Consensus guidelines for antifungal prophylaxis in haematological malignancy and haemopoietic stem cell transplantation, 2021

Introduction

In the seven years since the publication of the last guidelines, the treatment of malignancies has undergone a paradigm shift with the increasing use of targeted oral therapies (e.g. Bruton's tyrosine kinase [BTK] inhibitors) and the advancement of immune-based therapies (e.g. immune checkpoint inhibitors [ICI], chimeric antigen receptor [CAR] T cell therapy). These therapeutic advances have led to significant improvements in disease prognosis and survival; however, in haematological malignancy, they also appear to be associated with new potential risks for invasive fungal disease (IFD). Other well-established risks for IFD, such as prolonged neutropenia and graft-versus-host disease (GVHD), remain relevant and unchanged from previous guidelines.

The landscape for the prevention and treatment of IFD has also evolved with the development of new antifungal therapies, as well as new formulations of established agents, for treatment and prophylaxis.^{6, 7} Selection of the optimal prophylactic agent is now impacted by new challenges, such as the emergence of the multi-drug resistant fungal pathogen *Candida auris*, increasing rates of antifungal resistance in *Aspergillus* spp., and the changing epidemiology of invasive candidiasis and non-*Aspergillus* moulds.⁸⁻¹⁰

The current guidelines take all of these developments into account and serve as an update to the 2014 guidelines.⁵ Primary antifungal prophylaxis is recommended for disease groups associated with a high risk of IFD. Not surprisingly, the higher the baseline prevalence of

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IFD, the more marked the beneficial effect of prophylaxis is.¹¹ When implementing prophylaxis regimens, consideration must also be given to institutional epidemiology, and relevant adjustments made. Most of the available evidence for prophylaxis relates to haematological malignancies, and this is the primary focus of these guidelines. For solid organ tumours, the need for prophylaxis remains undefined and expert advice should be sought on an individual case-by-case basis.

Methodology

Questions asked

This update addresses the following questions:

- 1. What new therapies for haematological malignancies are associated with potential risk for IFD and what are the new risk groups?
- 2. Is there new evidence to support and guide the use of established prophylactic agents?
- 3. What are the new options for antifungal prophylaxis?
- 4. How does the use of mould-active antifungal prophylaxis impact diagnostic testing?
- 5. What critical drug-drug interactions do clinicians need to be aware of and what is the need for therapeutic drug monitoring (TDM) in the era of targeted and immune-based therapies?
- 6. How do antifungal resistance patterns affect choice of prophylaxis?
- 7. What special considerations are required in the paediatric setting?
- 8. How could antifungal prophylaxis recommendations be implemented into practice?

9. Are there new approaches to defining risk for IFD?

Search strategy

A comprehensive literature search was performed utilising PUBMED to address the questions previously outlined. The search encompassed studies published since 2013 and utilised the following terms in combination: 'haematology', 'malignancy', 'haemopoietic stem cell transplantation', 'haematopoietic stem cell transplantation', 'leukaemia', 'myeloma', 'lymphoma', 'risk factors', 'antifungal', 'prevention', 'prophylaxis', and 'invasive fungal infection'.

Question 1 What new therapies for haematological malignancies are associated with potential risk for IFD and what are the new risk groups?

Recommendations

- Due to the absence of high-level evidence, the routine use of antifungal prophylaxis is
 not recommended for the majority of patients undergoing treatment with new
 haematological treatments; rates of IFD with new therapies are summarised in Table 2.
 Antifungal prophylaxis should be considered on an individual patient risk model (see
 Table 3).
- For patients receiving new generation immunomodulatory, monoclonal antibody therapy for relapsed and refractory myeloma, prophylaxis with fluconazole could be considered [Marginal recommendation, Level III evidence].
- For patients undergoing CAR-T cell therapy, prophylaxis with fluconazole should be considered [Strong recommendation, Level II evidence].
- For patients deemed at higher risk of fungal infection (e.g. due to severe neutropenia or multiple lines of therapy, treatment of cytokine release syndrome), mould-active azole prophylaxis could be considered [Moderate recommendation, Level II evidence].
- For patients with a prior history of IFD, secondary prophylaxis should be administered
 [Marginal recommendation, Level III evidence].

Given recent advances in the treatment of haematological malignancies and the potential association between these newer agents and IFD risk, this section discusses new advances in the treatment of haematological malignancies, their potential association with risk for IFD

and the resultant recommended approach to prevention of IFD. Established risk groups for IFD are summarised in Table 1.

Targeted and immunomodulatory drug therapies

Since the publication of the previous guidelines, an increasing number of targeted agents have become available as standard of care options for the treatment of both haematology and medical oncology patients. Such agents, through their effects on immune function, may increase the risk of IFD.¹² Reported rates of IFD accompanying the use of these agents vary according to the patient group being treated (i.e. treatment naïve versus relapsed/refractory malignancy); previous treatments used, including number of lines of therapy; and whether these agents are used in combination with other therapies, especially conventional chemotherapy that induces mucositis or prolonged neutropenia.

Ibrutinib, a BTK inhibitor commonly used for the treatment of chronic lymphocytic leukaemia (CLL) and other B-cell lymphoproliferative disorders, interrupts B-cell receptor signalling and also results in hypogammaglobulinaemia. There is an association between ibrutinib therapy and risk of IFD. A retrospective study of 378 patients receiving ibrutinib (monotherapy in 84% of cases) reported an IFD rate of 4.2%, with the majority of IFD cases lacking classical risk factors such as neutropenia or corticosteroid usage.¹³ Local, real-world data suggests an IFD rate as high as 12.1% in patients treated with ibrutinib monotherapy in the setting of relapsed/refractory CLL.¹⁴ These findings contrast the 1% IFD rate reported in the randomised, phase 3 RESONATE study.¹⁵ The majority of fungal infections reported were invasive aspergillosis with a predilection for central nervous system (CNS) involvement (40%).¹⁶ Most patients developed IFD within three to six months of starting ibrutinib.

Substantially higher IFD rates of 38.9% (7/18) have been observed in the context of primary CNS lymphoma treated with ibrutinib, potentially due to the concomitant use of chemotherapy and corticosteroid agents.¹⁷ This emphasises the additional risk imposed when combining BTK inhibition with other immunosuppressive therapies.

Cases of IFD, including *Pneumocystis jirovecii* pneumonia, have been reported in association with the use of other tyrosine kinase inhibitors such as Janus kinase (JAK) inhibitors, ^{18, 19} phosphatidylinositol 3-kinase (PI3K) inhibitors, ²⁰ B-cell lymphoma 2 (BCL-2) inhibitors (venetoclax)²¹ and mammalian target of rapamycin (mTOR) inhibitors. ²² In retrospective studies, the use of hypomethylating agents such as azacitidine in patients with myelodysplastic syndromes and acute myeloid leukaemia (AML), has been associated with an IFD risk of up to 8.3% – consistent with the risk posed by treatment of the underlying disease. ^{23, 24} The greatest IFD risk was observed amongst patients who had received prior intensive chemotherapy.

In the setting of variable antifungal prophylaxis use (i.e. either an azole, echinocandin, or none), the use of hypomethylating agents in combination with the BCL-2 inhibitor venetoclax for newly-diagnosed and relapsed or refractory AML, was associated with an overall IFD rate of 12.6%.²⁵ The IFD rate in newly-diagnosed AML was 5.0%.²⁵ The addition of an fms-like tyrosine kinase (FLT3) inhibitor (midostaurin, gilterinib) to standard chemotherapy for the treatment of FLT3-positive AML has been demonstrated to improve survival.^{26, 27} However, the impact on IFD rates beyond that expected of the underlying AML remains undefined. In an early clinical trial, an IFD rate of 5% was reported.²⁶ These targeted agents, particularly BTK inhibitors and BCL-2 inhibitors, have significant CYP3A4 interactions with azole antifungals, necessitating their dose adjustment.³

IFD rates of 3.8–5.6% were reported in patients with multiple myeloma (MM) treated with first-generation immunomodulatory drugs and proteasome inhibitors.^{28, 29} The rate of invasive mould infection was less than 1.0%.²⁸ An IFD rate of 15% was reported in patients who had received three or more lines of therapy, including conventional chemotherapy.²⁸ In contrast, a recent study of a heavily-treated MM patient cohort (median five lines of therapy) treated with next-generation immunomodulatory drugs, proteasome inhibitors and new anti-CD38/SLAM-F7 monoclonal antibody therapies, reported a low overall IFD rate of 3.4%.³⁰ The rates ranged from 2.3–7.0% per the specific drug classes. Patients with IFD had received more lines of therapy and all proven IFD cases involved yeasts, i.e. cryptococcal infection and *Pichia kudriavzeveii* (formerly *Candida krusei*).³⁰

Adoptive T-cell therapies

Adoptive T-cell therapies such as CAR T cell therapy and bi-specific constructs such as bi-specific T-cell engagers (BiTE) (e.g. blinatumomab) have now become standard of care in relapsed, high-grade B-cell malignancies: CAR T cell therapy for diffuse large B-cell lymphoma and paediatric acute lymphoblastic leukaemia (ALL), and blinatumomab for ALL. Patients receiving these therapies have often had multiple prior lines of therapy, including HSCT, which may influence their fungal infection risk. Other agents such as fludarabine may be administered prior to CAR T cell infusion, which may further increase fungal infection risk.³¹⁻³⁵

CAR T cell therapy

Rates of infective toxicity appear similar in patients receiving CAR T cell therapy or other salvage therapies. Bacterial infections are a larger concern, occurring in 23% of patients receiving CAR T cell therapy, predominately during the first 28 days following CAR T cell infusion. IFD rates in trials of ALL and B-cell lymphoma have ranged between 5–13%

depending on the disease group.^{31, 32, 36} In a dedicated study of infectious complications with CAR T cell therapy, 5% of patients developed a fungal infection in the setting of fluconazole prophylaxis.⁴ An IFD rate of 7% has also been reported with the use of micafungin prophylaxis in ALL patients managed with CAR T cell therapy.³⁷ Patients with ALL appear to be at a higher risk of infection if they have received four or more prior lines of therapy, been treated with higher CAR T cell therapy doses, developed cytokine release syndrome (CRS) or used systemic corticosteroid agents during their CAR T cell therapy.^{4, 38, 39} Potentially, a subset of patients with prolonged neutropenia or with higher-grade CRS requiring additional immunosuppressive treatments (e.g. corticosteroid agents) may be at higher risk for mould infections.

Bi-specific immune engagers

At time of publication, blinatumomab is the only bi-specific immune engaging agent in clinical practice and is indicated for patients with ALL, both relapsed/refractory disease and minimal residual disease. Blinatumomab is only modestly myelosuppressive and does not cause mucotoxicity. However, it can be associated with neutropenia and risk of infection can be compounded by prior therapies received. Fungal infection is relatively rare; however, early development of neutropenia has been associated with an increased risk of possible fungal infection.^{35, 40}

Rates of IFD associated with these emerging cancer therapies and potential new risk groups are summarised in Table 2 and recommended approaches to prophylaxis are summarised in Table 3. There should be an increased awareness of the potential risk and clinicians should have a low threshold for initiating investigations for suspected IFD in the setting of compatible symptoms such as persistent fever or pulmonary infiltrates.

Question 2 Is there new evidence to support and guide the use of established prophylactic agents?

Recommendations

- Posaconazole remains recommended as a first-line agent for IFD prophylaxis in high-risk patients [Strong recommendation, Level I evidence].
- Voriconazole is considered to be an alternate agent for IFD prophylaxis due to higher rates of adverse events (e.g. liver function abnormalities) and variable metabolism
 [Strong recommendation, Level II evidence].
- Use of micafungin could be considered during periods of neutropenia in high-risk
 patients if use of azoles is contraindicated or in the setting of expected poor
 gastrointestinal absorption [Moderate recommendation, Level II evidence].
- Itraconazole is considered to be an alternate agent due to a lower number of clinical trials and cohort studies [Moderate recommendation, Level II evidence].
- Use of liposomal amphotericin B could be considered if use of azoles is contraindicated due to drug-drug interactions, adverse events or poor absorption [Moderate recommendation, Level II evidence].

Posaconazole

Posaconazole remains the preferred agent for prophylaxis against IFD in high-risk patients with AML or for those undergoing allogeneic HSCT.

Network meta-analyses of randomised controlled trials of triazole prophylaxis confirm posaconazole's efficacy for the prevention of proven or probable IFD and invasive

aspergillosis, reducing the requirement for empiric antifungal therapy and all-cause mortality compared to fluconazole and itraconazole.⁴¹⁻⁴³ Although posaconazole was not significantly better than voriconazole for prevention of IFD and invasive aspergillosis, it was the highest-ranked agent for achieving these outcomes in two published network meta-analyses.^{41, 43} When evaluated, its use appears to be cost-effective compared to voriconazole.⁴¹

Cohort studies evaluating posaconazole against voriconazole, itraconazole or micafungin consistently report lower rates of IFD with posaconazole ranging from 0–5% versus 5–11%. ⁴⁴⁻⁴⁸ For patients undergoing HSCT, observational cohort studies have shown that rates of breakthrough IFD during prophylaxis with posaconazole suspension remain low at between 3–8%. ^{49, 50} Cohort studies of AML patients report similarly low rates of proven or probable breakthrough IFD (0–7%). ^{45, 51, 52}

Voriconazole

Voriconazole is an alternate agent for IFD prophylaxis. Meta-analyses show no significant difference between posaconazole and voriconazole efficacy for the prevention of proven or probable IFD and invasive aspergillosis. 41-43 However, a significantly higher risk for treatment-related liver abnormalities was noted, compared to other azoles. 41, 43 In the same analysis, this agent is ranked second as an effective prophylaxis agent. 41 In cohort studies of AML patients, the use of voriconazole prophylaxis was associated with an IFD rate of 3–5%. 48, 53, 54 Due to variable metabolism, CYP2C19 testing prior to commencement could assist with dose selection (please refer to the accompanying optimising antifungal therapy and TDM guidelines by Chau *et al.* 2021, which can be found elsewhere in this supplement). 55, 56

Itraconazole

Since the 2014 guidelines, a new formulation of itraconazole has been introduced (see later discussion). The only new data supporting the use of intravenous itraconazole or its solution are from a few cohort studies reporting IFD rates of 1–7% for HSCT patients and 5% for patients with AML.^{53, 57, 58}

Micafungin

There have been further studies evaluating the use of micafungin during the neutropenic period in HSCT patients. In two trials, the rate of IFD was not significantly different when assessed against fluconazole and itraconazole at 7.3% and 4.4%, respectively.^{57, 59} Adverse event rates were significantly higher with itraconazole.⁵⁷ Different doses have been assessed but, in general, dosing with 100–150 mg intravenous (IV) daily followed by oral voriconazole or posaconazole on discharge, led to proven or probable IFD rates of between 1–4%.⁶⁰⁻⁶² In the AML cohort, the rate was 6.3%.⁴⁷ Overall, use of micafungin could be considered during the neutropenic period in high-risk patients if use of azoles is contraindicated or there are concerns about absorption.

Liposomal amphotericin B

Evidence supporting the use of liposomal amphotericin B (L-AMB) remains limited. A recent randomised trial of L-AMB at 5 mg/kg twice a week compared to placebo for prophylaxis in ALL reported no difference in the rate of proven or probable IFD (7.9% vs. 11.7%; P = 0.24); however, a significantly higher rate of adverse events led to interruption of L-AMB in 20.3% of patients.⁶³ *Post hoc* analysis did report a trend for lower IFD rates in patients who were administered L-AMB prophylaxis (7.6 vs. 14.4%; P = 0.07).⁶³ In a cohort study of Australian ALL patients, use of L-AMB at a median dose of 100 mg three times per week had a similar rate of IFD to posaconazole (6% vs. 7%), and a lower rate compared to both fluconazole (14.3%) and no prophylaxis (21%).⁶⁴ At this stage, evidence supporting its use

remains poor but this agent could be considered in the setting of azole intolerance or contraindication. Optimal dosing of this agent for prophylaxis requires further evaluation. Doses ranging from 50–200 mg, three times per week, have been used.

Recommendations by risk group and grading of evidence for the selection and dosing of an antifungal prophylactic agent are summarised in Table 4.

Question 3 What are the new options for antifungal prophylaxis?

Recommendations

- Isavuconazole is not recommended as a first-line agent for prophylaxis against IFD in high-risk patients due to higher reported rates of IFD in cohort studies. Its use can be considered if azoles are contraindicated (e.g. QTc prolongation) [Moderate recommendation, Level II evidence].
- There is insufficient evidence to support use of the new formulation of itraconazole as a
 first-line agent for prophylaxis against IFD [Moderate recommendation, Level II
 evidence]. However, it is used in several Australian centres.
- Oral posaconazole remains recommended as a first-line agent for IFD prophylaxis in high-risk patients (tablets are preferred but in some cases, suspension may be necessary, in which case, prescribers should consult with pharmacy regarding levels and dosing) [Strong recommendation, Level I evidence]. Intravenous formulation is an option for continuation of posaconazole prophylaxis in the setting of poor or limited oral intake.

Isavuconazole

Isavuconazole is a recently introduced broad-spectrum triazole antifungal agent. It has been licensed for the treatment of invasive mould disease on the basis of non-inferiority compared to voriconazole in the SECURE trial and for the treatment of mucormycosis, either as primary treatment or for treatment of infection refractory or intolerant to other antifungals, based on a mixed group of patients in the VITAL study.^{6, 65} Based on its broader spectrum of activity and improved side-effect profile, it is being evaluated as prophylaxis in

patients with AML and allogeneic HSCT in prospective trials. In an early phase 2 dose-escalation study, the rate of breakthrough invasive fungal infection was 10%.⁶⁶

Results from its use as prophylaxis, as reported in prospective studies, retrospective studies and case reports, have been variable. $^{67-71}$ In a mixed population of relapsed refractory and HSCT patients, a breakthrough rate of 5.8% was reported. 70 However, the use of isavuconazole as prophylaxis in newly-diagnosed AML was associated with a rate of 7.9%. This is higher than the rate reported with posaconazole (2.7%), but is not statistically significant (P = 0.06). 68 Posaconazole was used for a longer period in patients with a longer duration of neutropenia. 68 A prospective single-centre, single-arm, primary prophylaxis study in AML and myelodysplastic syndrome reported an overall breakthrough rate of 18.0% with a proven/probable rate of 6.0%. 71 Half the patients were receiving treatment with oral targeted anti-leukaemic agents such as venetoclax, which has CYP3A4-mediated drug-drug interactions when co-administered with other azoles. 71

Use of isavuconazole as prophylaxis in the setting of relapsed or refractory AML has been associated with breakthrough rates of between 12–18.5%, ^{68, 69} higher than that reported with posaconazole and voriconazole (5.5%). ⁶⁸ When performed in a subset of patients, isavuconazole levels appeared to be adequate. ^{68, 70, 71} In the majority of cases, breakthrough infections were due to *Aspergillus* spp. and *Mucor* spp. ^{68, 69} The use of isavuconazole following micafungin prophylaxis in HSCT patients has been associated with an IFD rate of 3.1%. In this study all IFD were bloodstream infections with *Candida parapsilosis* and *Candida glabrata*. ⁷² Tolerability appears to be good with a low risk of QTc prolongation in the setting of potential drug-drug interactions. ⁷¹ Evidence from trials being conducted may provide further clarity.

Currently, the use of isavuconazole as a first-line agent for prophylaxis in patients with AML and HSCT cannot be recommended due to higher observed IFD rates in uncontrolled cohort studies. Its use could be considered in the setting of intolerance or if use of other azoles is contraindicated.

New formulation of itraconazole

A novel formulation of itraconazole (SUper BioAvailability [SUBA]-itraconazole) has been approved in Australia since 2014.^{73, 74} At the time of the 2014 guidelines, only data from healthy volunteer studies were available on the SUBA-itraconazole formulation, which demonstrated that this formulation is not affected by gastric pH with dosing recommendations differing from the previous conventional itraconazole capsule formulation.⁷⁵ However, recently there have been a number of small cohort studies demonstrating good tolerability and levels in the therapeutic range using SUBA-itraconazole in haematology and HSCT recipients. 76,77 One small prospective cohort (n = 57) compared SUBA-itraconazole for primary prophylaxis in an allogeneic HSCT cohort to itraconazole oral solution. Therapeutic concentrations were achieved significantly more quickly in the SUBAitraconazole group (median of six days versus 14 days) with therapeutic concentrations achieved in 69% vs. 21% of patients (P < 0.01). Of note, there were no treatment failures due to gastrointestinal intolerance, which has previously limited the use of conventional itraconazole formulations. 76 In another small retrospective cohort study (n = 74) of myeloma, AML, ALL, autologous and allogeneic HSCT patients, therapeutic concentrations were achieved at a median of seven days by 87% of patients.⁷⁶ The incidence of IFD reported in both studies was low at 3% and 1% respectively; however, the studies were too small to accurately assess efficacy. It should be noted that despite the manufacturer's recommendation to use half the relative dose of the conventional capsules, both studies

used SUBA-itraconazole at the recommended dose range of the itraconazole oral solution (i.e. 200 mg twice daily with TDM used to monitor levels and dose-adjust accordingly).

Posaconazole: tablet and intravenous formulation

Posaconazole modified-release tablet and intravenous (IV) formulation have been licensed in Australia since 2014 and 2015, respectively. The tablet is listed on the Pharmaceutical Benefits Scheme (PBS) for prophylaxis of IFD in specific high-risk groups, namely patients with anticipated neutropenia while receiving chemotherapy for AML or myelodysplastic syndrome, and patients with acute (grade II–IV) or extensive-chronic GVHD receiving intensive immunosuppressive therapy following an allogenic HSCT. The evidence for its efficacy in these settings is extrapolated from the prophylaxis studies using posaconazole oral solution.^{78, 79} Advantages of the tablet formulation are once-daily dosing, higher drug exposure than with the oral solution,⁸⁰ and no requirement for concurrent intake of fatty food to improve absorption.⁸¹ The safety profile is similar between the two formulations.⁷ Although, gastric pH was not thought to influence absorption (which occurs in the small intestine), recent studies in patients with haematological malignancies or HSCT receiving posaconazole tablets as prophylaxis have indicated that proton pump inhibitors and corticosteroid agents (>0.7 mg/kg daily) may lead to lower plasma concentrations with the tablet formulation.⁸²

In real-life studies reporting trough levels in approximately 230 haematological malignancy or HSCT patients receiving posaconazole tablets, between 3–18% of patients had subtherapeutic levels, defined as <700 ng/ml with prophylaxis and <1000 ng/ml with treatment.⁸³⁻⁸⁶ These studies may be biased towards detecting lower levels, as levels were not routinely measured in all series reported and when performed, it was due to clinical concerns about absorption or failure. Correlations with higher posaconazole levels and

hepatotoxicity were not clearly seen in these real-world studies, although one small report linked trough levels >1,830 ng/ml and pre-existing liver damage with grade 3–4 liver injury.⁸⁴ In these studies, breakthrough infection did not always occur within the context of low levels, indicating that host factors continue to play a role in prophylaxis failure.

IV posaconazole remains an option for continuing posaconazole administration when oral medication cannot be taken (e.g. severe mucositis, nausea, vomiting, GVHD of the gut or a requirement to remain fasting). In one study, IV posaconazole was given as antifungal prophylaxis to 237 neutropenic patients with AML, myelodysplastic syndrome or recipients of allogeneic HSCT.⁸⁷ The dose was 300 mg of posaconazole IV twice daily on day 1, followed by 300 mg IV once daily for at least five days followed by a switch to the oral suspension. The mean posaconazole trough level on day 6 was 1320 ng/mL and posaconazole was well tolerated.⁸⁷ A real-life study of patients treated for haematological malignancy and HSCT recipients – the majority of whom received prophylaxis – reported a median trough level of 1.16 (0.69–2.06) mg/L, taken 3–7 days after commencement. The median duration for prophylaxis was 10 days. No severe adverse events specifically attributable to IV posaconazole were documented, although six courses were curtailed due to potential toxicity.⁸⁸

Question 4 How does the use of mould-active antifungal prophylaxis impact diagnostic testing?

The performance of diagnostic tests for fungal infection is highly dependent on pre-test probability and incidence of IFD.⁸⁹ Primary antifungal prophylaxis in high-risk patients successfully reduces the incidence of proven or probable IFD to around 5%, with a corresponding reduction in test performance, in particular galactomannan (GM) testing.^{44, 51, 90, 91} The utility of performing serum GM or *Aspergillus* PCR testing for IFD surveillance in the setting of primary antifungal prophylaxis appears to be limited.⁸⁹⁻⁹¹ However, these assays remain useful as part of a diagnostic-driven algorithm for suspected breakthrough IFD in high-risk patients.^{91, 92} The impact of antifungal prophylaxis on the performance of a range of diagnostic tests is discussed in further detail in subsequent sections of these guidelines.

Question 5 What critical drug-drug interactions should clinicians be aware of and what is the need for TDM in the era of targeted and immune-based therapies?

Recommendations

- Dose adjustment of targeted therapies is required in the setting of major CYP3A4 interaction [Strong recommendation, Level II evidence].
- Use of CYP testing may assist with ascertainment of target drug level (refer to the
 accompanying optimising antifungal therapy and TDM guidelines by Chau et al. 2021,
 which can be found elsewhere in this supplement).
- Due to real-world evidence of sub-therapeutic drug levels in up to 20–30% of patients,
 TDM is still important for new formulations of azole antifungal agents [Marginal recommendation, Level III evidence].

With the introduction of novel targeted therapies for hematological malignancies and HSCT, managing the drug-drug interactions associated with azole antifungals requires careful consideration. Of particular note, when using a strong CYP3A4 inhibitor, such as posaconazole, itraconazole or voriconazole, in combination with novel targeted therapies that are major CYP3A4 substrates such as venetoclax and ibrutinib, manufacturer recommendations of up to 75% dose reductions of the novel agents are indicated.^{3, 93, 94} However, other commonly used novel agents such as midostaurin and gilteritinib are considered weak/moderate CYP3A4 substrates and dose adjustment is not recommended.^{95, 96} Therefore, manufacturer recommendations and/or specialist advice should be considered before using azoles in combination with novel targeted therapies. There is also growing

evidence that agents such as letermovir and flucloxacillin, significantly reduce voriconazole concentrations. ^{97, 98} These potential interactions further emphasise the importance of TDM of azole antifungal agents, particularly during changes in concomitant medications, as well as during periods of critical illness such as intensive care unit (ICU) admissions. ⁹⁹ In addition, despite superior level attainment from new oral and IV formulations of posaconazole, ^{7, 100} post-marketing experience has reported subtherapeutic serum trough concentrations in approximately 20–30% of haematology patients. ^{84, 88, 101} Therefore, ongoing use of TDM for these new formulations should be considered. For further information, please refer to the accompanying optimising antifungal therapy and TDM guidelines by Chau *et al.* 2021, which can be found elsewhere in this supplement.

Question 6 How do antifungal resistance patterns affect choice of prophylaxis?

Recommendation

The antifungal prophylaxis recommendations in this guideline take into account currently
available data on antifungal resistance rates and patterns; however, a local
organisational approach for ongoing surveillance of the rates and epidemiology of IFD in
high-risk patients is important for guiding effective prophylaxis choice [Strong
recommendation, Level II evidence].

Clinicians continue to be challenged by the changing epidemiology of fungal infections over time, emergence of multi-drug resistant fungi such as *Candida auris*, and primary antifungal resistance in human and environmental isolates.

The epidemiology of *Candida* spp. infection continues to evolve with increasing use of azoles and echinocandins. A nationwide surveillance of candidaemia revealed a 1.7-fold increase in the proportion of candidaemia due to *Candida glabrata*. ¹⁰ *Candida glabrata* is now responsible for up to 30% of candidaemia¹⁰² and is the leading cause of candidaemia in haematology patients. ¹⁰³ Between 13–23% of *Candida glabrata* isolates are fluconazole-resistant, a key consideration in lower-risk patients receiving fluconazole prophylaxis. Multi-drug resistance remains uncommon in isolates responsible for candidaemia but azole resistance is at 17% and thus ongoing surveillance is vital. ¹⁰

In the last five years, *Candida auris* has emerged as a healthcare-associated, multi-drug resistant yeast, which has caused significant outbreaks in multiple countries around the world. It is an effective coloniser of the hospital environment and patients, causing invasive

infection (predominantly candidaemia).⁹ *Candida auris* is resistant to at least two antifungal drug classes in nearly 25% of cases.⁹ At last count, less than 10 cases of this infection have been reported in Australia.^{9, 104} An Australasian diagnostic, infection prevention and clinical management approach to this infection has recently been developed with multi-stakeholder involvement.⁹

Multi-triazole-resistant *Aspergillus fumigatus* have been isolated in up to 30% of clinical isolates in countries overseas. ¹⁰⁵ A 13-year review of *Aspergillus fumigatus* isolates in the National Mycology Reference Laboratory detected only two isolates carrying the TR34/L98H mutation of *CYP51A*. ¹⁰⁵ Fortunately, rates of azole resistance in *Aspergillus fumigatus* isolates in Australia appear to be low at 2% of clinical isolates in a limited screen of human, animal and environmental isolates. ¹⁰⁶ No azole-resistant isolates were detected in animal or environmental isolates. ¹⁰⁶ Testing of clinical isolates of non-*Aspergillus* fungal pathogens in Australia has confirmed expected susceptibility patterns for the isolated species. ¹⁰⁷

In the setting of antifungal prophylaxis, between 6–17% of IFD episodes were due to non-Aspergillus moulds, which have intrinsic resistance to a range of azole antifungals. ¹⁰⁸⁻¹¹⁰ In a national study, *Scedosporium* spp. and mucormycetes were the dominant species, contributing up to 80% of IFD caused by non-*Aspergillus* moulds, with underlying haematological malignancy associated with significantly higher odds of mortality. ¹¹⁰ Increasing use of mould-active antifungal prophylaxis will contribute to changing epidemiology of IFD in high-risk patients and active surveillance of breakthrough IFD should continue to guide choice of antifungal prophylaxis.

Question 7 What special considerations are required in the paediatric setting?

Recommendations

- Indications for antifungal prophylaxis in children (adapted from Lehrnbecher et al.,
 2020)¹¹ are provided in Table 6.
- Where antifungal prophylaxis is indicated, a mould-active agent is recommended [Strong recommendation, Level I evidence].
- The choice of mould-active agent will depend on age, potential drug interactions and
 patient location (inpatient versus outpatient), with preference given to mould-active
 azoles or echinocandins [Strong recommendation, Level II evidence].
- Intermittent liposomal amphotericin is an alternative option for children in whom azole
 prophylaxis is not tolerated or contraindicated and daily echinocandin administration is
 not feasible [Marginal recommendation, Level III evidence].
- Where antifungal prophylaxis is indicated, administer during periods of observed or expected severe neutropenia [Strong recommendation, Level II evidence].
- Secondary prophylaxis is recommended for children with proven or probable IFD
 undergoing subsequent immunosuppression, particularly during intensive phases of
 chemotherapy [Moderate recommendation, Level III evidence]. For agent selection and
 duration, specialist infectious diseases advice is recommended.
- Dosing recommendations in children are summarised in Table 7.

New therapies for haematological malignancy and new patient risk groups in children

Since the publication of the previous guidelines, the risk factors for IFD in children have been further refined (Table 5).⁵ In addition to previously recognised associations, including haematological malignancy, prolonged neutropenia, high-dose corticosteroids and severe-acute (grade II or above) or chronic GVHD, age (>7.5 years) has now also been identified as a risk factor.¹¹¹ In an Australian, multisite, 10-year cohort study of IFD in children, IFD prevalence in AML and HSCT cohorts was 28.2% and 11.7%, respectively.^{112, 113} IFD prevalence was <5% in autologous HSCT recipients (3.1%) and children with solid tumours (4.4%). Amongst ALL patients, IFD prevalence was 23.5% for relapsed/refractory ALL, 14.5% for high-risk ALL and 7.3% for standard-risk ALL, with IFDs more common during induction, consolidation and delayed intensification phases.¹¹² Across both relapsed and non-relapsed ALL, mould infections were more common than non-mould infections. These data suggest that a tailored prophylaxis approach is required for patients with ALL, taking into consideration disease risk status, chemotherapy intensity, remission status and phase of treatment.

There are limited data on risk of IFD with targeted and adoptive T-cell therapies in children. For an overview of adult data and approach to prophylaxis in both adults and children, please refer to earlier discussion on this topic, as well as Table 3. In a review of infective complications in 83 children and young adults receiving CAR T cell therapy, one patient (1.2%) developed a proven IFD in the first 30 days, although it is unclear if this was a new infection or progression of a previously documented probable pulmonary invasive mould infection.¹¹⁴

New evidence for established prophylactic agents in children

An international clinical practice guideline for antifungal prophylaxis in children with cancer or HSCT has been recently published.¹¹ Underpinning these recommendations were rigorous and comprehensive systematic reviews addressing: (i) which paediatric patients should routinely receive antifungal prophylaxis and (ii) what agents should be used. Only randomised clinical trials were considered and evidence rated according to the Grading of Recommendations Assessment, Development, and Evaluation (GRADE) approach.

Below is a summary of the new evidence for prophylaxis in children. For a review of the paediatric literature up to 2014, please see the previous Australian guidelines.⁵ For a comprehensive review of all published randomised control trials investigating antifungal prophylaxis in adult and paediatric patients, please refer to the recently published paediatric antifungal guidelines.¹¹

Mould-active azoles

In a systematic review and meta-analysis, predominantly adult pooled data found a significant reduction in proven or probable IFD, mould infection or invasive aspergillosis in patients receiving any mould-active azole compared to fluconazole. Compared to echinocandins, there was no difference in proven or probable infections or mortality, although there were more adverse effects in the group receiving mould-active azole prophylaxis. TDM is recommended for children receiving mould-active azoles. Please refer to the accompanying optimising antifungal therapy and TDM guidelines by Chau *et al.* 2021, which can be found elsewhere in this supplement.

Posaconazole

Since 2014, observational studies of children with cancer or following HSCT have reported a breakthrough proven/probable IFD rate between 0–4.3% with posaconazole. Many of these breakthrough IFDs occurred in the setting of subtherapeutic levels and all studies included children less than 12 years of age where data are specifically lacking.

In Australia, posaconazole suspension and modified-release tablets are approved for use in children \geq 13 years of age. There are emerging pharmacokinetic-population studies to guide dosing of both formulations in younger children (<13 years) (Table 7).^{100, 120} Poor attainment of target levels using the suspension are reported^{115, 119, 121-123} and tablets are preferred when this is possible (see discussion on new evidence for established agents in children).^{100, 121} Where suspension is necessary, higher doses, increasing dose fractionation (e.g. four times daily [QID] dosing rather than three times daily [TDS]) and co-administration with a fatty meal are recommended.¹²⁴

Voriconazole

Since 2014, observational studies of children with cancer or following HSCT have reported a breakthrough proven/probable IFD rate between 0–12% with voriconazole. ¹²⁵⁻¹²⁸ In the study reporting a 12% breakthrough rate, patients received voriconazole doses of between 5–10 mg/kg/day, ¹²⁸ which is below the currently recommended dose (Table 7). ¹²⁹

Itraconazole

Since 2014, observational studies of children with haematological malignancy or following HSCT have reported a breakthrough proven/probable IFD rate between 0–3.2%with itraconazole.^{116, 117, 130} No studies included patients receiving SUBA-itraconazole.

Amphotericin B

The aforementioned clinical practice guidelines for antifungal prophylaxis in children with cancer or HSCT includes a recommendation against the routine use of amphotericin B for prophylaxis based on a meta-analysis of randomised controlled trial data showing amphotericin B was not more effective than fluconazole in preventing IFD (risk ratio 0.99; 95% CI: 0.52–1.88) but was associated with more adverse effects (risk ratio 5.63; 95% CI: 1.17–27.02). However, while this recommendation includes both conventional and lipid formulations, only one randomised controlled trial investigated the lipid formulation and none assessed liposomal amphotericin B (L-AMB) specifically.

Liposomal amphotericin B

Despite the limited supporting data, intermittent L-AMB continues to be prescribed for prophylaxis, particularly where triazole prophylaxis is not tolerated or contraindicated and daily IV echinocandins are not feasible. Since 2014, observational studies report a breakthrough proven/probable IFD rate with L-AMB of between 0–8% and mild renal toxicity of between 8.8–22%. Optimal dosing for prophylaxis remains unknown, and varied doses, including 1 mg/kg thrice weekly, 2.5 mg/kg twice weekly and 3–5 mg/kg thrice weekly, have been used. 134-136

Echinocandins

Echinocandins are considered a suitable option, alongside mould-active triazoles, for primary prophylaxis with similar efficacy in preventing proven/probable IFD.¹¹

Caspofungin

A recent randomised controlled trial assessing the efficacy of echinocandin prophylaxis in 517 children and young adults with AML found a significant reduction in proven/probable

IFD with caspofungin compared with fluconazole (3.1% vs. 7.2%; P = 0.03). Both agents were well tolerated.

Micafungin

Since 2014, observational studies of micafungin prophylaxis (predominantly using 1 mg/kg/day) in children with cancer or undergoing HSCT, have reported breakthrough IFD rates of 1.5% and 12.8%. ^{125, 138-142} Serious drug-related adverse events were rare (0–3%). ¹³⁸⁻¹⁴² Higher micafungin doses (2 mg/kg and 3 mg/kg daily) ¹⁴³⁻¹⁴⁵ and intermittent high-dose micafungin (up to 5 mg/kg weekly) ^{146, 147} have been assessed in observational paediatric studies, yet comparative data to support widespread use of these strategies are still lacking. ¹⁴⁸

Evidence for newer antifungal prophylactic agents and formulations in children

Please refer to earlier discussion on new options for antifungal prophylaxis in adults, including isavuconazole, tablet and IV formulations of posaconazole and SUBA-itraconazole. Specific paediatric data follows.

Isavuconazole in children

Published data on paediatric isavuconazole dosing, efficacy and safety is limited to small case series predominantly reporting on its use in the treatment of IFD rather than prophylaxis. ¹⁴⁹⁻¹⁵² In the largest series to date, 29 children (median age 14.5 years) received isavuconazole 200 mg/day (>30 kg) or 100 mg/day (<30 kg) following initial loading (TDS dosing for 48 hours); no IFD occurred amongst five patients receiving isavuconazole for IFD prophylaxis. ¹⁴⁹ Weight-based isavuconazole dosing for children aged 2–17 years of 10 mg/kg (maximum 372 mg) TDS for 48 hours and once daily thereafter has been proposed. ¹⁴⁹

New formulation of itraconazole in children

There are also limited data on SUBA-itraconazole dosing, efficacy and safety in children. A prophylactic dose of 2.5 mg/kg/day has been proposed based on a small paediatric cohort study.¹⁵³

Posaconazole: tablet and intravenous formulation in children

In keeping with the adult literature, the administration of posaconazole modified-release tablets is associated with superior attainment of therapeutic levels in children compared to oral suspension.^{100, 154-157} Attainment of target level (>700 ng/ml) in 94% (32/34) of children receiving modified-release tablets using weight-banded dosing has been reported (Table 7).¹⁵⁷

In two small studies of IV posaconazole for IFD prophylaxis, attainment of target levels was reported in 95–100% of children using doses of 6–7 mg/kg/day.^{158, 159} A new posaconazole powder formulation for suspension (PFS) was also assessed in children in one study, with attainment of target levels in 89–94% of patients and no serious drug related adverse effects.¹⁵⁸ Although not yet widely available, the PFS could replace liquid posaconazole for children unable to swallow tablets.

Impact of antifungal prophylaxis on diagnostic testing in children

While adult studies have suggested a reduced utility of GM for IFD screening in patients on mould-active prophylaxis, 90, 160, 161 paediatric specific data are lacking. Paediatric guidelines suggest the GM assay, particularly on bronchoscopy specimens, remains useful in children on mould-active prophylaxis with clinically-suspected IFD. 162, 163

Significant drug-drug interactions and TDM in children

Vincristine and azole therapy remain problematic in children undergoing ALL treatment.

Concomitant vincristine use with itraconazole or voriconazole is more problematic than with fluconazole. With increasing use of posaconazole in the paediatric population, there are also reports of associated severe neuropathic pain, has myalgia and autonomic neuropathy in patients receiving concurrent vinka-alkaloids.

Awareness of potential interactions with novel agents in paediatric leukaemia treatment is required when prescribing mould-active triazole prophylaxis. Inotuzomab and gemtuzumab are included in open studies for ALL and AML treatment respectively. ¹⁶⁷⁻¹⁶⁹ Monitoring of QT interval is recommended in the setting of concomitant triazole prophylaxis. Venetoclax is increasingly used in children with relapsed or refractory leukaemia ^{170, 171} and dose reduction by up to 75% is recommended with concomitant triazole use. ³

Rates of antifungal resistance and choice of prophylaxis in children

For detailed discussion, please refer to *Question 6*.

Question 8 How could antifungal prophylaxis recommendations be implemented into practice?

The adult and paediatric recommendations for antifungal prophylaxis outlined in these guidelines take into account the latest available evidence. While agents are licensed for use by the Therapeutic Goods Administration (TGA), not all agents are reimbursable for use as prophylaxis for all at-risk groups under the PBS, which will impact availability and their use, particularly in the paediatric population. For successful implementation, these guidelines will need to be adapted for use at a local institutional level. Early consideration of potential needs and barriers will facilitate local adaptation and adoption. Factors for consideration include haematology patient population (leukaemia, HSCT), use of targeted agents, haematology clinical trials, local epidemiology (i.e. incidence of IFD, pattern of fungal infection, rates of fungal resistance), and pharmacy cost and budgets. Expertise in infectious diseases, microbiology and pharmacology are required to implement these guidelines to optimise the use of prophylaxis and diagnosis of breakthrough infection.

Question 9 Are there new approaches to defining risk for IFD?

Antifungal prophylaxis is effective in reducing incidence of IFD but its use should be optimised and targeted to derive the most benefit. Accurate assessment of risk for IFD remains an ongoing challenge with new, emerging haematological treatments that are not associated with known risk factors such as prolonged neutropenia.¹⁴

Genetic polymorphisms of pattern recognition receptors or soluble acute phase reactants, critical components of mounting an innate response against IFD, have been associated with risk for invasive fungal infection.¹⁷² In particular, single nucleotide polymorphisms (SNP) of genes that code for reduced expression or production of Toll-like receptor 4 (TLR4), Dectin-1, Pentraxin-3 and mannose-binding lectin (MBL), are significantly associated with increased risk for invasive aspergillosis in HSCT and patients with haematological malignancies (odds ratio between 2.8–7.3).^{172, 173} To date, testing for known SNPs has not been translated into clinical practice to help identify higher-risk patients that may benefit from antifungal prophylaxis.

In the last several years there have been further advances in platforms for the use of genetic and functional immune profiling. By integrating genome-wide analysis and systems-level immune profiling (RNA sequencing and cytokine analysis), several genes such as the SERPINA1 and MAP3K8 genes, have been identified as potential markers for increased susceptibility to candidaemia.¹⁷⁴ Cytokine analysis has revealed that higher baseline interlukin-2 receptor (IL-2R) and monocyte chemoattractant protein-1 (MCP-1/CCL2) levels have been associated with higher rates of invasive aspergillosis.¹⁷⁵

Due to increased understanding of intestinal microbiota, a relationship between stem cell transplant-related dysbiosis, expansion of pathogenic *Candida* species, and translocation and subsequent invasive bloodstream infection, has been established.¹⁷⁶ Progress has been made in identifying genetic markers, immune profiles and mycobiomes associated with increased risk for IFD. However, the use of testing for known SNPs associated with increased risk has not been evaluated for its impact in large prospective cohorts. Validation of potential newly identified biomarkers has yet to occur. While not yet ready for use in clinical practice, genetic and immune profiling offers great potential for guiding the optimal use of antifungal prophylaxis in the near future.

Conclusion

While there have been limited changes to recommendations for antifungal prophylaxis in adults since the last published guidelines, there has been a shift in the recognised risk groups and type of prophylaxis (mould versus non-mould) for children. New formulations have addressed some of the limitations of established agents, while the utility of newer antifungal agents for prophylaxis in high-risk groups remains unclear. New antifungal agents introduced for therapy such as rezafungin, are undergoing evaluation for use as prophylaxis.¹⁷⁷

Evidence gaps remain around the need for antifungal prophylaxis in the setting of new haematological treatments such as BTK inhibitors and CAR T cell therapy, the choice of antifungal prophylaxis for ALL in adults and children, dosing of the newer formulations in children, and the role of long-acting antifungal agents. These are active issues for future research.

The rapid advances in immune-based haematological therapies with non-classical impact on immunity and risk for IFD highlights the need for new approaches for IFD risk assessment and early, active systems-based detection of new at-risk groups. Emergence of drug-resistant fungal pathogens requires ongoing active surveillance and knowledge of local IFD epidemiology in order to help tailor recommendations for choice of prophylaxis. While challenges remain, novel antifungal therapies in development offer new options and will change the prophylaxis landscape in the near future.¹⁷⁸

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References

- Reinwald M, Silva JT, Mueller NJ, Fortun J, Garzoni C, de Fijter JW *et al.* ESCMID Study Group for Infections in Compromised Hosts (ESGICH) consensus document on the safety of targeted and biological therapies: an infectious diseases perspective (intracellular signaling pathways: tyrosine kinase and mTOR inhibitors). *Clin Microbiol Infect* 2018; **24** (Suppl 2): S53–S70.
- Teh BW, Tam CS, Handunnetti S, Worth LJ, Slavin MA. Infections in patients with chronic lymphocytic leukaemia: mitigating risk in the era of targeted therapies. *Blood Rev* 2018; **32**: 499–507.
- Lindsay J, Teh BW, Micklethwaite K, Slavin M. Azole antifungals and new targeted therapies for hematological malignancy. *Curr Opin Infect Dis* 2019; **32**: 538–545.
- Hill JA, Li D, Hay KA, Green ML, Cherian S, Chen X *et al.* Infectious complications of CD19-targeted chimeric antigen receptor-modified T-cell immunotherapy. *Blood* 2018;
 131: 121–130.
- Fleming S, Yannakou CK, Haeusler GM, Clark J, Grigg A, Heath CH *et al.* Consensus guidelines for antifungal prophylaxis in haematological malignancy and haemopoietic stem cell transplantation, 2014. *Intern Med J* 2014; **44**: 1283–1297.
- 6 Maertens JA, Raad, II, Marr KA, Patterson TF, Kontoyiannis DP, Cornely OA *et al.*Isavuconazole versus voriconazole for primary treatment of invasive mould disease

caused by *Aspergillus* and other filamentous fungi (SECURE): a phase 3, randomised-controlled, non-inferiority trial. *Lancet* 2016; **387**: 760–769.

- Cornely OA, Duarte RF, Haider S, Chandrasekar P, Helfgott D, Jimenez JL *et al.* Phase 3 pharmacokinetics and safety study of a posaconazole tablet formulation in patients at risk for invasive fungal disease. *J Antimicrob Chemother* 2016; **71**: 718–726.
- Beardsley J, Halliday CL, Chen SC, Sorrell TC. Responding to the emergence of antifungal drug resistance: perspectives from the bench and the bedside. *Future Microbiol* 2018; **13**: 1175–1191.
- 9 Ong CW, Chen SC, Clark JE, Halliday CL, Kidd SE, Marriott DJ *et al.* Diagnosis, management and prevention of *Candida auris* in hospitals: position statement of the Australasian Society for Infectious Diseases. *Intern Med J* 2019; **49**: 1229–1243.
- 10 Chapman B, Slavin M, Marriott D, Halliday C, Kidd S, Arthur I *et al.* Changing epidemiology of candidaemia in Australia. *J Antimicrob Chemother* 2017; **72**: 1103–1108.
- Lehrnbecher T, Fisher BT, Phillips B, Beauchemin M, Carlesse F, Castagnola E *et al.*Clinical practice guideline for systemic antifungal prophylaxis in pediatric patients with cancer and hematopoietic stem-cell transplantation recipients. *J Clin Oncol* 2020; **38**: 3205–3216.
- 12 Chamilos G, Lionakis MS, Kontoyiannis DP. Call for action: invasive fungal infections associated with ibrutinib and other small molecule kinase inhibitors targeting immune signaling pathways. *Clin Infect Dis* 2018; **66**: 140–148.

- Varughese T, Taur Y, Cohen N, Palomba ML, Seo SK, Hohl TM *et al.* Serious infections in patients receiving ibrutinib for treatment of lymphoid cancer. *Clin Infect Dis* 2018; **67**: 687–692.
- Teh BW, Chui W, Handunnetti S, Tam C, Worth LJ, Thursky KA *et al.* High rates of proven invasive fungal disease with the use of ibrutinib monotherapy for relapsed or refractory chronic lymphocytic leukemia. *Leuk Lymphoma* 2019; **60**: 1572–1575.
- Byrd JC, Brown JR, O'Brien S, Barrientos JC, Kay NE, Reddy NM *et al.* Ibrutinib versus ofatumumab in previously treated chronic lymphoid leukemia. *N Engl J Med* 2014; **371**: 213–223.
- Ghez D, Calleja A, Protin C, Baron M, Ledoux MP, Damaj G *et al.* Early-onset invasive aspergillosis and other fungal infections in patients treated with ibrutinib. *Blood* 2018; **131**: 1955–1959.
- Lionakis MS, Dunleavy K, Roschewski M, Widemann BC, Butman JA, Schmitz R *et al.*Inhibition of B cell receptor signaling by ibrutinib in primary CNS lymphoma. *Cancer Cell* 2017; **31**: 833–43 e5.
- Shanavas M, Popat U, Michaelis LC, Fauble V, McLornan D, Klisovic R *et al.* Outcomes of allogeneic hematopoietic cell transplantation in patients with myelofibrosis with prior exposure to janus kinase 1/2 inhibitors. *Biol Blood Marrow Transplant* 2016; **22**: 432–440.

- Mori Y, Ikeda K, Inomata T, Yoshimoto G, Fujii N, Ago H *et al.* Ruxolitinib treatment for GvHD in patients with myelofibrosis. *Bone Marrow Transplant* 2016; **51**: 1584–1587.
- Brown JR, Byrd JC, Coutre SE, Benson DM, Flinn IW, Wagner-Johnston ND *et al.*Idelalisib, an inhibitor of phosphatidylinositol 3-kinase p110delta, for relapsed/refractory chronic lymphocytic leukemia. *Blood* 2014; **123**: 3390–3397.
- Davids MS, Hallek M, Wierda W, Roberts AW, Stilgenbauer S, Jones JA *et al.*Comprehensive safety analysis of venetoclax monotherapy for patients with relapsed/refractory chronic lymphocytic leukemia. *Clin Cancer Res* 2018; **24**: 4371–4379.
- 22 Motzer RJ, Escudier B, Oudard S, Hutson TE, Porta C, Bracarda S *et al.* Phase 3 trial of everolimus for metastatic renal cell carcinoma: final results and analysis of prognostic factors. *Cancer* 2010; **116**: 4256–4265.
- Falantes JF, Calderon C, Marquez-Malaver FJ, Aguilar-Guisado M, Martin-Pena A, Martino ML *et al.* Patterns of infection in patients with myelodysplastic syndromes and acute myeloid leukemia receiving azacitidine as salvage therapy. Implications for primary antifungal prophylaxis. *Clin Lymphoma Myeloma Leuk* 2014; **14**: 80–86.
- Pomares H, Arnan M, Sanchez-Ortega I, Sureda A, Duarte RF. Invasive fungal infections in AML/MDS patients treated with azacitidine: a risk worth considering antifungal prophylaxis? *Mycoses* 2016; **59**: 516–519.

- Aldoss I, Dadwal S, Zhang J, Tegtmeier B, Mei M, Arslan S *et al.* Invasive fungal infections in acute myeloid leukemia treated with venetoclax and hypomethylating agents. *Blood Adv* 2019; **3**: 4043–4049.
- Perl AE, Altman JK, Cortes J, Smith C, Litzow M, Baer MR *et al.* Selective inhibition of FLT3 by gilteritinib in relapsed or refractory acute myeloid leukaemia: a multicentre, first-in-human, open-label, phase 1-2 study. *Lancet Oncol* 2017; **18**: 1061–1075.
- 27 Stone RM, Mandrekar SJ, Sanford BL, Laumann K, Geyer S, Bloomfield CD *et al.*Midostaurin plus chemotherapy for acute myeloid leukemia with a FLT3 mutation. *N Engl J Med* 2017; **377**: 454–464.
- Teh BW, Teng JC, Urbancic K, Grigg A, Harrison SJ, Worth LJ *et al.* Invasive fungal infections in patients with multiple myeloma: a multi-center study in the era of novel myeloma therapies. *Haematologica* 2015; **100**: e28–31.
- Liu J, Huang H, Li Y, Liu L, Li J, Liu Z *et al.* Epidemiology and treatment of invasive fungal diseases in patients with multiple myeloma: findings from a multicenter prospective study from China. *Tumour Biol* 2016; **37**: 7893–7900.
- Lim C, Sinha P, Harrison SJ, Quach H, Slavin MA, Teh BW. Low rates of invasive fungal disease in patients with multiple myeloma managed with new generation therapies: results from a multi-centre cohort study. *Mycoses* 2020: doi: 10.1111/myc.13178. Epub ahead of print. PMID: 32885525.

- Maude SL, Laetsch TW, Buechner J, Rives S, Boyer M, Bittencourt H *et al.*Tisagenlecleucel in children and young adults with B-cell lymphoblastic leukemia. *N Engl J Med* 2018; **378**: 439–448.
- 32 Schuster SJ, Bishop MR, Tam CS, Waller EK, Borchmann P, McGuirk JP *et al.*Tisagenlecleucel in adult relapsed or refractory diffuse large B-cell lymphoma. *N Engl J Med* 2019; **380**: 45–56.
- Nastoupil LJ, Jain MD, Spiegel JY, Ghobadi A, Lin Y, Dahiya S *et al.* Axicabtagene ciloleucel (Axi-cel) CD19 Chimeric Antigen Receptor (CAR) T-Cell therapy for relapsed/refractory large B-cell lymphoma: real world experience. *Blood* 2018; **132** (Suppl 1): 91.
- 34 Kantarjian H, Stein A, Gokbuget N, Fielding AK, Schuh AC, Ribera JM *et al.*Blinatumomab versus chemotherapy for advanced acute lymphoblastic leukemia. *N Engl J Med* 2017; **376**: 836–847.
- Gokbuget N, Zugmaier G, Klinger M, Kufer P, Stelljes M, Viardot A *et al.* Long-term relapse-free survival in a phase 2 study of blinatumomab for the treatment of patients with minimal residual disease in B-lineage acute lymphoblastic leukemia. *Haematologica* 2017; **102**: e132–e5.
- Neelapu SS, Locke FL, Bartlett NL, Lekakis LJ, Miklos DB, Jacobson CA *et al.*Axicabtagene ciloleucel CAR T-Cell therapy in refractory large B-Cell lymphoma. *N Engl J Med* 2017; **377**: 2531–2544.

- Park JH, Romero FA, Taur Y, Sadelain M, Brentjens RJ, Hohl TM *et al.* Cytokine release syndrome grade as a predictive marker for nfections in patients with relapsed or refractory B-cell acute lymphoblastic leukemia treated with chimeric antigen receptor T cells. *Clin Infect Dis* 2018; **67**: 533–540.
- Haidar G, Dorritie K, Farah R, Bogdanovich T, Nguyen MH, Samanta P. Invasive mold infections after chimeric antigen receptor-modified T-cell therapy: a case series, review of the literature, and implications for prophylaxis. *Clin Infect Dis* 2019.
- Wudhikarn K, Palomba ML, Pennisi M, Garcia-Recio M, Flynn JR, Devlin SM *et al.*Infection during the first year in patients treated with CD19 CAR T cells for diffuse large B cell lymphoma. *Blood Cancer J* 2020; **10**: 79.
- 40 So W, Pandya S, Quilitz R, Shah B, Greene JN. Infectious risks and complications in adult leukemic patients receiving blinatumomab. *Mediterr J Hematol Infect Dis* 2018; **10**: e2018029.
- 2hao YJ, Khoo AL, Tan G, Teng M, Tee C, Tan BH *et al.* Network meta-analysis and pharmacoeconomic evaluation of fluconazole, itraconazole, posaconazole, and voriconazole in invasive fungal infection prophylaxis. *Antimicrob Agents Chemother* 2016; **60**: 376–386.
- 42 Bow EJ, Vanness DJ, Slavin M, Cordonnier C, Cornely OA, Marks DI *et al.* Systematic review and mixed treatment comparison meta-analysis of randomized clinical trials of primary oral antifungal prophylaxis in allogeneic hematopoietic cell transplant recipients. *BMC Infect Dis* 2015; **15**: 128.

- Lee CH, Lin C, Ho CL, Lin JC. Primary fungal prophylaxis in hematological malignancy:

 A network meta-analysis of randomized controlled trials. *Antimicrob Agents Chemother*2018; **62**: e00355–18.
- Epstein DJ, Seo SK, Huang YT, Park JH, Klimek VM, Berman E *et al.* Micafungin versus posaconazole prophylaxis in acute leukemia or myelodysplastic syndrome: a randomized study. *J Infect* 2018; **77**: 227–234.
- Pagano L, Caira M, Candoni A, Aversa F, Castagnola C, Caramatti C *et al.* Evaluation of the practice of antifungal prophylaxis use in patients with newly diagnosed acute myeloid leukemia: results from the SEIFEM 2010-B registry. *Clin Infect Dis* 2012; **55**: 1515–1521.
- Hachem R, Assaf A, Numan Y, Shah P, Jiang Y, Chaftari AM *et al.* Comparing the safety and efficacy of voriconazole versus posaconazole in the prevention of invasive fungal infections in high-risk patients with hematological malignancies. *Int J Antimicrob Agents* 2017; **50**: 384–388.
- Park H, Youk J, Shin DY, Hong J, Kim I, Kim NJ *et al.* Micafungin prophylaxis for acute leukemia patients undergoing induction chemotherapy. *BMC Cancer* 2019; **19**: 358.
- Tang L, Yang XF, Qiao M, Zhang L, Tang XW, Qiu HY *et al.* Posaconazole vs. voriconazole in the prevention of invasive fungal diseases in patients with haematological malignancies: a retrospective study. *J Mycol Med* 2018; **28**: 379–383.
- 49 Vehreschild MJ, von Bergwelt-Baildon M, Tran L, Shimabukuro-Vornhagen A, Wisplinghoff H, Bangard C *et al.* Feasibility and effectiveness of posaconazole

prophylaxis in combination with micafungin bridging for patients undergoing allogeneic stem cell transplantation: a 6-yr analysis from the cologne cohort for neutropenic patients. *Eur J Haematol* 2014; **93**: 400–406.

- Wang CH, Kan LP, Lin HA, Chang FY, Wang NC, Lin TY *et al.* Clinical efficacy and safety of primary antifungal prophylaxis with posaconazole versus fluconazole in allogeneic blood hematopoietic stem cell transplantation recipients: a retrospective analysis of a single medical center in Taiwan. *J Microbiol Immunol Infect* 2016; **49**: 531–538.
- Pagano L, Verga L, Busca A, Martino B, Mitra ME, Fanci R *et al.* Systemic antifungal treatment after posaconazole prophylaxis: results from the SEIFEM 2010-C survey. *J Antimicrob Chemother* 2014; **69**: 3142–3147.
- Calmettes C, Gabriel F, Blanchard E, Servant V, Bouchet S, Kabore N *et al.*Breakthrough invasive aspergillosis and diagnostic accuracy of serum galactomannan enzyme immune assay during acute myeloid leukemia induction chemotherapy with posaconazole prophylaxis. *Oncotarget* 2018; **9**: 26724–26736.
- Rodriguez-Veiga R, Montesinos P, Boluda B, Lorenzo I, Martinez-Cuadron D, Salavert M *et al.* Incidence and outcome of invasive fungal disease after front-line intensive chemotherapy in patients with acute myeloid leukemia: impact of antifungal prophylaxis. *Ann Hematol* 2019; **98**: 2081–2088.
- Bui V, Walker SA, Elligsen M, Vyas A, Kiss A, Palmay L. Voriconazole prophylaxis in leukemic patients: a retrospective single-center study. *J Oncol Pharm Pract* 2020; **26**: 873–881.

- Hicks JK, Quilitz RE, Komrokji RS, Kubal TE, Lancet JE, Pasikhova Y *et al.* Prospective CYP2C19-guided voriconazole prophylaxis in patients with neutropenic acute myeloid leukemia reduces the incidence of subtherapeutic antifungal plasma concentrations. *Clin Pharmacol Ther* 2020; **107**: 563–570.
- Patel JN, Hamadeh IS, Robinson M, Shahid Z, Symanowski J, Steuerwald N *et al.*Evaluation of CYP2C19 genotype-guided voriconazole prophylaxis after allogeneic hematopoietic cell transplant. *Clin Pharmacol Ther* 2020; **107**: 571–579.
- Huang X, Chen H, Han M, Zou P, Wu D, Lai Y *et al.* Multicenter, randomized, open-label study comparing the efficacy and safety of micafungin versus itraconazole for prophylaxis of invasive fungal infections in patients undergoing hematopoietic stem cell transplant. *Biol Blood Marrow Transplant* 2012; **18**: 1509–1516.
- Lin R, Xu X, Li Y, Sun J, Fan Z, Jiang Q *et al.* Comparison of long-term and short-term administration of itraconazole for primary antifungal prophylaxis in recipients of allogeneic hematopoietic stem cell transplantation: a multicenter, randomized, openlabel trial. *Transpl Infect Dis* 2014; **16**: 286–294.
- Park S, Kim K, Jang JH, Kim SJ, Kim WS, Chung DR *et al.* Randomized trial of micafungin versus fluconazole as prophylaxis against invasive fungal infections in hematopoietic stem cell transplant recipients. *J Infect* 2016; **73**: 496–505.
- Rosillo C, Avila AM, Huang YT, Devlin S, Cho C, Montoro J *et al.* Sequential systematic anti-mold prophylaxis with micafungin and voriconazole results in very low incidence of invasive mold infections in patients undergoing allogeneic hematopoietic stem cell transplantation. *Transpl Infect Dis* 2018; **20**: e12897.

- Rothe A, Classen A, Carney J, Hallek M, Mellinghoff SC, Scheid C *et al.* Bridging antifungal prophylaxis with 50 mg or 100 mg micafungin in allogeneic stem cell transplantation: a retrospective analysis. *Eur J Haematol* 2019.
- Langebrake C, Rohde H, Lellek H, Wolschke C, Kroger NM. Micafungin as antifungal prophylaxis in recipients of allogeneic hematopoietic stem cell transplantation: results of different dosage levels in clinical practice. *Clin Transplant* 2014; **28**: 286–291.
- Cornely OA, Leguay T, Maertens J, Vehreschild M, Anagnostopoulos A, Castagnola C *et al.* Randomized comparison of liposomal amphotericin B versus placebo to prevent invasive mycoses in acute lymphoblastic leukaemia. *J Antimicrob Chemother* 2017; **72**: 2359–2367.
- Doan TN, Kirkpatrick CM, Walker P, Slavin MA, Ananda-Rajah MR, Morrissey CO *et al.*Primary antifungal prophylaxis in adult patients with acute lymphoblastic leukaemia: a multicentre audit. *J Antimicrob Chemother* 2016; **71**: 497–505.
- Marty FM, Ostrosky-Zeichner L, Cornely OA, Mullane KM, Perfect JR, Thompson GR, 3rd *et al.* Isavuconazole treatment for mucormycosis: a single-arm open-label trial and case-control analysis. *Lancet Infect Dis* 2016; **16**: 828–837.
- Cornely OA, Bohme A, Schmitt-Hoffmann A, Ullmann AJ. Safety and pharmacokinetics of isavuconazole as antifungal prophylaxis in acute myeloid leukemia patients with neutropenia: results of a phase 2, dose escalation study. *Antimicrob Agents Chemother* 2015; **59**: 2078–2085.

- Fung M, Schwartz BS, Doernberg SB, Langelier C, Lo M, Graff L *et al.* Breakthrough invasive fungal infections on isavuconazole prophylaxis and treatment: what is happening in the real-world setting? *Clin Infect Dis* 2018; **67**: 1142–1143.
- Fontana L, Perlin DS, Zhao Y, Noble BN, Lewis JS, Strasfeld L *et al.* Isavuconazole prophylaxis in patients with hematologic malignancies and hematopoietic-cell transplant recipients. *Clin Infect Dis* 2020; **70**: 723–730.
- Rausch CR, DiPippo AJ, Bose P, Kontoyiannis DP. Breakthrough fungal infections in patients with leukemia receiving isavuconazole. *Clin Infect Dis* 2018; **67**: 1610–1613.
- Bowen CD, Tallman GB, Hakki M, Lewis JS. Isavuconazole to prevent invasive fungal infection in immunocompromised adults: initial experience at an academic medical centre. *Mycoses* 2019; **62**: 665–672.
- Bose P, McCue D, Wurster S, Wiederhold NP, Konopleva M, Kadia TM *et al.*Isavuconazole as primary anti-fungal prophylaxis in patients with acute myeloid leukemia or myelodysplastic syndrome: an open-label, prospective, phase II study. *Clin Infect Dis* 2020.
- Stern A, Su Y, Lee YJ, Seo S, Shaffer B, Tamari R *et al.* A single-center, open-label trial of isavuconazole prophylaxis against invasive fungal infection in patients undergoing allogeneic hematopoietic cell transplantation. *Biol Blood Marrow Transplant* 2020; **26**: 1195–1202.
- 73 Mayne Pharma. LOZANOC (itraconazole capsules) US prescribing information, 2014.

- 74 Mayne Pharma. TOLSURA (itraconazole capsules) US prescribing information,

 December 2018.
- Abuhelwa AY, Foster DJ, Mudge S, Hayes D, Upton RN. Population pharmacokinetic modeling of itraconazole and hydroxyitraconazole for oral SUBA-itraconazole and sporanox capsule formulations in healthy subjects in fed and fasted states. *Antimicrob Agents Chemother* 2015; **59**: 5681–5696.
- Lindsay J, Sandaradura I, Wong K, Arthur C, Stevenson W, Kerridge I *et al.* Serum levels, safety and tolerability of new formulation SUBA-itraconazole prophylaxis in patients with haematological malignancy or undergoing allogeneic stem cell transplantation. *J Antimicrob Chemother* 2017; **72**: 3414–3419.
- Nield B, Larsen SR, van Hal SJ. Clinical experience with new formulation SUBA(R)itraconazole for prophylaxis in patients undergoing stem cell transplantation or
 treatment for haematological malignancies. *J Antimicrob Chemother* 2019; **74**: 3049–
 3055.
- Cornely OA, Maertens J, Winston DJ, Perfect J, Ullmann AJ, Walsh TJ *et al.*Posaconazole vs. fluconazole or itraconazole prophylaxis in patients with neutropenia.

 N Engl J Med 2007; **356**: 348–359.
- 79 Ullmann AJ, Lipton JH, Vesole DH, Chandrasekar P, Langston A, Tarantolo SR *et al.*Posaconazole or fluconazole for prophylaxis in severe graft-versus-host disease. *N Engl J Med* 2007; **356**: 335–347.

- Krishna G, Ma L, Martinho M, O'Mara E. Single-dose phase I study to evaluate the pharmacokinetics of posaconazole in new tablet and capsule formulations relative to oral suspension. *Antimicrob Agents Chemother* 2012; **56**: 4196–4201.
- Kersemaekers WM, Dogterom P, Xu J, Marcantonio EE, de Greef R, Waskin H *et al.*Effect of a high-fat meal on the pharmacokinetics of 300-milligram posaconazole in a solid oral tablet formulation. *Antimicrob Agents Chemother* 2015; **59**: 3385–3389.
- Cojutti PG, Candoni A, Lazzarotto D, Rabassi N, Fanin R, Hope W *et al.* Coadministration of proton pump inhibitors and/or of steroids may be a risk factor for low trough concentrations of posaconazole delayed-released tablets in adult patients with haematological malignancies. *Br J Clin Pharmacol* 2018; **84**: 2544–2550.
- Boglione-Kerrien C, Picard S, Tron C, Nimubona S, Gangneux JP, Lalanne S *et al.* Safety study and therapeutic drug monitoring of the oral tablet formulation of posaconazole in patients with haematological malignancies. *J Cancer Res Clin Oncol* 2018; **144**: 127–134.
- Tverdek FP, Heo ST, Aitken SL, Granwehr B, Kontoyiannis DP. Real-life assessment of the safety and effectiveness of the new tablet and intravenous formulations of posaconazole in the prophylaxis of invasive fungal infections via analysis of 343 courses. *Antimicrob Agents Chemother* 2017; **61**: e00188–17.
- Chin A, Pergam SA, Fredricks DN, Hoofnagle AN, Baker KK, Jain R. Evaluation of posaconazole serum concentrations from delayed-release tablets in patients at high risk for fungal infections. *Antimicrob Agents Chemother* 2017; **61**: e00569–17.

- Belling M, Kanate AS, Shillingburg A, Lu X, Wen S, Shah N *et al.* Evaluation of serum posaconazole concentrations in patients with hematological malignancies receiving posaconazole suspension compared to the delayed-release tablet formulation. *Leuk Res Treatment* 2017; **2017**: 3460892.
- 87 Cornely OA, Robertson MN, Haider S, Grigg A, Geddes M, Aoun M *et al.*Pharmacokinetics and safety results from the Phase 3 randomized, open-label, study of intravenous posaconazole in patients at risk of invasive fungal disease. *J Antimicrob Chemother* 2017; **72**: 3406–3413.
- Jeong W, Haywood P, Shanmuganathan N, Lindsay J, Urbancic K, Ananda-Rajah MR *et al.* Safety, clinical effectiveness and trough plasma concentrations of intravenous posaconazole in patients with haematological malignancies and/or undergoing allogeneic haematopoietic stem cell transplantation: off-trial experience. *J Antimicrob Chemother* 2016; **71**: 3540–3547.
- Barnes RA, White PL, Morton CO, Rogers TR, Cruciani M, Loeffler J *et al.* Diagnosis of aspergillosis by PCR: clinical considerations and technical tips. *Med Mycol* 2018; **56**: 60–72.
- Duarte RF, Sanchez-Ortega I, Cuesta I, Arnan M, Patino B, Fernandez de Sevilla A *et al.* Serum galactomannan-based early detection of invasive aspergillosis in hematology patients receiving effective antimold prophylaxis. *Clin Infect Dis* 2014; **59**: 1696–1702.
- Duarte RF, Sanchez-Ortega I, Arnan M, Patino B, Ayats J, Sureda A *et al.* Serum galactomannan surveillance may be safely withdrawn from antifungal management of

hematology patients on effective antimold prophylaxis: a pilot single-center study. Bone Marrow Transplant 2017; **52**: 326–329.

- 92 Morrissey CO, Chen SC, Sorrell TC, Milliken S, Bardy PG, Bradstock KF *et al.*Galactomannan and PCR versus culture and histology for directing use of antifungal treatment for invasive aspergillosis in high-risk haematology patients: a randomised controlled trial. *Lancet Infect Dis* 2013; **13**: 519–528.
- 93 AbbVie Inc. VENCLEXTA (venetoclax) US prescribing information, April 2016.
- Janssen Biotech Inc. IMBRUVICA (ibrutinib) Australian approved product information, February 2016.
- 95 Novartis Pharmaceuticals Corporation. RYDAPT (midostaurin) US prescribing information, April 2017.
- 96 Astellas Pharma US Inc. XOSPATA (gilteritinib) US prescribing information, November2018.
- 97 Muilwijk EW, Dekkers BGJ, Henriet SSV, Verweij PE, Witjes B, Lashof A *et al.*Flucloxacillin results in suboptimal plasma voriconazole concentrations. *Antimicrob Agents Chemother* 2017; **61**: e00915–17.
- Duong A, Sweet A, Jain R, Hill JA, Pergam SA, Boeckh M *et al.* Clinically significant drug interaction: letermovir and voriconazole. *J Antimicrob Chemother* 2020; **75**: 775–777.

- 99 Sime FB, Stuart J, Butler J, Starr T, Wallis SC, Pandey S *et al.* Pharmacokinetics of intravenous posaconazole in critically ill patients. *Antimicrob Agents Chemother* 2018; **62**: e00242–18.
- Boonsathorn S, Cheng I, Kloprogge F, Alonso C, Lee C, Doncheva B *et al.* Clinical pharmacokinetics and dose recommendations for posaconazole in infants and children. *Clinical Pharmacokinetics* 2019; **58**: 53–61.
- Tang LA, Marini BL, Benitez L, Nagel JL, Miceli M, Berglund C *et al.* Risk factors for subtherapeutic levels of posaconazole tablet. *J Antimicrob Chemother* 2017; **72**: 2902–2905.
- Boan P, Gardam D. Epidemiology and antifungal susceptibility patterns of candidemia from a tertiary centre in Western Australia. *J Chemother* 2019; **31**: 137–140.
- Trubiano JA, Leung VK, Worth LJ, Teh BW, Thursky KA, Slavin MA. *Candida glabrata* fungaemia at an Australian cancer centre: epidemiology, risk factors and therapy. *Leuk Lymphoma* 2015; **56**: 3442–3444.
- Worth LJ, Harrison SJ, Dickinson M, van Diemen A, Breen J, Harper S *et al. Candida auris* in an Australian health care facility: importance of screening high risk patients. *Med J Aust* 2020; **212**: 510–1.e1.
- Kidd SE, Goeman E, Meis JF, Slavin MA, Verweij PE. Multi-triazole-resistant *Aspergillus* fumigatus infections in Australia. *Mycoses* 2015; **58**: 350–355.

- Talbot JJ, Subedi S, Halliday CL, Hibbs DE, Lai F, Lopez-Ruiz FJ *et al.* Surveillance for azole resistance in clinical and environmental isolates of *Aspergillus fumigatus* in Australia and cyp51A homology modelling of azole-resistant isolates. *J Antimicrob Chemother* 2018; **73**: 2347–2351.
- Halliday CL, Chen SC, Kidd SE, van Hal S, Chapman B, Heath CH *et al.* Antifungal susceptibilities of non-*Aspergillus* filamentous fungi causing invasive infection in Australia: support for current antifungal guideline recommendations. *Int J Antimicrob Agents* 2016; **48**: 453–458.
- Paige E, Haywood P, Xie M, Worth L, Thursky K, Urbancic K *et al.* Auditing fungal disease in leukemia patients in a tertiary care center: opportunities and challenges for an antifungal stewardship program. *Leuk Lymphoma* 2019; **60**: 2373–2383.
- Corzo-Leon DE, Satlin MJ, Soave R, Shore TB, Schuetz AN, Jacobs SE *et al.*Epidemiology and outcomes of invasive fungal infections in allogeneic haematopoietic stem cell transplant recipients in the era of antifungal prophylaxis: a single-centre study with focus on emerging pathogens. *Mycoses* 2015; **58**: 325–336.
- Slavin M, van Hal S, Sorrell TC, Lee A, Marriott DJ, Daveson K *et al.* Invasive infections due to filamentous fungi other than *Aspergillus*: epidemiology and determinants of mortality. *Clin Microbiol Infect* 2015; **21**: 490 e1–10.
- Fisher BT, Roninson PD, Lehrnbecher T, Steinback wJ, Zaoutis TE, Phillips B *et al.* Risk factors for invasive fungal disease in pediatric cancer and hematopoietic stem cell transplantation: A systematic review. *J Pediatr Infect Dis Society* 2018; **7**: 191–198.

- Wang SS, Kotecha RS, Bernard A, Blyth CC, McMullan BJ, Cann MP *et al.* Invasive fungal infections in children with acute lymphoblastic leukaemia: results from four Australian centres, 2003–2013. *Pediatr Blood Cancer* 2019; **66**: e27915.
- Bartlett AW, Cann MP, Yeoh DK, Bernard A, Ryan AL, Blyth CC *et al.* Epidemiology of invasive fungal infections in immunocompromised children: an Australian national 10-year review. *Pediatr Blood Cancer* 2019; **66**: e27564.
- Vora SB, Waghmare A, Englund JA, Qu P, Gardner RA, Hill JA. Infectious complications following CD19 Chimeric antigen receptor T-cell therapy for children, adolescents, and young adults. *Open Forum Infect Dis* 2020; **7**: ofaa121.
- 115 Lai T, Alffenaar JW, Kesson A, Bandodkar S, Roberts JA. Evaluation of target attainment of oral posaconazole suspension in immunocompromised children. *J Antimicrob Chemother* 2020; **75**: 726–729.
- Doring M, Blume O, Haufe S, Hartmann U, Kimmig A, Schwarze CP *et al.* Comparison of itraconazole, voriconazole, and posaconazole as oral antifungal prophylaxis in pediatric patients following allogeneic hematopoietic stem cell transplantation. *Eur J Clin Microbiol Infect Dis* 2014; **33**: 629–638.
- Doring M, Eikemeier M, Cabanillas Stanchi KM, Hartmann U, Ebinger M, Schwarze CP *et al.* Antifungal prophylaxis with posaconazole vs. fluconazole or itraconazole in pediatric patients with neutropenia. *Eur J Clin Microbiol Infect Dis* 2015; **34**: 1189–1200.

- Doring M, Cabanillas Stanchi KM, Klinker H, Eikemeier M, Feucht J, Blaeschke F *et al.*Posaconazole plasma concentrations in pediatric patients receiving antifungal prophylaxis during neutropenia. *Med Mycol* 2017; **55**: 375–384.
- 119 Vicenzi EB, Calore E, Decembrino N, Berger M, Perruccio K, Carraro F *et al.*Posaconazole oral dose and plasma levels in pediatric hematology-oncology patients.

 *Eur J Haematol 2018; **100**: 315–322.
- Boonsathorn S, Cheng I, Kloprogge F, Alonso C, Lee C, Doncheva B *et al.* Correction to: Clinical pharmacokinetics and dose recommendations for posaconazole in infants and children. *Clin Pharmacokinet* 2019; **58**: 141.
- Arrieta AC, Sung L, Bradley JS, Zwaan CM, Gates D, Waskin H *et al.* A non-randomized trial to assess the safety, tolerability, and pharmacokinetics of posaconazole oral suspension in immunocompromised children with neutropenia. *PLoS One* 2019; **14**: e0212837.
- Jancel T, Shaw PA, Hallahan CW, Kim T, Freeman AF, Holland SM *et al.* Therapeutic drug monitoring of posaconazole oral suspension in paediatric patients younger than 13 years of age: a retrospective analysis and literature review. *J Clin Pharm Ther* 2017; **42**: 75–79.
- Heinz WJ, Cabanillas Stanchi KM, Klinker H, Blume O, Feucht J, Hartmann U *et al.*Posaconazole plasma concentration in pediatric patients receiving antifungal prophylaxis after allogeneic hematopoietic stem cell transplantation. *Med Mycol* 2016; **54**: 128–137.

- Gwee A, Cranswick N, Curtis N. Posaconazole: promising but problematic in practice in pediatric patients. *Pediatr Infect Dis J* 2015; **34**: 604–606.
- Bui A, Nguyen V, Hsu C, Hyde B, Simms-Waldrip T. Invasive fungal infections while on voriconazole, liposomal amphotericin B, or micafungin for antifungal prophylaxis in pediatric stem cell transplant patients. *J Pediatr Pharmacol Ther* 2019; **24**: 220–226.
- Pana ZD, Kourti M, Vikelouda K, Vlahou A, Katzilakis N, Papageorgiou M *et al.*Voriconazole antifungal prophylaxis in children with malignancies: a nationwide study. *J Pediatr Hematol Oncol* 2018; **40**: 22–26.
- 127 Aftandilian C, Weinberg K, Willert J, Kharbanda S, Porteus M, Maldonado Y *et al.*Invasive fungal disease in pediatric patients undergoing allogeneic hematopoietic stem cell transplant. *J Pediatr Hematol Oncol* 2016; **38**: 574–580.
- Sano H, Kobayashi R, Hori D, Kishimoto K, Suzuki D, Yasuda K *et al.* Prophylactic administration of voriconazole with two different doses for invasive fungal infection in children and adolescents with acute myeloid leukemia. *J Microbiol Immunol Infect* 2018; **51**: 260–266.
- Lehrnbecher T. Antifungal prophylaxis in pediatric patients undergoing therapy for cancer: drugs and dosing. *Curr Opin Infect Dis* 2015; **28**: 523–531.
- Leong YH, Boast A, Cranswick N, Curtis N, Gwee A. Itraconazole dosing and drug monitoring at a tertiary children's hospital. *Pediatr Infect Dis J* 2019; **38**: 60–64.

- Ferreras-Antolin L, Sharland M, Warris A. Management of invasive fungal disease in neonates and children. *Pediatr Infect Dis J* 2019; **38**: S2–S6.
- Mendoza-Palomar N, Soques E, Benitez-Carabante MI, Gonzalez-Amores M, Fernandez-Polo A, Renedo B *et al.* Low-dose liposomal amphotericin B for antifungal prophylaxis in paediatric allogeneic haematopoietic stem cell transplantation. *J Antimicrob Chemother* 2020; **75**: 2264–2271.
- Hand EO, Ramanathan MR. Safety and tolerability of high-dose weekly liposomal amphotericin B antifungal prophylaxis. *Pediatr Infect Dis J* 2014; **33**: 835–836.
- Meryk A, Kropshofer G, Hutter J, Fritz J, Salvador C, Lass-Flörl C *et al.* Benefits of risk-adapted and mould-specific antifungal prophylaxis in childhood leukaemia. *Br J Haematol* 2020; **191**: 816–824.
- Warris A, Lehrnbecher T, Roilides E, Castagnola E, Brüggemann RJM, Groll AH. ESCMID-ECMM guideline: diagnosis and management of invasive aspergillosis in neonates and children. *Clin Microbiol Infect* 2019; **25**: 1096–1113.
- Groll AH, Castagnola E, Cesaro S, Dalle JH, Engelhard D, Hope W *et al.* Fourth European Conference on Infections in Leukaemia (ECIL-4): guidelines for diagnosis, prevention, and treatment of invasive fungal diseases in paediatric patients with cancer or allogeneic haemopoietic stem-cell transplantation. *Lancet Oncol* 2014; **15**: e327–40.
- 137 Fisher BT, Zaoutis T, Dvorak CC, Nieder M, Zerr D, Wingard JR *et al.* Effect of caspofungin vs fluconazole prophylaxis on invasive fungal disease among children and

young adults with acute myeloid leukemia: a randomized clinical trial. *Jama* 2019; **322**: 1673–1681.

- El Cheikh J, Ceballos P, Dalle JH, Ducastelle-Lepretre S, Dulon E, Herbrecht R. Micafungin prophylaxis in routine medical practice in adult and pediatric patients with hematological malignancy: a prospective, observational study in France. *Diagn Microbiol Infect Dis* 2019; **94**: 268–273.
- Leverger G, Timsit JF, Milpied N, Gachot B. Use of micafungin for the prevention and treatment of invasive fungal infections in everyday pediatric care in France: results of the MYRIADE study. *Pediatr Infect Dis J* 2019; **38**: 716–721.
- 140 Kobayashi C, Hanadate T, Niwa T, Hirano Y, Yoshiyasu T, So M *et al.* Safety and efficacy of micafungin for prophylaxis against invasive fungal infections in Japanese patients undergoing hematopoietic stem cell transplantation: results of a post-marketing surveillance study. *J Infect Chemother* 2015; **21**: 438–443.
- Park HJ, Park M, Han M, Nam BH, Koh KN, Im HJ *et al.* Efficacy and safety of micafungin for the prophylaxis of invasive fungal infection during neutropenia in children and adolescents undergoing allogeneic hematopoietic SCT. *Bone Marrow Transplant* 2014; **49**: 1212–1216.
- Maximova N, Schillani G, Simeone R, Maestro A, Zanon D. Comparison of efficacy and safety of caspofungin versus micafungin in pediatric allogeneic stem cell transplant recipients: a retrospective analysis. *Adv Ther* 2017; **34**: 1184–1199.

- Yoshikawa K, Nakazawa Y, Katsuyama Y, Hirabayashi K, Saito S, Shigemura T *et al.*Safety, tolerability, and feasibility of antifungal prophylaxis with micafungin at 2 mg/kg daily in pediatric patients undergoing allogeneic hematopoietic stem cell transplantation. *Infection* 2014; **42**: 639–647.
- 144 Kusuki S, Hashii Y, Yoshida H, Takizawa S, Sato E, Tokimasa S *et al.* Antifungal prophylaxis with micafungin in patients treated for childhood cancer. *Pediatr Blood Cancer* 2009; **53**: 605–609.
- Long S. Incidence of breakthrough invasive fungal infections while on micafungin for antifungal prophylaxis in pediatric hematopoietic cell transplant patients [abstract]. Biol Blood Marrow Transplant 2020; **26** (Suppl): S387.
- 146 Chandra S, Fukuda T, Mizuno K, Davies SM, Teusink-Cross A, Tarin R *et al.* Micafungin antifungal prophylaxis in children undergoing HSCT: can we give higher doses, less frequently? A pharmacokinetic study. *J Antimicrob Chemother* 2018; **73**: 1651–1658.
- Bochennek K, Balan A, Muller-Scholden L, Becker M, Farowski F, Muller C *et al.*Micafungin twice weekly as antifungal prophylaxis in paediatric patients at high risk for invasive fungal disease. *J Antimicrob Chemother* 2015; **70**: 1527–1530.
- Lehrnbecher T, Bochennek K, Klingebiel T, Gastine S, Hempel G, Groll AH. Extended dosing regimens for fungal prophylaxis. *Clin Microbiol Rev* 2019; **32**: e00010–19.
- Decembrino N, Perruccio K, Zecca M, Colombini A, Calore E, Muggeo P *et al.* A case series and literature review of isavuconazole use in pediatric patients with hemato-

oncologic diseases and hematopoietic stem cell transplantation. *Antimicrobial Agents* and *Chemotherapy* 2020; **64**: e01783–19.

- 150 Cornu M, Bruno B, Loridant S, Navarin P, François N, Lanternier F *et al.* Successful outcome of disseminated mucormycosis in a 3-year-old child suffering from acute leukaemia: the role of isavuconazole? A case report. *BMC Pharmacol Toxicol* 2018; **19**: 81.
- Barg AA, Malkiel S, Bartuv M, Greenberg G, Toren A, Keller N. Successful treatment of invasive mucormycosis with isavuconazole in pediatric patients. *Pediatr Blood Cancer* 2018; **65**: e27281.
- De Leonardis F, Novielli C, Giannico B, Mariggiò MA, Castagnola E, Santoro N. Isavuconazole treatment of cerebral and pulmonary aspergillosis in a pediatric patient with acute lymphoblastic leukemia: case report and review of literature. *J Pediatr Hematol Oncol* 2020; **42**: e469–e71.
- Abbotsford J, Foley DA, Goff Z, Bowen AC, Blyth CC, Yeoh DK. Clinical experience with SUBA-itraconazole at a tertiary paediatric hospital. *J Antimicrob Chemother* 2021; **76**: 249–252.
- Döring M, Cabanillas Stanchi KM, Queudeville M, Feucht J, Blaeschke F, Schlegel P *et al.* Efficacy, safety and feasibility of antifungal prophylaxis with posaconazole tablet in paediatric patients after haematopoietic stem cell transplantation. *J Cancer Res Clin Oncol* 2017; **143**: 1281–1292.

- Mauro M, Colombini A, Perruccio K, Zama D, D'Amico MR, Calore E *et al.* Posaconazole delayed-release tablets in paediatric haematology-oncology patients. *Mycoses* 2020;
 63: 604–609.
- Bernardo V, Miles A, Fernandez AJ, Liverman R, Tippett A, Yildirim I. Initial posaconazole dosing to achieve therapeutic serum posaconazole concentrations among children, adolescents, and young adults receiving delayed-release tablet and intravenous posaconazole. *Pediatr Transplant* 2020: e13777.
- Tragiannidis A, Herbruggen H, Ahlmann M, Vasileiou E, Gastine S, Thorer H *et al.*Plasma exposures following posaconazole delayed-release tablets in immunocompromised children and adolescents. *J Antimicrob Chemother* 2019; **74**: 3573–3578.
- 158 Groll AH, Abdel-Azim H, Lehrnbecher T, Steinbach WJ, Paschke A, Mangin E *et al.*Pharmacokinetics and safety of posaconazole intravenous solution and powder for oral suspension in children with neutropenia: an open-label, sequential dose-escalation trial. *Int J Antimicrob Agents* 2020; **56**: 106084.
- Nickless JR, Bridger KE, Vora SB, Brothers AW. Evaluation of intravenous posaconazole dosing and pharmacokinetic target attainment in pediatric patients. *J Pediatric Infect Dis Soc* 2019; **8**: 365–367.
- Hope WW, Castagnola E, Groll AH, Roilides E, Akova M, Arendrup MC *et al.* ESCMID guideline for the diagnosis and management of *Candida* diseases 2012: prevention and management of invasive infections in neonates and children caused by *Candida* spp. *Clin Microbiol Infect* 2012; **18** (Suppl 7): 38–52.

- Vena A, Bouza E, Álvarez-Uría A, Gayoso J, Martín-Rabadán P, Cajuste F *et al.* The misleading effect of serum galactomannan testing in high-risk haematology patients receiving prophylaxis with micafungin. *Clin Microbiol Infect* 2017; **23**: 1000.e1–.e4.
- Huppler AR, Fisher BT, Lehrnbecher T, Walsh TJ, Steinbach WJ. Role of molecular biomarkers in the diagnosis of invasive fungal diseases in children. *Pediatr Infect Dis J* 2017; **6**: S32–S44.
- Lehrnbecher T, Hassler A, Groll AH, Bochennek K. Diagnostic approaches for invasive aspergillosis-specific considerations in the pediatric population. *Front Microbiol* 2018; **9**: 518.
- Yang L, Yu L, Chen X, Hu Y, Wang B. Clinical analysis of adverse drug reactions between vincristine and triazoles in children with acute lymphoblastic leukemia. *Med Sci Monit* 2015; **21**: 1656–1661.
- Lin MJ, Paul MR, Kuo DJ. Severe neuropathic pain With concomitant administration of vincristine and posaconazole. *J Pediatr Pharmacol Ther* 2018; **23**: 417–420.
- Pekpak E, İleri T, İnce E, Ertem M, Uysal Z. Toxicity of vincristine combined with posaconazole in children with acute lymphoblastic leukemia. *J Pediatr Hematol Oncol* 2018; **40**: e309–e10.
- 167 Tramsen L, Salzmann-Manrique E, Bochennek K, Klingebiel T, Reinhardt D, Creutzig U *et al.* Lack of effectiveness of neutropenic diet and social restrictions as anti-infective measures in children with acute myeloid leukemia: an analysis of the AML-BFM 2004 trial. *J Clin Oncol* 2016; **34**: 2776–2783.

- Abramson DH, Marr BP, Francis JH, Dunkel IJ, Fabius AW, Brodie SE *et al.*Simultaneous bilateral ophthalmic artery chemosurgery for bilateral retinoblastoma
 (Tandem Therapy). *PLoS One* 2016; **11**: e0156806.
- Neemann K, Yonts AB, Qiu F, Simonsen K, Lowas S, Freifeld A. Blood cultures for persistent fever in neutropenic pediatric patients are of low diagnostic yield. *J Pediatric Infect Dis Soc* 2016; **5**: 218–221.
- De Luca M, Donà D, Montagnani C, Lo Vecchio A, Romanengo M, Tagliabue C *et al.*Antibiotic prescriptions and prophylaxis in Italian children. Is it time to change? Data from the ARPEC project. *PLoS One* 2016; **11**: e0154662.
- 171 Ku BC, Bailey C, Balamuth F. Neutropenia in the febrile child. *Pediatr Emerg Care* 2016; **32**: 329–334.
- 172 Camargo JF, Husain S. Immune correlates of protection in human invasive aspergillosis. *Clin Infect Dis* 2014; **59**: 569–577.
- 173 Cunha C, Aversa F, Lacerda JF, Busca A, Kurzai O, Grube M *et al.* Genetic PTX3 deficiency and aspergillosis in stem-cell transplantation. *N Engl J Med* 2014; **370**: 421–432.
- 174 Matzaraki V, Gresnigt MS, Jaeger M, Ricano-Ponce I, Johnson MD, Oosting M *et al.* An integrative genomics approach identifies novel pathways that influence candidaemia susceptibility. *PLoS One* 2017; **12**: e0180824.

- 175 Ceesay MM, Kordasti S, Rufaie E, Lea N, Smith M, Wade J *et al.* Baseline cytokine profiling identifies novel risk factors for invasive fungal disease among haematology patients undergoing intensive chemotherapy or haematopoietic stem cell transplantation. *J Infect* 2016; **73**: 280–288.
- Zhai B, Ola M, Rolling T, Tosini NL, Joshowitz S, Littmann ER et al. High-resolution mycobiota analysis reveals dynamic intestinal translocation preceding invasive candidiasis. Nat Med 2020; 26: 59–64.
- 177 Van Daele R, Spriet I, Wauters J, Maertens J, Mercier T, Van Hecke S *et al.* Antifungal drugs: what brings the future? *Med Mycol* 2019; **57**: S328–S343.
- 178 Rauseo AM, Coler-Reilly A, Larson L, Spec A. Hope on the horizon: novel fungal treatments in development. *Open Forum Infect Dis* 2020; **7**: ofaa016.
- 179 Science M, Robinson PD, MacDonald T, Rassekh SR, Dupuis LL, Sung L. Guideline for primary antifungal prophylaxis for pediatric patients with cancer or hematopoietic stem cell transplant recipients. *Pediatr Blood Cancer* 2014; **61**: 393–400.
- Uhlenbrock S, Zimmermann M, Fegeler W, Jurgens H, Ritter J. Liposomal amphotericin
 B for prophylaxis of invasive fungal infections in high-risk paediatric patients with
 chemotherapy-related neutropenia: interim analysis of a prospective study. *Mycoses*2001; **44**: 455–463.
- Bochennek K, Tramsen L, Schedler N, Becker M, Klingebiel T, Groll AH *et al.* Liposomal amphotericin B twice weekly as antifungal prophylaxis in paediatric haematological malignancy patients. *Clin Microbiol Infect* 2011; **17**: 1868–1874.

Figure legends

Nil

Tables

Table 1 Established risk groups for IFD and recommended antifungal prophylaxis coverage in adults

Risk level	Risk groups	Recommended prophylaxis [†]	SoR	QoE
High risk >10% incidence of	Neutrophil <0.1 x 10^9 /L for >3 weeks or <0.5 x 10^9 /L for >5 weeks (e.g. allogeneic HSCT)	First line: Posaconazole	Α	I
IFD	Corticosteroids >1 mg/kg prednisolone equivalent and neutrophils <1 x 10 ⁹ /L for >1 week Corticosteroids >2 mg/kg prednisolone equivalent >2 weeks	Alternate agents: Voriconazole Itraconazole Micafungin Liposomal amphotericin		

		Isavuconazole		
	Unrelated, mismatched or cord blood allogeneic			
	HSCT			
	GVHD – extensive or severe			
	AML – induction/reinduction			
	ALL – induction/reinduction			
	MDS			
Low risk	Autologous HSCT (e.g. patients at high risk for	First line:	В	II (context
Less than 5%	mucositis)	Fluconazole		dependent;
incidence of				level I
IFD	Allogeneic HSCT with expected neutropenia <14 days	Alternate agents:		evidence in
		Alternate agents.		setting of
		Echinocandins		alloHSCT)

	Lymphoma (e.g. intensive/dose-escalated therapy)	Itraconazole		
Very low risk [‡]	Other lymphoproliferative neoplasms (e.g.	No prophylaxis	В	II
Less than 5%	standard chemotherapy for lymphoma, induction			
incidence of	therapy for myeloma, treatment-naïve CLL)			
IFD				
No mucositis	Other myeloproliferative neoplasms			
	Treatment for solid organ tumours			

†Please refer to Table 4 for summary of recommendations and level of evidence supporting choice of antifungal prophylaxis agents. ‡Consider that low and/or sporadic occurrence is not equal to no risk and is dependent on underlying treatment regimen, previous and cumulative treatments. ALL, acute lymphoblastic leukaemia; AML, acute myeloid leukaemia; CLL, chronic lymphocytic leukaemia; GVHD, graft vs. host disease; HSCT, haemopoietic stem cell transplantation; IFD, invasive fungal disease; MDS, myelodysplastic syndrome; QoE, quality of evidence; SoR, strength of recommendation

Table 2 Summary of IFD rates associated with emerging use of new generation cancer therapies

Therapy	Population	IFD rates	Comments
BTK inhibitor (e.g. ibrutinib)	Relapsed/refractory B-cell	3–12%	Rates of 1% reported in clinical
	lymphoproliferative disorder		trials of BTK inhibitors
			Invasive aspergillosis with CNS
			involvement up to 40%
			Cryptococcus spp.
			Pneumocystis jirovecii pneumonia

	Primary CNS lymphoma	5–44%	In combination with
			corticosteroids and conventional
			chemotherapy
PI3K inhibitor (e.g. idelalisib)	Relapsed/refractory B-cell	3%	Pneumocystis jirovecii pneumonia
	lymphoproliferative disorder		
BCL-2 inhibitor (e.g. venetoclax)	CLL	1%	Aspergillus spp., Pneumocystis
			<i>jirovecii</i> pneumonia
Hypomethylating agents (e.g.	MDS	5–13%	Rates higher in
azacitadine)	AML		relapsed/refractory disease
			versus its use as front-line
			therapy

			Rate of 13% when used in
			combination with BCL-2 inhibitor
			COMBINATION WITH BCL-2 INHIBITOR
			venetoclax
			Aspergillus spp., Candida spp.
FLT-3 inhibitors (e.g.	AML	5%	Limited data from clinical trial
midostaurin, gliteritinib)			
Second generation IMiD, PI	Relapsed/refractory myeloma	2–7%	Candida spp., Cryptococcus spp.
CD38 or SLAMF7 monoclonal			
antibodies			
CAR T cell therapy	Relapsed/refractory ALL	5–8%	In the setting of fluconazole or
			micafungin prophylaxis

	Relapsed/refractory NHL		
			Rates up to 13% in patients with
			ALL
			<i>Aspergillus</i> spp., <i>Candida</i> spp.,
			<i>Mucor</i> spp.
		201	
Bi-specific antibody therapies	Relapsed/refractory ALL	2%	Limited clinical trial data
(e.g. blinatumomab)	Relapsed/refractory NHL		
		· pg 2 p III I	O DTI(D /

ALL, acute lymphoblastic leukaemia; AML, acute myeloid leukaemia; BCL-2, B-cell lymphoma 2; BTK, Bruton's tyrosine kinase; CAR, chimeric antigen receptor; CLL, chronic lymphocytic leukaemia; CNS, central nervous system; FLT-3, fms-like tyrosine kinase; IMiD, immunomodulatory drug therapy; IFD, invasive fungal disease; MDS, myelodysplastic syndrome; NHL, non-Hodgkin lymphoma; PI, proteasome inhibitor; PI3K, phosphatidylinositol 3-kinase; SLAMF7, signalling lymphocytic activation molecule F7

Table 3 Summary of key new haematological treatments and recommended approaches to prophylaxis in adults and children

Therapy	Patient group	Infection	Measures	SoR	QoE
Targeted therapies	Relapsed/refractory	Yeast	Need for prophylaxis should be	В	II
(e.g. BTK	B-cell	and	determined taking into account recent		
inhibitors)	lymphoproliferative	mould	therapy (e.g. fludarabine-based), ongoing		
	disorders	infections	immune suppression and		
			presence/absence of previous IFD		
Immunomodulatory	Relapsed/refractory	Yeast	Need for prophylaxis should be	С	III
drug therapy,	myeloma	infection	determined by number of previous lines of		
monoclonal			therapy, risk factors for IFD such as		
antibody therapy			prolonged neutropenia and		
(CD38/SLAMF7)			presence/absence of previous IFD		

CAR T cell therapy	Relapsed/refractory	Yeast	Yeast prophylaxis with fluconazole or	A	II
	lymphoproliferative	and	micafungin		
	disorders	mould			
		infections	Consider mould prophylaxis in the setting of prolonged neutropenia or additional treatments for high-grade cytokine release syndrome following CAR T cell therapy		
			, , , , , , , , , , , , , , , , , , , ,		
			Previous therapies including recent		
			allogeneic or autologous HSCT should be		
			taken into account		
Bi-specific antibody	ALL	Yeast	Need for prophylaxis should be	С	III
therapies		and	determined taking into account recent therapy (e.g. fludarabine-based), ongoing		

	mould	immune suppression and
Being evaluated for	infections	presence/absence of previous IFD
other aggressive B-		
cell		
lymphoproliferative		
disorders		

ALL, acute lymphoblastic leukaemia; BTK, Bruton's tyrosine kinase; CAR, chimeric antigen receptor; HSCT, haemopoietic stem cell transplantation; IFD, invasive fungal disease; QoE, quality of evidence; SLAMF7, signalling lymphocytic activation molecule F7; SoR, strength of recommendation

Table 4 Recommendations for choice and dose of antifungal prophylaxis agent in adults

Risk group		Antifungal agent	SoR	QoE	Comments
High risk	First line	Posaconazole Oral (tablets) Loading with 300 mg twice daily on Day 1, followed by 300 mg daily	A	I	Intravenous formulation can be used to continue prophylaxis if poor oral intake/absorption
	Alternate agents	Voriconazole Oral or intravenous 4 mg/kg twice daily [†]	Α	II	High rates of adverse events (liver function abnormalities); variable CYP metabolism
		Micafungin Intravenous	В	II	Could be used during periods of neutropenia if azoles

100–150 mg daily			contraindicated, poor oral
			intake/absorption
Itraconazole	В	II	Less new data supporting its use
Oral			compared to other azoles
200 mg twice daily			
Liposomal amphotericin	В	II	Could be used if azoles
Intravenous			contraindicated due to drug-drug
50-200 mg three times per week			interactions, adverse events, poor oral intake/absorption
Isavuconazole	С	II	Higher rates of IFD in cohort
Oral			studies; could be used if other
200 mg three times per day for 48 hours			azoles contraindicated due to
followed by 200 mg daily			

					adverse events such as QTc prolongation
Low risk	First line	Fluconazole	Α	I	
		Oral			
		200–400 mg daily			
	Alternate agents	Echinocandin	Α	II	
		Intravenous			
		Dosing dependent on agent			
		Itraconazole	Α	II	
		Oral			
		200 mg twice daily			
Very low risk		No prophylaxis	В	II	

[†]Dose used in prophylaxis studies have been 200 mg twice daily; measure voriconazole levels to ensure achievement of target level (refer to accompanying optimising antifungal therapy and TDM guidelines by Chau *et al.* 2021, which can be found elsewhere in this supplement). CYP, cytochrome P450; IFD, invasive fungal disease; QoE, quality of evidence; QTc, corrected QT interval; SoR, strength of recommendation

Table 5 Updated risk groups for IFD in children (adapted from Groll *et al.* 2014,¹³⁶ Science *et al.* 2014,¹⁷⁹ Fisher *et al.* 2018¹¹¹ and Lehrnbecher *et al.* 2020¹¹)

Risk level	Clinical examples
High risk (>10%)	AML
	Recurrent/relapsed acute leukaemia
	High-risk ALL [†]
	Allogeneic HSCT
	Allogeneic with acute grade 2-4 GVHD or chronic extensive
	GVHD
Low risk (<5%) [‡]	Standard-risk or low-risk ALL [†]
	Non-Hodgkin lymphomas
	Autologous HSCT
Sporadic	Paediatric solid tumours
occurrence [‡]	Brain tumours
	Hodgkin's lymphoma
Unknown	CAR T cell therapy
	Other immunotherapy

[†]Key change from previous Australian antifungal prophylaxis guidelines based on TERIFIC study results.^{112, 113} Note, high-risk ALL includes T-cell ALL, infant ALL, Philadelphia-positive ALL and high-risk B-cell ALL. [‡]Consider that low and sporadic occurrence is not equal to no risk and dependant on underlying chemotherapy regimen. ALL, acute lymphoblastic leukaemia; AML, acute myeloid leukaemia; CAR,

chimeric antigen receptor; GVHD, graft-versus-host disease; HSCT, haemopoietic stem cell transplantation; IFD, invasive fungal disease

Table 6 Indications for primary antifungal prophylaxis in children based on clinical practice guideline by Lehrnbecher *et al.*, 2020.¹¹ Prophylaxis recommendations used with permission from lead authors and journal. Strength of recommendations adapted from Lehrnbecher *et al.*, 2020¹¹ to align with Australian guideline methodology. See in-text discussion for type of agent recommended.

Risk classification	Clinical example	Prophylaxis	SoR	QoE
		recommended		
High-risk IFD	AML (<i>de novo</i> or	Routine mould-	Α	I
	relapsed)	active prophylaxis		
		recommended [†]		
	ALL (high-risk or	Routine mould-	В	III
	relapsed)	active prophylaxis		
		should be		
		$considered^{\scriptscriptstyle \dagger}$		

	Allogeneic HSCT	Routine mould-	Α	II
	(pre-	active prophylaxis		
	engraftment [‡]	$recommended^{\dagger}$		
	and treatment			
	of GVHD)			
Low-risk IFD	ALL (standard	Routine	Α	III
	risk)§	prophylaxis not		
		recommended		
	Solid tumours§	Routine	Α	II
		prophylaxis not		
		recommended		
	Most	Routine	Α	II
	lymphomas§	prophylaxis not		
		recommended		

Au	tologous	Routine	С	III
HS	CT§	prophylaxis not		
		recommended		

†Choice of mould-active agent will depend on age, potential drug interactions and patient location (e.g. inpatient versus outpatient), with preference given to mould-active azoles or echinocandins. ‡Fluconazole is a reasonable alternative to a mould-active agent for patients undergoing allogeneic HSCT in the pre-engraftment phase, provided they do not have a history of proven or probable mould IFD and where local epidemiology supports its use. §Consider that low risk is not equal to no risk and dependant on underlying disease and chemotherapy regimen. For autologous HSCT, there is less certainty in the setting of tandem transplantations where the cumulative duration of neutropenia may be longer. ALL, acute lymphoblastic leukaemia; AML, acute myeloid leukaemia; GVHD, graft-versus-host disease; HSCT, haemopoietic stem cell transplantation; IFD, invasive fungal disease; N/A, not applicable; QoE, quality of evidence; SoR, strength of recommendation

Table 7 Suggested dosing for antifungal prophylaxis in children

Medication	Recommended dose	
Posaconazole [†]	Able to swallow tablets	<u>Unable to swallow tablets</u>
	Oral (modified-release tablets)	Oral (suspension)
	300 mg daily (>30 kg) [‡]	200 mg three times a day (≥13 years)
	For dosing options for children weighing <30 kg, please	For dosing options for children <13 years, please
	refer to Tragiannidis <i>et al</i> . 2019 ¹⁵⁷	refer to Boonsathorn <i>et al.</i> 2019 ¹⁰⁰
Itraconazole [†]	Oral liquid	
	2.5 mg/kg (max 200 mg) twice daily	
Voriconazole [†]	2 to <12 years of age OR	≥15 years of age OR
	12–14 years of age and <50 kg:	12–14 years of age and >50 kg:
	Oral	Oral
	9 mg/kg (max 350 mg) twice daily	200 mg twice daily

Intravenous Intravenous

8 mg/kg twice daily 4 mg/kg twice daily

(loading dose 9 mg/kg twice daily on Day 1) (loading dose 6 mg/kg twice daily on Day 1)

Micafungin Intravenous

1 mg/kg (max 50 mg) daily

Caspofungin Intravenous

50 mg/m² (max 50 mg) daily

(70 mg/m² loading dose on Day 1)

Liposomal amphotericin B	Intravenous
	1 mg/kg three times per week ¹⁸⁰
	or
	3 mg/kg three times per week ¹³⁴
	or
	2.5 mg/kg twice weekly ¹⁸¹

[†]Adjust based on therapeutic drug monitoring target trough levels: posaconazole 0.7 mg/L; itraconazole 0.5–4 mg/L; voriconazole 1–5.5 mg/L.

 $^{^{\}dagger}$ Not TGA-approved for use in children <13 years of age in Australia. TGA, Therapeutic Goods Administration

Figures

Nil

Appendices

Nil

Consensus guidelines for antifungal prophylaxis in haematological malignancy and haemopoietic stem cell transplantation, 2021

Short title

Antifungal prophylaxis guidelines 2021

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Conflicts of interest

The following working group members are consultants or advisory committee members or

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Abstract

Antifungal prophylaxis can reduce morbidity and mortality from invasive fungal disease

(IFD). However, its use needs to be optimised and appropriately targeted to patients at

highest risk to derive the most benefit. In addition to established risks for IFD, considerable

recent progress in the treatment of malignancies has resulted in the development of new

'at-risk' groups. The changing epidemiology of IFD and emergence of drug resistance

continue to impact choice of prophylaxis, highlighting the importance of active surveillance

and knowledge of local epidemiology. These guidelines aim to highlight emerging risk groups and review the evidence and limitations around new formulations of established agents and new antifungal drugs. It provides recommendations around use and choice of antifungal prophylaxis, discusses the potential impact of the changing epidemiology of IFD and emergence of drug resistance, and future directions for risk stratification to assist optimal management of highly-vulnerable patients.

Keywords

antifungal prophylaxis, *Aspergillus*, *Candida*, stem cell transplantation, haematological malignancy