1	
2	DR CAROLINE DONG (Orcid ID: 0000-0003-3352-4006)
3	
4	
5	Article type : Research Papers
6	
7	
8	RUNNING TITLE: Parental effects on hybrid fitness
9	
10	TITLE: Maternal reproductive output and F1 hybrid fitness may influence contact zone
11	dynamics
12	
13	AUTHORS: Caroline M. Dong ^{1*} , Katrina J. Rankin ² , Claire A. McLean ³ , Devi Stuart-Fox ⁴
14	
15	¹ School of BioSciences, The University of Melbourne, Parkville, Victoria, Australia
16	Sciences Department, Museums Victoria, Carlton Gardens, Victoria, Australia
17	Present address: Department of Ecology and Evolutionary Biology, Tulane University, New
18	Orleans, Louisiana, United States
19	Email: cdong1@tulane.edu
20	Twitter: colorfulagamids
21	*Author for correspondence
22	
23	² School of BioSciences, The University of Melbourne, Parkville, Victoria, Australia
24	Email: katrina.rankin@unimelb.edu.au
25	Twitter: kjrankin88
26	
27	³ School of BioSciences, The University of Melbourne, Parkville, Victoria, Australia
28	Sciences Department, Museums Victoria, Carlton Gardens, Victoria, Australia
29	Email: cmclean@museum.vic.gov.au
	This is the author manuscript accepted for publication and has undergone full peer review but has

not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the <u>Version of Record</u>. Please cite this article as <u>doi</u>: 10.1111/JEB.13772

30	Twitter:	Claire	A	McLean

31

32 ⁴School of BioSciences, The University of Melbourne, Parkville, Victoria, Australia

Email: d.stuart-fox@unimelb.edu.au

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

33

Abstract

The outcome of secondary contact between divergent lineages or species may be influenced by both the reproductive traits of parental species and the fitness of offspring; however, their relative contributions have rarely been evaluated, particularly in longer lived vertebrate species. We performed pure and reciprocal laboratory crosses between Ctenophorus decresii (tawny dragon) and C. modestus (swift dragon) to examine how parental reproductive traits and ecologically-relevant offspring fitness traits may explain contact zone dynamics in the wild. The two species meet in a contact zone of post-F1 hybrids with asymmetric backcrossing and predominantly C. modestus mtDNA haplotypes. We found no evidence for reduced parental fecundity or offspring fitness for F1 hybrid crosses. However, maternal reproductive strategy differed between species, irrespective of the species of their mate. Ctenophorus modestus females had higher fecundity and produced more and larger clutches with lower embryonic mortality. Parental lineage also influenced sex ratios and offspring traits, with C. modestus \mathcal{L} × C. decresii \(\frac{1}{2} \) hybrids exhibiting higher trait values for more fitness measures (growth rate, sprint speed, bite force) than offspring from all other pairings. Together, these patterns are consistent with the prevalence of C. modestus mtDNA in the contact zone, and asymmetric backcrossing likely reflects fitness effects that manifest in the F2 generation. Our results highlight how parental species can influence multiple offspring traits in different ways, which together may combine to influence offspring fitness and shape contact zone dynamics.

54

55

56

57

58

59

60

Keywords: captive-breeding, *Ctenophorus decresii*, *Ctenophorus modestus*, hybridisation, life history, lizard, reinforcement, reproductive isolation

Introduction

Hybridisation is an important evolutionary process that can influence the trajectory or persistence of one or both parental species. Although hybridisation occurs across a wide range of spatial and temporal contexts, there are three possible distinct outcomes: a stable equilibrium (i.e.

61 tension zone), the breakdown of barriers due to gene flow and recombination, or the 62 strengthening of barriers driving further divergence (reviewed in Abbott et al. 2013). During 63 secondary contact, the viability and fertility of first generation (F1) hybrids in particular may 64 determine the long-term consequences of hybridisation by precluding, reducing or facilitating 65 gene flow in one or either direction (Arnold et al., 2012). Numerous studies have investigated 66 genetic and phenotypic patterns across naturally occurring contact zones but few have 67 concurrently used laboratory studies of parental reproductive output and hybrid fitness to aid in interpreting patterns observed (but see MacGregor et al., 2017; Orr, 1996; Peterson et al., 2005; 68 69 Rank et al., 2020; Scribner, 1993; While et al., 2015). An interplay of intrinsic and extrinsic 70 factors governs F1 fitness, and laboratory studies (e.g. captive breeding) may enable stronger 71 inferences about the nature of reproductive isolating barriers. For example, the absence of post-72 F1s across a beetle hybrid zone (Chrysochus auratus and C. cobaltinus), in conjunction with the 73 reduced fertility of F1 hybrid females in a laboratory setting, suggested that the hybrid zone 74 represented a tension zone between these species (Peterson et al., 2005). 75 Intrinsic low hybrid fitness is generally attributed to Dobzhansky-Muller 76 incompatibilities (DMIs; Dobzhansky, 1937; Muller, 1942) between biparentally inherited 77 autosomal loci (nuclear-nuclear). Postzygotic barriers are expected to select for the formation of 78 prezygotic barriers to avoid maladaptive hybridisation, thereby promoting speciation 79 (Dobzhansky 1937; reviewed in Butlin 1987; Howard 1993; Noor 1999; Turelli et al. 2001; 80 Servedio and Noor 2003; Coyne and Orr 2004; Servedio 2004; Ortiz-Barrientos et al. 2009; 81 Abbott et al. 2013; Pfennig 2016). This process of reinforcement following secondary contact is 82 thought to complete speciation initiated in allopatry (Hoskin et al., 2005). Conversely, increased 83 hybrid fitness may result in hybrids outcompeting parental lineages (Huxel, 1999; Rhymer & Simberloff, 1996; Rosenfield et al., 2004; Taylor et al., 2006), the formation of a stable hybrid 84 85 zone when hybrids only have higher fitness in the distinct habitat of the contact zone (Nosil et 86 al., 2009) or in tension zones (Barton & Hewitt, 1985), or hybrid speciation (reviewed in Mallet 87 2007). Fitness disparities between reciprocal hybrids may arise from DMIs involving sex 88 chromosomes or cytoplasmic factors (e.g. mitochondria, chloroplasts; Arntzen et al., 2009; 89 Bolnick et al., 2008; Bolnick & Near, 2005; Rank et al., 2020; Tiffin et al., 2001), and may also 90 be shaped by exogenous selection (Campbell & Waser, 2001; Kimball et al., 2008; Muir et al.,

92 differences in both parental fecundity and hybrid fitness are essential. 93 Here, we investigate differences in parental reproductive output and the fitness of F1 94 hybrids produced from captive-breeding between Ctenophorus decresii (tawny dragon; Duméril 95 & Bibron, 1837) and C. modestus (swift dragon; Ahl, 1926), which were previously regarded as 96 divergent lineages of C. decresii sensu lato (Dong et al., 2021). Ctenophorus decresii sensu 97 stricto (previously southern lineage C. decresii) and C. modestus (previously northern lineage C. 98 decresii) differ in male coloration and meet naturally in a zone of secondary contact (Figure 1). 99 Ctenophorus modestus males are polymorphic with four co-occurring throat colour morphs 100 (orange, yellow, yellow with orange centre, and grey), whereas C. decresii males are 101 monomorphic with blue throats that reflect ultraviolet (UV) wavelengths (McLean et al., 2014; 102 Teasdale et al., 2013). Secondary contact between populations or lineages that differ in colour 103 morphs is predicted to generate genetic incompatibilities due to the role of colour signals in 104 reproductive isolation (Boughman, 2001; Jiggins et al., 2001; Mavárez et al., 2006; Sætre et al., 105 1997; Seehausen et al., 2008) and the genetic architecture of colour polymorphism (Sinervo & 106 Svensson, 2002). We recently showed that the polymorphic throat coloration is the target of 107 selection during incipient speciation between C. decresii and C. modestus (Dong et al., 2019), 108 consistent with the view that colour polymorphism can promote speciation (Corl, Davis, Kuchta, 109 & Sinervo, 2010; Forsman, Ahnesjö, Caesar, & Karlsson, 2008; Gray & McKinnon, 2007; 110 Hugall & Stuart-Fox, 2012; McLean & Stuart-Fox, 2014; West-Eberhard, 1986). The contact 111 zone represents a hybrid swarm and there is evidence of intrinsic incompatibilities, with admixed 112 individuals primarily having C. modestus mtDNA haplotypes and genomic patterns indicating 113 restricted backcrossing to C. decresii (Dong et al., 2019; McLean et al., 2014). Further, captive 114 behavioural trials indicate incomplete and asymmetric behavioural barriers to mating, with stronger barriers between C. decresii males and C. modestus females than the reciprocal 115 116 combination driven by both male and female mate preferences (McLean et al., 2020). Specifically, C. decresii males displayed a preference for conspecific individuals and C. 117 118 modestus females were generally more choosy (displayed more rejection behaviours), whereas 119 C. modestus males and C. decresii females did not discriminate between con- and heterospecific 120 individuals.

2015; Wesselingh et al., 2019). To distinguish these possibilities, laboratory crosses to assess

In order to assess the nature and extent of reproductive isolating barriers, we conducted a captive-breeding experiment to produce all pairwise combinations of crosses using genetically pure parental populations from either end of the contact zone. We compared parental reproductive life history traits and the viability and fitness of F1 hybrids relative to that of pure offspring. We measured offspring survival, growth rate, maximum bite force, and maximum sprint speed as proxies of competitive ability and fitness, as these traits strongly correlate with ecologically relevant tasks and are known to be the target of selection (reviewed in Irschick et al., 2007; Wainwright, 2007). Based on the structure of the contact zone, we predicted that F1 hybrids would have lower fitness than pure crosses, due to the low prevalence of F1 hybrids and near absence of individuals backcrossed to *C. decresii*. Furthermore, given the prevalence of *C. modestus* mtDNA haplotypes in the contact zone, we predicted greater reproductive output by *C. modestus* mothers and/or that hybrids with *C. decresii* mtDNA haplotypes (i.e. *C. decresii* mother) may suffer from reduced viability and/or fitness. We discuss the implications of our results for inferring the consequences of secondary contact and trajectories of parental populations.

Materials and Methods

Animals

Ctenophorus decresii and C. modestus are small (snout-vent-length [SVL] ≤ 90 mm) rock-dwelling agamid lizards endemic to South Australia. We captured individuals from the wild in 2015 and 2016 (September − November) using a standard lassoing protocol or by hand. This included 18 adult female and 21 adult male C. modestus from the vicinity of Caroona Creek Conservation Park (longitude: 139.103°, latitude: −33.443°, ca. 130 km north of the contact zone centre) and 17 adult female and 18 adult male C. decresii from the vicinity of Palmer (longitude: 139.159°, latitude: −34.851°, ca. 30 km south of the contact zone centre) in South Australia (Figure 1). These populations represent the nearest pure parental populations to the contact zone. Lizards were housed individually in an indoor facility at The University of Melbourne, Melbourne, Australia, in 55 cm × 34 cm × 38 cm (length [L] × width [W] × depth [D]) opaque plastic enclosures containing a layer of sand and two stacked ceramic tiles for shelter and basking. Lizards were fed live crickets and mealworms ad libitum three times per week and misted with water for hydration. The room was maintained at temperature and lighting regimes

152 that reflected natural seasonal variation, with appropriate UV lighting (Outback Max 10.0 153 UVA/UVB). Heat lamps were provided in each enclosure to generate a thermal gradient and 154 allow animals to attain their preferred body temperature (36°C; Gibbons, 1977; Walker, 155 unpublished data).

156

157

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

Captive breeding

Our objective was to produce pure offspring (C. modestus $\mathcal{D} \times C$. modestus \mathcal{D}, C . decresii $\mathcal{L} \times C$. decresii \mathcal{L}) and reciprocal cross hybrids (C. modestus $\mathcal{L} \times C$. decresii $\mathcal{L} \times C$. C. modestus 3). The sexes of each species were equally partitioned into two groups (when possible), all individuals of each group were paired with either a same-species mate or oppositespecies mate to produce a clutch of eggs, and this was reversed in the next reproductive cycle for up to three cycles within the breeding season. If a female did not become visibly gravid after two weeks, she was paired with an alternative equivalent-species male to increase the likelihood of copulation in each cycle. Females mate again after oviposition; therefore, for the next reproductive cycle, one to two days following oviposition, females were housed with a male of the opposite species from the previous mate. Males of each species followed the same partitioning and pairing procedure such that they encountered alternating species females. Breeding trials were conducted in October – December 2017, during their natural breeding season.

Nest boxes were provided in the form of 17 cm \times 13 cm \times 15 cm (L \times W \times D) clear plastic enclosed containers with a 3.5 cm circular entrance in a lower corner and damp 50:50 sand:peat moss mixture sloped upwards to the top of the box. For lizards, most females will bury their clutches when provided with a suitable nesting area but inexperienced individuals may lay clutches in the open (Sanger et al., 2008). To account for clutches laid in the open, where they are vulnerable to desiccation, gravid females were checked for evidence of oviposition up to five times daily. Eggs were collected following oviposition (typically within one hour) and individually weighed to the nearest 0.01 g using a digital balance. Eggs were incubated in sealed plastic containers filled two-thirds with moist vermiculite (volume ratio 5:1 vermiculite:water) and incubated at 28°C (± 0.12°C) until hatching. This incubation temperature was chosen for a 50:50 sex ratio (Rankin et al., 2016).

Measuring parental reproductive traits

Indicators of female reproductive output were: fecundity, clutch size, embryonic mortality, egg mass at oviposition, and offspring mass at hatching. Female fecundity was measured as total reproductive output across the breeding season (i.e. total mass of eggs produced across clutches). Embryonic mortality was scored based on the presence of calcification of the egg shell and/or failure to hatch (Olsson & Shine, 1997). Following the breeding season, we recorded male testes mass as this trait may be under sexual selection in lizards (Olsson & Madsen, 1998). Males were humanely killed, and testes were dissected and weighed to the nearest 0.01 g on a digital balance.

Offspring rearing conditions

Experimental crosses resulted in a total of 53 clutches comprising 259 eggs of which 211 were able to be incubated (i.e. laid in nest boxes or found before they had desiccated). Of these, 176 viable offspring hatched: 69 out of 73 pure *C. modestus* (94.5%), 14 out of 22 pure *C. decresii* (63.6%), 61 out of 67 hybrids sired by *C. decresii* males (91.0%), 32 out of 41 hybrids sired by *C. modestus* males (78.1%). Offspring were raised for a minimum of six months under laboratory conditions and housed individually in 30 cm × 20 cm × 10 cm (L × W × D) plastic enclosures with fly screen lids. Each enclosure contained two stacked ceramic tiles to provide shelter with paper substrate and a water dish. Ambient temperature was maintained at 24°C, UV lighting was provided, and a heat lamp was positioned at one end of the enclosure for a basking temperature of 31°C. Hatchlings were fed pinhead crickets and fruit flies (*Drosophila*) *ad libitum* twice daily and misted with water. At three months post-hatching, their diet was changed to small crickets *ad libitum* daily, supplemented with mealworms.

Measuring offspring fitness

We recorded offspring SVL (mm) with digital callipers and mass (g) with a digital balance, both to the nearest 0.1, immediately post-hatching and then at weekly intervals for 24 weeks. We used SVL as a measure of growth rate rather than mass because it does not vary with factors such as changes in stomach contents and hydration status. Growth rate from hatching to sexual maturity in reptiles is typically initially linear, before beginning to plateau (Andrews, 1982). This was the case for our data, enabling us to use a simple measure of growth rate:

 $\frac{SVL(24\ weeks)-SVL(hatching)}{2}$. Further, overall body condition (i.e. relative body mass) at hatching 214 elapsed weeks 215 and 24 weeks of age was calculated by taking the residuals from a linear model of SVL and 216 mass. We recorded deaths in the population for 24 weeks and subsequently quantified the 217 proportion of each group surviving at the conclusion of each week to estimate survival curves. 218 For both sprint speed and bite force, each lizard was tested at 16 weeks post-hatching 219 during two trials on separate days which consisted of at least two repeated measures per trial. 220 Individuals were not tested for both bite force and sprint speed on the same day. Trials were 221 conducted in the middle of the light cycle (i.e. afternoon) to maximise the likelihood that they 222 had reached their optimal body temperature and reduce variance in temperature. Body 223 temperature was recorded via a thermocouple in the cloaca and the small variation in body 224 temperature did not predict maximum bite force or sprint speed, therefore it was subsequently dropped from analyses (sprint speed: $\chi^2 = 104.34$, d.f. = 136, p = 0.98; bite force: $\chi^2 = 23.87$, d.f.225 = 145, p = 1). To quantify morphometric variation relevant to performance, we measured the 226 227 following morphological characters: head width at the widest point of the head, head length from 228 the tip of the snout to the posterior of the jaw bone, head depth at the deepest point, jaw length 229 from the tip of the snout to the last supralabial scale, hind limb length from the body wall to the 230 edge of the foot pad, and SVL. All measurements were made using digital callipers to the nearest 231 0.01 mm. Head dimensions (width, length, depth, jaw) and hind limb length values were 232 regressed against SVL and the residuals were retained for use in subsequent analyses. Principal 233 components analysis (PCA) was performed on the residuals of head dimension measurements to 234 obtain one dimension representing the greatest variance in head shape (principal component 1, 235 PC1). PC1 explained 39.0% of variance and was driven primarily by head length and width 236 (Table S1). 237 Sprint trials were conducted in a 115 cm \times 10 cm \times 20 cm (L \times W \times D) racetrack on a 238 30° incline with 80-grit sandpaper as substrate to provide traction and to resemble natural 239 substrate (i.e. rock). A shelter, identical to that provided in their enclosure, was provided at the 240 end of the racetrack. If reluctant to run, lizards were induced to run by gently tapping on the 241 hindquarters using a paintbrush. Trials were filmed using a Canon EOS 600D camera (Canon 242 U.S.A., Inc., Lake Success, NY) at 50 frames per second and subsequently analysed; all trials 243 were conducted and analysed by one observer (C.M.D.). Interval speeds were calculated to 0.01 244

cm/s over each 10 cm section of the track from trials where the lizard ran continuously for ≥50

cm. Maximum interval speed from between four to six replicates per individual was retained for analysis.

In vivo bite forces were measured using an isometric Kistler Type 9203 force transducer mounted on a vertical holder and connected to a Kistler Type 5995A charge amplifier (Kistler Inc., Winterthur, Switzerland). Thin metal plates (i.e. bite plates) were attached to the force transducer and padded with medical tape on the outer surfaces to prevent damage to the teeth. The mouth of the lizard was opened by gently pulling on the gular region. The open jaws were placed onto the bite plates at the midway point of jaw length to standardize the point of force exertion; the position and angle of the head was kept as similar as possible across individuals (Lappin & Jones, 2014). This provoked voluntary forceful biting, otherwise the side of the mouth was tapped softly to induce biting. At least three recordings were taken per individual and the highest value was retained as the maximum bite force (Herrel et al., 1999).

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

245

246

247

248

249

250

251

252

253

254

255

256

Confirmation of paternity

Maternity was known for all offspring; however, paternity was not known due to multiple mates, and the potential for sperm storage and multiple paternity within clutches in the genus Ctenophorus (Hacking et al., 2017; Lebas, 2001; Mats Olsson et al., 2007, 2009; Uller et al., 2013). To confirm paternity, we genotyped adults and offspring at five microsatellite loci previously developed for C. decresii sensu lato (Ctde45, Ctde05, Ctde12; McLean et al. 2014) or C. pictus and Amphibolurus muricatus (CP10 and AM41, respectively; Schwartz et al. 2007). We collected blood for genetic samples by venipuncture from the sinus angularis in the corner of the mouth. Genomic DNA was extracted using an E.Z.N.A. Tissue DNA Kit (Omega Bio-tek, Norcross, GA, USA) and sent to the Australian Genome Research Facility (Melbourne, Victoria, AUS) for PCR amplification, fragment visualization and size calling. Paternity was assigned using CERVUS v 3.0.7 (Kalinowski et al., 2007; Marshall et al., 1998) which employs a maximum likelihood approach to assign the most likely candidate sire based on the genotypes of parents and offspring by estimating a likelihood-of-difference (LOD) score. First, we conducted a simulation of parentage analysis with the parameters of 50,000 offspring, 1% error rate, 5 candidate fathers, 95% of loci typed, and a minimum of 4 loci typed. Following this, we conducted a paternity analysis based on the trio (mother, father, and offspring) LOD score and a strict 95% confidence level (following Rankin et al. 2016). These were manually confirmed as

likely fathers based on breeding pairings. To prevent researcher bias, paternity was confirmed following the conclusion of growth and performance data collection.

277278

279

280

281

282

283

284

285

286

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

276

Statistical analyses

We conducted a linear discriminant analysis (LDA) to assess differentiation between the four classes of offspring (i.e. pure *C. decresii*, pure *C. modestus*, hybrids sired by *C. decresii* males, hybrids sired by *C. modestus* males). Offspring class was used as a dependent variable with fitness traits as independent variables (offspring class ~ bite force + sprint speed + growth rate + body condition at 24 weeks + body condition at hatching). Data were centred, scaled, and analysed using the R package *MASS* (v7.3.50; Ripley, 2002).

We used linear mixed models to examine the effects of parental species on parental reproductive traits (clutch size, embryonic mortality, seasonality of mating, egg mass at oviposition), offspring fitness traits (offspring mass at hatching, growth rate, bite force, sprint speed, body condition), and offspring morphology (SVL, hind limb length residuals, PC1 for head dimensions). Maternal species, paternal species, and their interaction were included as fixed terms and unique identifiers for the mother, father, and clutch were included as random-effect terms (response ~ maternal species + paternal species + maternal species *paternal species + (1|maternal ID) + (1|paternal ID) + (1|clutch ID)). For body condition, growth rate, bite force, and sprint speed, offspring sex and its interactions with parental species were included as fixed terms (response ~ maternal species + paternal species + sex + maternal species*paternal species + maternal species*sex + paternal species*sex + maternal species*paternal species*sex + (1|maternal ID) + (1|paternal ID) + (1|clutch ID)). For model simplification, we removed nonsignificant interactions with sex. We examined the residuals of each model for significant departures from normality and transformed the data if necessary. A Tukey-adjusted least-squares means for pairwise comparisons post hoc analysis was performed if an interaction term was statistically significant. One outlier was removed for clutch size and mass (8 eggs from a southern female which required medical intervention to remove them).

To investigate differences in sex ratios, we fitted a generalised linear mixed model with a binomial distribution and checked the data for overdisperson. Offspring sex was the response variable, parental species (maternal and paternal) and their interaction were fixed terms, and

clutch ID was a random-effect term (sex \sim maternal species + paternal species + maternal species *paternal species + $(1|\text{clutch_ID})$).

We examined the relationship between egg mass and hatchling mass based on maternal species using linear regression. To estimate total investment in egg production (i.e. fecundity) over the breeding season, we used the sum of egg masses produced by a female over the breeding season and fit a linear model with fecundity as the response variable and species as a fixed term. Desiccated clutches were included in this analysis only; to extrapolate, we assumed egg mass was the mean egg mass per clutch for each species because maternal SVL was not correlated with average egg mass in *C. modestus* ($F_{1, 15} = 0.01$, p = 0.924, $r^2 = 0.001$, slope \pm SE = -0.001 ± 0.01) or *C. decresii* ($F_{0.3, 9} = 0.33$ p = 0.582, $r^2 = 0.035$, slope \pm SE = 0.013 ± 0.02).

To examine offspring survival, we estimated survival curves using the Kaplan-Meier method and compared using a log-rank test (i.e. Mantel-Haenszel test) using the R packages *survival* (Therneau, 2015), and *survminer* (Kassambara & Kosinki, 2018).

Lastly, to investigate differences in adult male (sire) testes mass between species, we ran a linear model with testes mass as the response variable and species and SVL, and their interaction, as predictors (testes ~ species + SVL + species*SVL). The interaction was non-significant and was removed for model simplification.

All statistical analyses were conducted in R v. 3.5.1 (R Core Team 2018) using packages *factoMineR* (Lê et al., 2008), *MASS* (Venables & Ripley, 2002), *lme4* (Bates et al., 2015), *lmerTest* (Kuznetsova et al., 2017), and *lsmeans* (Lenth, 2016) unless otherwise stated.

Results

Offspring fitness

Linear discriminant analyses (LDA) showed significant differentiation between the four offspring classes (pure *C. modestus*, pure *C. decresii*, *C. modestus* $\mathcal{L} \times C$. decresii $\mathcal{L} \times C$. decresii $\mathcal{L} \times C$. modestus $\mathcal{L} \times C$: Wilks' $\mathcal{L} \times C$: Wilks

significant main effects of maternal species on growth rate and sprint speed, and effects of paternal species on bite force and body condition at 24 weeks of age in offspring (Table 2). Offspring produced by C. modestus females had a higher growth rate in the initial 24 weeks (p < 0.001; Table 2; Figure 3a; Figure S1) and had higher maximum sprint speeds (p = 0.007; Table 2; Figure 3b; Figure S1). Offspring sired by C. decresii males had higher maximum bite forces (p = 0.025; Table 2; Figure 3c; Figure S1) whereas those sired by C. modestus males had higher body condition at 24 weeks of age (p = 0.004; Table 2; Figure 3d; Figure S1).

There were no significant interactions between parental species for offspring fitness traits (growth rate, survival to 24 weeks, bite force, sprint speed, body condition) or morphological characters (SVL, hind limb length residuals, PC1 for head dimensions), indicating that these traits did not differ for pure and hybrid offspring. Additionally, there were no significant effects of parental species on body condition at hatching (p > 0.05; Table 2). There was a significant effect of sex on growth rate, bite force, body condition at hatching where measures for males were higher than for females (all p < 0.05; Table 2, Figure S2). Survivorship curves were not significantly different between offspring classes or by any combination of parental species (p = 0.89; Figure 3e; Table S2). Parental species did not influence any morphological traits (p > 0.05; Table S3).

Parental reproductive traits

All females were reproductively active: *C. modestus* females produced 32 clutches comprising 169 eggs and *C. decresii* females produced 21 clutches comprising 90 eggs. Of these, 5 clutches (29 eggs) and 7 clutches (27 eggs) produced by *C. modestus* and *C. decresii* females respectively were found desiccated outside of the nest box and excluded from all analyses except of fecundity. A higher proportion of *C. modestus* females multi-clutched with 66.7% and 11.1% laying second and third clutches respectively, whereas 23.5% of *C. decresii* females laid a second clutch and none laid a third clutch.

There were no significant interactions between parental species for reproductive fitness traits (fecundity, clutch size, embryonic mortality, egg mass at oviposition, and offspring mass at hatching), indicating that these traits did not differ for pure and hybrid pairings (Table 2; Figure S3). However, maternal species affected all of these traits (Table 2). Female C. modestus had overall higher fecundity over the breeding season (p = 0.016; Figure 4a). Female C. modestus

also produced larger clutches (p = 0.042; Table 2; Figure 4b) with lower embryonic mortality (p = 0.021; Figure 4c). Female *C. decresii* produced heavier eggs (p < 0.0001; Figure 4d) and correspondingly heavier hatchlings (p < 0.0001; Figure 4e). Additionally, for the relationship between egg and hatchling mass, there was a steeper slope and stronger correlation for *C. decresii* females (slope \pm SE = 1.12 ± 0.09 , $r^2 = 0.84$) than for *C. modestus* females (slope \pm SE = 0.60 ± 0.06 , $r^2 = 0.72$, p < 0.0001; Figure 4f).

Overall, 76.2% and 52.4% of *C. modestus* and *C. decresii* males were reproductively successful, siring 26 and 27 clutches total, respectively. There was no significant difference in adult male testes mass between the species (Table 2; Figure 4g). We detected a 2.4% rate of multiple paternity, occurring in one out of the 41 clutches genotyped. Specifically, a clutch produced by a *C. modestus* female was sired by two *C. modestus* males which were paired with the female consecutively in one reproductive cycle and fathered 40% and 60% of the clutch respectively. This low rate of multiple paternity is comparable to the 4.3% rate of multiple paternity estimated from a wild population of *C. modestus* (one out of 23 clutches; Hacking et al. 2017). Additionally, there was a significant effect of paternal species on sex ratios of clutches (*p* = 0.003; Table 3; Figure 4h; Figure S3) where clutches sired by *C. modestus* males had a sex ratio skewed towards males (1:1.64 female:male) whereas clutches sired by *C. decresii* males had a sex ratio skewed towards females (1:0.69 female:male) at an incubation temperature of 28°C.

Discussion

Empirical assessments of reproductive incompatibility and hybrid fitness are essential to interpret genomic patterns observed across naturally occurring contact zones. Previous studies of the contact zone between *Ctenophorus decresii* and *C. modestus* revealed a hybrid swarm with predominantly *C. modestus* mtDNA haplotypes and genetic incompatibilities when backcrossing to *C. decresii* (Dong et al., 2019). In order to investigate intrinsic reproductive barriers, we conducted captive-breeding experiments between individuals from pure populations nearest to the contact zone and measured offspring fitness traits. We found that *C. modestus* and *C. decresii* females differed in maternal reproductive investment and strategy, irrespective of the species of their mate. *Ctenophorus modestus* females exhibited higher fecundity over the breeding season and produced larger clutches with lower individual egg masses and more clutches per season;

Differences in maternal reproductive output may have broad ecological and evolutionary effects on populations by influencing offspring survival and fitness. Females of C. modestus produced a greater number of smaller offspring relative to C. decresii females, and this difference in maternal reproductive strategies reflects a fundamental reproductive trade-off between the number and size of eggs, with total clutch mass functionally constrained (Brockelman, 1975; Kaplan & Cooper, 1984; Smith & Fretwell, 1974). Egg and offspring size are intertwined with a range of factors such as survival and growth rate, and have fitness consequences due to the size-dependent nature of performance traits (Sinervo, 1990b, 1993). Additional differences in embryonic mortality, egg and hatching mass relationship, and sex ratio likely reflect differences in other unmeasured underpinning traits (e.g. yolk volume and content, gestation period, selection on incubation conditions). The differing reproductive strategies of the species could be the result of climate-driven selection. Climatic conditions of the higher latitudinal range of C. decresii (i.e. colder, temperate conditions) may place seasonal time constraints on their breeding season, restricting the time for embryo development and resulting in a higher investment in the first clutch. Comparatively, the range of C. modestus extends to a lower latitude (i.e. warmer, semi-arid conditions), which may allow for multi-clutching.

Maternal and paternal reproductive traits together may contribute to the prevalence of the C. modestus mtDNA haplotype within the contact zone. Based on our data, we estimate that the number of offspring produced by C. modestus females is at least 2–3 fold higher and with lower rates of embryonic mortality than C. decresii females. Additionally, in the context of hybrid crosses within the contact zone, C. modestus $\mathcal{P} \times C.$ decresii \mathcal{P} crosses will result in a higher proportion of female offspring that will pass on C. modestus mtDNA; however, this may vary

with differing incubation conditions in the wild. A similar pattern was found following gene flow between fishes (*Gambusia affnis* and *G. holbrooki*) where differences in parental life-history traits confer a competitive advantage (i.e. body size, growth rates, age at sexual maturity) to progeny with *G. holbrooki* maternal parentage (pure and F1 offspring; Scribner 1993). This competitive advantage correlated to patterns in experimental and natural populations where *G. holbrooki* mtDNA predominated (Scribner, 1993; Scribner & Avise, 1994). Similarly, hybrids of mulberry species (*Morus rubra* and *M. alba*) with *M. alba* maternal genotypes have a strong fitness advantage, whereas paternal genotype has no effect on fitness (Burgess & Husband, 2004). Further exploration is needed, but these maternal effects are predicted to influence the genetic composition of the hybrid zone between *M. rubra* and *M. alba*.

Contrary to our predictions, hybrids did not differ from pure offspring in individual fitness traits. Nonetheless, there was some evidence of differentiation between reciprocal hybrid classes when considering the full complement of fitness traits. Hybrids sired by *C. decresii* males had higher growth rates and sprint speeds (*via* maternal influences) and higher bite forces (*via* paternal influence). Pure *C. modestus* offspring had higher growth rates and sprint speeds but lower bite forces; pure *C. decresii* offspring had lower growth rates and sprint speeds but higher bite forces; and hybrids sired by *C. modestus* males had lower growth rates, sprint speeds, and bite forces. Based on combined traits, pure *C. modestus* may have higher fitness than pure *C. decresii* (higher values of both axes in the multivariate analysis of fitness); however, the relative fitness of parental and hybrid classes ultimately depends on how the traits we measured during early ontogeny affect adult fitness in the wild. Exogenous factors (e.g. genotype-environment interactions) may significantly influence the survival of reciprocal F1 hybrids, even in the absence of genomic incompatibilities (Kimball et al., 2008).

The fitness traits we measured are ecologically relevant and indicative of survival and reproductive success. Higher growth rates are likely advantageous given the direct effects on factors such as body size and age at sexual maturity (Andrews, 1982; Ballinger, 1983). Differences in growth rates, exclusive of exogenous influences, are likely to reflect genetic differences in physiology and/or behaviour (Dawson, 1975; Hutchinson & Maness, 1979; Porter & Travy, 1983; Sinervo, 1990a; Sinervo & Adolph, 1989). Further, performance traits (i.e. bite force and sprint speed) are tightly linked with competitive ability in ecologically relevant tasks such as predator evasion (survivorship), foraging and prey capture, and the ability to acquire and

defend territories (social dominance and reproductive success) in reptiles (locomotor performance: Garland, Jr. et al. 1990; Jayne and Bennett 1990; Robson and Miles 2000; Lailvaux et al. 2004; Perry et al. 2004; Husak 2006; Husak et al. 2006; Peterson and Husak 2006; reviewed in Husak and Fox 2008; bite force: Lailvaux et al. 2004; Huyghe et al. 2005; Lappin and Husak 2005; Anderson et al. 2008; Husak et al. 2009). These traits may be especially important determinants of male fitness in species with a polyandrous mating system where there is little evidence of female choice, such as *C. decresii and C. modestus*.

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490

Ctenophorus modestus $\mathcal{L} \times C$. decresii $\mathcal{L} \times C$. The hybrids exhibited higher trait values for more fitness measures than remaining offspring including the reciprocal hybrid cross, C. decresii $\mathcal{L} \times C$. modestus \mathcal{L} . Given these patterns, the composition of the F2 generation may be dominated by C. modestus mtDNA (progeny of hybrids sired by C. decresii males and/or pure C. modestus offspring). This is consistent with the observed prevalence of hybrids with C. modestus mtDNA in the contact zone; however, this is speculative because our study was limited to F1 hybrids due to the generation time of the species (≥ 2 years until sexual maturity). However, F2 hybrids often suffer strong negative fitness effects (hybrid breakdown) when recessive DMIs are more likely to manifest and/or cytonuclear interactions are further disrupted (Burton et al., 2006; Edmands, 1999; Ellison et al., 2008; Stelkens et al., 2015; Xue et al., 2019). Fitness can be rescued by backcrossing to the maternal species to restore parental nuclear and mitochondrial combinations, whereas backcrossing to the paternal species perpetuates the mismatch (Ellison & Burton, 2008). This is a conceivable explanation for the observed extensive backcrossing to C. modestus by post-F1 hybrids which virtually all have C. modestus mtDNA (Dong et al., 2019). We cannot exclude the possibility of sterility or reduced fertility of F1 hybrids with C. decresii mtDNA and/or the contribution of exogenous selection. Another contributor could be the mate preference of F1 hybrid males for females with C. modestus mtDNA (pure or hybrid) and/or preference of *C. modestus* females for hybrid males; however, F1 hybrid mate preferences in this system are unknown. It is difficult to draw inferences about mating behaviours based on cytonuclear patterns in hybrids because a variety of factors may produce these patterns. In other systems, cytonuclear patterns in hybrids have been attributed to differences in the abundance of each sex (Malmos et al., 2001; Wirtz, 1999) or species (Burgess et al., 2005) in the hybrid zone, marked differences in body size which only allow certain combinations (Grant & Grant, 1997;

491	Karl et al., 1995), and postmating prezygotic barriers such as conspecific sperm precedence
492	(Monsen et al., 2007).
493	In summary, our data indicate that differences in maternal reproductive investment and
494	strategy, together with parental effects on offspring fitness and sex ratio differences, may
495	contribute to patterns of genetic variation across the contact zone between C. decresii and C.
496	modestus, particularly the prevalence of C. modestus mtDNA. Stronger intrinsic genetic
497	incompatibilities between the two species may manifest primarily in the F2 generation.
498	Nevertheless, our results highlight how divergence between parental species in reproductive
499	traits, as well as effects on offspring fitness, can combine to shape contact zone dynamics.
500	
501	Acknowledgements
502	We are grateful to Adam Elliott for fieldwork assistance and lizard husbandry advice; Eliza
503	Thompson, Emma Dragh, Hannah Edwards, Lucy Howell, Miriam Pastor, and Murraya Lane for
504	assistance with captive animal care; Po Peng for statistical support; private landowners for access
505	to their property for data collection; Adnan Moussalli and two anonymous reviewers whose
506	comments helped to improve the manuscript. This research was conducted with permits from the
507	Department of Environment, Water and Natural Resources, South Australia (permit Q26428-3),
508	the Department of Environment, Land, Water and Planning, Victoria (permit 10007751) and
509	with approval by The University of Melbourne Animals Ethics Committee (approval
510	1413220.4), and the Wildlife Ethics Committee of South Australia (approval 25/2015). DS-F
511	was supported by the Australian Research Council (DP150101048) and CMD was supported by
512	The David Lachlan Hay Memorial Fund. The authors declare no competing or financial interests
513	
514	Data Accessibility
515	Data are available from the Dryad Digital Repository: https://doi.org/10.5061/dryad.p5hqbzkp1
516	
517	Literature Cited
518	Abbott, R., Albach, D., Ansell, S., Arntzen, J. W., Baird, S. J. E., Bierne, N., Boughman, J.,
519	Brelsford, A., Buerkle, C. A., Buggs, R., Butlin, R. K., Dieckmann, U., Eroukhmanoff, F.,
520	Grill, A., Cahan, S. H., Hermansen, J. S., Hewitt, G., Hudson, A. G., Jiggins, C., Zinner,
521	D. (2013). Hybridization and speciation. <i>Journal of Evolutionary Biology</i> , 26(2), 229–246.

522	https://doi.org/10.1111/j.1420-9101.2012.02599.x
523	Anderson, R. A., McBrayer, L. D., & Herrel, A. (2008). Bite force in vertebrates: Opportunities
524	and caveats for use of a nonpareil whole-animal performance measure. Biological Journal
525	of the Linnean Society, 93(4), 709–720. https://doi.org/10.1111/j.1095-8312.2007.00905.x
526	Andrews, R. M. (1982). Patterns of growth in reptiles. In C. Gans & F. H. Pough (Eds.), Biology
527	of the Reptilia (Vol. 13, pp. 273-320). Academic Press.
528	Arnold, M. L., Ballerini, E. S., & Brothers, A. N. (2012). Hybrid fitness, adaptation and
529	evolutionary diversification: Lessons learned from Louisiana Irises. Heredity, 108(3), 159-
530	166. https://doi.org/10.1038/hdy.2011.65
531	Arntzen, J. W., Jehle, R., Bardakci, F., Burke, T., & Wallis, G. P. (2009). Asymmetric viability
532	of reciprocal-cross hybrids between crested and marbled newts (Triturus cristatus and T.
533	marmoratus). Evolution, 63(5), 1191–1202. https://doi.org/10.1111/j.1558-
534	5646.2009.00611.x
535	Ballinger, R. (1983). Life-history variations. In R. B. Huey, E. R. Pianka, & T. W. Schoener
536	(Eds.), Lizard ecology: studies of a model organism (pp. 241-260). Harvard University
537	Press.
538	Barton, N. H., & Hewitt, G. M. (1985). Analysis of hybrid zones. Annual Review of Ecology and
539	Systematics, 16(1), 113-148. https://doi.org/10.1146/annurev.ecolsys.16.1.113
540	Bates, D., Mächler, M., Bolker, B. M., & Walker, S. C. (2015). Fitting linear mixed-effects
541	models using lme4. Journal of Statistical Software, 67(1), 1-48.
542	https://doi.org/10.18637/jss.v067.i01
543	Bolnick, D. I., & Near, T. J. (2005). Tempo of hybrid inviability in centrarchid fishes (Teleostei:
544	Centrarchidae). Evolution, 59(8), 1754-1767. https://doi.org/10.1111/j.0014-
545	3820.2005.tb01824.x
546	Bolnick, D. I., Turelli, M., López-Fernández, H., Wainwright, P. C., & Near, T. J. (2008).
547	Accelerated mitochondrial evolution and "Darwin's corollary": Asymmetric viability of
548	reciprocal F1 hybrids in centrarchid fishes. Genetics, 178(2), 1037-1048.
549	https://doi.org/10.1534/genetics.107.081364

Brockelman, W. Y. (1975). Competition, the fitness of offspring, and optimal clutch size. *The*

Boughman, J. W. (2001). Divergent sexual selection enhances reproductive isolation in

sticklebacks. Nature, 411(6840), 944-948. https://doi.org/10.1038/35082064

550

553	American I	Naturalist.	109(970).	677–699.

- Burgess, K. S., Morgan, M., Deverno, L., & Husband, B. C. (2005). Asymmetrical introgression
- between two *Morus* species (M. alba, M. rubra) that differ in abundance. *Molecular*
- 556 Ecology, 14(11), 3471–3483. https://doi.org/10.1111/j.1365-294X.2005.02670.x
- Burgess, Kevin S., & Husband, B. C. (2004). Maternal and paternal contributions to the fitness
- of hybrids between red and white mulberry (Morus, Moraceae). American Journal of
- 559 Botany, 91(11), 1802–1808. https://doi.org/10.3732/ajb.91.11.1802
- Burton, C. K. Ellison, & Harrison. (2006). The sorry state of F2 hybrids: consequences of rapid
- mitochondrial DNA evolution in allopatric populations. *The American Naturalist*, 168(6),
- 562 S14–S24. https://doi.org/10.2307/4122277
- Butlin, R. (1987). Speciation by reinforcement. *Trends in Ecology and Evolution*, 2(1), 8–13.
- 564 https://doi.org/10.1016/0169-5347(87)90193-5
- 565 Campbell, D. R., & Waser, N. M. (2001). Genotype-by-environment interaction and the fitness
- of plant hybrids in the wild. *Evolution*, 55(4), 669. https://doi.org/10.1554/0014-
- 567 3820(2001)055[0669:gbeiat]2.0.co;2
- 568 Corl, A., Davis, A. R., Kuchta, S. R., & Sinervo, B. (2010). Selective loss of polymorphic mating
- 569 types is associated with rapid phenotypic evolution during morphic speciation. *Proceedings*
- of the National Academy of Sciences, 107(9), 4254–4259.
- 571 https://doi.org/10.1073/pnas.0909480107
- 572 Coyne, J. A., & Orr, H. A. (2004). Speciation. In *Sinauer Associates*. Sinauer Associates.
- Dawson, W. R. (1975). On the physiological significance of the preferred temperatures of
- reptiles. In D. M. Gates & R. B. Schmerl (Eds.), *Perspectives in Biophysical Ecology*.
- 575 Ecological Studies (Analysis and Synthesis) (12th ed., pp. 443–473). Springer.
- Dobzhansky, T. (1937). Genetics the Origin and of Species. In Genetics and the origin of
- 577 *species*. Columbia University Press.
- 578 Dong, C. M., Johnston, G. R., Stuart-Fox, D., Moussalli, A., Rankin, K. J., & McLean, C. A.
- 579 (2021). Elevation of divergent color polymorphic and monomorphic lizard lineages
- (Squamata: Agamidae) to species level. *Ichthyology and Herpetology*, 109(1).
- 581 https://doi.org/10.1643/h2020064
- Dong, C. M., McLean, C. A., Elliott, A., Moussalli, A., & Stuart-Fox, D. (2019). When
- polymorphism and monomorphism meet: discordant genomic and phenotypic clines across

584	a lizard contact zone.	BioRxiv.	840678, htt	tps://doi.org/	10.1101/840678

- 585 Edmands, S. (1999). Heterosis and outbreeding depression in interpopulation crosses spanning a
- wide range of divergence. *Evolution*, 53(6), 1757. https://doi.org/10.2307/2640438
- 587 Ellison, C. K., Niehuis, O., & Gadau, J. (2008). Hybrid breakdown and mitochondrial
- dysfunction in hybrids of *Nasonia* parasitoid wasps. *Journal of Evolutionary Biology*, 21(6),
- 589 1844–1851. https://doi.org/10.1111/j.1420-9101.2008.01608.x
- 590 Ellison, Christopher K., & Burton, R. S. (2008). Interpopulation hybrid breakdown maps to the
- mitochondrial genome. *Evolution*, 62(3), 631–638. https://doi.org/10.1111/j.1558-
- 592 5646.2007.00305.x
- 593 Forsman, A., Ahnesjö, J., Caesar, S., & Karlsson, M. (2008). A model of ecological and
- evolutionary consequences of color polymorphism. *Ecology*, 89(1), 34–40.
- 595 https://doi.org/10.1890/07-0572.1
- Garland, Jr., T., Hankins, E., & Huey, R. B. (1990). Locomotor capacity and social dominance in
- male lizards. Functional Ecology, 4(2), 243–250.
- 598 Gibbons, J. R. H. (1977). Comparative ecology and behaviour of lizards of the Amphibolurus
- decresii *species complex*. The University of Adelaide.
- 600 Grant, P. R., & Grant, R. B. (1997). Hybridization, sexual imprinting, and mate choice. *The*
- 601 *American Naturalist*, 149(1), 1–28.
- 602 Gray, S. M., & McKinnon, J. S. (2007). Linking color polymorphism maintenance and
- speciation. Trends in Ecology & Evolution, 22(2), 71–79.
- https://doi.org/10.1016/j.tree.2006.10.005
- Hacking, J., Stuart-Fox, D., & Gardner, M. (2017). Very low rate of multiple paternity detected
- in clutches of a wild agamid lizard. Australian Journal of Zoology, 65(5), 328–334.
- 607 https://doi.org/10.1071/ZO18006
- Herrel, A., Spithoven, L., Van Damme, R., & De Vree, F. (1999). Sexual dimorphism of head
- size in *Gallotia galloti*: Testing the niche divergence hypothesis by functional analyses.
- *Functional Ecology*, 13(3), 289–297. https://doi.org/10.1046/j.1365-2435.1999.00305.x
- Hoskin, C. J., Higgie, M., McDonald, K. R., & Moritz, C. (2005). Reinforcement drives rapid
- allopatric speciation. *Nature*, 437, 1353–1356. https://doi.org/doi:10.1038/nature04004
- Howard, D. J. (1993). Reinforcement: Origin, dynamics, and fate of an evolutionary hypothesis.
- In R. G. Harrison (Ed.), Hybrid Zones and the Evolutionary Process (pp. 46–69). Oxford

- University Press.
- Hugall, A. F., & Stuart-Fox, D. (2012). Accelerated speciation in colour-polymorphic birds.
- 617 *Nature*, 485, 631–634. https://doi.org/10.1038/nature11050
- Husak, J F. (2006). Does speed help you survive? A test with collared lizards. Functional
- 619 Ecology, 20, 174–179. https://doi.org/10.1111/j.1365-2435.2006.01069.x
- Husak, Jerry F., & Fox, S. F. (2008). Sexual selection on locomotor performance. *Evolutionary*
- 621 Ecology Research, 10(2), 213–228.
- Husak, Jerry F., Fox, S. F., Lovern, M. B., & Van Den Bussche, R. A. (2006). Faster lizards sire
- more offspring: sexual selection on whole-animal performance. *Evolution*, 60(10), 2122.
- 624 https://doi.org/10.1554/05-647.1
- Husak, Jerry F., Lappin, K. A., Van Den Bussche, R. A., Lappin, A. K., & Van Den Bussche, R.
- A. (2009). The fitness advantage of a high-performance weapon. *Biological Journal of the*
- 627 Linnean Society, 96(4), 840–845. https://doi.org/10.1111/j.1095-8312.2008.01176.x
- Hutchinson, V. H., & Maness, J. H. (1979). The role of behaviour in temperature acclimation
- and tolerance in ectotherms. *American Zoologist*, 19, 367–384.
- Huxel, G. R. (1999). Rapid displacement of native species by invasive species: Effects of
- hybridization. *Biological Conservation*, 89(2), 143–152. https://doi.org/10.1016/S0006-
- 632 3207(98)00153-0
- Huyghe, K., Vanhooydonck, B., Scheers, H., Molina-Borja, M., & Van Damme, R. (2005).
- Morphology, performance and fighting capacity in male lizards, *Gallotia galloti. Functional*
- 635 Ecology, 19(5), 800–807. https://doi.org/10.1111/j.1365-2435.2005.01038.x
- 636 Irschick, D. J., Herrel, A., Vanhooydonck, B., & Van Damme, R. (2007). A functional approach
- 637 to sexual selection. Functional Ecology, 21(4), 621–626. https://doi.org/10.1111/j.1365-
- 638 2435.2007.01281.x
- Jayne, B. C., & Bennett, A. F. (1990). Selection on locomotor performance capacity in a natural
- population of garter snakes. *Evolution*, 44(5), 1204–1229. https://doi.org/10.2307/2409283
- Jiggins, C. D., Naisbit, R. E., Coe, R. L., & Mallet, J. (2001). Reproductive isolation caused by
- colour pattern mimicry. *Nature*, 411(6835), 302–305. https://doi.org/10.1038/35077075
- Kalinowski, S. T., Taper, M. L., & Marshall, T. C. (2007). Revising how the computer program
- 644 CERVUS accommodates genotyping error increases success in paternity assignment.
- 645 Molecular Ecology, 16(5), 1099–1106. https://doi.org/10.1111/j.1365-294X.2007.03089.x

- Kaplan, R. H., & Cooper, W. S. (1984). The evolution of developmental plasticity in
- reproductive characteristics: an application of the "adaptive coin-flipping" principle. *The*
- 648 *American Naturalist*, 123(3), 393–410.
- Karl, S. A., Bowen, B. W., & Avise, J. C. (1995). Hybridization among the ancient mariners:
- 650 Characterization of marine turtle hybrids with molecular genetic assays. *Journal of*
- 651 Heredity, 86, 262–268. https://doi.org/10.1142/9789814317764 0004
- Kassambara, A., & Kosinki, M. (2018). Drawing survival curves using "ggplot2."
- Kimball, S., Campbell, D. R., & Lessin, C. (2008). Differential performance of reciprocal
- 654 hybrids in multiple environments. *Journal of Ecology*, 96(6), 1306–1318.
- https://doi.org/10.1111/j.1365-2745.2008.01432.x
- Kuznetsova, A., Brockhoff, P. B., & Christensen, R. H. B. (2017). lmerTest Package: tests in
- linear mixed effects models. *Journal of Statistical Software*, 82(13), 1–26.
- 658 https://doi.org/10.18637/jss.v082.i13
- Lailvaux, S. P., Herrel, A., VanHooydonck, B., Meyers, J. J., & Irschick, D. J. (2004).
- Performance capacity, fighting tactics and the evolution of life-stage male morphs in the
- green anole lizard (Anolis carolinensis). Proceedings of the Royal Society B: Biological
- Sciences, 271(1556), 2501–2508. https://doi.org/10.1098/rspb.2004.2891
- Lappin, A. K., & Jones, M. E. H. (2014). Reliable quantification of bite-force performance
- requires use of appropriate biting substrate and standardization of bite out-lever. *Journal of*
- 665 Experimental Biology, 217(24), 4303–4312. https://doi.org/10.1242/jeb.106385
- 666 Lappin, K. A., & Husak, J. F. (2005). Weapon performance, not size, determines mating success
- and potential reproductive output in the collared lizard (*Crotaphytus collaris*). The
- 668 American Naturalist, 166(3), 426–436. https://doi.org/10.1086/432564
- 669 Lê, S., Josse, J., & Husson, F. F. (2008). FactoMineR: An R Package for Multivariate Analysis.
- 670 Journal of Statistical Software, 25(1), 1–18. https://doi.org/10.1016/j.envint.2008.06.007
- 671 Lebas, N. R. (2001). Microsatellite determination of male reproductive success in a natural
- population of the territorial ornate dragon lizard, Ctenophorus ornatus. Molecular Ecology,
- 673 10(1), 193–203. https://doi.org/10.1046/j.1365-294X.2001.01174.x
- 674 Lenth, R. V. (2016). Least-squares means: The R package Ismeans. *Journal of Statistical*
- 675 Software, 69(1), 1–33. https://doi.org/10.18637/jss.v069.i01
- MacGregor, H. E. A., While, G. M., Barrett, J., Pérez i de Lanuza, G., Carazo, P., Michaelides,

- S., & Uller, T. (2017). Experimental contact zones reveal causes and targets of sexual
- selection in hybridizing lizards. *Functional Ecology*, 31(3), 742–752.
- 679 https://doi.org/10.1111/1365-2435.12767
- 680 Mallet, J. (2007). Hybrid speciation. *Nature*, 446(7133), 279–283.
- 681 https://doi.org/10.1038/nature05706
- Malmos, K. B., Sullivan, B. K., & Lamb, T. (2001). Calling behavior and directional
- 683 hybridization between two toads (*Bufo microscaphus* x *B. woodhousii*) in Arizon.
- 684 Evolution, 55(3), 626–630.
- Marshall, T. C., Slate, J., Kruuk, L. E. B., & Pemberton, J. M. (1998). Statistical confidence for
- likelihood-based paternity inference in natural populations. *Molecular Ecology*, 7, 639–655.
- Mavárez, J., Salazar, C. A., Bermingham, E., Salcedo, C., Jiggins, C. D., & Linares, M. (2006).
- Speciation by hybridization in *Heliconius* butterflies. *Nature*, 441(7095), 868–871.
- 689 https://doi.org/10.1038/nature04738
- McLean, C. A., Bartle, R. A., Dong, C. M., Rankin, K. J., & Stuart-Fox, D. (2020). Divergent
- male and female mate preferences do not explain incipient speciation between lizard
- lineages. Current Zoology, March, 1–8. https://doi.org/10.1093/cz/zoaa010
- McLean, C. A., & Stuart-Fox, D. (2014). Geographic variation in animal colour polymorphisms
- and its role in speciation. *Biological Reviews*, 89(4), 860–873.
- 695 https://doi.org/10.1111/brv.12083
- McLean, C. A., Stuart-Fox, D., & Moussalli, A. (2014). Phylogeographic structure, demographic
- history and morph composition in a colour polymorphic lizard. *Journal of Evolutionary*
- 698 *Biology*, 27(10), 2123–2137. https://doi.org/10.1111/jeb.12464
- Monsen, K. J., Honchak, B. M., Locke, S. E., & Peterson, M. A. (2007). Cytonuclear
- disequilibrium in Chrysochus hybrids is not due to patterns of mate choice. *Journal of*
- 701 *Heredity*, 98(4), 325–330. https://doi.org/10.1093/jhered/esm039
- Muir, G., Ruiz-Duarte, P., Hohmann, N., Mable, B. K., Novikova, P., Schmickl, R., Guggisberg,
- A., & Koch, M. A. (2015). Exogenous selection rather than cytonuclear incompatibilities
- shapes asymmetrical fitness of reciprocal Arabidopsis hybrids. Ecology and Evolution, 5(8),
- 705 1734–1745. https://doi.org/10.1002/ece3.1474
- Muller, H. J. (1942). Isolating mechanisms, evolution, and temperature. *Biol Symp*, 6, 71–125.
- Noor, M. A. F. F. (1999). Reinforcement and other consequences of sympatry. *Heredity*, 83(5),

- 708 503–508. https://doi.org/10.1038/sj.hdy.6886320
- Nosil, P., Harmon, L. J., & Seehausen, O. (2009). Ecological explanations for (incomplete)
- speciation. Trends in Ecology and Evolution, 24(3), 145–156.
- 711 https://doi.org/10.1016/j.tree.2008.10.011
- 712 Olsson, M, & Madsen, T. (1998). Sexual selection and sperm competition in reptiles. In T. R.
- Birkhead & A. P. Moller (Eds.), *Sperm competition and sexual selection* (pp. 503–578).
- 714 Academic Press.
- Olsson, Mats, Schwartz, T., Uller, T., & Healey, M. (2007). Sons are made from old stores:
- sperm storage effects on sex ratio in a lizard. *Biology Letters*, 3(5), 491–493.
- 717 https://doi.org/10.1098/rsbl.2007.0196
- 718 Olsson, Mats, Schwartz, T., Uller, T., & Healey, M. (2009). Effects of sperm storage and male
- 719 colour on probability of paternity in a polychromatic lizard. Animal Behaviour, 77(2), 419–
- 720 424. https://doi.org/10.1016/j.anbehav.2008.10.017
- Olsson, Mats, & Shine, R. (1997). Advantages of multiple matings to females: a test of the
- 722 infertility hypothesis using lizards. *Evolution*, 51(5), 1684–1688.
- 723 https://www.jstor.org/stable/2411220
- Orr, M. R. (1996). Life-history adaptation and reproductive isolation in a grasshopper hybrid
- 725 zone. *Evolution*, 50(2), 704–716.
- Ortiz-Barrientos, D., Grealy, A., & Nosil, P. (2009). The genetics and ecology of reinforcement:
- 727 implications for the evolution of prezygotic isolation in sympatry and beyond. *Annals of the*
- 728 New York Academy of Sciences, 1168, 158–182. https://doi.org/10.1111/j.1749-
- 729 6632.2009.04919.x
- Perry, G., Levering, K., Girard, I., Garland, T., & Garland, Jr., T. (2004). Locomotor
- performance and social dominance in male *Anolis cristatellus*. *Animal Behaviour*, 67(1),
- 732 37–47. https://doi.org/10.1016/j.anbehav.2003.02.003
- Peterson, C. C., & Husak, J. F. (2006). Locomotor performance and sexual selection: individual
- variation in sprint speed of collared lizards (*Crotaphytus collaris*). *Copeia*, 3(4), 216–224.
- 735 https://doi.org/10.1643/0045-8511(2006)6
- Peterson, M. A., Monsen, K. J., Pedersen, H., McFarland, T., & Bearden, J. (2005). Direct and
- 737 indirect analysis of the fitness of *Chrysochus* (Coleoptera: Chrysomelidae) hybrids.
- 738 Biological Journal of the Linnean Society, 84(2), 273–286. https://doi.org/10.1111/j.1095-

- 739 8312.2004.00429.x
- 740 Pfennig, K. S. (2016). Reinforcement as an initiator of population divergence and speciation.
- 741 *Current Zoology*, 62(2), 145–154. https://doi.org/10.1093/cz/zow033
- Porter, W. P., & Travy, C. R. (1983). Biophysical analyses of energetics, time-space utilization,
- and distributional limits. In R. B. Huey, E. R. Pianka, & T. W. Schoener (Eds.), *Lizard*
- 744 *ecology: studies of a model organism* (pp. 55–83). Harvard University Press.
- Rank, N. E., Mardulyn, P., Heidl, S. J., Roberts, K. T., Zavala, N. A., Smiley, J. T., & Dahlhoff,
- E. P. (2020). Mitonuclear mismatch alters performance and reproductive success in
- naturally introgressed populations of a montane leaf beetle. *Evolution*, 1–17.
- 748 https://doi.org/10.1111/evo.13962
- Rankin, K. J., McLean, C. A., Kemp, D. J., & Stuart-Fox, D. (2016). The genetic basis of
- discrete and quantitative colour variation in the polymorphic lizard, *Ctenophorus decresii*.
- 751 *BMC Evolutionary Biology*, 16(1), 1–14. https://doi.org/10.1186/s12862-016-0757-2
- Rhymer, J. M., & Simberloff, D. (1996). Extinction by hybridization and introgression. *Annual*
- 753 Review of Ecology and Systematics, 27, 83–109.
- 754 https://doi.org/10.1146/annurev.ecolsys.27.1.83
- Robson, M. A., & Miles, D. B. (2000). Locomotor performance and dominance in male Tree
- 756 Lizards, *Urosaurus ornatus*. Functional Ecology, 14, 338–344.
- Rosenfield, J. A., Nolasco, S., Lindauer, S., Sandoval, C., & Kodric-Brown, A. (2004). The role
- of hybrid vigor in the replacement of pecos pupfish by its hybrids with sheepshead minnow.
- 759 Conservation Biology, 18(6), 1589–1598. https://doi.org/10.1111/j.1523-1739.2004.00356.x
- Sætre, G. P., Moum, T., Bureš, S., Král, M., Adamjan, M., Moreno, J., Ultrahigh, S. M.,
- Metamorphism, P., Sætre, G. P., Moum, T., Saetre, G.-P., Moum, T., Bures, S., Král, M.,
- Adamjan, M., & Moreno, J. (1997). A sexually selected character displacement in
- flycatchers reinforces premating isolation. *Nature*, 387(June), 589–592.
- 764 https://doi.org/10.1038/42451
- Sanger, T. J., Hime, P. M., Johnson, M. A., Diani, J., & Losos, J. B. (2008). Laboratory protocols
- for husbandry and embryo collection of *Anolis* lizards. *Herpetological Review*, 39(1), 58–
- 767 63.
- Schwartz, T. S., Warner, D. A., Beheregaray, L. B., Olsson, M., & Note, P. (2007).
- Microsatellite loci for Australian agamid lizards. *Molecular Ecology Notes*, 7(3), 528–531.

- 770 https://doi.org/10.1111/j.1471-8286.2006.01644.x
- 771 Scribner, K. I. M. T. (1993). Hybrid zone dynamics are influenced by genotype-specific
- variation in life-history traits: experimental evidence from hybridizing *Gambusia* species.
- 773 Evolution, 47(2), 632–646.
- Scribner, K. T., & Avise, J. C. (1994). Population cage experiments with a vertebrate: the
- temporal demography and cytonuclear genetics of hybridization in *Gambusia* fishes.
- 776 Evolution, 48(1), 155. https://doi.org/10.2307/2410011
- Seehausen, O., Terai, Y., Magalhaes, I. S., Carleton, K. L., Mrosso, H. D. J., Miyagi, R., van der
- Sluijs, I., Schneider, M. V, Maan, M. E., Tachida, H., Imai, H., & Okada, N. (2008).
- Speciation through sensory drive in cichlid fish. *Nature*, 455(7213), 620–627.
- 780 https://doi.org/10.1038/nature07285
- Servedio, M. R. (2004). The what and why of research on reinforcement. *PLoS Biology*, 2(12),
- 782 2032–2035. https://doi.org/10.1371/journal.pbio.0020420
- Servedio, M. R., & Noor, M. A. F. F. (2003). The role of reinforcement in speciation: theory and
- data. Annual Review of Ecology, Evolution, and Systematics, 34(Noor 1999), 339–364.
- 785 https://doi.org/10.1146/annurev.ecolsys.34.011802.132412
- Sinervo, B., & Svensson, E. (2002). Correlational selection and the evolution of genomic
- 787 architecture. *Heredity*, 89(5), 329–338. https://doi.org/10.1038/sj.hdy.6800148
- Sinervo, Barry. (1990a). Evolution of thermal physiology and growth rate between populations
- of the western fence lizard (Sceloporus occidentalis). Oecologia, 83(2), 228–237.
- 790 https://doi.org/10.1007/BF00317757
- 791 Sinervo, Barry. (1990b). The evolution of maternal investment in lizards: an experimental and
- comparative analysis of egg size and its effects on offspring performance. Evolution, 44(2),
- 793 279–294.
- Sinervo, Barry. (1993). The effect of offspring size on physiology and life history. *BioScience*,
- 795 *43*(4), 210–218. https://doi.org/10.2307/1312121
- 796 Sinervo, Barry, & Adolph, S. C. (1989). Thermal Sensitivity of Growth Rate in Hatchling
- 797 Sceloporus Lizards. Oecologica, 78(3), 411–419. http://www.jstor.org/stable/4218883
- 798 Accessed:
- 799 Smith, C. C., & Fretwell, S. D. (1974). The optimal balance between size and number of
- offspring. The American Naturalist, 108(962), 499–506. https://doi.org/10.2307/2459681

Stelkens, R. B., Schmid, C., & Seehausen, O. (2015). Hybrid breakdown in cichl	d fish. <i>PLoS</i>
--	---------------------

- 802 *ONE*, 10(5), 1–11. https://doi.org/10.1371/journal.pone.0127207
- Taylor, E. B., Boughman, J. W., Groenenboom, M., Sniatynski, M., Schluter, D., & Gow, J. L.
- 804 (2006). Speciation in reverse: Morphological and genetic evidence of the collapse of a
- three-spined stickleback (Gasterosteus aculeatus) species pair. Molecular Ecology, 15(2),
- 806 343–355. https://doi.org/10.1111/j.1365-294X.2005.02794.x
- Teasdale, L. C., Stevens, M., & Stuart-Fox, D. (2013). Discrete colour polymorphism in the
- tawny dragon lizard (*Ctenophorus decresii*) and differences in signal conspicuousness
- among morphs. *Journal of Evolutionary Biology*, 26(5), 1035–1046.
- 810 https://doi.org/10.1111/jeb.12115
- Therneau, T. M. (2015). A package for survival analysis in S. version 2.3.8, https://cran.r-
- project.org/package=survival (2.38). https://cran.r-project.org/package=survival
- Tiffin, P., Olson, M. S., & Moyle, L. C. (2001). Asymmetrical crossing barriers in angiosperms.
- *Proceedings of the Royal Society B: Biological Sciences*, 268(1469), 861–867.
- https://doi.org/10.1098/rspb.2000.1578
- Turelli, M., Barton, N. H., & Coyne, J. A. (2001). Theory and speciation. *Trends in Ecology &*
- 817 Evolution, 16(7), 330–343. https://doi.org/10.1016/S0169-5347(01)02177-2
- Uller, T., Schwartz, T., Koglin, T., & Olsson, M. (2013). Sperm storage and sperm competition
- across ovarian cycles in the dragon lizard, Ctenophorus fordi. Journal of Experimental
- 820 Zoology, 319A, 404–408. https://doi.org/10.1002/jez.1803
- Venables, W. N., & Ripley, B. D. (2002). *Modern Applied Statistics with S* (Fourth). Springer.
- http://www.stats.ox.ac.uk/pub/MASS4
- Wainwright, P. C. (2007). Functional versus morphological diversity in macroevolution. *Annual*
- *Review of Ecology, Evolution, and Systematics*, 38(1), 381–401.
- https://doi.org/10.1146/annurev.ecolsys.38.091206.095706
- Wesselingh, R. A., Hořčicová, Š., & Mirzaei, K. (2019). Fitness of reciprocal F1 hybrids
- between *Rhinanthus minor* and *Rhinanthus major* under controlled conditions and in the
- field. Journal of Evolutionary Biology, 32, 931–942. https://doi.org/10.1111/jeb.13492
- West-Eberhard, M. J. (1986). Alternative adaptations, speciation, and phylogeny (A Review).
- *Proceedings of the National Academy of Sciences*, 83, 1388–1392.
- https://doi.org/10.1073/pnas.83.5.1388

Ę	
	2
	-
	<

832	While, G. M., Michaelides, S., Heathcote, R. J. P., Macgregor, H. E. A., Zajac, N., Beninde, J.,
833	Carazo, P., Pérez i de Lanuza, G., Sacchi, R., Zuffi, M. A. L., Horváthová, T., Fresnillo, B
834	Schulte, U., Veith, M., Hochkirch, A., & Uller, T. (2015). Sexual selection drives
835	asymmetric introgression in wall lizards. Ecology Letters, 18(12), 1366–1375.
836	https://doi.org/10.1111/ele.12531
837	Wirtz, P. (1999). Mother species-father species: Unidirectional hybridization in animals with
838	female choice. Animal Behaviour, 58(1), 1-12. https://doi.org/10.1006/anbe.1999.1144
839	Xue, W., Anderson, S. N., Wang, X., Yang, L., Crisp, P. A., Li, Q., Noshay, J., Albert, P. S.,
840	Birchler, J. A., Bilinski, P., Stitzer, M. C., Ross-Ibarra, J., Flint-Garcia, S., Chen, X.,
841	Springer, N. M., & Doebley, J. F. (2019). Hybrid decay: a transgenerational epigenetic
842	decline in vigor and viability triggered in backcross populations of teosinte with maize.
843	Genetics, 213, 143-160. https://doi.org/10.1534/genetics.119.302378
844	

848

849

	LD1	LD2
Bite Force	-0.531	-0.107
Sprint Speed	0.385	0.609
Growth Rate	0.013	0.620
Body Condition		
at 24 Weeks	0.831	0.111
Body Condition		
at Hatching	0.550	-0.577
Proportion of		
variance	0.533	0.439

851

852

853

854

855

856

857

Table 2. Results of a linear model of female fecundity and species, and linear mixed-effects models investigating the effects of parental species on reproductive traits (embryonic mortality, egg mass at oviposition, offspring mass at hatching) and on offspring fitness traits (growth rate, maximum sprint speed, maximum bite force, body condition at hatching and 24 weeks of age). Statistically significant values are indicated with asterisks (* = p < 0.05, ** = p < 0.01, *** = p < 0.001).

Response	Fixed Term	Sum Sq	Mean Sq	d.f.	DenDF	F	p
Growth Rate	Maternal	0.13	0.13	1	36.37	5.14	0.029*
	Paternal	0.00	0.00	1	26.77	0.03	0.854
_	$M \times P$	0.01	0.01	1	39.16	0.41	0.527
	Sex	0.12	0.12	1	125.19	4.59	0.034*
Sprint Speed	Maternal	8370.10	8370.10	1	30.27	9.45	0.004**
	Paternal	2402.10	2402.10	1	25.08	2.71	0.112
	$\mathbf{M} \times \mathbf{P}$	408.10	408.10	1	32.62	0.46	0.502
	Sex	1.30	1.30	1	114.35	0.00	0.970
Bite Force	Maternal	0.47	0.47	1	35.89	1.01	0.323
	Paternal	2.43	2.43	1	34.08	5.24	0.028*

	$\mathbf{M} \times \mathbf{P}$	0.24	0.24	1	38.72	0.51	0.478
	Sex	2.70	2.70	1	128.55	5.82	0.017*
Body Condition	Maternal	0.02	0.02	1	16.12	1.96	0.180
at Hatching	Paternal	0.01	0.01	1	25.74	1.09	0.305
	$\mathbf{M} \times \mathbf{P}$	0.00	0.00	1	24.94	0.37	0.547
+	Sex	0.06	0.06	1	124.30	6.88	0.010**
Body Condition	Maternal	0.02	0.02	1	19.49	0.12	0.737
at 24 Weeks	Paternal	1.30	1.30	1	22.40	9.91	0.005**
	$\mathbf{M} \times \mathbf{P}$	0.06	0.06	1	57.72	0.46	0.501
	Sex	0.06	0.06	1	125.24	0.43	0.511
Fecundity	Species	99.71	99.713	1	-	6.46	0.016*
Clutch Size	Maternal	4.01	4.01	1	16.62	4.83	0.042*
	Paternal	0.50	0.50	1	19.95	0.60	0.448
	$M \times P$	0.71	0.71	1	33.29	0.85	0.363
Embryonic	Maternal	0.32	0.32	1	22.81	6.11	0.021*
	Paternal	0.06	0.06	1	31.37	1.14	0.295
	$M \times P$	0.03	0.03	1	31.37	0.50	0.486
Egg Mass	Maternal	0.11	0.11	1	28.37	21.72	< 0.0001***
	Paternal	0.00	0.00	1	40.69	0.88	0.355
	$-M \times P$	0.00	0.00	1	40.69	0.22	0.642
Hatchling Mass	Maternal	0.06	0.06	1	26.53	21.11	< 0.0001***
	Paternal	0.00	0.00	1	40.44	0.49	0.489
	$M \times P$	0.00	0.00	1	40.44	0.06	0.808
Testes Mass	SVL	0.03	0.03	1	-	7.15	0.012*
	Species	0.0003	0.0003	1	-	0.06	0.812

†M x P = Maternal x Paternal

Table 3. Results of a generalised linear model investigating the effects of parental species on offspring sex ratio. Statistically significant values are indicated with asterisks (* = p < 0.05, ** = p < 0.01, *** = p < 0.001).

Response	Fixed Term	d.f.	χ^2	p
Sex	Maternal	1	0.424	0.515
	Paternal	1	8.637	0.003**
	Maternal x Paternal	1	2.362	0.124

Figure legends

Figure 1. The distributions of *Ctenophorus modestus* (shaded orange; previously northern lineage *C. decresii sensu lato*) and *C. decresii* (shaded blue; previously southern lineage *C.*

869 decresii sensu lato) and an enlarged map section of the contact zone. Pie charts show mean 870 proportion of genome inherited from each parental species (i.e. hybrid index) for 14 sites along a 871 linear transect through the contact zone. An approximately 12-km density trough where multi-872 year surveys recovered no individuals, and whether sites contain C. modestus or C. decresii 873 mitochondrial (mtDNA) haplotypes, is indicated. Genetically pure sites from which individuals 874 were sourced for captive breeding anchor each end of the contact zone transect: Caroona Creek 875 Conservation Park (CP; C. modestus) and Palmer (C. decresii). 876 877 Figure 2. Linear discriminant analyses of offspring classes based on five fitness variables: sprint 878 speed, bite force, growth rate, body condition at 24 weeks, body condition at hatching. Linear 879 discriminant 1 (LD1) is positively associated with body condition (at hatching and 24 weeks of 880 age) and negatively associated with bite force; linear discriminant 2 (LD2) is positively 881 associated with growth rate and sprint speed, and negatively associated with body condition at 882 hatching (Table 1). 95% confidence ellipses are shown. 883 Figure 3. Species differences in measures of F1 hybrid offspring fitness. (a) Growth rate over 884 885 the initial 24 weeks and (b) maximum sprint speed by maternal species; (c) maximum bite force 886 and (d) body condition at 24 weeks of age by paternal species; (e) survival curves over the initial 887 24 weeks estimated using the Kaplan-Meier method by offspring class [H (m-d) = C. modestus \mathcal{L} 888 \times C. decresii \lozenge hybrids, H (d-m) = C. decresii $\lozenge \times$ C. modestus \lozenge hybrids]. Statistically significant values are indicated with asterisks (* = p < 0.05, ** = p < 0.01, *** = p < 0.001). 889 890 891 Figure 4. Species differences in parental reproductive traits: (a) fecundity (i.e. total reproductive 892 output during the breeding season), (b) clutch size, (c) embryonic mortality, (d) egg mass, (e) 893 hatchling mass, and (f) relationship between egg and hatchling masses by maternal species with 894 regression line, (g) testes mass adjusted for SVL by male species, and (h) sex ratio by paternal

species. Statistically significant values are indicated with asterisks (* = p < 0.05, ** = p < 0.01,

895

896

*** = p < 0.001).







