescent bacteria and commercially available quantum dots leads to an increase in red photon production. Two spectrophotometric cuvettes were filled with solutions containing 100 μ l aliquots of fresh *V. fischeri* culture with an OD_{600} of 1-1.5, 895 μ l of PS, and either 5 μ l of QD705 or physiological saline (PS), before being placed into an IVIS Spectrum and the emission spectrum acquired. A significant increase in red signal is achieved due to the presence of the QD705, as can be noted by the increase in detected photons \cdot sec $^{-1}\cdot$ cm $^{-2}$ (p \cdot sec $^{-1}\cdot$ cm $^{-2}$) compared to the control under the 710-730 nm emission filter (A). Here, the dual membrane of the bacteria excludes the interaction of the luminescent moieties (blue circles) and the QD705 (black circles). The former are found only in the bacterial cytoplasm while the latter are freely distributed in the bulk solution (B). N=3 for each bacterial solution. [Please click here to view a larger version of this figure.](#)

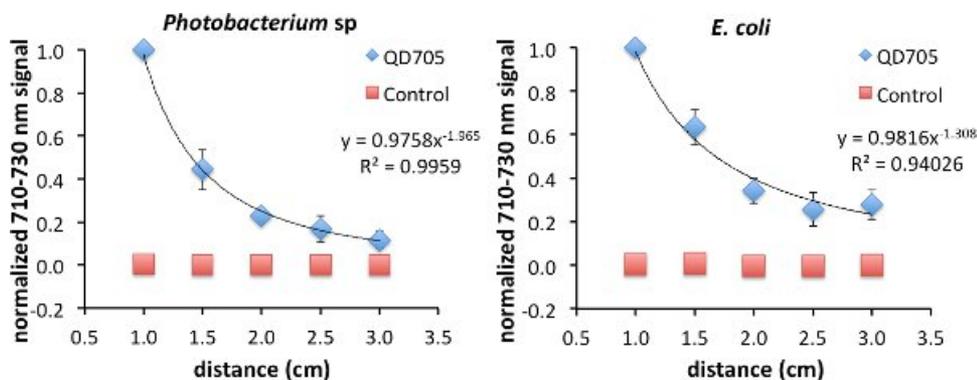


Figure 2. Evidence of FUEL occurring over distances completely excluding resonance energy transfer. Spectrophotometric cuvettes were filled with solutions containing either 50 μ l of QD705 and 950 μ l of physiological saline (PS) or 97.2 μ l of non-fluorescent 48 nm polystyrene microspheres and 902.8 μ l of PS, and placed equidistantly on opposite sides of a central black cuvette containing 1 ml of fresh luminescent *Photobacterium* sp at an OD₆₀₀ of 1-1.5. The central cuvette had two identical optical windows allowing the emitted photons to freely disperse. Acquiring images at distances (D) ranging from 1.0 cm to 3 cm, the observed production of red light decreased as a function of distance displaying evidence that FUEL can occur at distances not achievable by resonance energy transfer. The intensity appeared to have D⁻² dependence, similar to the inverse square law (left). The same protocol was followed for *E. coli* (right). The resulting trend lines hold form to the concept that the bacteria are acting as point sources of light. A total of three independent distance measurements were acquired with each using a unique subculture. Error bars are present at each point. In some cases the error bars were smaller than the symbols used that indicate the normalized intensity. [Please click here to view a larger version of this figure.](#)

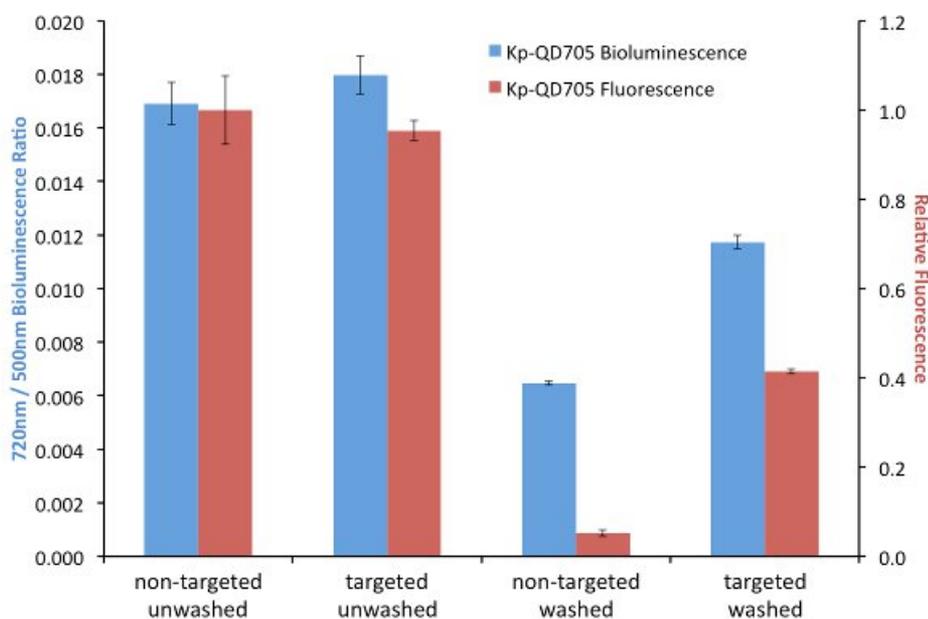


Figure 3. A comparison between targeted (specific) and non-targeted (bulk solution) FUEL. *K. pneumoniae* were functionalized with biotinylated antibodies and then exposed to either streptavidin-labeled (Targeted) or non-labeled (Non-Targeted) QD705. The resulting solutions were equally divided and one set washed three times with PBS before being resuspended into its initial volume in PBS. The solutions were then dispensed into individual wells of a black 96-well plate and the bioluminescence observed under the 490-510 nm (500 nm) and 710-730 nm (720 nm) emission filters (indicated as blue bars). The $\text{p}\cdot\text{sec}^{-1}\cdot\text{cm}^{-2}$ found from the 720 nm filter for each well was normalized by the respective $\text{p}\cdot\text{sec}^{-1}\cdot\text{cm}^{-2}$ bioluminescence intensity from the 500 nm of the same well. The relative fluorescence intensity resulting from an epifluorescent excitation was determined using 450-480 nm excitation and 710-730 nm emission filters (indicated as red bars). Little to no difference in bioluminescent or fluorescent signal was observed under the unwashed conditions (Non-Targeted Unwashed and Targeted Unwashed). This suggests that the bound and free-floating QD705 were equally excited by the bioluminescent photons. Upon washing, nearly a two-fold difference in relative red signal was observed for the QD705-labeled bacteria (Targeted Washed) compared to the control (Non-Targeted Washed). The absence of QD705 in the Non-Targeted Washed was confirmed by the lack of fluorescent signal and verified the QD705 labeling in the Targeted Washed state. The data is from four independent cultures of *K. pneumoniae*. For clarity, the legend indicates the source of the QD705 excitation. [Please click here to view a larger version of this figure.](#)

Fluorophore	Concentration	μ l	PS (μ l)	λ_{max} ex/em (nm)	Emission Filter (nm)
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Alexa 555	37.2 μ M	4.77	995.23	555/565	570 – 590
Alexa 568	37.2 μ M	4.77	995.23	578/603	590 – 610
Alexa 633	37.2 μ M	4.77	995.23	632/637	650 – 670
Alexa 700	37.2 μ M	4.77	995.23	702/723	710 – 730
Nonfluorescent	2.62% solids	9.72	990.28		
μ Sph Pink	5% solids	4.77	995.23	580/605	610 – 630
μ Sph Yellow	5% solids	4.77	995.23	505/515	510 – 530
QD705	2 μ M	5	995	465/705	710 – 730
QD800	2 μ M	5	995	465/705	790 – 810

Table 1. Properties of fluorophores used throughout the FUEL demonstrations.

	Control	filter	fluorophore		Control		p value
			$\text{p}\cdot\text{sec}^{-1}\cdot\text{cm}^{-2}$	SD	$\text{p}\cdot\text{sec}^{-1}\cdot\text{cm}^{-2}$	SD	
A555	PS	580	1.08×10^7	3.31×10^6	9.09×10^6	1.75×10^6	0.233
A568	PS	600	8.47×10^6	4.23×10^6	4.49×10^6	9.71×10^5	0.094
A633	PS	660	2.40×10^6	1.25×10^6	7.85×10^5	2.39×10^5	0.046
A700	PS	720	5.53×10^5	2.46×10^5	1.54×10^5	6.05×10^4	0.026
MSph Yellow	non-fluorescent μ spheres	520	1.19×10^8	4.85×10^7	5.79×10^7	1.99×10^7	0.057
MSph Pink	non-fluorescent μ spheres	620	2.37×10^7	1.36×10^7	2.16×10^6	8.00×10^5	0.026
QD705	non-fluorescent μ spheres	720	1.76×10^7	7.33×10^6	2.08×10^5	7.16×10^4	0.007
QD800	non-fluorescent μ spheres	800	7.79×10^6	4.72×10^6	3.60×10^4	1.52×10^4	0.023

Table 2. Identification of fluorophores and fluorescent nanoparticles compatible with FUEL.

Discussion

The fundamental demonstration of FUEL can be achieved simply by mixing luminescent bacteria with fluorescent nanoparticles or QDs. The two entities will be physically separated and remain beyond any efficient RET distance. More difficult is the FUEL signal optimization both *in vitro* and *in vivo*. Under *in vitro* conditions, both with and without an optical absorber present, usually the addition of excess fluorophore will be sufficient to maximize the FUEL response. However, at high concentrations phenomena such as static or collisional quenching can lead to a loss of fluorescent signal. Performing a dilution series by independently varying the concentration of the luminescent source and the fluorophore will help to optimize the desired concentrations. The establishment and optimization of FUEL under *in vivo* concentrations is much more difficult and needs to be addressed on a case-by-case basis. It can be difficult to create a condition where the fluorescent entity can be accessed optically by the luminescent source. As such, beginning with direct co-injections of the two moieties can provide information regarding the success of FUEL under optimal conditions.

Standard protocols exist for labeling bacteria and eukaryotic cells with fluorescent entities such as the Alexa series and QDs. Often this requires surface functionalization or activation with antibodies, which can lead to unwanted effects like reduced cell viability or altered metabolic activity. To overcome this, it is important to determine the optimal amount of antibody or activation agent needed that minimizes cellular perturbations while maximizing the fluorescent labeling. The use of QDs is advantageous because of their characteristically broad excitation spectra, narrow and tunable emission spectra, and the possibility of a large Stokes shift. However, QDs can be cytotoxic and may not be desirable in some cases.

FUEL is a phenomenon that is present in many BRET experiments¹³ and is applicable to a variety of luminescent and fluorescent sources. Until now, the photons resulting from FUEL were considered the product of non-specific interactions or an unfortunate background signal resulting from poorly designed BRET experiments. It is only with the type of experiments demonstrated here that we were able to identify the utility of this unwanted signal. In the shown examples, the luminescent bacteria act as a diffuse excitation source capable of eliciting a standard fluorescent response from a wide variety of fluorescent entities. Furthermore, due to the substantial working distance, it is safe to conclude that while FUEL can be constructed without the occurrence of BRET, in general BRET cannot be observed without a contribution from FUEL. Importantly, due to the lack of a targeting requirement, FUEL can be used to cover the spatial gap that exists between BRET and conventional whole-animal imaging techniques.

Disclosures

The authors declare competing financial interests. J.D., S.B., K.L.R. and S.L.S. contributed to European Patent EP10290158.4 and World Intellectual Property Organization Patent Application WO/2011/117847.

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