

1 **Evaporation reduction and salinity control in microalgae production ponds**  
2 **using chemical monolayers**

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14 **Abstract**

15 Algae have great potential as a hyper-productive crop to produce food, fuels, and chemicals.  
16 However, freshwater availability limits their widespread application. Here we investigate  
17 whether chemical monolayers can reduce evaporation in microalgae cultures, and whether algal  
18 growth is affected. Thin-film monolayers were formed on the surface of freshwater (*Chlorella*  
19 *vulgaris*) and marine (*Nannochloropsis salina*) algae cultures using ethylene glycol  
20 monooctadecyl ether. Monolayers applied daily reduced evaporation in both cultures by 70%  
21 on the first day, and ~50% by day 3. The cause of the reduced performance was investigated  
22 but could not be directly attributed to any particular cellular activity or chemical change.  
23 *Nannochloropsis* was uninhibited by the monolayer, while the growth of *Chlorella* decreased  
24 by 38% over 3 days. There was no evidence that the monolayer reduced gas exchange  
25 (CO<sub>2</sub>/O<sub>2</sub>), but the reduced growth of *Chlorella* could have been caused by direct chemical  
26 inhibition by the monolayer or the slightly elevated temperature (1–2 °C) resulting from the  
27 reduction in evaporative cooling. A techno-economic analysis indicated that water savings  
28 could make monolayers economically beneficial, especially in arid climates suited to algae  
29 production. In addition, monolayers enable control of salinity in marine production systems.  
30 Overall, the application of monolayers to reduce evaporation from outdoor algae cultures has  
31 great promise, with testing in outdoor trials an obvious next step.

32 **Keywords:** Microalgae production; monolayer film; evaporation reduction; *Chlorella*  
33 *vulgaris*; *Nannochloropsis*; biofuel.

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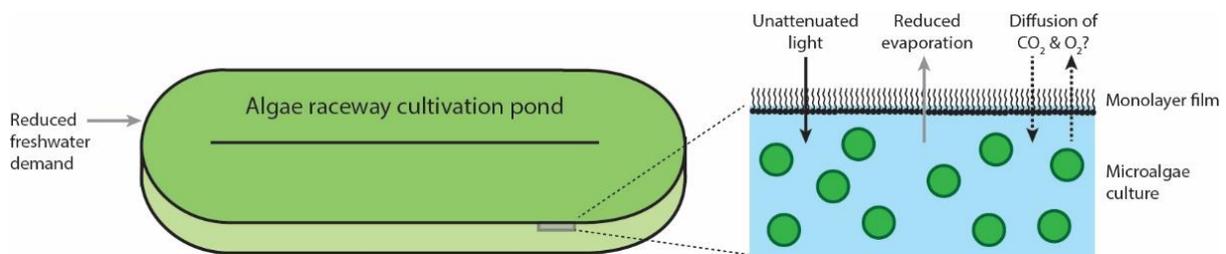
## 35 1. INTRODUCTION

36 Microalgae are currently grown commercially at a limited scale to produce high-value  
37 antioxidant pigments and health food supplements. In the future, microalgae have great  
38 promise for mass production of bulk commodity products such as animal and aquaculture feed,  
39 biofuel, and chemical feedstocks owing to their ability to be cultivated in low-cost open ponds  
40 on non-arable land at much greater productivities than terrestrial plants. Cultivation of algae at  
41 such a large scale could provide major environmental benefits in terms of land and nutrient  
42 utilisation. However, economically viable production of low-value commodity products from  
43 microalgae is a major challenge that requires further research efforts to reduce the cost of  
44 cultivation and biomass processing [1]. One of the major factors currently limiting mass  
45 production of microalgae is their requirement for water [2]. As microalgae are cultivated as  
46 highly dilute suspensions (0.5 – 1 g/L), very large amounts of water are required relative to the  
47 amount of biomass that is produced. While the overall demand for water can be reduced  
48 considerably by recycling the growth media following harvest of the biomass, evaporation from  
49 open pond systems represents a major and unavoidable loss of water. For example, it has been  
50 estimated that approximately 530 L of water is lost by evaporation per litre of microalgal lipid  
51 produced in a large-scale open pond system producing  $4 \times 10^6$  L/year of algal oil [1]. Although  
52 different species of microalgae can grow in fresh water, seawater or brackish water, the  
53 addition of fresh water is always required to compensate for the water lost from evaporation to  
54 maintain salinity levels for optimum algal growth. This requirement for fresh water to  
55 compensate for evaporation effectively nullifies one apparent advantage of cultivating marine  
56 microalgae. In addition to the cost of the fresh water itself, there is a cost associated with  
57 pumping this make-up water into algal ponds [3], and therefore a need for mass cultivation  
58 sites to be situated near an abundant source of fresh water, even if a marine alga is grown near  
59 the coast using seawater. Unfortunately, many of the locations otherwise most suited for mass

60 cultivation of algae (i.e., regions with an abundance of cheap, flat land with high solar radiation  
61 per unit area), often have limited supplies of fresh water [4]. Owing to these challenges, it is  
62 recognised that water consumption is a major bottleneck to large-scale algae production [2].  
63 While much research has been devoted to reducing the water requirements for algal cultivation  
64 by water recycling [5, 6] and high-density cultivation [7], it does not appear that anyone has  
65 yet tried to mitigate evaporation, which is seen as an inevitable consequence of cultivation  
66 using open systems [2].

67 The aim of this study was to investigate for the first time, the use of chemical monolayers to  
68 reduce evaporation rates in open microalgae production systems. Beyond microalgae  
69 cultivation, evaporation is a practical issue in the context of water storage in reservoirs and  
70 dams [8]. As such, a number of strategies have been developed to reduce evaporation from  
71 freshwater storage bodies, including mechanical methods such as floating spheres and  
72 suspended covers [9]. While these systems can be quite effective at reducing evaporation, the  
73 capital cost can be high, particularly when applied over a large surface area. Even more  
74 importantly in the context of algae cultivation, these devices block solar radiation into the water  
75 body, which would thereby reduce photosynthetic algal growth. Therefore, to be applied in  
76 algal cultivation, any water evaporation technology must not reduce the light available to the  
77 algae. Based on this requirement, it is proposed here that optically transparent chemical  
78 monolayer films could be a promising method to reduce water evaporation for large-scale algae  
79 cultivation (Fig. 1). In the context of evaporation reduction, a 'monolayer' is a closely-packed,  
80 single-molecule thin film of amphiphilic compounds, which spreads spontaneously to cover a  
81 water surface [10]. Such monolayers have been widely investigated for freshwater storage  
82 applications, including in numerous field trials that demonstrate the potential to reduce water  
83 evaporation by as much as 40% [8]. In addition to their effectiveness and light transparency,  
84 advantageous features of monolayers for algae applications are the low capital costs and their

85 ability to be applied only when required, specifically in times of high evaporation. Various  
86 amphiphilic chemicals have been used in monolayer applications, including hexadecanol,  
87 octadecanol and ethylene glycol monooleyl ether (referred to as C18E1). As C18E1 has  
88 been shown to be particularly effective for evaporation reduction [11], it was chosen for this  
89 study.



91 **Fig. 1** Illustration of the proposed approach of applying a monolayer film to reduce evaporation  
92 from large-scale outdoor algae cultivation ponds.

93 The novel application of chemical monolayers to algae cultures presents new research  
94 questions that have not been addressed in previous studies for pure water applications such as  
95 reservoir preservation [12]. In particular, the aim of the current study was to investigate  
96 whether monolayers could successfully reduce evaporation in the presence of concentrated  
97 algae cultures, and conversely, whether the presence of a monolayer adversely affects the  
98 growth of algae. It is possible that the algae and associated bacteria could degrade the  
99 monolayer chemicals or interfere with the intermolecular packing to reduce performance. On  
100 the other hand, the monolayer could directly or indirectly inhibit algal growth. For instance, as  
101 photosynthetic organisms, microalgae assimilate atmospheric CO<sub>2</sub> and release oxygen during  
102 photosynthesis. The exchange of gaseous CO<sub>2</sub> and oxygen between the atmosphere and culture  
103 is important, in particular to facilitate photosynthesis and to avoid the build-up of dissolved  
104 oxygen which can be toxic to algae [13, 14]. In this respect, one important physical effect of  
105 monolayers is an increase in the boundary layer thickness at the air/water interface, which could  
106 decrease the rate of gas diffusion [8].

107 The present study investigates the evaporation reduction performance and potential impacts of  
108 C18E1 monolayers during the growth of both a marine and freshwater algal species  
109 (*Nannochloropsis salina* and *Chlorella vulgaris*, respectively) of industrial relevance. The  
110 economic feasibility of using monolayers to control evaporation in algal ponds is also  
111 investigated and a sensitivity analysis is applied to explore the impact of variations in water  
112 price and climate on the cost savings. The results from the present study provide insights into  
113 a novel application of monolayer films as an innovative means for reducing water requirements  
114 for commercial algal cultivation.

## 115 **2. MATERIALS AND METHODS**

### 116 **2.1 Preparation of algae seed cultures and chemical monolayer**

117 A seed culture of *Nannochloropsis salina* (CCMP 1776, obtained from the University of  
118 Melbourne Culture Collection) was grown in an aerated 2 L Schott bottle containing 1500 mL  
119 of f/2 medium [15] at  $21 \pm 2$  °C under a light:dark cycle of 14:10 h with a light intensity of 60  
120 – 70  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$ . Similarly, a seed culture of a freshwater species, *Chlorella vulgaris*  
121 (obtained from CSIRO, Australia), was grown in 1500 mL of MLA medium[16]. The nitrogen  
122 source was sodium nitrate for both f/2 and MLA media, provided at a concentration of 0.1 g/L  
123 or 0.17 g/L, respectively. The molar concentrations of the individual nutrient stock solutions  
124 of f/2 and MLA media are provided in Supplementary File S.1-a.

125 Monolayer solutions were prepared by dissolving ethylene glycol monoctadecyl ether  
126 (C18E1) in chloroform (RCI Labscan Ltd) at a final concentration of 10 mg/mL, which was  
127 applied to the surface daily unless specified to form a film.

### 128 **2.2 Experimental set-up**

129 Evaporation experiments (with or without monolayer) were conducted in duplicate algae  
130 cultures grown in open round glass dishes (diameter = 11.5 cm, height = 6.5 cm). For this, the

131 algae seed cultures (Section 2.1) of *N. salina* and *C. vulgaris* were centrifuged at 2000 g for 5  
132 min and the resulting pellets were resuspended into 100 mL of fresh f/2 or MLA media,  
133 respectively to obtain an initial biomass concentration of 0.16 g/L or 0.13 g/L, respectively.  
134 These 100 mL cultures were then added to the open round glass dishes, which were placed on  
135 individual digital balances (Mettler-Toledo Limited). These algae cultures were then grown for  
136 3 days under constant light (24:0 h) with an intensity of approximately 70  $\mu\text{mol photons m}^{-2}$   
137  $\text{s}^{-1}$ . At the start of each experiment, 36.4  $\mu\text{L}$  of the monolayer solution was added to the test  
138 cultures, and this addition was repeated daily over the course of the experiment. Parallel  
139 duplicate cultures without monolayer films were included as controls. The experimental design  
140 is depicted in Supplementary File S.1-b.

### 141 **2.3 Evaporation analysis**

142 Evaporation rates were measured gravimetrically by recording the mass loss from the cultures  
143 over time (the cultures remained in place on individual digital balances during the 3-day  
144 cultivation period). In the cultures with monolayer film, the mass was recorded automatically  
145 every minute using a weight reading program (BalanceLink®). Due to a limited availability of  
146 automatic recording, the mass loss from cultures without monolayer (i.e., the control cultures)  
147 was manually recorded intermittently during the cultivation period. To reduce the cumulative  
148 concentrating effect of evaporation, deionised water was added daily to replenish the water lost  
149 due to evaporation. Immediately following the daily water replenishment, 36.4  $\mu\text{L}$  of the  
150 monolayer solution was reapplied. Subsequently, the average daily evaporation rates in the  
151 control and monolayer cultures were calculated.

152 A separate evaporation analysis (with or without monolayer) was conducted using algal culture  
153 supernatants. Briefly, 400 mL of *N. salina* (in f/2 media) and *C. vulgaris* (in MLA media)  
154 cultures were grown for 3 days in 225  $\text{cm}^2$  Corning flasks having 0.2  $\mu\text{m}$  vented caps, under

155 the same growth conditions as those described above and shaken at 100 rpm using an orbital  
156 shaker. On day 3, *N. salina* and *C. vulgaris* cultures were centrifuged at 2000 g for 5 min and  
157 each of the obtained supernatants (i.e. used f/2 or MLA media) was divided into four and placed  
158 in open round glass dishes (duplicate controls and monolayer dishes), which were placed on  
159 individual digital balances for determination of the evaporation rate. As with the algae culture  
160 evaporation experiments, monolayer was reapplied daily over the 3 day experiment.

## 161 **2.4 Measurement of algal growth and culture pH, temperature, and dissolved oxygen**

162 To monitor algal growth, culture samples (3 mL) were collected daily and the absorbance at  
163 750 nm measured using a Cary 3E UV-Vis spectrophotometer (Agilent Technologies,  
164 Australia). The absorbance was converted into dry weight biomass concentrations (g/L) using  
165 a standard curve prepared separately for each species. As the absorbance was measured prior  
166 to replenishing the evaporated water, the biomass concentrations were adjusted based on the  
167 amounts of water evaporated in the monolayer and control dishes. Also prior to replacement of  
168 evaporated water, the culture pH (Metler-Toledo InLab® Science), dissolved oxygen  
169 concentration (LDO™ probe, HQ40d, Hach) and temperature (infrared thermometer) were  
170 measured periodically. Results are presented as the average of biological duplicate culture  
171 dishes.

## 172 **2.5 Techno-economic analysis**

### 173 ***2.5.1 Net cost savings calculations and assumptions***

174 Calculations of cost savings resulting from the use of monolayer films were based on the cost  
175 of purchasing replacement water and associated pumping costs. The cost associated with  
176 pumping the replacement water was based on the economic evaluation by Rogers et al. [3]  
177 (\$2.07 million/year, for 5 mm evaporation/day over an area of 4875 ha, which works out to be  
178 \$0.23/(year.ha.mm)). Water demand was based on the amount of evaporation (mm/year) with

179 and without the film, as determined experimentally in this study. Estimates of the evaporation  
180 rate were obtained for different climate regions from Guieysse et al. [17]. The different regions  
181 used in this study were ‘Tropical’, ‘Sub-tropical’, ‘Temperate’, ‘Mediterranean’, and ‘Arid’  
182 climates. The allocated rate of process water disposal (3230 mm/year) and the leak loss (860  
183 mm/year) were also obtained from Guieysse et al. [17]. For the cost of the monolayer, a price  
184 of \$10 per kg was used in the calculations at the same loading rate as the lab  
185 trials (388 g/(ha.day)) based on the total surface area of the water. Variation in water depth will  
186 not impact the amount required, as the monolayer forms on the water surface. Chloroform was  
187 used in these small-scale experiments to improve the spreading of monolayer film and to  
188 facilitate accurate application of small amounts. Chloroform would not be used in field trials  
189 or large-scale applications, with the monolayer added directly to the surface in these cases.  
190 Therefore, chloroform is not considered in the techno-economic analysis. For economic  
191 calculations, an inflation rate of 1.5% was assumed.

### 192 **2.5.2 Salt concentration analysis and assumptions**

193 An analysis of the effect of monolayer evaporation reduction on the salinity of marine algae  
194 growth ponds was performed using *Nannochloropsis* as a reference marine alga, where 35 g/L  
195 is the optimum salinity for growth and 45 g/L was used as the upper range for growth [18,  
196 19]. The algae media solution was assumed to be withdrawn at a rate of 10% of the pond  
197 volume per day to balance new algae growth with algae removed to maintain the concentration  
198 of algae at 0.5 g/L [3]. The algae stream was then concentrated via a two-stage process going  
199 through a clarifier where it was concentrated to 30 g/L [3, 18]. This stream was then  
200 concentrated to 200 g/L via a centrifuge. The water removed was then collected and sent back  
201 to the pond with a fraction being purged at varying rates. Salt water was used as the inlet water  
202 supply to make up for water lost to evaporation and removed with the algae. This was set at 35  
203 g/L in the simulation unless otherwise stated.

### 204 ***2.5.3 Impact of evaporation suppression on pond temperature***

205 Calculations of the increase in pond temperature due to the reduced evaporation by the  
206 monolayer were performed on the basis of a pond with a depth of 30 cm. Energy lost from the  
207 system through evaporation was calculated based on the latent heat capacity of water at 25 °C  
208 (2442 kJ/kg) and the difference in evaporation through use of the monolayer. The  
209 corresponding temperature change was calculated based on the equivalent energy embedded in  
210 water at 25 °C with a specific heat capacity of 4.19 kJ/(kg.K).

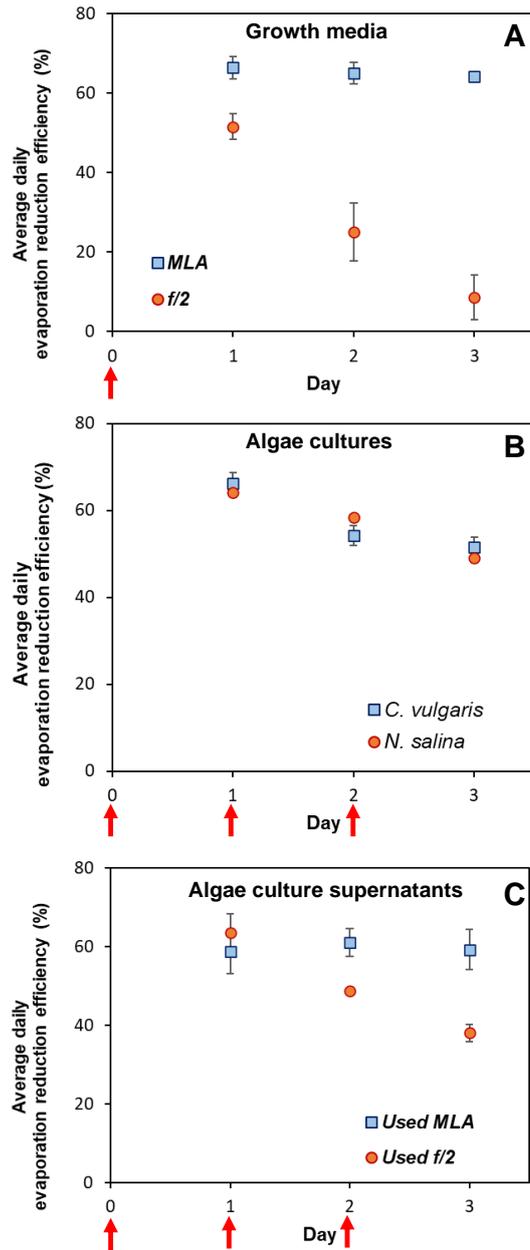
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## 212 **3. RESULTS AND DISCUSSION**

### 213 **3.1 Evaporation reduction in fresh water and marine algae cultures**

214 While chemical monolayers are usually applied to fresh water, monolayer dissociation can be  
215 affected by environmental conditions such as pH, temperature, salt type and concentration [20].  
216 As such, the evaporation reduction performance of a single dose of C18E1 monolayer was first  
217 tested on freshwater and marine algae growth media in the absence of algae cells to determine  
218 the influence of salt and media components (Fig. 2A). In the freshwater medium (MLA),  
219 evaporation was reduced by approximately 66% without decline over the three days of testing  
220 (Fig. 2A), representing similar performance to that on deionised water (approximately 64%,  
221 data not shown). However, in the marine growth medium (f/2), evaporation performance  
222 progressively declined from >50% over the first day to <10% during the third day. The  
223 reduction in evaporation performance was not found to occur in the presence of either the sea  
224 salt (30 g/L) alone or with the other major medium components included individually  
225 (Supplementary File S.2), suggesting some complex interactions with the monolayer occurred.  
226 To address this issue in algae cultures, the monolayer was replenished each day, consistent  
227 with standard practice in large-scale water reservoir applications [21]. As discussed later, daily

228 reapplication of a monolayer is economically feasible. The reduction in water evaporation in  
229 freshwater *C. vulgaris* and marine *N. salina* cultures was tested, with the average daily  
230 evaporation rate measured over three days (Fig. 2B). The impact of the monolayer on algal  
231 growth was investigated in parallel (Fig. 3). Overall, the monolayer film was found to  
232 significantly reduce the evaporative water loss from both freshwater and marine algae culture  
233 (by 50 – 70%) (Fig. 2B), consistent with previously published results for pure water systems  
234 [22]. There was no apparent difference in evaporation performance between the two cultures,  
235 despite the different algae and the different longer-term performance of the freshwater and  
236 marine media (Fig. 2A).



237

238 **Fig. 2** Average daily evaporation reduction efficiency (percentage of evaporation prevented by  
 239 the presence of the monolayer relative to controls) in (A) freshwater (MLA) and marine (f/2)  
 240 growth media (blue square and orange circles, respectively), (B) *Nannochloropsis salina*  
 241 (orange circles) and freshwater *Chlorella vulgaris* (blue squares) cultures, and (C) cell-free,  
 242 used media (supernatants) from *Nannochloropsis salina* (orange circles) and freshwater  
 243 *Chlorella vulgaris* (blue squares) cultures. As indicated by the red arrows, monolayer solution  
 244 (36.4  $\mu$ L; 10 mg/mL) was added at day 0, and for (B) and (C) reapplied on days 1 and 2. Results  
 245 are presented as the average and standard error of biological duplicate cultures.

246 While evaporation reduction remained high over three days for both cultures, there was an  
247 observable decrease in the reduction efficiency from 70% to 50% during this period. The  
248 decline in performance (Fig. 2B) despite daily replenishment of the monolayer on the surface  
249 of algal cultures could be due to various reasons. An increase in the concentration of algal cells  
250 during batch cultivation would result in more frequent collisions of the cells with the water  
251 surface, which could temporarily create localised interruptions in the film. However, a separate  
252 experiment using *N. salina* cultures at a range of initial biomass concentrations (0.12 – 0.57  
253 g/L) showed a negligible effect of cell concentration on monolayer performance (data not  
254 shown). During batch cultivation, extracellular products released by algal cells could also  
255 accumulate, increasing the surface pressure [23] and reducing the packing density and  
256 performance of the monolayer film [22]. To investigate this, the performance of monolayer  
257 was tested in cell-free f/2 and MLA media (i.e., supernatants taken from *N. salina* and *C.*  
258 *vulgaris* cultures, respectively; without monolayer) (Fig. 2C). Interestingly, the *N. salina*  
259 supernatant resulted in a progressive decline in the evaporation reduction efficiency of the  
260 monolayer film, despite daily replenishment, while the impact of the *C. vulgaris* supernatant  
261 was negligible. These trends (Fig. 2C) are consistent with the differential impact of the fresh  
262 and marine growth media (Fig. 2A) rather than the cultures (Fig. 2B), suggesting that  
263 extracellular algae products cannot explain the diminishing monolayer performance in both  
264 cultures.

265 The results from the experiments using culture supernatants would also have been influenced  
266 by other culture-induced changes to the growth medium such as pH shifts or nutrient uptake.  
267 As such, these factors do not appear sufficient to explain the differential decline in monolayer  
268 performance in *N. salina* and *C. vulgaris* supernatants. However, interestingly monolayer  
269 performance in the first day was worse in the marine growth media (Fig. 2A) than in the *N.*  
270 *salina* supernatant (Fig. 2C), suggesting that some growth media components impair

271 monolayer packing and that these media components are reduced by the algal growth. The  
272 better performance of monolayer film with *N. salina* cultures (Fig. 2B) than in the  
273 corresponding supernatant (Fig. 2C) could also indicate a potential protective behaviour of  
274 algal cells in shielding the monolayer from these interfering components.

275 In the current study, the pH increased from approximately 8.5 to 10 and 10.5 in *N. salina* and  
276 *C. vulgaris* cultures, respectively (Fig. 3). While previously published literature has shown that  
277 the structure, phase behaviour, and function of monolayers with acidic head groups can be  
278 negatively impacted by high pH conditions such as pH 10 [24-26], the monolayer used here  
279 (C18E1) does not contain an acidic head group and should not therefore be severely affected  
280 by pH. This was verified in a separate experiment in which a monolayer on f/2 media (free of  
281 algal cells) was adjusted to pH 8.5 and 10.5, with no apparent difference in the evaporation  
282 rates (data not shown). As discussed below, the presence of the monolayer slightly increased  
283 the temperature of the cultures (1–2 °C) due to suppression of evaporative cooling. However,  
284 this does not appear to be a factor in the gradual decline in performance, as the elevation in  
285 temperature remained quite constant throughout the experiments.

286 It has previously been suggested that some chemical monolayer compounds such as  
287 hexadecanol and octadecanol could be degraded by bacteria [10, 12], the rate of which could  
288 increase over time. However, microscopic analysis revealed no obvious increase in the  
289 abundance of bacterial cells in the cultures with monolayer film present (Supplementary File  
290 S.3), consistent with previously published field trials using C18E1 monolayer film [23].  
291 Monoctadecyl ether-based monolayers have been previously shown to be at risk of  
292 photodegradation [23], although only a very low light intensity (compared to sunlight) of 70  
293  $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$  was applied. In addition, the monolayer was replenished each day,  
294 meaning that the impact of bacterial or light degradation of monolayer films is not likely to

295 account for the decline in performance observed here. However, in environments with higher  
296 light intensities, the rate of photodegradation of the monolayer will likely be higher. Pittaway  
297 et al., demonstrated that photodegradation was a function of cumulative solar radiation and the  
298 concentration of components in the media (e.g. iron and aromatic compounds) [12]. The rate  
299 of addition of monolayer may need to be adjusted for these environments.

300 In summary, the reason for the decline in monolayer performance in both cultures over 3 days  
301 cannot be attributed to any obvious singular cause. Although the performance declined, the  
302 evaporation reduction was still around 50% after 3 days of cultivation, which is consistent with  
303 outdoor trials on pure water and highly significant from a practical standpoint as discussed  
304 below. The rate of decline is sufficiently slow in relation to periodic harvesting of the cells,  
305 which would remove interfering cell materials and thus restore monolayer performance.  
306 Nonetheless, detailed research into the direct interactions between algal cell materials and  
307 monolayers is recommended to improve stability and performance. In addition, given the  
308 decline observed over 3 days, it would be worthwhile investigating monolayer performance  
309 over prolonged periods in future studies, which are of relevance to extended batch cultivations  
310 and implementation of nutrient deprivation strategies for lipid accumulation.

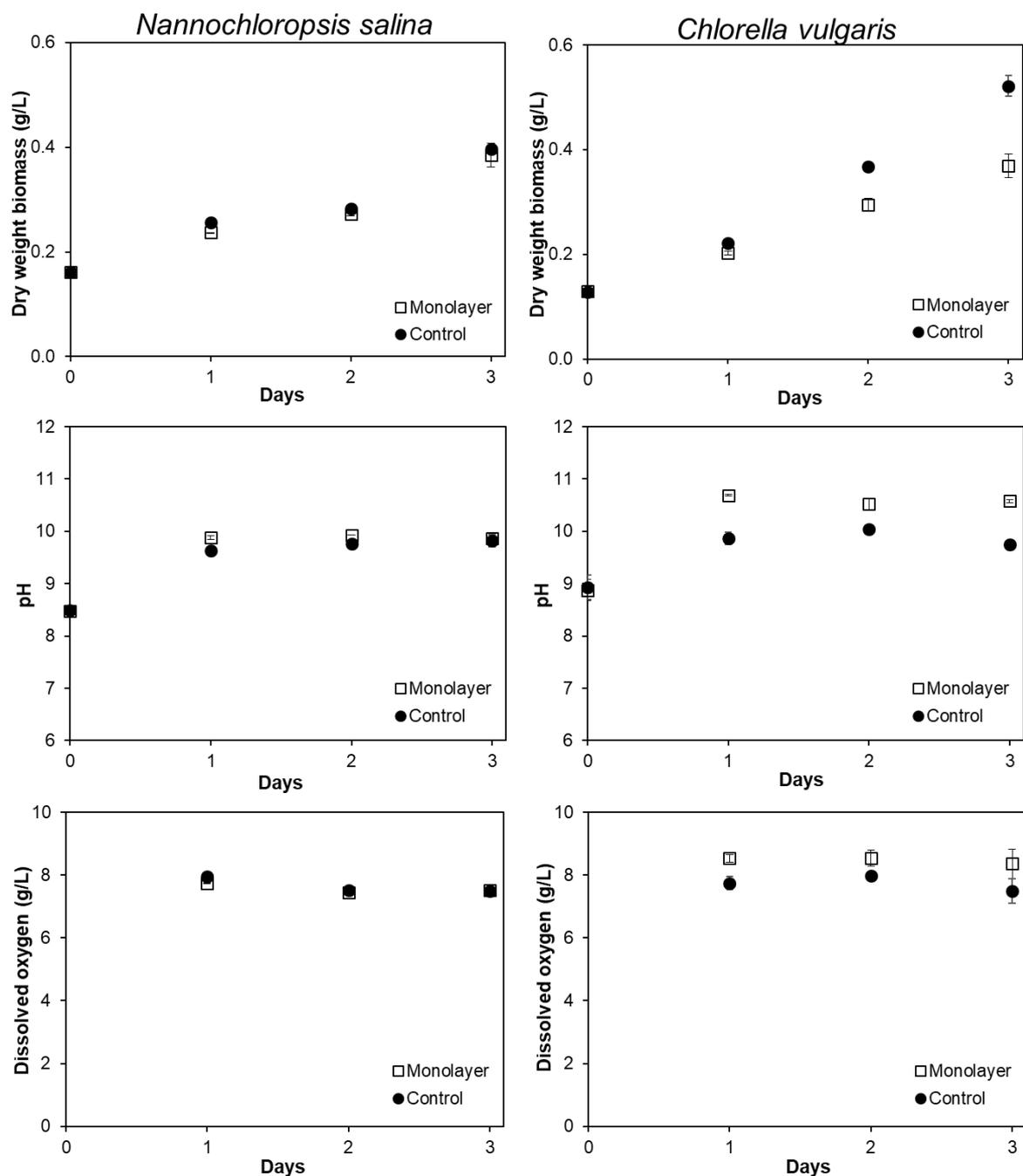
### 311 **3.2 Algae growth in the presence of monolayer**

312 Similar experiments were used to assess the potential impact of the monolayer on algal growth  
313 (Fig. 3). For *N. salina* cultures grown in marine f/2 media, the biomass productivities in both  
314 the monolayer and control cultures were not significantly different over the 3-d growth period  
315 ( $75 \pm 7$  and  $78.5 \pm 0.3$  mg/L/d, respectively). The pH increased from 8.5 to 9.8 in both the  
316 monolayer and control cultures (Fig. 3), due to the consumption of nitrate and CO<sub>2</sub> in the media.  
317 Monolayers have previously been reported to reduce oxygen diffusion (by 10 - 15%) [27].  
318 Here, the dissolved oxygen concentrations were similar in the control and monolayer cultures,

319 at around 7.5 - 8 mg/L, indicating that the monolayer did not prevent oxygen egress from these  
320 cultures. The lack of any appreciable effect of the monolayer on the pH and dissolved oxygen  
321 concentration are consistent with the unimpaired productivity in these cultures.

322 The reduction in evaporation by the monolayers also reduces evaporative cooling, which can  
323 increase the temperature of the water body [28]. The temperature in the monolayer cultures  
324 (21.2– 24.2 °C) of *N. salina* was on average  $1.1 \pm 0.5$  °C higher than the control cultures (21.2  
325 – 23 °C), the impact of which on algal growth is likely negligible (Fig. 3), consistent with the  
326 constant growth rate of the *Nannochloropsis* cultures at elevated temperatures [29, 30].

327



328  
 329  
 330 **Fig. 3** Dry weight biomass, pH, and dissolved oxygen concentration in *Nannochloropsis salina*  
 331 (left hand panels) and *Chlorella vulgaris* (right hand panels) cultures grown in open cultures  
 332 for 3 days in the presence (monolayer; empty square) or absence (control; filled circle) of a  
 333 monolayer film. All the parameters were measured before compensating for evaporated water.  
 334 Biomass is normalised to the proportional water evaporation in the monolayer and control

335 cultures. Results are presented as the average and standard error of biological duplicate  
336 cultures.

337 Continual growth was observed in both the monolayer and control cultures of freshwater *C.*  
338 *vulgaris*; however, the biomass productivity in the control cultures was higher than that of the  
339 cultures containing a monolayer ( $131 \pm 7$  and  $80 \pm 8$  mg/L/d, respectively). There are a number  
340 of possible reasons why *Chlorella* growth was negatively impacted by the presence of the  
341 monolayer while *Nannochloropsis* was not. Firstly, it is conceivable that *Chlorella* was directly  
342 inhibited by either the chloroform or the monolayer chemical, while *Nannochloropsis* was not.  
343 However, chloroform was only applied to the surface of the cultures in very small amounts  
344 ( $\sim 36.4$   $\mu$ L on 100 mL cultures). As a water-immiscible and volatile solvent it would not have  
345 mixed into the cultures but rather would have been rapidly evaporated from the surface,  
346 preventing any possible inhibition of the algae. On the other hand, the monolayer was present  
347 for the duration of the experiments, and although predominantly on the surface, a proportion  
348 of the molecules could partition into the culture. The differential effect on the two algae  
349 indicates that if chemical inhibition was a factor, it is species specific. Direct chemical  
350 inhibition of algae will also be dependent on the chemical monolayer used, meaning this is a  
351 complex and important topic that warrants future investigation.

352 Considering other factors, the temperature of *C. vulgaris* monolayer cultures ( $22.6 - 24$  °C)  
353 was on average  $1.4 \pm 0.5$  °C higher than that of the control cultures ( $20 - 23$  °C), similar to the  
354 *N. salina* cultures. Although the growth of *Nannochloropsis* was evidently not affected by the  
355 small increase in temperature, some algal species like *C. vulgaris* (Fig. 2) have a strict  
356 temperature range, outside of which growth efficiency can drop significantly [31]. It is  
357 therefore possible that the increase in temperature of just a few degrees Celsius could have  
358 negatively impacted the growth rate of *Chlorella*. The ability to start and stop the application

359 of the monolayer provides a means of controlling evaporative cooling, and thereby influencing  
360 the culture temperature. For example, in cooler periods the monolayer could increase the  
361 temperature to enhance growth, whereas it may be preferable to not apply monolayer on  
362 extremely hot days to avoid reducing evaporative cooling and to keep the temperature within  
363 the preferred limits of the desired algae strain.

364 In the current experiments, the cultures were not aerated and relied solely on CO<sub>2</sub> uptake from  
365 the atmosphere. The pH increased from 8.9 to 9.8 - 10.2 in the *Chlorella* culture without  
366 monolayer and increased to >10.5 in the first 24 hours of cultivation with the monolayer.  
367 Meanwhile, the dissolved oxygen was approximately 8.5 mg/L and 8 mg/L in the monolayer  
368 and control cultures, respectively (Fig. 3). The slightly higher pH and decreased biomass  
369 productivity in the freshwater monolayer cultures could reflect a reduction in CO<sub>2</sub> mass transfer  
370 into the media caused by the monolayer, while the increase in dissolved oxygen could similarly  
371 result from a reduction in outgoing O<sub>2</sub> mass transfer. Decreased gas diffusion resulting from  
372 the presence of a monolayer has previously been proposed in freshwater systems [8]. To  
373 investigate whether the monolayer film reduced the rate of CO<sub>2</sub> mass transfer from the  
374 atmosphere, separate experiments were performed using a novel method based on the Wilbur  
375 Anderson assay (Supplementary File S.4). The results showed no reduction in the rate of CO<sub>2</sub>  
376 mass transfer due to the presence of monolayer film for either freshwater or marine growth  
377 media. The reduced growth rate of *C. vulgaris* (Fig. 2) in the presence of a monolayer cannot  
378 therefore be explained by a reduction in gas exchange. The most likely remaining factors  
379 appear to be direct chemical inhibition by the monolayer or a reduced growth rate due to the  
380 slightly elevated temperature resulting from reduced evaporation. Future research to  
381 understand the potential direct (e.g. chemical) and indirect (e.g. thermal) effects of this and  
382 other monolayer chemicals on *Chlorella* and other algae species is warranted. In particular, the  
383 current study showed that the ethylene glycol mono-octadecyl ether monolayer may have

384 negatively impacted *Chlorella* but not *Nannochloropsis*, suggesting the value of further  
 385 research involving different combinations of monolayer chemicals and algal species.

### 386 3.3 Techno-economic analysis of monolayer films for algae cultivation

#### 387 3.3.1 Impact of monolayers on water demand for large-scale algae cultivation

388 The experimental results from this study show that a monolayer can reduce water evaporation  
 389 from algae ponds by 50 - 70%. This information was used to investigate the effect of this  
 390 reduction on the overall water demand in large-scale algae cultivation (including contributions  
 391 from evaporation, leak losses, and process water disposal [17]) in relation to climate. Assuming  
 392 a 60% reduction in evaporation it was found that monolayer films can reduce the overall water  
 393 demand of freshwater algae cultivation by 6 - 21% depending on the climate (Table 1), with  
 394 arid regions seeing larger benefits from using the monolayer. This is due to evaporation losses  
 395 making up a smaller percentage of the water demand (10.4%) in regions with low evaporation  
 396 rates and high rainfall, such as tropical climates, compared to arid regions (35.7%). The impact  
 397 in arid regions is likely to be further enhanced given higher water prices associated with lower  
 398 water availability. This is particularly relevant to algae cultivation, as warm, sunny, arid regions  
 399 are typically most suited for large-scale algae cultivation.

400 **Table 1** Comparison of the water demand of standard algae production pond operation (WD  
 401 base) with that of using a monolayer film (WD film)\*.

Climate	Evaporation (mm/year)		WD base (mm/year)	WD film (mm/year)	Reduction in WD (%)	Rainfall (mm/year)
	Base	With film				
Tropical	476	190	4,566	4,280	6.3	4,250
Temperate	740	296	4,830	4386	9.2	1,150

Sub-tropical	1,150	460	5,240	4550	13.2	1,010
Mediterranean	1,320	528	5,410	4618	14.6	1,280
Arid	2,275	910	6,365	5000	21.4	54.9
*WD base and WD film both include process water disposal (3,230 mm/year) and leak losses (860 mm/year). Data for base evaporation rates, rainfall, and other water losses were obtained from <sup>[17]</sup> .						

### 402 3.3.2 Impact of monolayer on pond temperature

403 The application of a monolayer was seen to increase the temperature of the experimental algae  
404 cultures, which can be attributed to the reduction in thermal energy lost through evaporation.  
405 To investigate the temperature impact of reduced evaporation in different climates, calculations  
406 were performed based on the change in energy lost via evaporation and the corresponding  
407 temperature change assuming this energy was present as the specific heat of water in the pond,  
408 again assuming a 60% reduction in evaporation. The results are summarised in **Table 2**. For  
409 tropical and temperate climates, the calculated increase in pond temperature is between 1.5 to  
410 2.4 °C, which is similar to the temperature increase seen in the experimental results in this  
411 study. For regions with higher evaporation rates, the temperature increases. For example, in  
412 arid areas the temperature increase could be as high as 7.3 °C. Further trials would need to be  
413 conducted in these areas to demonstrate the impact of evaporation suppression on pond  
414 temperatures and algal growth rates in higher evaporation climates. As mentioned above,  
415 periodic application of the monolayer may be needed in these situations to minimise  
416 temperature stress.

417 **Table 2** Comparison of the impact of evaporation suppression on the rate of evaporative energy  
418 loss and pond temperature in different climates for a pond with a depth of 30 cm.

<i>Climate</i>	<i>Water evaporation</i> ( $m^3/day$ )		<i>Change in conditions</i>	
	<i>No monolayer</i>	<i>Monolayer</i>	<i>Rate of energy lost</i> ( $W/m^2$ )	<i>Pond temperature</i> ( $^{\circ}C$ )
<i>Tropical</i>	13.0	5.2	-22.0	1.5
<i>Temperate</i>	20.3	8.1	-34.3	2.4
<i>Sub-tropical</i>	31.5	12.6	-53.3	3.7
<i>Mediterranean</i>	36.2	14.5	-61.1	4.2
<i>Arid</i>	62.3	24.9	-105.4	7.3

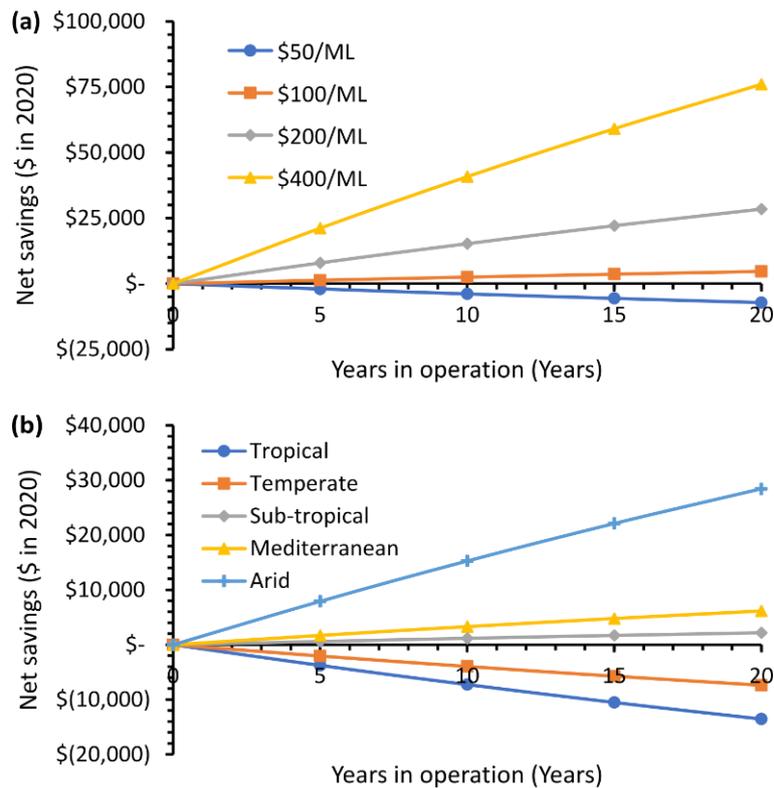
419

### 420 **3.3.2 Economic evaluation**

421 The application of monolayers to reduce evaporation in algae production ponds could have  
422 both economic and operational benefits. Firstly, the economic benefit was assessed for the  
423 reduced amount of water required to replace water lost and a reduction in associated pumping  
424 costs. A monolayer price of \$10/kg was assumed, with the same loading as the small-scale  
425 laboratory trials applied daily. The cost of applying the monolayer was not included, based on  
426 the assumption that the ponds will be actively managed, and monolayer application could be  
427 simply integrated into daily operations. Based on the performance of the monolayer in the algae  
428 trials, an evaporation reduction of 60% was assumed.

429 The first major factor that was considered was the price of water, which can range from \$50/ML  
430 up to \$400/ML in arid regions (Fig. 4a). The analysis shows that even without considering any  
431 operational benefits, monolayer application could be profitable for water prices >\$80/ML in  
432 arid regions. At a water price of \$200/ML, net savings were estimated at \$15,280 per hectare  
433 after 10 years. For a hypothetical commercial algae cultivation facility of 800 ha this would  
434 result in \$12.2 million in net savings over the life of a project [1].

435 The economic viability of the monolayer film was also considered for a range of different  
436 climates to show the impact of different evaporation and rainfall rates (Fig. 4b). Assuming a  
437 water price of \$200/ML, the economic benefit of monolayer films depends on the climate and  
438 is highest in regions where annual rainfall is below annual evaporation losses. Net cost savings  
439 are predicted for regions with medium to high rates of evaporation, such as sub-tropical and  
440 arid regions, which importantly are typically the regions most suited to algae production. It is  
441 also possible that monolayer addition could be avoided during times of the year when  
442 evaporation losses are below the economically viable rates, further improving the savings.



443

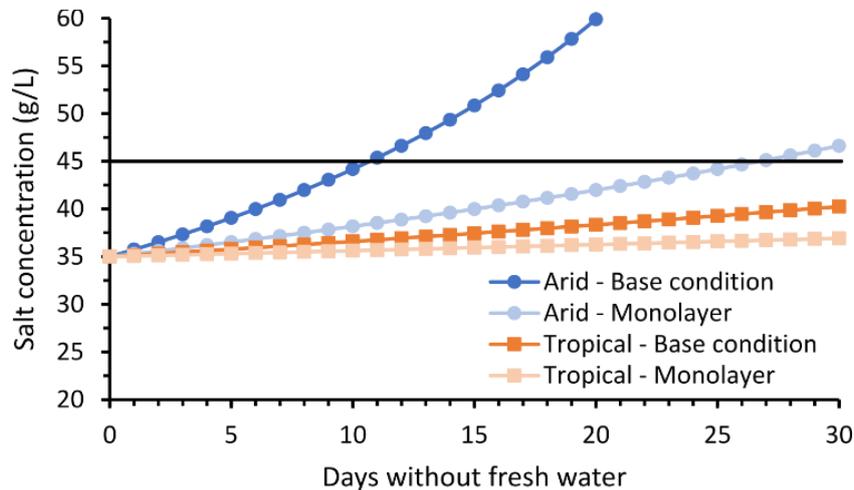
444 **Fig. 4** Economic analysis of using monolayer films on a one hectare basis over a 20 year period:  
 445 a) in arid regions with water prices between \$50/ML and \$400/ML, b) in regions with different  
 446 climates at a water price of \$200/ML.

447 **3.3.3 Operational benefits of applying monolayers for marine algae cultivation**

448 Beyond direct water savings, the ability to reduce evaporation using monolayers has the  
 449 potential to positively impact the conditions and operation of the algae ponds. In particular,  
 450 evaporation during the cultivation of marine microalgae either results in increases in salinity  
 451 or the requirement for a freshwater source to maintain salinity. In this analysis, we investigate  
 452 the impact of monolayer application on the salt concentration in batch and continuous  
 453 operations.

454 First, the increase in salt concentration caused by evaporation during batch algae cultivation  
 455 was calculated with and without a monolayer in arid and tropical regions (Fig. 5). In the arid

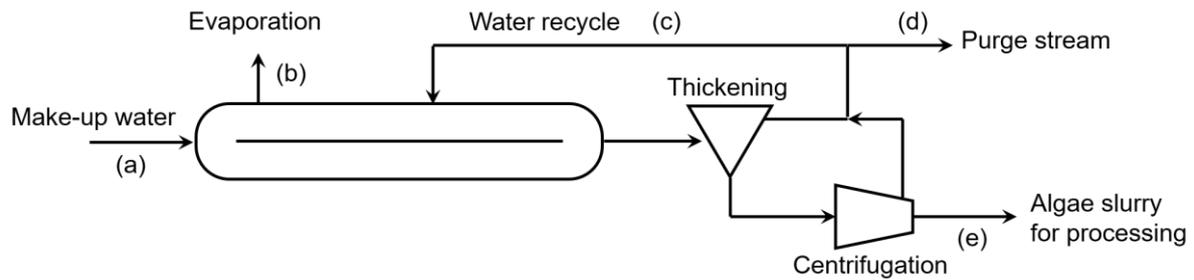
456 climate base case, the salt concentration reached 45 g/L (a potentially inhibitory level of salts  
 457 for some species) within 11 days, while the addition of monolayer increased this time to 27  
 458 days. This indicates that the addition of monolayer would help to keep the ponds closer to  
 459 optimal salt concentrations when there is limited or intermittent fresh water available.



460

461 **Fig. 5** Concentration of salt in a marine algae pond in arid and tropical climates with and  
 462 without monolayer. The initial salt concentration is 35 g/L (seawater), and operation is  
 463 conducted without any outflow or water top-up. An indicative upper threshold for algae growth  
 464 (45 g/L) is shown for reference.

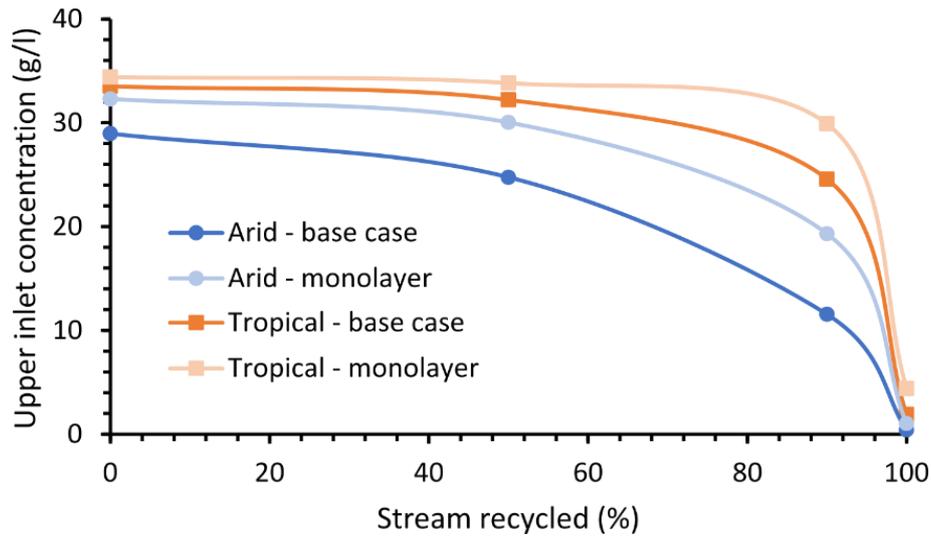
465 More commonly, algae will be cultivated in a continuous or semi-continuous manner (Fig. 6),  
 466 which includes streams of make-up water/growth medium (a), evaporated water (b), recycled  
 467 growth medium (c), a high-salt purge (d), and harvested and thickened algae (e). Without a  
 468 purge stream the salt level will increase as with batch operation, with the monolayer reducing  
 469 the rate of increase. Incorporating a purge stream into the system is more practical for  
 470 continuous algae growth as it allows the salt concentration to be maintained at a desired level  
 471 [32].



472

473 **Fig. 6** Process flow diagram of a raceway pond with concentration steps and a water recycle  
 474 stream.

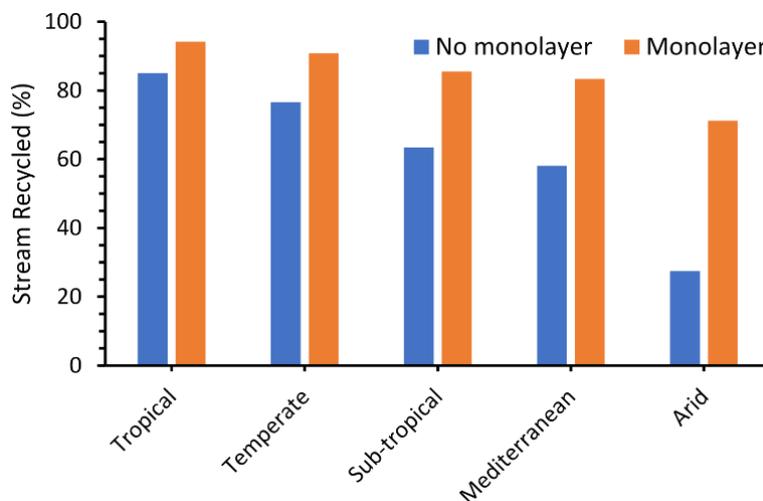
475 The impact of evaporation and water recycle on the maximum salt concentration in the make-  
 476 up water (stream (a) in Fig. 6) was investigated to determine the effect of the monolayer on  
 477 pond operation. The results (calculated based on an upper limit salt concentration in the pond  
 478 of 45 g/L) showed that at higher recycled stream percentages the upper limit salt concentration  
 479 in the make-up water stream must be reduced (Fig. 7). Higher evaporation rates (i.e., in arid  
 480 regions or without the monolayer) reduce the concentration of salt that can be included in the  
 481 make-up, reflective of the need to replace evaporated water with fresh water. For example, at  
 482 90% recycle rates the base case for an arid region required a make-up stream concentration less  
 483 than 12 g/L (about 1/3 the concentration of seawater), whereas in a tropical region up to 25 g/L  
 484 can be tolerated. Adding the monolayer to the surface significantly increased the maximum  
 485 make-up salt concentration. For example, in the arid region, this salt concentration increased  
 486 from 12 to 19 g/L. For regions where the make-up water is from salty or brackish water sources,  
 487 lower levels of purification will be required before use, potentially reducing processing costs  
 488 or even enabling algae cultivation in otherwise non-viable locations.



489

490 **Fig. 7** Upper inlet stream salt concentration (g/L) as a function of the percentage stream  
 491 recycled comparing the use of monolayer and no monolayer in arid and tropical climates. The  
 492 maximum salt concentration in the pond was set at 45 g/L.

493 As a result of adding a monolayer the amount of water recycled for a given inlet salt  
 494 concentration can be increased. In Fig. 8, this impact is illustrated for all climate regions. The  
 495 impact of the monolayer is significant in arid regions with more evaporation, again otherwise  
 496 suited for algae production. With an inlet salt concentration of 35 g/L, the addition of a  
 497 monolayer enables 71% of the recovered water to be recycled compared with 28% in the base  
 498 case.



499

500 **Fig. 8** Percent of recycle stream to maintain an equilibrium operation with a pond concentration  
501 at 45 g/L and an inlet saltwater stream of 35 g/L comparing operation with and without a  
502 monolayer.

### 503 ***3.3.4 Additional considerations for large-scale implementation***

504 The implementation of monolayer technology in large-scale, outdoor algae production  
505 facilities, will present a range of practical challenges that are beyond the scope of this study  
506 and will require future outdoor testing. For instance, the effects of turbulent mixing and CO<sub>2</sub>  
507 sparging on monolayer performance can be investigated in pilot raceways. In addition, the  
508 potential impact of the monolayer material on the need for post-treatment prior to water  
509 discharge, recycling or biomass utilisation should be considered. Studies on the use of  
510 monolayers for freshwater storage dams [10][11][21, 23] have concluded that the chemicals  
511 had no potential toxicity on the biodiversity of phytoplankton, and chemical monolayers such  
512 as cetyl alcohol and stearyl alcohol were found to be naturally biodegraded by bacteria [10].  
513 Nonetheless, depending on the choice of monolayer and the system used, there will need to be  
514 studies on the potential degradation products from the monolayer materials. The effect of the  
515 presence of monolayer compounds in the resulting algal products may also require additional  
516 consideration for certain applications such as animal and aquaculture feed.

517

## 518 **4. CONCLUSIONS**

519 This study has shown that an ethylene glycol monoctadecyl ether monolayer could reduce  
520 evaporation by 50-70% in both freshwater and marine algae cultures. Although performance  
521 declined, this evaporation reduction efficiency remained above 50% for 3 days. Despite  
522 extensive investigations, the reduction in performance could not be attributed to any singular  
523 cause. Freshwater *Chlorella* and marine *Nannochloropsis* algae were able to grow effectively

524 in the presence of the monolayer. The growth rate of *Chlorella* was somewhat lower in the  
525 presence of a monolayer, the reasons for which remain unconfirmed but could be due to  
526 chemical inhibition by the monolayer or a slight increase in culture temperature (1–2 °C).  
527 Importantly, CO<sub>2</sub> mass transfer from the atmosphere was shown to not be reduced by the  
528 monolayer. A techno-economic analysis indicated that the application of the monolayer could  
529 be economically beneficial, solely on the basis of water savings, particularly in arid climates  
530 in which algae are most suited for cultivation. In addition, reduction of evaporation using  
531 monolayers offers a means of controlling salinity in marine algae production systems. It is  
532 suggested that future investigations are conducted to understand the performance of different  
533 combinations of algae and monolayer chemicals, and that performance is tested in outdoor  
534 raceway ponds trials considering other operating factors such as turbulent mixing and excess  
535 CO<sub>2</sub> supply.

## 536 **ASSOCIATED CONTENT**

### 537 **Supporting Information**

538 Composition of f/2 and MLA media, performance of monolayer on sea salt and major  
539 components of f/2 medium, microscopic images of *N. salina* and *C. vulgaris* cultures with or  
540 without monolayer film, and measurement of surface CO<sub>2</sub> mass transfer.

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## 544 **AUTHOR STATEMENT**

545 The authors declare no commercial or proprietary interest in any product or concept discussed  
546 in this article.

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#### 550 **ABBREVIATIONS USED**

551 CO<sub>2</sub>, carbon dioxide; O<sub>2</sub>, oxygen; C18E1, ethylene glycol mono-octadecyl ether; f/2, marine  
552 growth media; MLA, freshwater growth media.

553

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