

TITLE: NEISSERIA GONORRHOEAE VACCINES – A CONTEMPORARY OVERVIEW

Eloise Williams^{a,b#}, Kate L Seib^c, Christopher K Fairley^{d,e}, Georgina L Pollock^a, Jane S Hocking^g,
James S McCarthy^{a,f,h}, Deborah A Williamson^{a,b,f*}

- a. Department of Infectious Diseases, The University of Melbourne at the Peter Doherty
Institute for Infection and Immunity, Melbourne, Victoria, Australia*
- b. Victorian Infectious Diseases Reference Laboratory at the Peter Doherty Institute for Infection
and Immunity, Melbourne, Victoria, Australia*
- c. Institute for Glycomics, Griffith University, Gold Coast, Queensland, Australia*
- d. Melbourne Sexual Health Centre, Alfred Health, Melbourne, Victoria, Australia*
- e. Central Clinical School, Monash University, Melbourne, Victoria, Australia*
- f. The Walter and Eliza Hall Institute of Medical Research, Melbourne, Victoria, Australia*
- g. Centre for Epidemiology and Biostatistics, Melbourne School of Population and Global
Health, University of Melbourne, Melbourne, Victoria, Australia*
- h. Victorian Infectious Diseases Service, Royal Melbourne Hospital at the Peter Doherty Institute
for Infection and Immunity, Melbourne, Victoria, Australia*

Running title: *Neisseria gonorrhoeae* vaccines

#Address correspondence to: Eloise Williams, eloise.williams@mh.org.au

26	SUMMARY.....	3
27	INTRODUCTION	4
28	Epidemiology and Clinical Manifestations of <i>Neisseria gonorrhoeae</i> Infection	4
29	The Need for a <i>Neisseria gonorrhoeae</i> Vaccine	6
30	NEISSERIA GONORRHOEAE VACCINE CHALLENGES	8
31	HISTORICAL NEISSERIA GONORRHOEAE VACCINE STUDIES	11
32	POTENTIAL VACCINE TARGETS FOR NEISSERIA GONORRHOEAE VACCINES	14
33	<i>Neisseria gonorrhoeae</i> Vaccine Antigens	14
34	Adherence and invasion of mucosal epithelial cells	16
35	Nutrient acquisition and metabolism.....	17
36	Immune evasion and intracellular survival	18
37	Protection from oxidative stress and antimicrobial substances	19
38	Key reverse vaccinology antigen discoveries	20
39	Novel Vaccine Delivery Systems	21
40	Meningococcal Outer Membrane Vesicle Vaccines.....	23
41	Route of Immunization	24
42	THE IMPACT OF NEISSERIA MENINGITIDIS OUTER MEMBRANE VESICLE VACCINES ON	
43	GONORRHOEA INFECTION	25
44	Observational studies	27
45	Randomised studies.....	31
46	Biological plausibility	34
47	IN THE PIPELINE: NEISSERIA GONORRHOEAE OUTER MEMBRANE VESICLE VACCINES	37
48	POTENTIAL PUBLIC HEALTH IMPACT OF A NEISSERIA GONORRHOEAE VACCINE	39
49	Modelling The Impact of <i>Neisseria gonorrhoeae</i> Vaccines in Heterosexual Populations	40

50	Modelling the Impact of <i>Neisseria gonorrhoeae</i> Vaccines in Men Who Have Sex With Men	
51	Populations	42
52	Modelling The Impact of <i>Neisseria gonorrhoeae</i> Vaccines In Low- and Middle-Income Settings ...	45
53	QUESTIONS REMAINING: RESEARCH PRIORITIES FOR GONOCOCCAL VACCINES	47
54	CONCLUSION	48
55	ACKNOWLEDGMENTS.....	49
56	REFERENCES	49
57	AUTHOR BIOGRAPHIES	83
58	TABLES	87

59

60 SUMMARY

61 *Neisseria gonorrhoeae* infection is an important public health issue, with an annual global
62 incidence of 87 million. *N. gonorrhoeae* infection causes significant morbidity and can have
63 serious long-term impacts on reproductive and neonatal health and may rarely cause life-
64 threatening disease. Global rates of *N. gonorrhoeae* infection have increased over the past
65 20 years. Importantly, rates of antimicrobial resistance to key antimicrobials also continue
66 to increase, with the United States Centers for Disease Control and Prevention identifying
67 drug-resistant *N. gonorrhoeae* as an urgent threat to public health. This review summarises
68 the current evidence for *N. gonorrhoeae* vaccines, including historical clinical trials, key *N.*
69 *gonorrhoeae* vaccine preclinical studies and studies of the impact of *Neisseria meningitidis*
70 vaccines on *N. gonorrhoeae* infection. A comprehensive survey of potential vaccine
71 antigens, including those identified through traditional vaccine immunogenicity approaches,
72 as well as those identified using more contemporary reverse vaccinology approaches are

also described. Finally, the potential epidemiological impacts of a *N. gonorrhoeae* vaccine and research priorities for further vaccine development are described.

INTRODUCTION

Epidemiology and Clinical Manifestations of *Neisseria gonorrhoeae* Infection

Infection with *Neisseria gonorrhoeae* is an important public health issue, with an estimated annual global incidence of 87 million (1). Reported global rates of *N. gonorrhoeae* infection have significantly increased over the past 20 years (1, 2). In the United States (US), rates of *N. gonorrhoeae* infection increased 111% between 2009 and 2020 (3); in Europe, rates increased by 218% between 2009 and 2018 (4); while in Australia, rates increased 127% between 2012 and 2019 (5). *N. gonorrhoeae* infection disproportionately affects vulnerable populations, with over 90% of cases occurring in low- and middle-income (LMIC) settings (1). Within high-income countries, *N. gonorrhoeae* infection is more prevalent in certain populations, including men who have sex with men (MSM) (6, 7), transgender persons, sex workers, racial/ethnic minorities and indigenous populations (8).

N. gonorrhoeae infection causes a wide range of disease, including symptomatic urogenital disease, asymptomatic mucosal infection and infrequently, disseminated gonococcal infection (9). Urogenital infection most commonly manifests as lower genital tract infection, usually presenting as purulent anterior urethritis in men, and as cervicitis in women (10). Up to 40% of cases of urogenital *N. gonorrhoeae* infections in women are asymptomatic (11, 12). If urogenital infection is not diagnosed and treated early, severe sequelae can ensue. In women, infection can ascend to the upper genital tract to cause salpingitis and pelvic inflammatory disease. Tubal infection can result in ectopic pregnancy and infertility, and

97 infection during pregnancy is associated with preterm birth and low birthweight (9, 10, 13).
98 Neonatal infection most commonly presents as ophthalmia neonatorum, a purulent
99 conjunctivitis that may result in blindness (14). In men, ascending infection can cause
100 epididymitis, and untreated infection may result in male infertility and urethral strictures
101 (15, 16). Extragenital mucosal infections in the oropharynx, rectum and conjunctiva also
102 occur. Oropharyngeal and rectal *N. gonorrhoeae* infections are more prevalent than urethral
103 infections in certain high-risk populations, such as MSM in high-income settings, where
104 regular asymptomatic screening with nucleic acid amplification testing (NAAT) at multiple
105 anatomical sites is recommended (17). While infections of the oropharynx and rectum are
106 often asymptomatic (18), they may represent a significant reservoir for *N. gonorrhoeae*
107 transmission (19). Manifestations of disseminated gonococcal infection include purulent
108 arthritis, tenosynovitis, dermatitis, polyarthritis and osteomyelitis. Rare life-threatening
109 complications of *N. gonorrhoeae* infection include meningitis and endocarditis (20). *N.*
110 *gonorrhoeae* infection also promotes the transmission and susceptibility to human
111 immunodeficiency virus (HIV) by causing local inflammation (21).
112
113 Importantly, resistance to all prior and currently recommended antimicrobials for treatment
114 of *N. gonorrhoeae* has been described (22). *N. gonorrhoeae* has the ability to develop
115 antimicrobial resistance (AMR) through numerous mechanisms (22). Consequently, the
116 World Health Organization (WHO) and the US Centers for Disease Control and Prevention
117 (CDC) have identified antimicrobial-resistant *N. gonorrhoeae* as an urgent threat to public
118 health (23, 24). *N. gonorrhoeae* has therefore been classified as a high priority pathogen on
119 the WHO *Global Priority List of Antibiotic-Resistant Bacteria to Guide Research, Discovery*
120 *and Development of New Antibiotics* (23). The first case of treatment failure due to an

extensively-drug-resistant (XDR) *N. gonorrhoeae* strain (resistant to both current first-line antimicrobials, ceftriaxone and azithromycin) was reported in the United Kingdom (UK) in 2016 (25); XDR *N. gonorrhoeae* with high-level resistance to both ceftriaxone and azithromycin has now been reported in the UK and Australia (26, 27). These cases demonstrate the growing global threat of untreatable *N. gonorrhoeae* infection. A number of novel gonococcal antimicrobial therapies have recently been tested in phase two and three trials, including solithromycin, zoliflodacin and gepotidacin. These studies have demonstrated several limitations of these new anti-gonococcal antimicrobials (28); in brief, a randomised trial found solithromycin to be inferior to standard-of-care dual ceftriaxone and azithromycin therapy (29); the efficacy of zoliflodacin was suboptimal for pharyngeal infection (30); and current data on the performance of gepotidacin for extragenital infections is sparse (31). As the spectre of untreatable *N. gonorrhoeae* infection looms, preventative strategies that overcome the extraordinary ability of *N. gonorrhoeae* to evade killing by antimicrobial therapy are therefore urgently required.

The Need for a *Neisseria gonorrhoeae* Vaccine

An effective and accessible *N. gonorrhoeae* vaccine could have a wide range of benefits, including: i) reduction of the individual and healthcare impact of urogenital infection; ii) improvement in reproductive and neonatal health; iii) reduction of individual and population antimicrobial usage and the unintended consequences arising from this, including the potential to drive further *N. gonorrhoeae* antimicrobial resistance; and iv) reduction in the healthcare costs associated with frequent screening for *N. gonorrhoeae* infection in asymptomatic individuals. However, there are multiple significant barriers to the development of a *N. gonorrhoeae* vaccine, including i) antigenic and phase variation of

potential vaccine targets; ii) the absence of protective immunity following natural infection; iii) the lack of a known immune correlate of protection; and iv) exclusive human host restriction, with limited appropriate animal models of infection (32). Encouragingly, the successes of vaccines for other sexually-transmitted infections (STIs) such as human papillomavirus (HPV), hepatitis A virus (HAV) and hepatitis B virus (HBV) (33), as well as closely-related pathogens, such as *Neisseria meningitidis*, have paved the way for further progress in *N. gonorrhoeae* vaccine development (34).

The development and implementation of safe and efficacious vaccines for HPV, HAV and HBV has had a significant impact on the incidence and resulting complications of these diseases (33). These successes have provided additional motivation for the development of new STI vaccines. In 2014, the WHO and National Institutes of Health (NIH) announced a comprehensive roadmap to accelerate the STI vaccine development (35). This roadmap comprised of nine areas of focus, including obtaining improved epidemiological data, modelling vaccine impact, accelerating basic science research, outlining preferred product characteristics and encouraging investment (35, 36). The WHO subsequently assembled a panel of international experts to define the potential public health value of and preferred product characteristics of a *N. gonorrhoeae* vaccine to inform vaccine development (37, 38). The WHO Global Health Sector Strategy on STIs has set a target of a 90% reduction in worldwide *N. gonorrhoeae* infection incidence by 2030. Given the rising incidence of *N. gonorrhoeae* infection worldwide and the limitations of current preventative interventions, this WHO strategy highlights *N. gonorrhoeae* vaccine development as a priority innovation to support this ambitious aim (39).

In this review, we examine the evidence for a *N. gonorrhoeae* vaccine, including i) historical clinical trials; ii) key *N. gonorrhoeae* vaccine preclinical studies; iii) observational and randomised studies of the impact of *N. meningitidis* vaccines on *N. gonorrhoeae* infection and iv) clinical trials currently underway. In addition, we present a comprehensive survey of potential vaccine antigens, including those identified through traditional vaccine immunogenicity approaches, as well as those identified using more contemporary approaches, such as bioinformatics, transcriptomics and proteomics. Finally, we review the potential epidemiological impacts of a *N. gonorrhoeae* vaccine, and outline research priorities for *N. gonorrhoeae* vaccine development.

References for this review were identified on the basis of the topics described above, with literature search conducted through PubMed and ClinicalTrials.gov. The websites of the WHO and US CDC were also reviewed and an online search engine was used to access press releases, conference abstracts and commercial information. Search terms included, “gonorrhoea*”, “gonococcal”, “Neisseria”, “vaccine”, “antigen”, “meningococcal”, “outer membrane vesicle”, “OMV”, “model*”, “impact”, “cost” and “economic”. In addition, a search was undertaken for each vaccine antigen listed in column 1 of Table 2. Relevant articles published between Jan 1, 1900 and March 1, 2023 were included. Articles published in English resulting from these searches and their relevant references were reviewed.

NEISSERIA GONORRHOEAE VACCINE CHALLENGES

A number of obstacles have impeded progress towards the development of an effective vaccine against *N. gonorrhoeae* (32). First, *N. gonorrhoeae* demonstrates significant surface antigen variability, such that key surface antigens have variable genomic sequences and

protein composition (antigenic variation) and/or change their protein expression (through phase variation). Second, there is no epidemiologic evidence that *N. gonorrhoeae* infection results in protective immunity against recurrent infections; indeed, repeated infections are relatively common in high-risk populations. Third, given the lack of protective immunity against reinfection, it has not been possible to define correlates of immunity that can be measured using immunologic methods (32).

As an exclusive human pathogen, establishment of an appropriate animal infection model to study the pathogenesis and preclinical immune responses to *N. gonorrhoeae* infection and vaccines has been difficult. A 17- β -estradiol-treated mouse model (40, 41), using inbred mice and recently modified by the use of transgenic mice with additional human host-cell receptors such as human carcinoembryonic antigen cellular adhesion molecules (42, 43) and human transferrin (44) or supplementation of inbred mice with human transferrin (45, 46) have partially overcome this host-specific barrier. Although chimpanzees were also used in early infection models (47), they are no longer available or ethically appropriate for this work. A number of experimental systems have been used to assist drug development, however these models are not appropriate for vaccine development. These include a hollow fiber infection model that is well suited to characterize the pharmacodynamic and pharmacokinetic responses of novel antimicrobials for treatment of *N. gonorrhoeae* (48, 49), and an invertebrate *Galleria mellonella* greater wax moth of gonococcal infection (50). These latter models, however, lack the essential host immunity components required to test gonococcal vaccines.

A *N. gonorrhoeae* male urethritis controlled human infection model (CHIM) was developed by investigators at Walter Reed Army Institute of Research and the University of North Carolina at Chapel Hill in the US in the 1980s (51). Over 200 individuals have participated in *N. gonorrhoeae* urethritis CHIM studies. These studies have been reviewed for safety and compliance with modern ethical standards, and have been undertaken without serious or unexpected adverse events (51). There are a number of advantages to a *N. gonorrhoeae* CHIM, compared to alternative study designs. In particular, compared to animal studies, the model not only assesses microbiological outcomes, but also clinical disease and immune responses. In addition, CHIM studies provide a model that has the power to test for statistically significant vaccine efficacy in a much smaller study population (<100 participants) compared to an efficacy trial conducted in a population with high risk for gonorrhoea infection (>1000 participants)(52). Notably, the only *N. gonorrhoeae* CHIM that is currently available is a male urethritis model, which could limit the generalizability of vaccine efficacy findings to oropharyngeal, rectal and cervical *N. gonorrhoeae* infections (52). *N. gonorrhoeae* male urethritis CHIM studies have already advanced the understanding of the complex pathogenesis and immune responses to *N. gonorrhoeae* infection (51). As promising vaccine candidates become available, *N. gonorrhoeae* CHIM studies may offer a safe and effective model for testing these novel vaccines, particularly if models of extragenital infection, such as an oropharyngeal *N. gonorrhoeae* CHIM become available (52).

Although these obstacles may have slowed the progress of *N. gonorrhoeae* vaccines, evidence suggesting partial effectiveness of the *N. meningitidis* serogroup B outer membrane vesicle (OMV) vaccines against *N. gonorrhoeae* infection (34, 53-62) has

reinvigorated the field, with an increased international focus on the development an effective *N. gonorrhoeae* vaccine.

HISTORICAL NEISSERIA GONORRHOEAE VACCINE STUDIES

The aim of developing a gonococcal vaccine has been pursued since the turn of the twentieth century. Initially, these vaccines were designed as a therapeutic strategy for persistent *N. gonorrhoeae* infection, rather than as a preventative measure. At this time there were numerous attempts made by different groups to immunize patients with symptomatic gonorrhoea with various whole cell vaccines to promote opsonophagocytosis (63). With the development of effective antimicrobial therapy, therapeutic vaccine discovery stalled. Further efforts were made in the 1970s and 1980s, when three different preventative *N. gonorrhoeae* vaccines were developed and trialled in humans, all of which were unsuccessful (64-66). These vaccine studies are described in Table 1. The first vaccine, a partially inactivated whole-cell vaccine prepared from two pooled *N. gonorrhoeae* strains elicited specific antibody responses among the majority of the 54 participants included in the initial phase I study (67). A subsequent placebo-controlled double-blind field trial of this vaccine was undertaken in 1972-1973, this time using whole cell preparations from three pooled *N. gonorrhoeae* strains. This study involved 62 participants from an Aboriginal Inuit population in the northern Canada village of Inuvik, with the immunization schedule comprising three 1ml intramuscular injections of vaccine or placebo at weekly intervals. No significant difference between the groups in the cumulative incidence of laboratory-confirmed *N. gonorrhoeae* infection was observed over the 12-month period following vaccination (30% incidence in the vaccinated group versus 24% in the placebo group; $p=0.78$)(64).

264

265 The second vaccine, a *N. gonorrhoeae* pilus vaccine, elicited serum and genital anti-pilus
266 antibody responses against heterologous strains and demonstrated efficacy in an initial *N.*
267 *gonorrhoeae* urethral CHIM study after challenge with a homologous strain (68, 69). This was
268 followed by a placebo-controlled, double-blind trial in Korea in the 1980s, involving 3,250
269 high-risk US military personnel, using an immunization schedule comprising two 0.1ml
270 intradermal injections of vaccine or placebo, two weeks apart. There was no significant
271 difference in cumulative incidence of laboratory-confirmed *N. gonorrhoeae* infection in the
272 8-week period following vaccination between the two groups, with a cumulative incidence
273 of 6.9% observed in the vaccinated group, compared with 6.5% in the placebo group (65). In
274 a subsequent *N. gonorrhoeae* urethral CHIM study, no protection was observed against a
275 heterologous *N. gonorrhoeae* strain expressing antigenically different pili (70), suggesting
276 that despite the production of anti-pilus antibody responses against heterologous pili, these
277 responses were insufficient to prevent infection with *N. gonorrhoeae* strains expressing
278 antigenically different pili. Pilus antigen heterogeneity, a characteristic of circulating strains
279 of *N. gonorrhoeae*, was the most likely explanation for the unsuccessful field trial.

280

281 The most recent *N. gonorrhoeae* vaccine to be trialled in humans was a *N. gonorrhoeae*
282 outer membrane vaccine prepared from a single strain. In a randomized placebo-controlled
283 *N. gonorrhoeae* urethral CHIM undertaken in 1985, 63 male participants received a single
284 dose of intramuscular vaccine or placebo and underwent intraurethral challenge with a
285 homologous *N. gonorrhoeae* strain two to four weeks later. No significant difference in
286 infection was observed between the two groups. Infection rates were unexpectedly low in
287 this study, with 46% of those vaccinated and 36% of placebo recipients remaining

uninfected (66). Although designed to enrich for the Porin (Por) outer membrane protein, this vaccine was contaminated with other membrane antigens, including lipooligosaccharide (LOS) and reduction modifiable protein (Rmp). Later studies demonstrated that anti-Rmp antibodies downregulate the bactericidal activity of antibodies against other antigens (71). The contamination of this vaccine by Rmp therefore likely resulted in anti-Rmp antibodies that may have antagonized the bactericidal effect of anti-Por and anti-LOS antibodies. This hypothesis was supported by a retrospective analysis of the vaccine trial data which incorporated data on risk for pre-existing immunity. This analysis demonstrated that the ratio of Por and LOS antibody concentration to Rmp antibody concentration correlated with protection from *N. gonorrhoeae* infection in both vaccine and placebo recipients (66).

These early studies demonstrate three key considerations for future gonococcal vaccine trials. Firstly, CHIM trials that are appropriately designed to test investigational vaccines may serve as go-no-go measure using a relatively small number of participants before more resource-intensive, larger-scale efficacy trials are undertaken. Secondly, pre-existing immunity should be incorporated into the design and analysis of future gonococcal vaccine trials by documenting baseline antibody levels and previous exposure. Finally, the heterogeneity of circulating *N. gonorrhoeae* strains must be considered both in the selection of potential vaccine antigens and the selection of challenge strains for future gonococcal CHIM vaccine trials.

POTENTIAL VACCINE TARGETS FOR NEISSERIA GONORRHOEAE VACCINES

Neisseria gonorrhoeae Vaccine Antigens

A number of potential *N. gonorrhoeae* vaccine candidates have been evaluated in pre-clinical testing including *in vitro*, in animal models and occasionally, early phase human studies. Key features of an ideal *N. gonorrhoeae* vaccine antigen include i) surface exposure; ii) conservation (lack of phase or antigenic variation); iii) high prevalence among globally-circulating strains; iv) immunogenicity; and v) evidence that the antigen plays an important role in virulence or survival. In the absence of known immune correlates of protection against *N. gonorrhoeae* infection, a widely used approach has been to identify surface antigens that elicit an antibody response that confers complement-dependent bactericidal activity, and/or mediates opsonophagocytosis (72), hypothesising that these may be surrogate predictors of prevention. However, antibody responses to natural uncomplicated *N. gonorrhoeae* infection are typically described as weak and short-lived (32). In addition, in early vaccine studies where the pilus, Por and LOS antigens (described above) were evaluated, no correlates of protective immunity were apparent. The bactericidal and opsonophagocytic activity of antibodies induced by natural reinfection is influenced by a number of factors, including downregulation by blocking antibodies (e.g. anti-Rmp antibodies)(71) and soluble complement regulators (e.g. Factor H and C4b-binding protein)(66), as well as poor cross-protection of antibodies to polymorphic antigens (e.g. pilus and Por)(65, 71). Given the complex humoral immune responses to *N. gonorrhoeae* infection and the lack of protective immunity induced by natural gonococcal infection (32), an optimal *N. gonorrhoeae* vaccine will need to induce immune responses that are qualitatively and quantitatively different to that induced by natural immunity.

333 A number of novel strategies have informed the contemporary approach to gonococcal
334 vaccine antigen discovery. Reverse vaccinology is a process of vaccine antigen discovery that
335 harnesses genomics, proteomics, immunoproteomics, transcriptomics and bioinformatics to
336 identify highly conserved, widely distributed and surface exposed antigens that may
337 represent promising vaccine antigens. A reverse vaccinology approach has been used to
338 develop highly successful vaccines for other pathogens, such as *N. meningitidis* serogroup B
339 (4CMenB; GlaxoSmithKline)(73). Identification of novel surface-exposed antigens of *N.*
340 *gonorrhoeae* have used proteomic techniques to characterise *N. gonorrhoeae* membrane
341 vesicle and cell envelope proteins (74). Such an approach can be coupled with a range of
342 bioinformatic tools to predict function, subcellular localization, post-translational
343 modification and immunogenicity (74).

344

345 Using a proteomic-based approach, Zielke et al identified 305 cell envelope and 46
346 membrane vesicle proteins that were uniformly present among four well-characterized *N.*
347 *gonorrhoeae* strains, many of which were newly discovered proteins or proteins that had
348 not previously been characterized in *N. gonorrhoeae* (75). Using such proteomic approaches
349 it has been possible to identify candidate vaccine antigens with a range of attractive
350 characteristics, such as expression in physiologically-relevant environmental conditions,
351 including both aerobic and anaerobic, iron deprivation, exposure to normal human serum
352 and exposure to extended spectrum cephalosporins (76-78). Analysis of the genes expressed
353 during natural human mucosal infection, coupled with immune characterisation, has also
354 led to the discovery of a number of novel putative vaccine antigen targets (79). The
355 availability of public genomic databases, such as Neisseria PubMLST, has also enabled

assessment of the presence and conservation of putative vaccine antigens across globally diverse strains (80).

Another strategy to improve vaccine discovery efforts for *N. gonorrhoeae* has been to target antigens that not only elicit an antibody response with bactericidal and opsonophagocytic activity, but also those that elicit a functional antibody response that inhibits important physiological functions in the pathogenesis of gonococcal infection (81-83). These physiological functions include i) adherence to and invasion of mucosal epithelial cells; ii) nutrient acquisition and metabolism; iii) immune evasion; iv) intracellular survival and iv) protection from oxidative stress or antimicrobial substances. Promising vaccine antigens from each of these categories will be briefly highlighted below, and a comprehensive summary of potential *N. gonorrhoeae* vaccine antigens is presented in Table 2.

Adherence and invasion of mucosal epithelial cells

The potential of targeting with a vaccine a number of key mediators of attachment and invasion, such as type IV pili, LOS and the opacity-associated outer membrane proteins (Opa), has been confounded by the high levels of antigenic variation and/or phase variation in these antigens. For example, although the gonococcal porin protein, PorB is the most highly abundant outer membrane protein and constitutively expressed, targeting it with a vaccine is confounded by a high level of antigenic variation within the eight surface-exposed loops in different gonococcal strains (32, 84). However, PorB is an essential protein that plays a key role in host cellular attachment, invasion, nutrient acquisition, apoptosis and serum complement resistance (85) and has immune enhancing activity, making it a promising vaccine adjuvant (86). Preclinical studies of putative PorB vaccines are described

below. Alternative targets include mediators of host cell adherence such as the type IV pilus-associated outer membrane proteins PilC (87, 88), involved in pilus biogenesis and attachment; and PilQ, the secretin through which pili are extruded (89-91). Phospholipase D, which participates in host cell invasion and survival, is another potential outer membrane protein vaccine target (92, 93). In addition, the *Neisseria* heparin binding antigen (NHBA), which is also involved in host cell adherence and survival, has recently been demonstrated to be a promising vaccine antigen candidate, as it is widely distributed, highly conserved and induces bactericidal and opsonophagocytic antibodies (94-96).

Nutrient acquisition and metabolism

A number of antigens involved in nutrient acquisition through iron and zinc uptake have shown promise as potential vaccine antigen targets. The transferrin receptor proteins transferrin binding protein A (TbpA) and transferrin binding protein B (TbpB) facilitate iron acquisition, and are essential for experimental urethral infection of male volunteers when alternative iron acquisition mechanisms are not available (97). The transferrin receptor proteins are immunogenic, with the intranasal immunization of mice with TbpA and TbpB proteins fused to cholera toxin subunit B inducing serum and vaginal mucosal anti-TbpA and anti-TbpB bactericidal antibodies (98). However preliminary evidence suggests that antibodies to gonococcal TbpA have only a modest inhibitory effect on ligand binding (81). Nitrate reductase (AniA) is required for anaerobic growth and biofilm formation of *N. gonorrhoeae* (99, 100). Antibodies against AniA protein inhibit nitrite reductase activity (101, 102), suggesting this may be another promising function-blocking vaccine target.

402

403 **Immune evasion and intracellular survival**

404 Potential vaccine antigens involved in immune evasion and intracellular survival of *N.*
405 *gonorrhoeae* include alpha-2,3-sialyltransferase (Lst) and Neisserial surface protein A
406 (NspA). Lst expressed by gonococci scavenge sialic acid from the host and sialylate the
407 gonococcal LOS, thereby inhibiting complement-mediated and polymorphonuclear
408 leukocyte-mediated killing (103, 104). However recent evidence suggests that Lst is a
409 cytoplasmic rather than surface-exposed protein (105). NspA plays an important role in
410 complement evasion by binding to complement regulator human factor H and factor H-like
411 protein 1 (106). Immunization of mice with plasmid DNA containing the NspA gene followed
412 by boosting with recombinant NspA protein induced serum and mucosal antibodies with
413 bactericidal and opsonophagocytic activity (107).

414

415 The conserved LOS epitope 2C7, defined by lactose substitutions at HcpI and HcpII in the
416 LOS core promotes gonococcal colonization and survival, and is another important *N.*
417 *gonorrhoeae* vaccine target. Although this epitope is phase variable, the *lgtG*
418 glycosyltransferase gene that controls this phase variation is expressed in 95% of gonococci
419 in human infection (108, 109). Monoclonal antibodies against this epitope are bactericidal
420 and opsonophagocytic (108). In an intraperitoneal mouse immunization study of a multi-
421 antigenic 27C peptide mimic (MAP1) with a T helper type 1 (Th1)-inducing adjuvant
422 (Monophosphoryl lipid A; MPL), immunization induced Th-1 biased anti-LOS antibodies that
423 were also bactericidal. Immunization also reduced the length of gonococcal carriage and
424 bacterial burden in experimentally infected mice (110). Further studies of a LOS 2C7 vaccine
425 candidate with greater potential for scalability and economic production comprising a

426 stable, homogenous tetrapeptide 2C7 mimitope (TMCP2), administered with a
427 glucopyranosyl lipid A adjuvant in a stable oil-in-water nanoemulsion replicated these
428 findings (111). Anti-Rmp antibodies have been demonstrated to inhibit the efficacy of 2C7
429 monoclonal antibody in mice in a dose-dependent fashion. Therefore, an effective LOS 2C7
430 vaccine would likely need to produce concentrations of protective antibody sufficient to
431 overcome this inhibitory effect in individuals with pre-existing anti-Rmp antibodies (112).

432

433 **Protection from oxidative stress and antimicrobial substances**

434 Proteins that protect *N. gonorrhoeae* from the threats of oxidative stress and antimicrobial
435 substances play an important role in the pathogenesis of *N. gonorrhoeae*. A number of
436 these proteins have recently been identified as promising vaccine targets. Gonococcal
437 methionine sulfoxide reductase protein (MsrA/B) reduces methionine sulfoxide to
438 methionine to protect the organism from oxidative stress (83). MsrA/B is surface-exposed
439 and the gene encoding MsrA/B is highly conserved. Immunisation of mice with an
440 adjuvanted recombinant MsrA/B vaccine results in the production of function-blocking
441 antibodies with bactericidal and opsonophagocytic activity (83). Another promising vaccine
442 antigen is multiple transferable resistance protein E (MtrE), an outer membrane channel of
443 a multidrug transporter system (MtrCDE) which mediates export of hydrophobic
444 antimicrobial substances (fatty acids, long-chain lipids, bile salts and antimicrobials) from
445 the cell and survival after neutrophil exposure (113). It also plays a key role in the FarA-FarB-
446 MtrE active efflux pump, an additional efflux pump system that mediates resistance to
447 hydrophobic agents (114). Mice immunized with an adjuvanted recombinant MtrE vaccine
448 produce anti-MtrE antibodies that are bactericidal and reduce the activity of the MtrCDE
449 efflux pump in the presence of hydrophobic compounds (82).

Key reverse vaccinology antigen discoveries

Finally, a number of promising vaccine antigens have also been discovered using reverse vaccinology approaches described above. These include several antigens involved in cell envelope homeostasis and translocation, including beta-barrel assembly machinery protein A (BamA), lipopolysaccharide assembly protein D (LptD) and translocation and assembly module A (TamA) as well as two human lysozyme inhibitors, adhesin complex protein (ACP) and surface-exposed lysozyme inhibitor of ϵ -type lysozyme (SliC). BamA, LptD and TamA are surface-exposed, highly conserved, stably expressed and immunisation with them elicits antibodies with bactericidal activity (76). Both lysozyme inhibitor antigens ACP and SliC are highly conserved and stably expressed (115-117); antibodies to ACP are both bactericidal and inhibit binding to human lysozyme (116).

Another promising vaccine candidate discovered by reverse vaccinology is MetQ, the methionine binding component of an ATP-binding cassette transporter system (118). MetQ plays a role in epithelial cell adherence and survival. It is a highly conserved surface-exposed protein that is constitutively expressed (76, 118). Anti-MetQ antibodies are bactericidal and reduce adherence of *N. gonorrhoeae* to cervical epithelial cells (118). Mice immunized with a recombinant MetQ lipoprotein formulated with a Th1-stimulating adjuvant (cytosine phosphoguanine; CpG) developed robust Th1-biased serum and vaginal antibodies. After vaginal challenge, the immunized mice demonstrated accelerated clearance of gonococcal infection and a lower bacterial burden (119).

Novel Vaccine Delivery Systems

In addition to novel vaccine targets, there has been significant development in vaccine adjuvants that augment vaccine antigen immune responses. These include nanoparticle technologies such as liposome-based adjuvants which contain immunogens such as toll-like receptor ligands; and oil-in-water emulsions which activate myeloid cells to stimulate innate and adaptive immune responses (120). Novel adjuvants that have been assessed in preclinical *N. gonorrhoeae* vaccine studies include the Th1-stimulating adjuvants, microencapsulated interleukin-12 (IL-12) (121) and CpG oligodeoxynucleotides (119, 122). In recent mouse model studies, *N. gonorrhoeae* has been shown to be able to suppress the development of Th1 and T helper type 2 (Th2) T cell response, and to induce a T helper type 17- (Th17) driven immune response that facilitates immune evasion (123-125). Elevated levels of the Th17 cytokine, interleukin-17 (IL-17), have also been demonstrated in serum and genital secretions of patients with *N. gonorrhoeae* infection compared to healthy subjects or those with non-bacterial STIs, suggesting that the experimental observations in mice of a Th17-driven immune response may also apply in human *N. gonorrhoeae* infection (126, 127). Rational vaccine design using Th-1 stimulating adjuvants harnesses this key discovery. Th1-stimulating adjuvants have been shown to induce a Th1-driven response, generate anti-gonococcal antibodies and gamma-interferon secreting CD4+ T cells and accelerated clearance of *N. gonorrhoeae* infection in preclinical mouse model studies (119, 121).

The past decade has seen a number of newly licensed vaccines for important infectious diseases which use novel vaccine delivery systems, including nucleic acid vaccines (such as those used in messenger ribonucleic acid (mRNA) severe acute respiratory syndrome

497 coronavirus 2 (SARS-CoV-2) vaccines), virus-like particles (as used in HPV vaccines) and OMV
498 vaccines (used in serogroup B meningococcal vaccines) (128). A number of these novel
499 vaccine delivery systems have been studied in preclinical mouse models of *N. gonorrhoeae*
500 vaccines. Nucleic acid vaccines, viral replicon particles and recombinant vaccines are
501 particularly attractive for putative PorB gonococcal vaccines as these techniques avoid
502 potential problems of contamination by Rmp and inadvertent stimulation of anti-Rmp
503 blocking antibodies by the putative vaccine. Zhu et al have undertaken a number of studies
504 investigating various vaccine delivery techniques and prime-boost schedules for putative
505 PorB vaccines, including PorB deoxyribonucleic acid (DNA), renatured recombinant PorB
506 (rrPorB), PorB expressed from Venezuelan equine encephalitis virus replicon particles (PorB
507 VRPs) and OMV vaccines (in which the major constituent antigen is PorB)(129, 130). These
508 studies have demonstrated that different immune responses are triggered by various
509 vaccine antigen delivery systems and sites of inoculation. For example, mice immunized
510 subcutaneously with a rrPorB vaccine developed high levels of PorB-specific IgG antibodies,
511 with immunization administered in the hind footpad inducing a Th1 response and
512 immunization administered in the dorsal area inducing a Th2 response. In this study,
513 immunization with PorB VRPs induced a Th1 response while an intranasal OMV vaccine was
514 the only vaccine that generated serum bactericidal antibodies (129). Antibodies induced by
515 a PorB DNA vaccine alone were modest, however boosting by either rrPorB or PorB VRPs
516 significantly increased PorB-specific serum antibody levels (130).

517

518 Other novel vaccine delivery technologies such as bacterial ghosts have also been explored
519 in preclinical *N. gonorrhoeae* vaccines. Bacterial ghosts are empty Gram-negative bacterial
520 cell envelopes that retain the cellular morphology and antigenic determinants of the cell

envelope and provide a promising system for the delivery of nucleic acid (DNA or RNA) vaccines. This delivery system provides intrinsic adjuvant activity due to the enhanced immune responses produced against cell envelope antigens, including T cell activation and mucosal immunity (131). Jiao et al have demonstrated that *N. gonorrhoeae* PorB and NspA DNA vaccines delivered using *Salmonella enteritidis* ghosts induce serum IgG antibodies that are bactericidal in an experimental mouse model (132, 133).

Meningococcal Outer Membrane Vesicle Vaccines

Meningococcal OMV vaccines have been a key focus of both observational and preclinical *N. gonorrhoeae* vaccine studies. OMVs are spherical lipid bi-layer membrane structures that are released spontaneously from the outer membrane of Gram-negative bacteria and contain surface-exposed phospholipids, lipopolysaccharide/LOS and membrane proteins as well as RNA, DNA, proteins and peptidoglycans within the lumen of the vesicle (134-136). The role of OMVs in bacterial pathogenesis includes modulation of host immune response, nutrient acquisition, and biofilm formation (134-136). OMVs present a number of advantages as a novel vaccine platform, including the ability to enter lymphatic vessels for uptake by antigen-presenting cells and presentation of membrane surface antigens in their native configuration, thereby evoking humoral and cell-mediated responses (134-136). The association between immunization with currently-available meningococcal B vaccines and *N. gonorrhoeae* infection and preclinical studies of these vaccines will be discussed in further detail below. The focus in preclinical studies of novel OMV vaccines has recently shifted to optimizing meningococcal OMV design based on known features of gonococcal pathogenesis, such as the use of meningococcal isolates lacking Rmp proteins and avoiding detergent-based preparation of outer membranes (137, 138). Although detergent-based

preparation of outer membrane vesicles extracts LOS and decreases endotoxin activity, it also removes key meningococcal antigens, including factor H binding protein (fHbp) (137). A detoxified meningococcal OMV vaccine (lacking PorA, PorB and Rmp) has been shown to improve gonococcal clearance in a murine model (137). In addition, a meningococcal native OMV vaccine with attenuated endotoxin and overexpressed fHbp has been shown to induce high levels of serum immunoglobulin G (IgG) anti-FHbp as well as serum bactericidal antibodies against heterologous gonococcal strains (138). Furthermore, the next generation of OMV vaccines developed from *N. gonorrhoeae* strains, and designed specifically to induce protection against *N. gonorrhoeae* infection are under study, with a number of preclinical studies demonstrating promising results, including production of serum and vaginal antibodies and accelerated clearance of gonococcal infection in the estradiol-treated female mouse model (139-141). These include the dmGC_0817560 (140) and NGoXIM (141) native OMV vaccines described in further detail below.

Route of Immunization

The route of immunization may also play a significant role in determining the immunogenicity of a *N. gonorrhoeae* vaccine. It has been observed that the ability of parenteral immunization to induce mucosal immunoglobulin A (IgA) antibodies for other sexually transmitted pathogens is limited (142). By contrast, mucosal administration of vaccines via intranasal immunization has demonstrated relatively higher mucosal IgA and IgG antibodies compared with parenteral vaccines (98, 142). This has been shown in mouse model studies of a number of different experimental *N. gonorrhoeae* vaccines (98, 119, 129, 139, 141, 143). For a number of these vaccines, accelerated clearance of gonococcal infection has been observed in intranasally immunized mice, including a gonococcal OMV

preparation (139) and a recombinant MetQ-CpG adjuvant vaccine (119). In addition, intravaginal and intranasal immunization using a native gonococcal OMV plus microencapsulated IL-12 vaccine (NGoXIM) in a female mouse model induced serum and vaginal IgG and IgA antibodies and accelerated clearance of gonococcal infection (121, 141). Other novel routes of *N. gonorrhoeae* vaccine delivery studied in preclinical settings include a transdermal microneedle skin patch that enables slow release of antigens using a formalin-inactivated whole-cell gonococcal microparticle vaccine formulation. Mouse model studies of this vaccine demonstrated that the transdermal skin patch vaccine induced increased IgG antibody titres compared with the comparator subcutaneously administered vaccine (144).

Table 3 provides a summary of *N. gonorrhoeae* vaccines that have proceeded to contemporary preclinical studies in the experimental mouse model, many of which have included novel vaccine antigens, vaccine delivery systems or routes of immunization.

THE IMPACT OF NEISSERIA MENINGITIDIS OUTER MEMBRANE VESICLE VACCINES ON GONORRHOEA INFECTION

The most significant step in *N. gonorrhoeae* vaccine progress in the past decade was a landmark study that demonstrated 31% vaccine efficacy of a *N. meningitis* serogroup B outer membrane vesicle (OMV) vaccine (MeNZB) against *N. gonorrhoeae* infection in a retrospective observational case-control study of 15-30-year-olds attending sexual health clinics in New Zealand (34). This finding demonstrated the biological plausibility of vaccine-mediated protective immunity against *N. gonorrhoeae*, and provided a proof-of-concept that an effective *N. gonorrhoeae* vaccine may be possible (145). This observation was

supported by evidence from ecological studies in Cuba, Norway and Canada where an association between the introduction of *N. meningitidis* serogroup B OMV-containing vaccines and reduced rates of gonorrhoea infection were apparent (58-62). The impact of 4CMenB (Bexsero; GSK) has been assessed in further retrospective observational case-control and cohort studies. 4CMenB is a *N. meningitidis* serogroup B OMV-containing vaccine which incorporates the OMV included in MenZB, as well as three recombinant antigens, Neisseria adhesin A (NadA), fHbp and NHBA, as well as two accessory proteins (GNA2091 fused with fHbp and GNA1030 fused with NHBA) that increase the immunogenicity of the target recombinant antigens (146). These studies have also demonstrated a protective effect of 4CMenB on *N. gonorrhoeae* infection, with estimated vaccine effectiveness for a two-dose schedule ranging between 33-46% in various settings across the world, including in the US, Australia and Italy (53-56). These studies are summarised in Table 4.

Several studies are currently recruiting participants into randomized placebo-controlled trials of the 4CMenB vaccine to assess efficacy against *N. gonorrhoeae* infection (<https://clinicaltrials.gov/study/NCT04415424>; <https://clinicaltrials.gov/study/NCT04350138>; <https://clinicaltrials.gov/study/NCT05766904>; <https://clinicaltrials.gov/study/NCT05294588>; 147). Furthermore, in recently reported interim analysis of a randomised, open-label factorial study of the 4CMenB vaccine coupled with doxycycline post-exposure prophylaxis in MSM on HIV pre-exposure prophylaxis (PrEP) (DOXYVAC), a reduced incidence of first-episode *N. gonorrhoeae* infection in the 4CMenB group was observed compared to the no vaccine group (adjusted hazard ratio 0.49; 95% CI 0.27-0.88)(148). However, the final study

report is awaited, as review of the study data indicates that a number of *N. gonorrhoeae* infections were not included in the interim analysis (149). The randomised studies of 4CMenB will provide further high-level evidence of the protective efficacy of *N. meningitidis* serogroup B OMV vaccines against *N. gonorrhoeae* infection. Here we describe the clinical and basic science studies of meningococcal serogroup B OMV vaccines in further detail.

Observational studies

The first studies to suggest an association between various meningococcal serogroup B OMV vaccines and *N. gonorrhoeae* incidence were ecological analyses of the impact of mass serogroup B meningococcal OMV vaccination programs on *N. gonorrhoeae* infection rates in Cuba, Norway and Canada (58-62). In Cuba, a *N. meningitidis* serogroup B OMV-containing meningococcal vaccine, VA-MENGOC-BC, was used in a national mass vaccination program of individuals aged 3 months to 24 years between 1989 and 1990, and subsequently incorporated into the national infant immunization schedule (150). Reported vaccine coverage of the mass vaccination program in the target population was 95% (150). In the years immediately after the program (1989-1993), the incidence of gonorrhoea decreased from 381.9 to 190.3 cases per 100,000 ($r=0.9607$, $p=0.001$), despite an increase in other sexually transmitted infections such as syphilis (58-60).

In a second ecological study undertaken in Norway using trial registry data of a *N. meningitidis* serogroup B OMV-containing vaccine, MenBvac, delivered to 63% of 13-15-year-olds between 1988-1992, a reduced incidence rate ratio (IRR) of gonorrhoea was observed in the subsequent years 1993-2008 among 20 to 24-year-olds in the vaccinated cohort compared to the pre-vaccination cohort (IRR 0.58, 95% CI 0.42-0.8 for women and

adjusted IRR 0.68, 95% CI 0.51-0.93 for men)(62). In a third an ecological study the incidence of *N. gonorrhoeae* infection was studied in the context of a mass vaccination campaign undertaken in Canada in 2014 among individuals aged 6 months to 20 years vaccinated with the *N. meningitidis* serogroup B OMV-containing 4CMenB vaccine in the Sanguenay-Lac-Saint-Jean region of Quebec. Although an association between vaccination and reduced gonorrhoea incidence of 59% (95% CI -22% to 84%; $p=0.1$) was observed, this finding was not statistically significant.(61)

A landmark retrospective observational case-control study of the MeNZB *N. meningitidis* serogroup B OMV vaccine was the first to describe vaccine efficacy against *N. gonorrhoeae* infection among 14,730 sexual health clinic patients aged 15-30 years who were eligible to receive MeNZB vaccination through a mass vaccination program of individuals aged 6 weeks to 20 years implemented in New Zealand between 2004-2006 (34). This study demonstrated that vaccinated individuals were significantly less likely to be cases (*N. gonorrhoeae* mono-infection) than controls (*Chlamydia trachomatis* mono-infection); 41% vs 51%; adjusted odds ratio (OR) 0.69 (95% CI 0.61-0.79; $p<0.0001$). After adjustment for ethnicity, deprivation status, geographical area and sex, the estimated vaccine effectiveness of MeNZB against *N. gonorrhoeae* infection was 31% (95% CI 21-39; $p<0.0001$)(34). Further study of individuals vaccinated with MeNZB during New Zealand's mass vaccination program demonstrated that vaccinated individuals were also significantly less likely to be hospitalized due to *N. gonorrhoeae* infection, with an estimated vaccine effectiveness against *N. gonorrhoeae*-related hospitalization of 24% (95% CI 1-42%)(57).

664 A similar association between the *N. meningitidis* serogroup B OMV-containing vaccine
665 4CMenB and reduced risk of *N. gonorrhoeae* infection has been reported in subsequent
666 retrospective observational case-control studies using jurisdictional health registry and
667 immunization data in various populations in the US, Australia and Italy, with vaccine
668 effectiveness of a two-dose schedule ranging between 33% and 46% (53-56). In a
669 retrospective matched cohort study of 15-30 year-olds resident in Southern California, the
670 incidence of *N. gonorrhoeae* infection among individuals who received 4CMenB (with or
671 without a MenACWY *N. meningitidis* serogroup A, C, W, Y polysaccharide conjugate vaccine)
672 was compared with the incidence of *N. gonorrhoeae* infection among individuals who
673 received the MenACWY vaccine alone; the hazard ratio (HR) of incident *N. gonorrhoeae*
674 infection was 0.54 (95% CI 0.34-0.86)(55).

675

676 In contrast, no association has been noted between receipt of an alternative *N. meningitidis*
677 serogroup B vaccine, MenB-fHbp (Trumenba; Pfizer) and *N. gonorrhoeae* infection. The
678 MenB-fHbp vaccine contains recombinant fHbp, but does not contain OMVs. Importantly,
679 the homologue of fHbp in *N. gonorrhoeae* is not surface exposed, does not bind factor H
680 and is therefore not predicted to be protective against *N. gonorrhoeae* infection (151). In a
681 retrospective, observational case-control study of 96,235 persons aged 16-23 years of age
682 with a diagnosis of *N. gonorrhoeae* or *C. trachomatis* infection between 2016 and 2018 in
683 New York City and Philadelphia, no significant association between MenB-fHbp vaccination
684 and *N. gonorrhoeae* mono-infection was observed after adjustment for ethnicity, gender
685 and jurisdiction (adjusted prevalence ratio 0.97, 95% CI=0.79-1.19)(151). This suggests that
686 healthy vaccinee bias (where persons who adopt preventive vaccinations may be more likely
687 to adopt other protective behaviours and therefore have reduced risk of disease acquisition)

688 has not played a significant role in the association between meningococcal serogroup B
689 OMV vaccine and *N. gonorrhoeae* protection.

690

691 Collectively, these retrospective, observational studies are limited by potential biases
692 resulting from possible missing data associated with the use of health and immunization
693 registry data. In addition, this non-randomised data may be confounded by differences in
694 risk behaviour between vaccinated and non-vaccinated persons, such that those who adopt
695 a preventative meningococcal vaccine may also be more likely to adopt preventative
696 behaviours that reduce the risk of *N. gonorrhoeae* infection. Further data are also required
697 to determine vaccine effectiveness against *N. gonorrhoeae* infection in subpopulations at
698 high risk of *N. gonorrhoeae* infection, such as people living with HIV (PLHIV) and men who
699 have sex with men (MSM). Encouragingly, the first study investigating the impact of
700 4CMenB in PLHIV demonstrated promising results. This retrospective case-control study
701 comprised of 1,051 MSM living with HIV in Milan, Italy and demonstrated vaccine
702 effectiveness of 4CMenB against *N. gonorrhoeae* infection of 42% (95% CI 6-64, $p=0.027$), a
703 figure that remained significant after adjustment in multivariable analysis (56). A further
704 uncertainty remains regarding whether there are any differences in the protective efficacy
705 of meningococcal serogroup B OMV-containing vaccines against *N. gonorrhoeae* infection at
706 specific anatomical sites (ie. genital, anorectal or oropharyngeal infections) as well as the
707 duration of vaccine-induced protection against *N. gonorrhoeae* infection.

708

709 Finally, the evidence regarding vaccine effectiveness against *N. gonorrhoeae* and *C.*
710 *trachomatis* co-infection in published studies is mixed. In the initial New Zealand
711 retrospective case-control study of MeNZB, vaccine effectiveness was observed against *N.*

gonorrhoeae/C. trachomatis co-infection, albeit with a lower effect size. The estimated vaccine effectiveness against *N. gonorrhoeae/C. trachomatis* co-infection compared to *C. trachomatis*-only controls was 14% (95% CI, 1-26%) in this study, while estimated vaccine effectiveness against *N. gonorrhoeae*-only infection was 31% (95% CI, 21-39%)(34). In contrast, in the large retrospective case-control study described above of 109,737 individuals aged 16-23 years with *C. trachomatis* or *N. gonorrhoeae* infection in New York City and Philadelphia between 2016 and 2018, vaccination with 4CMenB was not protective against *N. gonorrhoeae/C. trachomatis* co-infection, despite an estimated two-dose vaccine effectiveness against *N. gonorrhoeae*-only infection of 40% (95% CI 23-53%)(53).

Randomised studies

The interim findings of the first randomised study of a meningococcal serogroup B OMV vaccine with 4CMenB were made available in February 2023. The French National Agency for AIDS Research (ANRS) DOXYVAC trial was a phase III randomised open-label factorial design trial of MSM on HIV PrEP with a history of STI in the previous 12 months (148). In this study participants were randomised to two interventions, i) two doses of 4CMenB or no vaccine (randomised 1:1); and ii) doxycycline post-exposure prophylaxis (PEP) (200mg within 72 hours of condomless sex) or no PEP (randomised 2:1). Participants underwent testing for *N. gonorrhoeae* infection at baseline, every three months and whenever they had symptoms of STIs. Testing for *N. gonorrhoeae* infection comprised NAAT (nucleic acid amplification test) of urine, oropharyngeal and anorectal swabs every three months. The primary endpoint of the study was the incidence of a first episode of *N. gonorrhoeae* infection one month after the second dose using an intention-to-treat analysis. Of 546 MSM enrolled, 502 were included in the intention-to-treat analysis. The interim findings reported a significant

reduction in incident *N. gonorrhoeae* infection between the two-dose 4CMenB recipients and unimmunized participants followed for 9 months, with the incidence of first episode *N. gonorrhoeae* infection 9.8 and 19.7 per 100 person-years in the 4CMenB arm and no vaccine arms, respectively (adjusted hazard ratio 0.49; 95% CI 0.27-0.88). There was no interaction for the primary endpoints between the doxycycline PEP and 4CMenB vaccination. No vaccine-related serious adverse events were reported. However, the results of this study are now under independent review due to a discrepancy between the results of the reported interim and final results, explained by the omission of a number of *N. gonorrhoeae* infections from the interim analysis (149). As the first reported randomised trial of a meningococcal B OMV vaccine, the final results of this trial and independent review are highly anticipated.

Further randomised studies of a two-dose schedule of 4CMenB are currently underway and are described in Table 5. Notably, four double-blind randomised-controlled trials are actively enrolling participants, including two large placebo-controlled multi-centre clinical trials and a CHIM study (<https://clinicaltrials.gov/study/NCT04415424>; <https://clinicaltrials.gov/study/NCT04350138>; <https://clinicaltrials.gov/study/NCT05766904>; <https://clinicaltrials.gov/study/NCT05294588>). In addition, a randomised, open-label, single-site trial of 18-50 year-old gay and bisexual men on HIV pre-exposure prophylaxis or recent *N. gonorrhoeae* infection is planned (147). The three double-blind, randomised, placebo-controlled trials evaluating the impact of 4CMenB on natural infection will recruit from different populations, including i) a multi-site Australian study of 18-50 year-old men (cis and trans), transsexual women and non-binary people who have sex with men(<https://clinicaltrials.gov/study/NCT04415424>); ii) a multi-site American study of 18-50

year-old healthy men and women (<https://clinicaltrials.gov/study/NCT04350138>); and iii) a single-site Hong Kong study of MSM aged 18 or above with risk factors for gonorrhoea infection (<https://clinicaltrials.gov/study/NCT05766904>). The randomised-controlled CHIM study is a single-site, double-blind randomised controlled trial where two doses of 4CMenB are compared to quadrivalent influenza and tetanus/diphtheria vaccination. The aim is to recruit up to 140 male participants who will undergo urethral challenge with *N. gonorrhoeae* strain FA1090 after immunization with 4CMenB or the comparator vaccine. Participants will be randomised 1:1 to the 4CMenB or control vaccine arm, and receive two immunizations prior to the anterior urethral bacterial challenge with 10^6 colony-forming units of *N. gonorrhoeae* strain FA1090 in suspension. The primary outcome measured will be microbiological confirmation of urethral infection via detection of *N. gonorrhoeae* by culture or NAAT of urine or urethral swab (<https://clinicaltrials.gov/study/NCT05294588>). In addition, this CHIM will measure the proportion of participants that develop symptomatic disease and also presents an opportunity for intensive biological sampling and immunological characterisation of responses in those who have received 4CMenB compared to the control group. Furthermore, subgroup analysis of this data regarding anatomical site-specific risk of *N. gonorrhoeae* infection (eg. genital, anorectal and oropharyngeal) between the vaccinated and non-vaccinated groups in these randomised studies will be important to inform the potential impact of the 4CMenB vaccine on *N. gonorrhoeae* transmission at a population level. Increasing evidence suggests that oropharyngeal *N. gonorrhoeae* infection may play a significant role in *N. gonorrhoeae* transmission (19) and modelling studies suggest that the impact of a *N. gonorrhoeae* vaccine will be significantly reduced if the vaccine is not effective at the oropharynx (152).

Biological plausibility

Biological plausibility for the association between meningococcal serogroup B OMV vaccines and protection against *N. gonorrhoeae* infection has been strengthened by basic science studies demonstrating a high level of genomic sequence identity between *N. gonorrhoeae* and the serogroup B *N. meningitidis* OMV protein antigens present in the MenZB and 4CMenB vaccines (94, 153). Bioinformatic analysis of twenty-two proteins that comprise >90% of 4CMenB OMV content resulted in the identification of twenty orthologues of these proteins in *N. gonorrhoeae* strain FA1090, including sixteen with >90% identity and two with >80% identity (153). Of the OMV proteins that have an orthologue in *N. gonorrhoeae*, fourteen of these also have a high level of sequence identity with >400 *N. gonorrhoeae* strains available on GenBank (153). A further study comprising bioinformatic analysis of abundant 4CMenB OMV vaccine antigens among 940 *N. gonorrhoeae* strains from the US, found that of all the predicted outer membrane proteins, OpcA (45%) and PorB (70%) had the lowest mean sequence similarity between the NZ98/254 *N. meningitidis* strain from which the 4CMenB OMV is derived and *N. gonorrhoeae*. In addition, although the *porA* gene was identified in 99.5% of *N. gonorrhoeae* isolates in this study, inactivating mutations render PorA a pseudogene in *N. gonorrhoeae* (94). Analysis of the additional recombinant antigens present in 4CMenB indicates that NadA is absent in *N. gonorrhoeae* (94) and although orthologues of NHBA, fHbp, GNA2091 and GNA1030 are present in *N. gonorrhoeae* strains, fHbp (154), GNA2091 (155) and GNA1030 (156) are not thought to be surface exposed. Importantly, the 4CMenB NZ98/254 *N. meningitidis* strain NHBA antigen shares 67% mean amino acid sequence similarity to *N. gonorrhoeae* (153), suggesting the presence of this antigen in the 4CMenB vaccine may provide an additive protective effect against *N. gonorrhoeae* infection.

808

809 In addition, analysis of the antibody response of rabbits, mice and humans after
810 immunization with 4CMenB, or the OMV present in 4CMenB has demonstrated the
811 induction of cross-reacting gonorrhoea-specific antibodies (153, 157). For example, in
812 rabbits immunized with the OMV present in 4CMenB, several cross reactive proteins were
813 detected by Western blot analysis of whole-cell lysates comprising three different *N.*
814 *gonorrhoeae* strains, and an elevated ELISA titre to *N. gonorrhoeae* strain 1291 OMVs was
815 observed (153). Similar findings were observed in a serology study of humans who had
816 received three doses of 4CMenB, with a significant rise in the enzyme-linked
817 immunosorbent assay (ELISA) titre against *N. gonorrhoeae* whole-cell lysates between pre-
818 and post-vaccination. Western blot analysis of human post-vaccination sera also
819 demonstrated reactivity to several gonococcal and meningococcal proteins (153). Further
820 investigation in the estradiol-treated female mouse model demonstrated that subcutaneous
821 and intraperitoneal immunization of mice with 4CMenB induced serum and vaginal
822 antibodies to whole-cell lysates of six different *N. gonorrhoeae* strains, as well as serum and
823 vaginal antibodies that cross-react with several OMV proteins, including promising *N.*
824 *gonorrhoeae* vaccine targets such as MtrE and PilQ. Furthermore, vaccination with 4CMenB
825 significantly reduced *N. gonorrhoeae* bacterial load and accelerated clearance of infection
826 after *N. gonorrhoeae* vaginal inoculation in the estradiol-treated mouse model (157).

827

828 A number of investigators are currently undertaking studies to further characterise the
829 immunological responses to a two-dose schedule of 4CMenB vaccine. These include a study
830 comprising up to 15 male and female participants conducted at the University of North
831 Carolina, Chapel Hill in which the change in anti-gonococcal OMV- specific IgG, IgA and

immunoglobulin M (IgM) concentrations and the mean change in the proportion of CD4+ T lymphocytes expressing at least two different activation markers (interferon-gamma, tumour necrosis factor-alpha and interleukin-2) will be measured after *in vitro* stimulation with *N. gonorrhoeae* strain FA1090 OMVs in participants after vaccination with two doses of 4CMenB (<https://clinicaltrials.gov/study/NCT04094883>). In another study, investigators at the University of Oxford and KEMRI-Wellcome Trust Collaborative Research Program aim to recruit approximately 50 male and female participants, including HIV-uninfected and HIV-infected individuals from existing follow-up cohorts in Mtwapa, Kenya. These investigators will also measure serum humoral and T cell responses to *N. gonorrhoeae* before and after two-doses of 4CMenB (<https://clinicaltrials.gov/study/NCT04297436>). Furthermore, in a study at the National Institute of Allergy and Infectious Diseases, 50 male and female participants will be recruited and serum and mucosal antibody responses at oropharyngeal, rectal and vaginal sites will be measured before and after vaccination with two-dose of 4CMenB (<https://clinicaltrials.gov/study/NCT04722003>). In addition, a number of the randomised two-dose 4CMen B vaccine efficacy studies described above will investigate serum (<https://clinicaltrials.gov/study/NCT04350138>) or serum and mucosal immune responses (<https://clinicaltrials.gov/study/NCT04415424>, 147).

849

In summary, there is substantial evidence of an association between meningococcal serogroup B OMV vaccines and reduced *N. gonorrhoeae* infection. This includes human ecological and observational trial data, evidence of overlap in important vaccine targets in meningococcal serogroup B OMV vaccines and *N. gonorrhoeae* and induction of cross-reactive antibody responses. Lacking are data defining the impact of meningococcal serogroup B OMV vaccines on *N. gonorrhoeae* infection at various anatomical sites and in

different population groups. With a number of randomised-controlled studies assessing the vaccine efficacy of 4CMenB currently underway, further information will become available. Given the promising findings of meningococcal serogroup B OMV vaccines against *N. gonorrhoeae* infection to date, as well as the widespread availability and demonstrated safety data of vaccines such as 4CMenB (158), implementation of this vaccine in settings with particularly high *N. gonorrhoeae* prevalence should be considered.

IN THE PIPELINE: NEISSERIA GONORRHOEAE OUTER MEMBRANE VESICLE VACCINES

Importantly, several *N. gonorrhoeae*-specific OMV vaccines are in preclinical or clinical development (141, 159, <https://clinicaltrials.gov/study/NCT05630859>). These include the NGoXIM (141) and dmGC_0817560 (140, 159) native OMV vaccine candidates which are in the late stages of preclinical development (160), and a generalized modules for membrane antigens (GMMA) vaccine, which is currently recruiting participants into a phase 1/2 study (<https://clinicaltrials.gov/study/NCT05630859>). The NGoXIM vaccine is being developed by Intravacc and TherapyX in the Netherlands and the United States, with funding from the US National Institute of Allergy and Infectious Diseases (161). This vaccine is a *N. gonorrhoeae* native OMV vaccine formulated for intranasal mucosal delivery combined with a sustained-release microsphere encapsulated IL-12 adjuvant (160). Studies have demonstrated that intravaginal and intranasal administration of this vaccine induced Th1-driven responses that accelerated clearance of *N. gonorrhoeae* genital tract infection in mice (121, 141). Intranasal administration of this experimental vaccine generated antgonococcal serum IgG, salivary IgA and vaginal IgG and IgA antibodies in female mice and antgonococcal serum IgG and salivary IgA antibodies in male mice. In addition, female mice that received intranasal immunization with this experimental vaccine demonstrated accelerated clearance of

880 homologous and heterologous strains of *N. gonorrhoeae* infection. Further, intranasal
881 immunization with vaccines comprising various adaptations to this vaccine include
882 detergent-extracted OMVs to reduce LOS content, and OMVs from *N. gonorrhoeae* strains
883 with deleted *rmp* and *lpxL1* genes to eliminate anti-Rmp blocking antibodies and reduce LOS
884 endotoxicity. These have shown accelerated clearance of vaginal gonococcal infection in the
885 female mouse model (141).

886

887 The dmGC_0817560 vaccine is being developed by the Jenner Institute and Oxford Vaccine
888 Group in the United Kingdom, with funding from CARB-X (160). This vaccine is also a native
889 OMV vaccine formulated from a Chilean *N. gonorrhoeae* strain in which genes for Rmp and
890 LpxL1 have deleted, combined with an aluminium hydroxide adjuvant. Preclinical studies
891 demonstrate that parenteral delivery of this experimental vaccine induced anti-gonococcal
892 serum and vaginal mucosal IgG and IgA antibodies and gonococcal-specific Th1/Th17 CD4+
893 T-cell responses in the female mouse model. In addition, female mice immunized with the
894 candidate vaccine demonstrated accelerated clearance of genital *N. gonorrhoeae* infection
895 with a heterologous strain and cleared infection significantly faster than mice immunized
896 with 4CMenB (140).

897

898 The intramuscular NgG generalized modules for membrane antigens (GMMA) vaccine is
899 being developed by GlaxoSmithKline in the US
900 (<https://clinicaltrials.gov/study/NCT05630859>). GMMA vaccines are OMV vaccines that have
901 been produced from bacterial strains that have been genetically modified to increase
902 production of OMVs and reduce endotoxin levels (162). To our knowledge, preclinical studies
903 of this experimental vaccine have not been published, however a phase 1/2 study of this

904 experimental vaccine has commenced recruitment, aiming to evaluate the safety,
905 reactogenicity, immunogenicity and efficacy of this experimental vaccine in a randomised,
906 observer-blind, placebo-controlled multicentre study in an estimated 774 participants aged
907 18-50 years of age (<https://clinicaltrials.gov/study/NCT05630859>). The phase 1 dose-
908 escalation safety study for this vaccine is now complete and the study has entered phase 2;
909 furthermore, the US Food and Drug Administration (FDA) has granted a Fast Track
910 designation to accelerate its path to US FDA submission (163).

911

912 These vaccines represent the next generation of anti-gonococcal OMV vaccines that have
913 been specifically engineered to build on the scientific advances in understanding of *N.*
914 *gonorrhoeae* pathogenesis and host immune response, as well as the significant progress
915 made in the past decade to explore the association between serogroup B meningococcal
916 OMV vaccines and reduced *N. gonorrhoeae* infection. These include i) use of a *N.*
917 *gonorrhoeae* strain to produce OMVs for use in next-generation vaccines, potentially
918 increasing the specificity of the immune responses induced by this multi-antigen vaccine
919 technology; ii) inclusion of adjuvants that stimulate a Th1 response; iii) genetically modifying
920 selected gonococcal strains to reduce the endotoxicity associated with LOS and blocking
921 antibodies induced by Rmp; and iv) evaluation of mucosal administration to increase the
922 immune response at the mucosal sites of gonorrhoea infection.

923

924 **POTENTIAL PUBLIC HEALTH IMPACT OF A NEISSERIA GONORRHOEAE VACCINE**

925 Determining the potential public health impact of a *N. gonorrhoeae* vaccine requires
926 consideration of the health, economic and societal value of future *N. gonorrhoeae* vaccines.
927 The WHO convened an international panel of experts in 2019 to define the public health

value and preferred product characteristics of *N. gonorrhoeae* vaccines (37, 38). At this meeting, prevention of poor sexual and reproductive health outcomes and addressing the threat of AMR were identified as the key goals of future *N. gonorrhoeae* vaccines. Important considerations to define the target product profile of a *N. gonorrhoeae* vaccine include i) defining the target endpoint for assessment of vaccine efficacy (eg. prevention of infection, versus prevention of symptomatic disease, versus prevention of AMR); ii) the target population for the vaccine (eg. all individuals prior to sexual activity or high risk populations; whether to include both females and males) and iii) the target programmatic delivery program (eg. schools or sexual health clinics). Notably, the preferred product characteristics of a potential vaccine may also vary according to the epidemiology of *N. gonorrhoeae* infection and AMR in the target population. The promotion of a vaccine against a sexually transmitted infection may also require adaptation to the specific socio-cultural context in order to maximise acceptability.

Modelling studies are important to understanding the potential impact of *N. gonorrhoeae* vaccines on gonococcal infection and AMR, and to aid policy development and programme delivery. The public health impact and cost-effectiveness of potential *N. gonorrhoeae* vaccines have been modelled in several studies, including various target population groups, vaccine program strategies and levels of vaccine coverage. In addition, the effects of various levels of vaccine efficacy and duration of protection have been investigated.

Modelling The Impact of *Neisseria gonorrhoeae* Vaccines in Heterosexual Populations

The impact of gonococcal vaccines delivered prior to commencement of sexual activity has been estimated in a number of heterosexual population model studies. Craig et al used an

individual-based, epidemiological simulation model of a *N. gonorrhoeae* vaccine delivered prior to commencement of sexual activity in a heterosexual population of 100,000 individuals using theoretical vaccines of 10-100% efficacy and 2.5-20 year duration of protection (164). The model output predicted that *N. gonorrhoeae* prevalence could be reduced by at least 90% after 20 years by a non-waning vaccine with 50% efficacy and universal vaccination coverage. The duration of protection of a theoretical vaccine had a significant effect on the prevalence of *N. gonorrhoeae* in the model; a vaccine with 100% efficacy that waned after 7.5 years was predicted to reduce *N. gonorrhoeae* prevalence by at least 90% after 20 years, one whose protection waned after 5 years by 50% and one with 2.5 years protection having minimal impact on prevalence. Similarly, vaccine coverage played a key role in predicted vaccine impact, with 50% vaccine coverage of a *N. gonorrhoeae* vaccine with 50% efficacy predicted to reduce *N. gonorrhoeae* prevalence by 50% after 20 years, compared to at least 90% reduction if the same vaccine had universal vaccine coverage (164).

The impact of a 4CMenB adolescent vaccine on *N. gonorrhoeae* prevalence has been estimated in a number of studies using transmission models of *N. gonorrhoeae* infection among heterosexual populations. Carey et al developed a heterosexual transmission model of 15-24 year-olds in the US using Approximate Bayesian Computation analysis to account for uncertainty in key transmission factors (rates of natural clearance, rates of screening, proportion of symptomatic infections and annual number of sexual contacts). The results of this analysis estimated that a vaccine with 30% efficacy and 2-year duration of protection would result in a 12.2-39.4% reduction in *N. gonorrhoeae* prevalence if 50% vaccine coverage was achieved in this population, and 4.8-14.3% reduction in prevalence if 20%

vaccine coverage was achieved (165). Looker et al developed a developed a deterministic transmission-dynamic model of heterosexual 13-64 year-olds in England and estimated the impact of a vaccinating 14-year-olds with a vaccine with 31% efficacy, 6-year duration of protection and 85% vaccine uptake (166). The results of this analysis indicated that 10% (95%CrI 8-13%), 18% (95%CrI 13-23%) and 25% (95%CrI 17-33%) of cases of *N. gonorrhoeae* infections would be prevented in this population over a 10-, 20- and 70-year period, respectively (166). Regnier et al modelled the potential health and economic impact of a 4CMenB adolescent vaccination on *N. gonorrhoeae* infection with an estimated 20% vaccine efficacy, 10-year duration of protection and 70.5% vaccination rate using a decision-analysis model developed using published US healthcare utilization and cost data (167). This model predicted that vaccination could prevent 83,167 lifetime *N. gonorrhoeae* infections and 55 lifetime HIV infections per vaccinated birth cohort in the US. This was predicted to reduce the direct medical costs of *N. gonorrhoeae* infection by US\$28.7 million and reduce income and productivity losses by US\$40.0 million (167).

Modelling the Impact of *Neisseria gonorrhoeae* Vaccines in Men Who Have Sex With Men Populations

The impact of gonococcal vaccines within a male population of MSM has been modelled in four studies. Using a stochastic transmission-dynamic model that incorporated heterogenous sexual behaviour and symptomatic and asymptomatic infection in an MSM population based on surveillance data from England, Whittles et al assessed potential *N. gonorrhoeae* vaccination impact and the feasibility of achieving the WHO target of reducing *N. gonorrhoeae* incidence by 90% by 2030 (168). This study estimated that the WHO target is achievable even if the worst-case scenario where untreatable AMR infection emerges, if

1000 all MSM attending sexual health clinics receive a vaccine with $\geq 52\%$ efficacy and ≥ 6 years or
1001 vaccination; or $\geq 70\%$ efficacy and ≥ 3 years protection (168). Heinje et al developed a
1002 compartmental model of *N. gonorrhoeae* transmission among a population of MSM with
1003 heterogenous sexual behaviour and symptomatic and asymptomatic infection. This model
1004 also incorporated AMR as a stepwise increase in minimum inhibitory concentration (MIC)
1005 and eventual resistance to ceftriaxone. The impact of a partially protective vaccine with 30%
1006 efficacy that provided 2 years of protection delivered to high risk MSM (with baseline
1007 gonorrhoea prevalence of 12.5%) on *N. gonorrhoeae* prevalence and AMR was assessed.
1008 The modelling output indicated that a vaccine with 30% vaccine effectiveness could not
1009 prevent AMR despite high uptake or long-term protection, but would increase time to
1010 development of AMR by several years (169).

1011

1012 More recent modelling studies of *N. gonorrhoeae* vaccines within male populations of MSM
1013 have added increasing layers of complexity to their models. Hui et al simulated anatomical
1014 site-specific data into their individual-based mathematical model of *N. gonorrhoeae*
1015 transmission in an urban population of 10,000 MSM with heterogenous sexual behaviour
1016 and symptomatic and asymptomatic infection (152). Three types of vaccine efficacy were
1017 investigated, including i) 'protective efficacy', the protection of a vaccinated individual
1018 against acquiring *N. gonorrhoeae* infection; ii) 'transmission suppression efficacy', the
1019 reduction of *N. gonorrhoeae* transmission from a vaccinated individual; and iii) 'symptom
1020 suppression efficacy', the reduction of symptoms of *N. gonorrhoeae* infection in the setting
1021 of infection in a vaccinated individual. It was estimated that *N. gonorrhoeae* elimination may
1022 be possible within the population in this model in eight years with vaccines with $\geq 50\%$
1023 efficacy and two years of protection if 30% of MSM presenting for sexually transmitted

1024 infection testing were vaccinated and underwent a booster vaccination every three years.
1025 Importantly, it was estimated that vaccine impact may be substantially reduced if a *N.*
1026 *gonorrhoeae* vaccine is not effective at the oropharynx and that prevalence may actually
1027 increase if a vaccine prevents symptoms but does not prevent infection or transmission. In
1028 addition, this study estimated that *N. gonorrhoeae* vaccines that reduced transmission
1029 without conferring protection from *N. gonorrhoeae* infection would have a similar impact on
1030 *N. gonorrhoeae* prevalence as vaccines with protective efficacy, and that the impact of
1031 vaccines with both transmission suppression and protective efficacy would be additive
1032 (152).

1033

1034 Whittles et al's most recent study incorporated a cost-effectiveness analysis into their
1035 transmission-dynamic model that incorporated heterogenous sexual behaviour and
1036 symptomatic and asymptomatic infection in an MSM population based on surveillance data
1037 from England (170). The impact and cost-effectiveness of four different vaccination
1038 strategies were assessed in this study. It was estimated that vaccination of adolescents in
1039 schools would have little impact on *N. gonorrhoeae* prevalence, whereas vaccination of
1040 individuals on attendance for STI testing at sexual health clinics would have the largest
1041 impact. Vaccination on diagnosis of *N. gonorrhoeae* infection at sexual health clinics would
1042 have a moderate impact but require fewer doses than a vaccination on attendance
1043 approach, while vaccination of sexual health clinic attendees according to risk (defined as
1044 individuals diagnosed with *N. gonorrhoeae* infection in the past 12 months or with >5 sexual
1045 partners per year) was estimated to have a similar impact as vaccination of all STI clinic
1046 attendees, however required administration of fewer vaccine doses. The most cost-effective
1047 strategy for vaccines with moderate efficacy or duration of protection was vaccination

1048 according to risk, whereas vaccination on diagnosis of *N. gonorrhoeae* infection was most
1049 cost-effective for highly efficacious and long-lasting vaccines. The impact of 4CMenB
1050 vaccination against *N. gonorrhoeae* infection, assuming a vaccine efficacy of 31% and
1051 protection lasting 18 months after two-dose primary vaccination and 36 months after
1052 single-dose booster vaccination, was also evaluated. A strategy comprising 4CMenB
1053 vaccination administered according to risk was estimated to prevent 110,200 cases, gaining
1054 a mean of 100.3 QALYs and save a mean £7.9 million over 10 years (170).

1055

1056 **Modelling The Impact of *Neisseria gonorrhoeae* Vaccines In Low- and Middle-Income** 1057 **Settings**

1058 The use of modelling to assess the impact of *N. gonorrhoeae* vaccines in a high prevalence
1059 LMIC setting was reported in a recent study (171). Using a compartmental model of *N.*
1060 *gonorrhoeae* transmission among a 15-49-year-old heterosexual population in a high
1061 prevalence LMIC setting similar to South Africa, Padeniya et al modelled the impact of
1062 vaccines with varying levels of protective and transmission suppression efficacy on the
1063 prevalence *N. gonorrhoeae* infection. In addition, the impact of vaccination programs
1064 delivered to various age- and sexual-activity groups was assessed. Vaccination of 15-49-
1065 year-olds with a vaccine with protective efficacy of 25%, a 5 year duration of protection and
1066 10% annual vaccine uptake would have the greatest impact on *N. gonorrhoeae* prevalence,
1067 with the model predicting that a 50% reduction in prevalence would be achieved, compared
1068 to 25% reduction in prevalence if only 15-24-year-olds were vaccinated. Vaccination of only
1069 individuals with high sexual-activity was predicted to achieve an almost equivalent
1070 reduction in *N. gonorrhoeae* prevalence to vaccinating the entire 15-49-year-old population
1071 using theoretical vaccines with same efficacy, duration of protection and uptake

1072 characteristics over the same time period, but was able to achieve this more efficiently,
1073 requiring approximately 3 times fewer vaccinations. Similar to the findings of the modelling
1074 study by Hui et al's of an urban MSM population, this study estimated that a vaccine with
1075 both protective and transmission suppression efficacy would have an additive impact on
1076 reducing *N. gonorrhoeae* prevalence (171).

1077

1078 In summary, modelling studies undertaken in both heterosexual and MSM populations using
1079 data from various international settings have demonstrated that delivery of vaccines with
1080 efficacy and duration of protection derived from estimates of the currently-available
1081 4CMenB vaccine could have a significant impact on *N. gonorrhoeae* prevalence, and even be
1082 cost-saving when implemented in select high-risk populations (167, 170). In addition, such
1083 vaccines could delay the development of AMR, providing time for more efficacious vaccines
1084 and novel antimicrobials to be developed (169). Furthermore, even moderate
1085 improvements in *N. gonorrhoeae* vaccine efficacy and duration of protection may have a
1086 significant impact on *N. gonorrhoeae* infection prevalence, with some studies estimating
1087 that *N. gonorrhoeae* infection may be eliminated or prevalence reduced by 90% through the
1088 implementation of vaccines with approximately 50% efficacy and 2-6 years duration of
1089 protection (152, 168). Given the prediction that vaccine impact may be reduced if a vaccine
1090 is not effective at the oropharynx (152), further data regarding vaccine efficacy at different
1091 anatomical sites is pivotal in informing current and future vaccine implementation
1092 strategies. In addition, modelling studies simulating the epidemiological characteristics of *N.*
1093 *gonorrhoeae* infection in LMIC settings, where the burden of gonorrhoea infection is
1094 greatest, should be prioritised.

1095

1096 **QUESTIONS REMAINING: RESEARCH PRIORITIES FOR GONOCOCCAL VACCINES**

1097 This is an exciting time for *N. gonorrhoeae* vaccine development, with evidence from
1098 observational studies suggesting that meningococcal B OMV vaccines may have efficacy
1099 against *N. gonorrhoeae* infection and multiple randomised trials underway. However, there
1100 are several key questions that remain unanswered about the currently available serogroup
1101 B meningococcal vaccines. These include i) the major effector antigen/s responsible for the
1102 efficacy of OMV vaccines; ii) the efficacy of vaccination on infection at various anatomical
1103 sites; iii) the duration of protective immunity; and iv) whether there is an immune correlate
1104 of protection that can be measured by laboratory tests. A number of these knowledge gaps,
1105 such as efficacy at various anatomical sites and further data on duration of protective
1106 immunity may be informed by currently recruiting clinical trials of the 4CMenB vaccine. In
1107 addition, a randomised trial of 4CMenB in a male urethritis gonorrhoea CHIM may provide
1108 more detailed data regarding immune responses to key serogroup B meningococcal OMV
1109 vaccine antigens.

1110

1111 The priority research areas outlined in the WHO Global STI Vaccine Roadmap and recently
1112 reviewed in the WHO stakeholder consultation regarding public health value and preferred
1113 product characteristics of gonococcal vaccines in 2019 remain pertinent today. These
1114 include i) improving access to quality epidemiological data regarding infection including
1115 AMR ii) advancing the understanding of the natural history of gonorrhoea infection; iii)
1116 modelling predicted gonorrhoea vaccine impact and cost-effectiveness; iv) accelerating
1117 basic science, translational, immunobiologic and clinical research; and v), advocating for
1118 investment and planning for policy and implementation decisions (35, 37). Although there is

1119 much work to be done, there is significant momentum in *N. gonorrhoeae* vaccine
1120 development that is being fuelled by the bench to bedside research described in this review.

1121

1122 **CONCLUSION**

1123 In this review, we have described the unique challenges involved in development of a *N.*
1124 *gonorrhoeae* vaccine. We have reviewed the breadth of data pertaining to *N. gonorrhoeae*
1125 vaccines, ranging from an overview of historical vaccines; to multi-omics vaccine antigen
1126 discovery and preclinical vaccine research; as well as contemporary clinical trials and
1127 modelling studies to inform potential vaccine implementation strategies. As we approach an
1128 important inflection point, with the imminent release of the results of six randomised trials
1129 of the efficacy of 4CMenB against *N. gonorrhoeae* infection, it is important to consider first
1130 how to best implement vaccination programs using currently available vaccines to protect
1131 against *N. gonorrhoeae* infection and secondly how to improve upon these technologies to
1132 develop the next generation of *N. gonorrhoeae* vaccines. The next generation of *N.*
1133 *gonorrhoeae*-specific OMV vaccines that include modifications of currently available
1134 vaccines may improve efficacy. However, alternative vaccines utilizing a range of gonococcal
1135 antigens that have shown promise in preclinical studies should also be pursued. Although
1136 these vaccine candidates are at a much earlier stage of development and their safety and
1137 efficacy in humans has not yet been demonstrated, there is good reason to hold optimism
1138 that they will confer improved protection over those currently available. As *N. gonorrhoeae*
1139 prevalence continues to increase and the threat of AMR to treatment of gonorrhoea
1140 becomes increasingly urgent, expediting the development of highly efficacious *N.*
1141 *gonorrhoeae* vaccines and implementing high-coverage vaccine programs is a key priority
1142 for sexual and reproductive health.

1143

1144 **ACKNOWLEDGMENTS**

1145 This project was supported by a Medical Research Future Fund Clinician Researcher Grant
1146 (MRFAR000354). E.W. is supported by a Postgraduate Scholarship from the National Health
1147 and Medical Research Council (NHMRC) (GNT2005380). K.L.S is supported by an NHMRC
1148 Leadership Investigator Grant (GNT2017383). C.K.F. is supported by an NHMRC Leadership
1149 Investigator Grant (GNT1172900). J.S.M is supported by an NHMRC Leadership Investigator
1150 Grant (GNT2016396). D.A.W. is supported by an NHMRC Investigator Grant (GNT1174555).

1151

1152 **REFERENCES**

1153

- 1154 1. Rowley J, Vander Hoorn S, Korenromp E, Low N, Unemo M, Abu-Raddad LJ, Chico
1155 RM, Smolak A, Newman L, Gottlieb S, Thwin SS, Broutet N, Taylor MM. 2019.
1156 Chlamydia, gonorrhoea, trichomoniasis and syphilis: global prevalence and incidence
1157 estimates, 2016. Bull World Health Organ 97:548-562P.
- 1158 2. Newman L, Rowley J, Vander Hoorn S, Wijesooriya NS, Unemo M, Low N, Stevens G,
1159 Gottlieb S, Kiarie J, Temmerman M. 2015. Global estimates of the prevalence and
1160 incidence of four curable sexually transmitted infections in 2012 based on systematic
1161 review and global reporting. PLoS One 10:e0143304.
- 1162 3. United States Centers for Disease Control and Prevention. 2022. Sexually
1163 transmitted disease surveillance 2020. US Department of Health and Human
1164 Services, Atlanta.
- 1165 4. European Centre for Disease Prevention and Control. 2020. Gonorrhoea. In: ECDC.
1166 Annual epidemiological report for 2018. ECDC, Stockholm.

- 1167 5. Kirby Institute. 2021. HIV, viral hepatitis and sexually transmissible infections in
1168 Australia: annual surveillance report 2021. Kirby Institute, Sydney.
- 1169 6. Unemo M, Bradshaw CS, Hocking JS, de Vries HJC, Francis SC, Mabey D, Marrazzo JM,
1170 Sonder GJB, Schwebke JR, Hoornenborg E, Peeling RW, Philip SS, Low N, Fairley CK.
1171 2017. Sexually transmitted infections: challenges ahead. *Lancet Infect Dis* 17:e235-
1172 e279.
- 1173 7. Chow EPF, Grulich AE, Fairley CK. 2019. Epidemiology and prevention of sexually
1174 transmitted infections in men who have sex with men at risk of HIV. *Lancet HIV*
1175 6:e396-e405.
- 1176 8. Kirkcaldy RD, Weston E, Segurado AC, Hughes G. 2019. Epidemiology of gonorrhoea:
1177 a global perspective. *Sex Health* 16:401-411.
- 1178 9. Unemo M, Seifert HS, Hook EW, 3rd, Hawkes S, Ndowa F, Dillon JR. 2019.
1179 Gonorrhoea. *Nat Rev Dis Primers* 5:79.
- 1180 10. Lovett A, Duncan JA. 2018. Human immune responses and the natural history of
1181 *Neisseria gonorrhoeae* infection. *Front Immunol* 9:3187.
- 1182 11. McCormack WM, Stumacher RJ, Johnson K, Donner A. 1977. Clinical spectrum of
1183 gonococcal infection in women. *Lancet* 1:1182-5.
- 1184 12. Barlow D, Phillips I. 1978. Gonorrhoea in women. Diagnostic, clinical, and laboratory
1185 aspects. *Lancet* 1:761-4.
- 1186 13. Vallely LM, Egli-Gany D, Wand H, Pomat WS, Homer CSE, Guy R, Silver B, Rumbold
1187 AR, Kaldor JM, Vallely AJ, Low N. 2021. Adverse pregnancy and neonatal outcomes
1188 associated with *Neisseria gonorrhoeae*: systematic review and meta-analysis. *Sex*
1189 *Transm Infect* 97:104-111.

- 1190 14. Desenclos JC, Garrity D, Scaggs M, Wroten JE. 1992. Gonococcal infection of the
1191 newborn in Florida, 1984-1989. *Sex Transm Dis* 19:105-10.
- 1192 15. Fall B, Sow Y, Mansouri I, Sarr A, Thiam A, Diao B, Fall PA, Ndoeye AK, Ba M, Diagne
1193 BA. 2011. Etiology and current clinical characteristics of male urethral stricture
1194 disease: experience from a public teaching hospital in Senegal. *Int Urol Nephrol*
1195 43:969-74.
- 1196 16. Ochsendorf FR. 2008. Sexually transmitted infections: impact on male fertility.
1197 *Andrologia* 40:72-5.
- 1198 17. Tabesh M, Fairley CK, Hocking JS, Williamson DA, Zhang L, Xu X, Bradshaw CS, Chen
1199 MY, Chow EP. 2022. Comparison of the patterns of chlamydia and gonorrhoea at the
1200 oropharynx, anorectum and urethra among men who have sex with men. *Sex*
1201 *Transm Infect* 98:11-16.
- 1202 18. Chan PA, Robinette A, Montgomery M, Almonte A, Cu-Uvin S, Lonks JR, Chapin KC,
1203 Kojic EM, Hardy EJ. 2016. Extragenital infections caused by *Chlamydia trachomatis*
1204 and *Neisseria gonorrhoeae*: a review of the literature. *Infect Dis Obstet Gynecol*
1205 2016:5758387.
- 1206 19. Fairley CK, Cornelisse VJ, Hocking JS, Chow EPF. 2019. Models of gonorrhoea
1207 transmission from the mouth and saliva. *Lancet Infect Dis* 19:e360-e366.
- 1208 20. Sawatzky P, Martin I, Thorington R, Alexander D. 2022. Disseminated gonococcal
1209 infections in Manitoba, Canada: 2013 to 2020. *Sex Transm Dis* 49:831-837.
- 1210 21. Ward H, Rönn M. 2010. Contribution of sexually transmitted infections to the sexual
1211 transmission of HIV. *Curr Opin HIV AIDS* 5:305-310.
- 1212 22. Unemo M, Shafer WM. 2014. Antimicrobial Resistance in *Neisseria gonorrhoeae* in
1213 the 21st Century: Past, Evolution, and Future. *Clin Microbiol Rev* 27:587.

- 1214 23. World Health Organization. 2017. Global priority list of antibiotic-resistant bacteria
1215 to guide research, discovery, and development of new antibiotics. World Health
1216 Organization, Geneva.
- 1217 24. Centers for Disease Control and Prevention. 2019. Antibiotic resistance threats in the
1218 United States, 2019. US Department of Health and Human Services, Atlanta.
- 1219 25. Fifer H, Natarajan U, Jones L, Alexander S, Hughes G, Golparian D, Unemo M. 2016.
1220 Failure of dual antimicrobial therapy in treatment of gonorrhea. *N Engl J Med*
1221 374:2504-6.
- 1222 26. Whiley DM, Jennison A, Pearson J, Lahra MM. 2018. Genetic characterisation of
1223 *Neisseria gonorrhoeae* resistant to both ceftriaxone and azithromycin. *Lancet Infect*
1224 *Dis* 18:717-718.
- 1225 27. Eyre DW, Sanderson ND, Lord E, Regisford-Reimmer N, Chau K, Barker L, Morgan M,
1226 Newnham R, Golparian D, Unemo M, Crook DW, Peto TE, Hughes G, Cole MJ, Fifer H,
1227 Edwards A, Andersson MI. 2018. Gonorrhoea treatment failure caused by a *Neisseria*
1228 *gonorrhoeae* strain with combined ceftriaxone and high-level azithromycin
1229 resistance, England, February 2018. *Euro Surveill* 23(27):1800323.
- 1230 28. Williams E, Fairley CK, Williamson D. 2021. Novel strategies for prevention and
1231 treatment of antimicrobial resistance in sexually-transmitted infections. *Curr Opin*
1232 *Infect Dis* 34:591-598.
- 1233 29. Chen MY, McNulty A, Avery A, Whiley D, Tabrizi SN, Hardy D, Das AF, Nenninger A,
1234 Fairley CK, Hocking JS, Bradshaw CS, Donovan B, Howden BP, Oldach D. 2019.
1235 Solithromycin versus ceftriaxone plus azithromycin for the treatment of
1236 uncomplicated genital gonorrhoea (SOLITAIRE-U): a randomised phase 3 non-
1237 inferiority trial. *Lancet Infect Dis* 19:833-842.

- 1238 30. Taylor SN, Marrazzo J, Batteiger BE, Hook EW, Seña AC, Long J, Wierzbicki MR, Kwak
1239 H, Johnson SM, Lawrence K, Mueller J. 2018. Single-dose zoliflodacin (ETX0914) for
1240 treatment of urogenital gonorrhea. *N Engl J Med* 379:1835-1845.
- 1241 31. Taylor SN, Morris DH, Avery AK, Workowski KA, Batteiger BE, Tiffany CA, Perry CR,
1242 Raychaudhuri A, Scangarella-Oman NE, Hossain M, Dumont EF. 2018. Gepotidacin
1243 for the treatment of uncomplicated urogenital gonorrhea: a phase 2, randomized,
1244 dose-ranging, single-oral dose evaluation. *Clin Infect Dis* 67:504-512.
- 1245 32. Russell MW, Jerse AE, Gray-Owen SD. 2019. Progress toward a gonococcal vaccine:
1246 the way forward. *Front Immunol* 10:2417.
- 1247 33. Fairley CK, Read TR. 2012. Vaccination against sexually transmitted infections. *Curr*
1248 *Opin Infect Dis* 25:66-72.
- 1249 34. Petousis-Harris H, Paynter J, Morgan J, Saxton P, McArdle B, Goodyear-Smith F, Black
1250 S. 2017. Effectiveness of a group B outer membrane vesicle meningococcal vaccine
1251 against gonorrhoea in New Zealand: a retrospective case-control study. *Lancet*
1252 390:1603-1610.
- 1253 35. Broutet N, Fruth U, Deal C, Gottlieb SL, Rees H, participants of the STIVTC. 2014.
1254 Vaccines against sexually transmitted infections: the way forward. *Vaccine* 32:1630-
1255 7.
- 1256 36. Gottlieb SL, Deal CD, Giersing B, Rees H, Bolan G, Johnston C, Timms P, Gray-Owen
1257 SD, Jerse AE, Cameron CE, Moorthy VS, Kiarie J, Broutet N. 2016. The global roadmap
1258 for advancing development of vaccines against sexually transmitted infections:
1259 Update and next steps. *Vaccine* 34:2939-2947.
- 1260 37. Gottlieb SL, Ndowa F, Hook EW, 3rd, Deal C, Bachmann L, Abu-Raddad L, Chen XS,
1261 Jerse A, Low N, MacLennan CA, Petousis-Harris H, Seib KL, Unemo M, Vincent L,

1262 Giersing BK. 2020. Gonococcal vaccines: Public health value and preferred product
 1263 characteristics; report of a WHO global stakeholder consultation, January 2019.
 1264 Vaccine 38:4362-4373.

1265 38. World Health Organization. 2021. WHO preferred product characteristics for
 1266 gonococcal vaccines. WHO, Geneva.

1267 39. World Health Organization. 2016. Global health sector strategy on sexually
 1268 transmitted infections 2016-2021: towards ending STIs.WHO, Geneva.

1269 40. Taylor-Robinson D, Furr PM, Hetherington CM. 1990. *Neisseria gonorrhoeae*
 1270 colonises the genital tract of oestradiol-treated germ-free female mice. *Microb*
 1271 *Pathog* 9:369-73.

1272 41. Jerse AE. 1999. Experimental gonococcal genital tract infection and opacity protein
 1273 expression in estradiol-treated mice. *Infect Immun* 67:5699-708.

1274 42. Islam EA, Anipindi VC, Francis I, Shaik-Dasthagirisahab Y, Xu S, Leung N, Sintsova A,
 1275 Amin M, Kaushic C, Wetzler LM, Gray-Owen SD. 2018. Specific binding to
 1276 differentially expressed human carcinoembryonic antigen-related cell adhesion
 1277 molecules determines the outcome of *Neisseria gonorrhoeae* infections along the
 1278 female reproductive tract. *Infect Immun* 86:e00092-18.

1279 43. Li G, Jiao H, Yan H, Wang J, Wang X, Ji M. 2011. Establishment of a human CEACAM1
 1280 transgenic mouse model for the study of gonococcal infections. *J Microbiol Methods*
 1281 87:350-4.

1282 44. Zarantonelli ML, Szatanik M, Giorgini D, Hong E, Huerre M, Guillou F, Alonso JM,
 1283 Taha MK. 2007. Transgenic mice expressing human transferrin as a model for
 1284 meningococcal infection. *Infect Immun* 75:5609-14.

1285 45. Perera Y, Cobas K, Garrido Y, Nazabal C, Brown E, Pajon R. 2006. Determination of
1286 human transferrin concentrations in mouse models of Neisserial infection. *J Immunol*
1287 *Methods* 311:153-63.

1288 46. Connolly KL, Pilligua-Lucas M, Gomez C, Costenoble-Caherty AC, Soc A, Underwood
1289 K, Macintyre AN, Sempowski GD, Jerse AE. 2021. preclinical testing of vaccines and
1290 therapeutics for gonorrhea in female mouse models of lower and upper
1291 reproductive tract infection. *J Infect Dis* 224:S152-s160.

1292 47. Arko RJ, Kraus SJ, Brown WJ, Buchanan TM, Kuhn US. 1974. *Neisseria gonorrhoeae*:
1293 effects of systemic immunization on resistance of chimpanzees to urethral infection.
1294 *J Infect Dis* 130:160-4.

1295 48. VanScoy BD, Scangarella-Oman NE, Fikes S, Min S, Huang J, Ingraham K, Bhavnani
1296 SM, Conde H, Ambrose PG. 2020. Relationship between gepotidacin exposure and
1297 prevention of on-therapy resistance amplification in a *Neisseria gonorrhoeae* hollow-
1298 fiber in vitro infection model. *Antimicrob Agents Chemother* 64:e00521-20.

1299 49. Jacobsson S, Golparian D, Oxelbark J, Alirol E, Franceschi F, Gustafsson TN, Brown D,
1300 Louie A, Drusano G, Unemo M. 2021. pharmacodynamic evaluation of dosing,
1301 bacterial kill, and resistance suppression for zoliflodacin against *Neisseria*
1302 *gonorrhoeae* in a dynamic hollow fiber infection model. *Front Pharmacol* 12:682135.

1303 50. Dijokaite A, Humbert MV, Borkowski E, La Ragione RM, Christodoulides M. 2021.
1304 Establishing an invertebrate *Galleria mellonella* greater wax moth larval model of
1305 *Neisseria gonorrhoeae* infection. *Virulence* 12:1900-1920.

1306 51. Hobbs MM, Sparling PF, Cohen MS, Shafer WM, Deal CD, Jerse AE. 2011.
1307 Experimental gonococcal infection in male volunteers: cumulative experience with
1308 *Neisseria gonorrhoeae* strains FA1090 and MS11mkC. *Front Microbiol* 2:123-123.

1309 52. Waltmann A, Duncan JA, Pier GB, Cywes-Bentley C, Cohen MS, Hobbs MM. 2022.
1310 Experimental urethral infection with *Neisseria gonorrhoeae*. *Curr Top Microbiol*
1311 *Immunol.* 10.1007/82_2021_250.

1312 53. Abara WE, Bernstein KT, Lewis FMT, Schillinger JA, Feemster K, Pathela P, Hariri S,
1313 Islam A, Eberhart M, Cheng I, Ternier A, Slutsker JS, Mbaeyi S, Madera R, Kirkcaldy
1314 RD. 2022. Effectiveness of a serogroup B outer membrane vesicle meningococcal
1315 vaccine against gonorrhoea: a retrospective observational study. *Lancet Infect Dis*
1316 22:1021-1029.

1317 54. Wang B, Giles L, Andraweera P, McMillan M, Almond S, Beazley R, Mitchell J, Lally N,
1318 Ahoure M, Denehy E, Koehler A, Flood L, Marshall H. 2022. Effectiveness and impact
1319 of the 4CMenB vaccine against invasive serogroup B meningococcal disease and
1320 gonorrhoea in an infant, child, and adolescent programme: an observational cohort
1321 and case-control study. *Lancet Infect Dis* 22:1011-1020.

1322 55. Bruxvoort KJ, Lewnard JA, Chen LH, Tseng HF, Chang J, Marrazzo J, Qian L. 2022.
1323 Prevention of *Neisseria gonorrhoeae* with meningococcal B vaccine: a matched
1324 cohort study in Southern California. *Clin Infect Dis.* 76:e1341-e1349.

1325 56. Raccagni AR, Galli L, Spagnuolo V, Bruzzesi E, Muccini C, Bossolasco S, Ranzenigo M,
1326 Gianotti N, Lolatto R, Castagna A, Nozza S. 2023. Meningococcus B vaccination
1327 effectiveness against *Neisseria gonorrhoeae* infection in people living with HIV: a
1328 case-control study. *Sex Transm Dis.* 50:247-251.

1329 57. Paynter J, Goodyear-Smith F, Morgan J, Saxton P, Black S, Petousis-Harris H. 2019.
1330 Effectiveness of a group b outer membrane vesicle meningococcal vaccine in
1331 preventing hospitalization from gonorrhea in New Zealand: a retrospective cohort
1332 study. *Vaccines (Basel)* 7:5.

- 1333 58. Azze RFO. 2019. A meningococcal B vaccine induces cross-protection against
1334 gonorrhea. Clin Exp Vaccine Res 8:110-115.
- 1335 59. Reyes Díaz LM, Lastre González M, Cuello M, Sierra-González VG, Ramos Pupo R,
1336 Lantero MI, Harandi AM, Black S, Pérez O. 2021. VA-MENGOC-BC vaccination induces
1337 serum and mucosal anti-Neisseria gonorrhoeae immune responses and reduces the
1338 incidence of gonorrhea. Pediatr Infect Dis J 40:375-381.
- 1339 60. Pérez O, del Campo J, Cuello M, González E, Nuñez N, Osmir C, Llanes R, Acevedo R,
1340 Zayas C, Balboa J, Romeu B, Baró M, Campa C, Lantero M, Sierra G, Galindo MA,
1341 Harandi A, Lastre M. 2009. Mucosal approaches in Neisseria vaccinology.
1342 VacciMonitor. 18:53-55
- 1343 61. Longtin J, Dion R, Simard M, Betala Belinga JF, Longtin Y, Lefebvre B, Labbé AC,
1344 Deceuninck G, De Wals P. 2017. Possible impact of wide-scale vaccination against
1345 serogroup B Neisseria meningitidis on gonorrhea incidence rates in one region of
1346 Quebec, Canada. Open Forum Infect Dis 4:S734-5.
- 1347 62. Whelan J, Klovstad H, Haugen IL, Holle MR, Storsaeter J. 2016. Ecologic study of
1348 meningococcal B vaccine and Neisseria gonorrhoeae infection, Norway. Emerg Infect
1349 Dis 22:1137-9.
- 1350 63. Eyre JH, Stewart B. 1909. The treatment of gonococcus infections by vaccines. Lancet
1351 174:76-81.
- 1352 64. Greenberg L, Diena BB, Ashton FA, Wallace R, Kenny CP, Znamirowski R, Ferrari H,
1353 Atkinson J. 1974. Gonococcal vaccine studies in Inuvik. Can J Public Health 65:29-33.
- 1354 65. Boslego JW, Tramont EC, Chung RC, McChesney DG, Ciak J, Sadoff JC, Piziak MV,
1355 Brown JD, Brinton CC, Jr., Wood SW, et al. 1991. Efficacy trial of a parenteral
1356 gonococcal pilus vaccine in men. Vaccine 9:154-62.

1357 66. Rice PA, Shafer WM, Ram S, Jerse AE. 2017. *Neisseria gonorrhoeae*: drug resistance,
1358 mouse models, and vaccine development. *Annu Rev Microbiol* 71:665-686.

1359 67. Greenberg I, Diena BB, Kenny CP, Znamirowski R. 1971. Preliminary studies on the
1360 development of a gonococcal vaccine. *Bull World Health Organ* 45:531-5.

1361 68. CC B, SW W, A B, AM L, JR B, SW L, SE P, EC T, J S, W Z. 1982. The development of a
1362 *Neisserial pilus* vaccine for gonorrhoea and meningococcal meningitis. *In* Weinstein L
1363 (ed), *Seminars in infectious diseases*, vol IV: bacterial vaccines. Thieme-Stratton, New
1364 York.

1365 69. McChesney D, Tramont EC, Boslego JW, Ciak J, Sadoff J, Brinton CC. 1982. Genital
1366 antibody response to a parenteral gonococcal pilus vaccine. *Infect Immun* 36:1006-
1367 12.

1368 70. Tramont EC, Boslego JW. 1985. Pilus vaccines. *Vaccine* 3:3-10.

1369 71. Rice PA, Vayo HE, Tam MR, Blake MS. 1986. Immunoglobulin G antibodies directed
1370 against protein III block killing of serum-resistant *Neisseria gonorrhoeae* by immune
1371 serum. *J Exp Med* 164:1735-48.

1372 72. Semchenko EA, Jen FE, Jennings MP, Seib KL. 2022. Assessment of serum bactericidal
1373 and opsonophagocytic activity of antibodies to gonococcal vaccine targets. *Methods*
1374 *Mol Biol* 2414:363-372.

1375 73. Serruto D, Bottomley MJ, Ram S, Giuliani MM, Rappuoli R. 2012. The new
1376 multicomponent vaccine against meningococcal serogroup B, 4CMenB:
1377 immunological, functional and structural characterization of the antigens. *Vaccine* 30
1378 Suppl 2:B87-97.

1379 74. Baarda BI, Martinez FG, Sikora AE. 2018. Proteomics, bioinformatics and structure-
1380 function antigen mining for gonorrhea vaccines. *Front Immunol* 9:2793.

1381 75. Zielke RA, Wierzbicki IH, Weber JV, Gafken PR, Sikora AE. 2014. Quantitative
1382 proteomics of the *Neisseria gonorrhoeae* cell envelope and membrane vesicles for
1383 the discovery of potential therapeutic targets. *Mol Cell Proteomics* 13:1299-317.

1384 76. Zielke RA, Wierzbicki IH, Baarda BI, Gafken PR, Soge OO, Holmes KK, Jerse AE, Unemo
1385 M, Sikora AE. 2016. Proteomics-driven antigen discovery for development of
1386 vaccines against gonorrhea. *Mol Cell Proteomics* 15:2338-55.

1387 77. El-Rami FE, Zielke RA, Wi T, Sikora AE, Unemo M. 2019. Quantitative proteomics of
1388 the 2016 WHO *Neisseria gonorrhoeae* reference strains surveys vaccine candidates
1389 and antimicrobial resistance determinants. *Mol Cell Proteomics* 18:127-150.

1390 78. Nabu S, Lawung R, Isarankura-Na-Ayudhya P, Roytrakul S, Dolprasit S, Sengyee S,
1391 Isarankura-Na-Ayudhya C, Prachayasittikul V. 2017. Comparative proteomics analysis
1392 of *Neisseria gonorrhoeae* strains in response to extended-spectrum cephalosporins.
1393 *EXCLI J* 16:1207-1229.

1394 79. Zhu T, McClure R, Harrison OB, Genco C, Massari P. 2019. Integrated bioinformatic
1395 analyses and immune characterization of new *Neisseria gonorrhoeae* vaccine
1396 antigens expressed during natural mucosal infection. *Vaccines (Basel)* 7:153.

1397 80. Baarda BI, Zielke RA, Holm AK, Sikora AE. 2021. Comprehensive bioinformatic
1398 assessments of the variability of *Neisseria gonorrhoeae* vaccine candidates. *mSphere*
1399 6:e00977-20.

1400 81. Cash DR, Noinaj N, Buchanan SK, Cornelissen CN. 2015. Beyond the crystal structure:
1401 insight into the function and vaccine potential of *tbpa* expressed by *Neisseria*
1402 *gonorrhoeae*. *Infect Immun* 83:4438-49.

1403 82. Wang S, Xue J, Lu P, Ni C, Cheng H, Han R, van der Veen S. 2018. Gonococcal MtrE
1404 and its surface-expressed loop 2 are immunogenic and elicit bactericidal antibodies. J
1405 Infect 77:191-204.

1406 83. Jen FE, Semchenko EA, Day CJ, Seib KL, Jennings MP. 2019. The *Neisseria*
1407 gonorrhoeae methionine sulfoxide reductase (MsrA/B) is a surface exposed,
1408 immunogenic, vaccine candidate. Front Immunol 10:137.

1409 84. McKnew DL, Lynn F, Zenilman JM, Bash MC. 2003. Porin variation among clinical
1410 isolates of *Neisseria gonorrhoeae* over a 10-year period, as determined by Por
1411 variable region typing. J Infect Dis 187:1213-22.

1412 85. Zhu W, Chen CJ, Thomas CE, Anderson JE, Jerse AE, Sparling PF. 2011. Vaccines for
1413 gonorrhea: can we rise to the challenge? Front Microbiol 2:124.

1414 86. Yuen R, Kuniholm J, Lisk C, Wetzler LM. 2019. Neisserial PorB immune enhancing
1415 activity and use as a vaccine adjuvant. Hum Vaccin Immunother 15:2778-2781.

1416 87. Bäckman M, Källström H, Jonsson AB. 1998. The phase-variable pilus-associated
1417 protein PilC is commonly expressed in clinical isolates of *Neisseria gonorrhoeae*, and
1418 shows sequence variability among strains. Microbiology (Reading) 144 (Pt 1):149-
1419 156.

1420 88. Morand PC, Tattevin P, Eugene E, Beretti JL, Nassif X. 2001. The adhesive property of
1421 the type IV pilus-associated component PilC1 of pathogenic *Neisseria* is supported by
1422 the conformational structure of the N-terminal part of the molecule. Mol Microbiol
1423 40:846-56.

1424 89. Drake SL, Koomey M. 1995. The product of the pilQ gene is essential for the
1425 biogenesis of type IV pili in *Neisseria gonorrhoeae*. Mol Microbiol 18:975-86.

- 1426 90. Helm RA, Barnhart MM, Seifert HS. 2007. pilQ Missense mutations have diverse
1427 effects on PilQ multimer formation, piliation, and pilus function in *Neisseria*
1428 *gonorrhoeae*. *J Bacteriol* 189:3198-3207.
- 1429 91. Haghi F, Peerayeh SN, Siadat SD, Zeighami H. 2012. Recombinant outer membrane
1430 secretin PilQ(406-770) as a vaccine candidate for serogroup B *Neisseria meningitidis*.
1431 *Vaccine* 30:1710-4.
- 1432 92. Edwards JL, Entz DD, Apicella MA. 2003. Gonococcal phospholipase d modulates the
1433 expression and function of complement receptor 3 in primary cervical epithelial cells.
1434 *Infect Immun* 71:6381-91.
- 1435 93. Edwards JL, Apicella MA. 2006. *Neisseria gonorrhoeae* PLD directly interacts with Akt
1436 kinase upon infection of primary, human, cervical epithelial cells. *Cell Microbiol*
1437 8:1253-1271.
- 1438 94. Marjuki H, Topaz N, Joseph SJ, Gernert KM, Kersh EN, Wang X. 2019. Genetic
1439 similarity of gonococcal homologs to meningococcal outer membrane proteins of
1440 serogroup B vaccine. *mBio* 10:e01668-19.
- 1441 95. Semchenko EA, Day CJ, Seib KL. 2020. The *Neisseria gonorrhoeae* vaccine candidate
1442 NHBA elicits antibodies that are bactericidal, opsonophagocytic and that reduce
1443 gonococcal adherence to epithelial cells. *Vaccines (Basel)* 8:219.
- 1444 96. Semchenko EA, Mubaiwa TD, Day CJ, Seib KL. 2020. Role of the gonococcal Neisserial
1445 heparin binding antigen in microcolony formation, and serum resistance and
1446 adherence to epithelial cells. *J Infect Dis* 221:1612-1622.
- 1447 97. Cornelissen CN, Kelley M, Hobbs MM, Anderson JE, Cannon JG, Cohen MS, Sparling
1448 PF. 1998. The transferrin receptor expressed by gonococcal strain FA1090 is required
1449 for the experimental infection of human male volunteers. *Mol Microbiol* 27:611-6.

1450 98. Price GA, Russell MW, Cornelissen CN. 2005. Intranasal administration of
1451 recombinant *Neisseria gonorrhoeae* transferrin binding proteins A and B conjugated
1452 to the cholera toxin B subunit induces systemic and vaginal antibodies in mice. *Infect*
1453 *Immun* 73:3945-53.

1454 99. Falsetta ML, Bair TB, Ku SC, Vanden Hoven RN, Steichen CT, McEwan AG, Jennings
1455 MP, Apicella MA. 2009. Transcriptional profiling identifies the metabolic phenotype
1456 of gonococcal biofilms. *Infect Immun* 77:3522-32.

1457 100. Mellies J, Jose J, Meyer TF. 1997. The *Neisseria gonorrhoeae* gene *aniA* encodes an
1458 inducible nitrite reductase. *Mol Gen Genet* 256:525-32.

1459 101. Shewell LK, Jen FE, Jennings MP. 2017. Refinement of immunizing antigens to
1460 produce functional blocking antibodies against the AniA nitrite reductase of
1461 *Neisseria gonorrhoeae*. *PLoS One* 12:e0182555.

1462 102. Shewell LK, Ku SC, Schulz BL, Jen FE, Mubaiwa TD, Ketterer MR, Apicella MA,
1463 Jennings MP. 2013. Recombinant truncated AniA of pathogenic *Neisseria* elicits a
1464 non-native immune response and functional blocking antibodies. *Biochem Biophys*
1465 *Res Commun* 431:215-20.

1466 103. Lewis LA, Gulati S, Burrowes E, Zheng B, Ram S, Rice PA. 2015. α -2,3-sialyltransferase
1467 expression level impacts the kinetics of lipooligosaccharide sialylation, complement
1468 resistance, and the ability of *Neisseria gonorrhoeae* to colonize the murine genital
1469 tract. *mBio* 6:e02565-14.

1470 104. Wu H, Jerse AE. 2006. Alpha-2,3-sialyltransferase enhances *Neisseria gonorrhoeae*
1471 survival during experimental murine genital tract infection. *Infect Immun* 74:4094-
1472 103.

1473 105. Jen FE, Ketterer MR, Semchenko EA, Day CJ, Seib KL, Apicella MA, Jennings MP. 2021.
1474 The Lst sialyltransferase of *Neisseria gonorrhoeae* can transfer keto-deoxyoctanoate
1475 as the terminal sugar of lipooligosaccharide: a glyco-achilles heel that provides a new
1476 strategy for vaccines to prevent gonorrhea. *mBio* 12:e03666-20.

1477 106. Lewis LA, Rice PA, Ram S. 2019. Role of gonococcal Neisserial surface protein A
1478 (NspA) in serum resistance and comparison of its factor h binding properties with
1479 those of its meningococcal counterpart. *Infect Immun* 87:2006858-18.

1480 107. Li G, Jiao H, Jiang G, Wang J, Zhu L, Xie R, Yan H, Chen H, Ji M. 2011. *Neisseria*
1481 *gonorrhoeae* NspA induces specific bactericidal and opsonic antibodies in mice. *Clin*
1482 *Vaccine Immunol* 18:1817-22.

1483 108. Gulati S, McQuillen DP, Mandrell RE, Jani DB, Rice PA. 1996. Immunogenicity of
1484 *Neisseria gonorrhoeae* lipooligosaccharide epitope 2C7, widely expressed in vivo
1485 with no immunochemical similarity to human glycosphingolipids. *J Infect Dis*
1486 174:1223-37.

1487 109. Gulati S, Shaughnessy J, Ram S, Rice PA. 2019. Targeting Lipooligosaccharide (LOS)
1488 for a Gonococcal Vaccine. *Front Immunol* 10:321.

1489 110. Gulati S, Zheng B, Reed GW, Su X, Cox AD, St Michael F, Stupak J, Lewis LA, Ram S,
1490 Rice PA. 2013. Immunization against a saccharide epitope accelerates clearance of
1491 experimental gonococcal infection. *PLoS Pathog* 9:e1003559.

1492 111. Gulati S, Pennington MW, Czerwinski A, Carter D, Zheng B, Nowak NA, DeOliveira RB,
1493 Shaughnessy J, Reed GW, Ram S, Rice PA. 2019. Preclinical efficacy of a
1494 lipooligosaccharide peptide mimic candidate gonococcal vaccine. *mBio* 10:e02552-
1495 19.

1496 112. Gulati S, Mu X, Zheng B, Reed GW, Ram S, Rice PA. 2015. Antibody to reduction
1497 modifiable protein increases the bacterial burden and the duration of gonococcal
1498 infection in a mouse model. *J Infect Dis* 212:311-5.

1499 113. Handing JW, Ragland SA, Bharathan UV, Criss AK. 2018. The MtrCDE efflux pump
1500 contributes to survival of *Neisseria gonorrhoeae* from human neutrophils and their
1501 antimicrobial components. *Front Microbiol* 9:2688.

1502 114. Jerse AE, Deal CD. 2013. Vaccine research for gonococcal infections: where are we?
1503 *Sex Transm Infect* 89 Suppl 4:iv63-8.

1504 115. Humbert MV, Awanye AM, Lian LY, Derrick JP, Christodoulides M. 2017. Structure of
1505 the *Neisseria* Adhesin Complex Protein (ACP) and its role as a novel lysozyme
1506 inhibitor. *PLoS Pathog* 13:e1006448.

1507 116. Almonacid-Mendoza HL, Humbert MV, Dijokaite A, Cleary DW, Soo Y, Hung MC, Orr
1508 CM, Machelett MM, Tews I, Christodoulides M. 2018. Structure of the recombinant
1509 *Neisseria gonorrhoeae* Adhesin Complex Protein (rNg-ACP) and generation of murine
1510 antibodies with bactericidal activity against gonococci. *mSphere* 3:e00331-18.

1511 117. Zielke RA, Le Van A, Baarda BI, Herrera MF, Acosta CJ, Jerse AE, Sikora AE. 2018. SliC
1512 is a surface-displayed lipoprotein that is required for the anti-lysozyme strategy
1513 during *Neisseria gonorrhoeae* infection. *PLoS Pathog* 14:e1007081.

1514 118. Semchenko EA, Day CJ, Seib KL. 2017. MetQ of *Neisseria gonorrhoeae* Is a surface-
1515 expressed antigen that elicits bactericidal and functional blocking antibodies. *Infect*
1516 *Immun* 85:ee00898-16.

1517 119. Sikora AE, Gomez C, Le Van A, Baarda BI, Darnell S, Martinez FG, Zielke RA,
1518 Bonventre JA, Jerse AE. 2020. A novel gonorrhea vaccine composed of MetQ

lipoprotein formulated with CpG shortens experimental murine infection. *Vaccine* 38:8175-8184.

120. Pulendran B, S. Arunachalam P, O'Hagan DT. 2021. Emerging concepts in the science of vaccine adjuvants. *Nat Rev Drug Discov* 20:454-475.

121. Liu Y, Hammer LA, Liu W, Hobbs MM, Zielke RA, Sikora AE, Jerse AE, Egilmez NK, Russell MW. 2017. Experimental vaccine induces Th1-driven immune responses and resistance to *Neisseria gonorrhoeae* infection in a murine model. *Mucosal Immunol* 10:1594-1608.

122. Bode C, Zhao G, Steinhagen F, Kinjo T, Klinman DM. 2011. CpG DNA as a vaccine adjuvant. *Expert Rev Vaccines* 10:499-511.

123. Feinen B, Jerse AE, Gaffen SL, Russell MW. 2010. Critical role of Th17 responses in a murine model of *Neisseria gonorrhoeae* genital infection. *Mucosal Immunol* 3:312-321.

124. Liu Y, Feinen B, Russell MW. 2011. New concepts in immunity to *Neisseria gonorrhoeae*: innate responses and suppression of adaptive immunity favor the pathogen, not the host. *Front Microbiol* 2:52.

125. Liu Y, Islam EA, Jarvis GA, Gray-Owen SD, Russell MW. 2012. *Neisseria gonorrhoeae* selectively suppresses the development of Th1 and Th2 cells, and enhances Th17 cell responses, through TGF- β -dependent mechanisms. *Mucosal Immunol* 5:320-31.

126. Gagliardi MC, Starnino S, Teloni R, Mariotti S, Dal Conte I, Di Carlo A, Stefanelli P. 2011. Circulating levels of interleukin-17A and interleukin-23 are increased in patients with gonococcal infection. *FEMS Immunol Med Microbiol* 61:129-32.

127. Masson L, Salkinder AL, Olivier AJ, McKinnon LR, Gamielien H, Mlisana K, Scriba TJ, Lewis DA, Little F, Jaspan HB, Ronacher K, Denny L, Abdool Karim SS, Passmore JA.

1543 2015. Relationship between female genital tract infections, mucosal interleukin-17
1544 production and local T helper type 17 cells. *Immunology* 146:557-67.

1545 128. Pollard AJ, Bijker EM. 2021. A guide to vaccinology: from basic principles to new
1546 developments. *Nat Rev Immunol* 21:83-100.

1547 129. Zhu W, Thomas CE, Chen CJ, Van Dam CN, Johnston RE, Davis NL, Sparling PF. 2005.
1548 Comparison of immune responses to gonococcal PorB delivered as outer membrane
1549 vesicles, recombinant protein, or Venezuelan equine encephalitis virus replicon
1550 particles. *Infect Immun* 73:7558-68.

1551 130. Zhu W, Thomas CE, Sparling PF. 2004. DNA immunization of mice with a plasmid
1552 encoding *Neisseria gonorrhoea* PorB protein by intramuscular injection and epidermal
1553 particle bombardment. *Vaccine* 22:660-9.

1554 131. Hoffelner H, Haas R. 2004. Recombinant bacterial ghosts: versatile targeting vehicles
1555 and promising vaccine candidates. *Int J Med Microbiol* 294:303-311.

1556 132. Jiao H, Yang H, Zhao D, Chen J, Zhang Q, Liang J, Yin Y, Kong G, Li G. 2018. Design and
1557 immune characterization of a novel *Neisseria gonorrhoeae* DNA vaccine using
1558 bacterial ghosts as vector and adjuvant. *Vaccine* 36:4532-4539.

1559 133. Jiao H, Yang H, Zheng W, Zhang Q, Zhao D, Li G. 2021. Enhancement of immune
1560 responses by co-administration of bacterial ghosts-mediated *Neisseria gonorrhoeae*
1561 DNA vaccines. *J Appl Microbiol* 130:1770-1777.

1562 134. van der Pol L, Stork M, van der Ley P. 2015. Outer membrane vesicles as platform
1563 vaccine technology. *Biotechnol J* 10:1689-706.

1564 135. Chbib C, Shah SM, Gala RP, Uddin MN. 2021. Potential applications of
1565 microparticulate-based bacterial outer membrane vesicles (OMVs) vaccine platform

1566 for sexually transmitted diseases (STDs): gonorrhea, chlamydia, and syphilis.

1567 Vaccines (Basel) 9.

1568 136. Kaparakis-Liaskos M, Ferrero RL. 2015. Immune modulation by bacterial outer
1569 membrane vesicles. *Nat Rev Immunol* 15:375-387.

1570 137. Matthias KA, Connolly KL, Begum AA, Jerse AE, Macintyre AN, Sempowski GD, Bash
1571 MC. 2022. meningococcal detoxified outer membrane vesicle vaccines enhance
1572 gonococcal clearance in a murine infection model. *J Infect Dis* 225:650-660.

1573 138. Beernink PT, Ispasanie E, Lewis LA, Ram S, Moe GR, Granoff DM. 2019. A
1574 Meningococcal native outer membrane vesicle vaccine with attenuated endotoxin
1575 and overexpressed factor H binding protein elicits gonococcal bactericidal
1576 antibodies. *J Infect Dis* 219:1130-1137.

1577 139. Plante M, Jerse A, Hamel J, Couture F, Rioux CR, Brodeur BR, Martin D. 2000.
1578 Intranasal immunization with gonococcal outer membrane preparations reduces the
1579 duration of vaginal colonization of mice by *Neisseria gonorrhoeae*. *J Infect Dis*
1580 182:848-55.

1581 140. MacLennan CA. 2022. GonoVac, a candidate parenteral NOMV gonococcal vaccine
1582 that clears gonococci faster than Bexsero in the mouse vaginal infection model.
1583 Abstr 22nd International Pathogenic *Neisseria* Conference, abstr 214.

1584 141. Liu Y, Hammer LA, Daamen J, Stork M, Egilmez NK, Russell MW. 2023.
1585 Microencapsulated IL-12 drives genital tract immune responses to intranasal
1586 gonococcal outer membrane vesicle vaccine and induces resistance to vaginal
1587 infection with diverse strains of *Neisseria gonorrhoeae*. *mSphere* 8:e0038822.

1588 142. Rosenthal KL, Gallichan WS. 1997. Challenges for vaccination against sexually-
1589 transmitted diseases: induction and long-term maintenance of mucosal immune
1590 responses in the female genital tract. *Semin Immunol* 9:303-314.

1591 143. de Jonge MI, Hamstra HJ, Jiskoot W, Roholl P, Williams NA, Dankert J, van Alphen L,
1592 van der Ley P. 2004. Intranasal immunisation of mice with liposomes containing
1593 recombinant meningococcal OpaB and OpaJ proteins. *Vaccine* 22:4021-8.

1594 144. Gala RP, Zaman RU, D'Souza MJ, Zughaier SM. 2018. Novel whole-cell inactivated
1595 *Neisseria gonorrhoeae* microparticles as vaccine formulation in microneedle-based
1596 transdermal immunization. *Vaccines (Basel)* 6:60.

1597 145. Edwards JL, Jennings MP, Seib KL. 2018. *Neisseria gonorrhoeae* vaccine
1598 development: hope on the horizon? *Curr Opin Infect Dis* 31:246-250.

1599 146. Ruiz García Y, Sohn WY, Seib KL, Taha MK, Vázquez JA, de Lemos APS, Vadivelu K,
1600 Pizza M, Rappuoli R, Bekkat-Berkani R. 2021. Looking beyond meningococcal B with
1601 the 4CMenB vaccine: the *Neisseria* effect. *NPJ Vaccines* 6:130.

1602 147. Thng C, Semchenko EA, Hughes I, O'Sullivan M, Seib KL. 2023. An open-label
1603 randomised controlled trial evaluating the efficacy of a meningococcal serogroup B
1604 (4CMenB) vaccine on *Neisseria gonorrhoeae* infection in gay and bisexual men: the
1605 MenGO study protocol. *BMC Public Health* 23:607.

1606 148. Molina J-MB, B; Assoumou L; Michele, I-G; Rubenstein, E; Pialoux, G; Katlama, C;
1607 Surgers, L; Bebear, C; Dupin, N; Viard, J-P; Pavie, J; Duvivier, C; Ghosn, J; Costagliola,
1608 D. ANRS 174 Doxyvac: an open-label randomised trial to prevent STIs in MSM on
1609 PrEP, Abstr 29th Conference on Retroviruses and Opportunistic Infections, abstr 119 .

1610 149. European AIDS Treatment Group. 2023. ANRS DOXYVAC: final analysis may modify
1611 interim results of this trial assessing the effectiveness of meningococcal B

1612 vaccination in preventing gonococcal infections. European AIDS Treatment Group,
 1613 Brussels. [https://www.eatg.org/hiv-news/anrs-doxyvac-final-analysis-may-modify-](https://www.eatg.org/hiv-news/anrs-doxyvac-final-analysis-may-modify-interim-results-of-this-trial-assessing-the-effectiveness-of-meningococcal-b-vaccination-in-preventing-gonococcal-infections/)
 1614 [interim-results-of-this-trial-assessing-the-effectiveness-of-meningococcal-b-](https://www.eatg.org/hiv-news/anrs-doxyvac-final-analysis-may-modify-interim-results-of-this-trial-assessing-the-effectiveness-of-meningococcal-b-vaccination-in-preventing-gonococcal-infections/)
 1615 [vaccination-in-preventing-gonococcal-infections/](https://www.eatg.org/hiv-news/anrs-doxyvac-final-analysis-may-modify-interim-results-of-this-trial-assessing-the-effectiveness-of-meningococcal-b-vaccination-in-preventing-gonococcal-infections/)

1616 150. Sotolongo F, Campa C, Casanueva V, Fajardo EM, Cuevas IE, González N. 2007. Cuban
 1617 Meningococcal BC Vaccine: Experiences & Contributions from 20 Years of
 1618 Application. MEDICC Rev 9:16-22.

1619 151. Abara WE, Bernstein KT, Lewis FMT, Pathela P, Islam A, Eberhart M, Cheng I, Ternier
 1620 A, Sanderson Slutsker J, Madera R, Kirkcaldy R. 2023. Healthy vaccinee bias and
 1621 MenB-FHbp vaccine effectiveness against gonorrhea. Sex Transm Dis 50:e8-10.

1622 152. Hui BB, Padeniya TN, Rebuli N, Gray RT, Wood JG, Donovan B, Duan Q, Guy R,
 1623 Hocking JS, Lahra MM, Lewis DA, Whiley DM, Regan DG, Seib KL. 2022. A gonococcal
 1624 vaccine has the potential to rapidly reduce the incidence of *Neisseria gonorrhoeae*
 1625 infection among urban men who have sex with men. J Infect Dis 225:983-993.

1626 153. Semchenko EA, Tan A, Borrow R, Seib KL. 2019. The serogroup B meningococcal
 1627 vaccine bexsero elicits antibodies to *Neisseria gonorrhoeae*. Clin Infect Dis 69:1101-
 1628 1111.

1629 154. Jongerius I, Lavender H, Tan L, Ruivo N, Exley RM, Caesar JJ, Lea SM, Johnson S, Tang
 1630 CM. 2013. Distinct binding and immunogenic properties of the gonococcal
 1631 homologue of meningococcal factor h binding protein. PLoS Pathog 9:e1003528.

1632 155. Bos MP, Grijpstra J, Tommassen-van Boxtel R, Tommassen J. 2014. Involvement of
 1633 *Neisseria meningitidis* lipoprotein GNA2091 in the assembly of a subset of outer
 1634 membrane proteins. J Biol Chem 289:15602-10.

1635 156. Donnarumma D, Golfieri G, Brier S, Castagnini M, Veggi D, Bottomley MJ, Delany I,
1636 Norais N. 2015. Neisseria meningitis GNA1030 is a ubiquinone-8 binding protein.
1637 FASEB J 29:2260-7.

1638 157. Leduc I, Connolly KL, Begum A, Underwood K, Darnell S, Shafer WM, Balthazar JT,
1639 Macintyre AN, Sempowski GD, Duncan JA, Little MB, Rahman N, Garges EC, Jerse AE.
1640 2020. The serogroup B meningococcal outer membrane vesicle-based vaccine
1641 4CMenB induces cross-species protection against Neisseria gonorrhoeae. PLOS
1642 Pathogens 16:e1008602.

1643 158. Rappuoli R, Pizza M, Masignani V, Vadivelu K. 2018. Meningococcal B vaccine
1644 (4CMenB): the journey from research to real world experience. Expert Rev Vaccines
1645 17:1111-1121.

1646 159. MacLennan CA. 2023. Advancing a native outer membrane vesicle vaccine against
1647 gonorrhoea towards clinical development. University of Birmingham, Birmingham.
1648 [https://www.birmingham.ac.uk/research/immunology-](https://www.birmingham.ac.uk/research/immunology-immunotherapy/research/bactivac/funded-pump-priming-projects-awardees/prof-calman-maclennan.aspx)
1649 [immunotherapy/research/bactivac/funded-pump-priming-projects-awardees/prof-](https://www.birmingham.ac.uk/research/immunology-immunotherapy/research/bactivac/funded-pump-priming-projects-awardees/prof-calman-maclennan.aspx)
1650 [calman-maclennan.aspx](https://www.birmingham.ac.uk/research/immunology-immunotherapy/research/bactivac/funded-pump-priming-projects-awardees/prof-calman-maclennan.aspx)

1651 160. World Health Organization. 2022. Bacterial vaccines in clinical and preclinical
1652 development: an overview and analysis. WHO, Geneva.

1653 161. Pharmaceutical Technology. 2022. Intravacc gets NIAID contract for intranasal
1654 gonorrhoea vaccine development. [https://www.pharmaceutical-](https://www.pharmaceutical-technology.com/news/intravacc-contract-gonorrhoea-vaccine/)
1655 [technology.com/news/intravacc-contract-gonorrhoea-vaccine/](https://www.pharmaceutical-technology.com/news/intravacc-contract-gonorrhoea-vaccine/).

1656 162. Mancini F, Micoli F, Necchi F, Pizza M, Berlanda Scorza F, Rossi O. 2021. GMMA-
1657 Based Vaccines: The Known and The Unknown. Front Immunol 12:715393.

1658 163. GlaxoSmithKline. 2023. GSK receives US FDA Fast Track designation for
1659 investigational vaccine against gonorrhoea. GSK, Middlesex.
1660 [https://www.gsk.com/en-gb/media/press-releases/gsk-receives-us-fda-fast-track-](https://www.gsk.com/en-gb/media/press-releases/gsk-receives-us-fda-fast-track-designation-for-investigational-vaccine-against-gonorrhoea/)
1661 [designa-
tion-for-investigational-vaccine-against-gonorrhoea/](https://www.gsk.com/en-gb/media/press-releases/gsk-receives-us-fda-fast-track-designation-for-investigational-vaccine-against-gonorrhoea/)

1662 164. Craig AP, Gray RT, Edwards JL, Apicella MA, Jennings MP, Wilson DP, Seib KL. 2015.
1663 The potential impact of vaccination on the prevalence of gonorrhea. *Vaccine*
1664 33:4520-4525.

1665 165. Carey KA, Newman LM, Spicknall IH. 2022. Estimating the population level impact of
1666 a gonococcal vaccine candidate: predictions from a simple mathematical model.
1667 *Vaccine* 40:7176-7181.

1668 166. Looker KJ, Booton R, Begum N, Beck E, Shen J, Turner KME, Christensen H. 2023. The
1669 potential public health impact of adolescent 4CMenB vaccination on *Neisseria*
1670 *gonorrhoeae* infection in England: a modelling study. *BMC Public Health* 23:1.

1671 167. Regnier SA, Huels J. 2014. Potential impact of vaccination against *Neisseria*
1672 *meningitidis* on *Neisseria gonorrhoeae* in the United States: results from a decision-
1673 analysis model. *Hum Vaccin Immunother* 10:3737-45.

1674 168. Whittles LK, White PJ, Didelot X. 2020. Assessment of the potential of vaccination to
1675 combat antibiotic resistance in gonorrhea: a modeling analysis to determine
1676 preferred product characteristics. *Clin Infect Dis* 71:1912-1919.

1677 169. Heijne JCM, Xiridou M, Turner KME, Basten M, Visser M, Benthem Bv, Low N. 2020.
1678 The impact of vaccination on *Neisseria gonorrhoeae* antimicrobial resistance and
1679 prevalence in men who have sex with men: a mathematical modelling study.
1680 medRxiv doi:10.1101/2020.09.14.20192062:2020.09.14.20192062.

1681 170. Whittles LK, Didelot X, White PJ. 2022. Public health impact and cost-effectiveness of
1682 gonorrhoea vaccination: an integrated transmission-dynamic health-economic
1683 modelling analysis. *Lancet Infect Dis* 22:1030-1041.

1684 171. Padeniya TN, Hui BB, Wood JG, Seib KL, Regan DG. 2023. The potential impact of a
1685 vaccine on *Neisseria gonorrhoeae* prevalence among heterosexuals living in a high
1686 prevalence setting. *Vaccine* 41:5553-5561.

1687 172. Senff LM, Wegener WS, Brooks GF, Finnerty WR, Makula RA. 1976. Phospholipid
1688 composition and phospholipase A activity of *Neisseria gonorrhoeae*. *J Bacteriol*
1689 127:874-80.

1690 173. Cacciapuoti AF, Wegener WS, Morse SA. 1978. Cell envelope of *Neisseria*
1691 *gonorrhoeae*: phospholipase activity and its relationship to autolysis. *Infect Immun*
1692 20:418-20.

1693 174. Bos MP, Tefsen B, Voet P, Weynants V, van Putten JP, Tommassen J. 2005. Function
1694 of *Neisseria* outer membrane phospholipase A in autolysis and assessment of its
1695 vaccine potential. *Infect Immun* 73:2222-31.

1696 175. Edwards JL, Jennings MP, Apicella MA, Seib KL. 2016. Is gonococcal disease
1697 preventable? The importance of understanding immunity and pathogenesis in
1698 vaccine development. *Crit Rev Microbiol* 42:928-41.

1699 176. Tramont EC, Sadoff JC, Boslego JW, Ciak J, McChesney D, Brinton CC, Wood S,
1700 Takafuji E. 1981. Gonococcal pilus vaccine. Studies of antigenicity and inhibition of
1701 attachment. *J Clin Invest* 68:881-8.

1702 177. Siegel M, Olsen D, Critchlow C, Buchanan TM. 1982. Gonococcal pili: safety and
1703 immunogenicity in humans and antibody function in vitro. *J Infect Dis* 145:300-10.

- 1704 178. Brinton CW, Wood SW, Brown A, Labik AM, Bryan JR, Lee SW, Polen SE, Tramont EC,
1705 Sadoff J, Zollinger W. 1982. The development of a *Neisseria pilus* vaccine for
1706 gonorrhoea and meningococcal meningitis, p 140-159. *In*: Robbins JB, Hill JC, Sandoff
1707 JC (eds), Seminars in Infectious Diseases, vol IV: bacterial vaccines. Thieme-Stratton,
1708 New York.
- 1709 179. Schoolnik GK, Tai JY, Gotschlich EC. 1983. A pilus peptide vaccine for the prevention
1710 of gonorrhea. *Prog Allergy* 33:314-31.
- 1711 180. Virji M, Heckels JE. 1984. The role of common and type-specific pilus antigenic
1712 domains in adhesion and virulence of gonococci for human epithelial cells. *J Gen*
1713 *Microbiol* 130:1089-95.
- 1714 181. Rothbard JB, Fernandez R, Wang L, Teng NN, Schoolnik GK. 1985. Antibodies to
1715 peptides corresponding to a conserved sequence of gonococcal pilins block bacterial
1716 adhesion. *Proc Natl Acad Sci U S A* 82:915-9.
- 1717 182. Hook EW, 3rd, Olsen DA, Buchanan TM. 1984. Analysis of the antigen specificity of
1718 the human serum immunoglobulin G immune response to complicated gonococcal
1719 infection. *Infect Immun* 43:706-9.
- 1720 183. Heckels JE, Virji M, Tinsley CR. 1990. Vaccination against gonorrhoea: the potential
1721 protective effect of immunization with a synthetic peptide containing a conserved
1722 epitope of gonococcal outer membrane protein IB. *Vaccine* 8:225-30.
- 1723 184. Christodoulides M, McGuinness BT, Heckels JE. 1993. Immunization with synthetic
1724 peptides containing epitopes of the class 1 outer-membrane protein of *Neisseria*
1725 *meningitidis*: production of bactericidal antibodies on immunization with a cyclic
1726 peptide. *J Gen Microbiol* 139:1729-38.

1727 185. Ram S, McQuillen DP, Gulati S, Elkins C, Pangburn MK, Rice PA. 1998. Binding of
1728 complement factor H to loop 5 of porin protein 1A: a molecular mechanism of serum
1729 resistance of nonsialylated *Neisseria gonorrhoeae*. *J Exp Med* 188:671-80.

1730 186. Ram S, Sharma AK, Simpson SD, Gulati S, McQuillen DP, Pangburn MK, Rice PA. 1998.
1731 A novel sialic acid binding site on factor H mediates serum resistance of sialylated
1732 *Neisseria gonorrhoeae*. *J Exp Med* 187:743-52.

1733 187. Ram S, Cullinane M, Blom AM, Gulati S, McQuillen DP, Monks BG, O'Connell C, Boden
1734 R, Elkins C, Pangburn MK, Dahlbäck B, Rice PA. 2001. Binding of C4b-binding protein
1735 to porin: a molecular mechanism of serum resistance of *Neisseria gonorrhoeae*. *J Exp*
1736 *Med* 193:281-296.

1737 188. Edwards JL, Brown EJ, Uk-Nham S, Cannon JG, Blake MS, Apicella MA. 2002. A co-
1738 operative interaction between *Neisseria gonorrhoeae* and complement receptor 3
1739 mediates infection of primary cervical epithelial cells. *Cell Microbiol* 4:571-84.

1740 189. Kühlewein C, Rechner C, Meyer TF, Rudel T. 2006. Low-phosphate-dependent
1741 invasion resembles a general way for *Neisseria gonorrhoeae* to enter host cells.
1742 *Infect Immun* 74:4266-4273.

1743 190. Garvin LE, Bash MC, Keys C, Warner DM, Ram S, Shafer WM, Jerse AE. 2008.
1744 Phenotypic and genotypic analyses of *Neisseria gonorrhoeae* isolates that express
1745 frequently recovered PorB PIA variable region types suggest that certain P1a porin
1746 sequences confer a selective advantage for urogenital tract infection. *Infect Immun*
1747 76:3700-9.

1748 191. Faulstich M, Böttcher JP, Meyer TF, Fraunholz M, Rudel T. 2013. Pilus phase variation
1749 switches gonococcal adherence to invasion by caveolin-1-dependent host cell
1750 signaling. *PLoS Pathog* 9:e1003373.

1751 192. Virji M, Makepeace K, Ferguson DJ, Achtman M, Moxon ER. 1993. Meningococcal
1752 Opa and Opc proteins: their role in colonization and invasion of human epithelial and
1753 endothelial cells. *Mol Microbiol* 10:499-510.

1754 193. Plummer FA, Chubb H, Simonsen JN, Bosire M, Slaney L, Nagelkerke NJ, Maclean I,
1755 Ndinya-Achola JO, Waiyaki P, Brunham RC. 1994. Antibodies to opacity proteins
1756 (Opa) correlate with a reduced risk of gonococcal salpingitis. *J Clin Invest* 93:1748-55.

1757 194. Chen T, Grunert F, Medina-Marino A, Gotschlich EC. 1997. Several carcinoembryonic
1758 antigens (CD66) serve as receptors for gonococcal opacity proteins. *J Exp Med*
1759 185:1557-1564.

1760 195. Cole JG, Jerse AE. 2009. Functional characterization of antibodies against *Neisseria*
1761 *gonorrhoeae* opacity protein loops. *PLoS One* 4:e8108.

1762 196. Callaghan MJ, Lewis S, Sadarangani M, Bailey SE, Chan H, Ferguson DJ, Derrick JP,
1763 Feavers I, Maiden MC, Pollard AJ. 2011. Potential of recombinant opa proteins as
1764 vaccine candidates against hyperinvasive meningococci. *Infect Immun* 79:2810-8.

1765 197. Sadarangani M, Pollard AJ, Gray-Owen SD. 2011. Opa proteins and CEACAMs:
1766 pathways of immune engagement for pathogenic *Neisseria*. *FEMS Microbiol Rev*
1767 35:498-514.

1768 198. Zhu P, Klutch MJ, Derrick JP, Prince SM, Tsang RS, Tsai CM. 2003. Identification of
1769 *opcA* gene in *Neisseria polysaccharea*: interspecies diversity of Opc protein family.
1770 *Gene* 307:31-40.

1771 199. Moore J, Bailey SE, Benmechernene Z, Tzitzilonis C, Griffiths NJ, Virji M, Derrick JP.
1772 2005. Recognition of saccharides by the OpcA, OpaD, and OpaB outer membrane
1773 proteins from *Neisseria meningitidis*. *J Biol Chem* 280:31489-97.

1774 200. Keiser PB, Gibbs BT, Coster TS, Moran EE, Stoddard MB, Labrie JE, 3rd, Schmiel DH,
1775 Pinto V, Chen P, Zollinger WD. 2010. A phase 1 study of a group B meningococcal
1776 native outer membrane vesicle vaccine made from a strain with deleted lpxL2 and
1777 synX and stable expression of opcA. *Vaccine* 28:6970-6.

1778 201. Serino L, Nesta B, Leuzzi R, Fontana MR, Monaci E, Mocca BT, Cartocci E, Massignani
1779 V, Jerse AE, Rappuoli R, Pizza M. 2007. Identification of a new OmpA-like protein in
1780 *Neisseria gonorrhoeae* involved in the binding to human epithelial cells and in vivo
1781 colonization. *Mol Microbiol* 64:1391-403.

1782 202. Starnino S, Leuzzi R, Ghisetti V, De Francesco MA, Cusini M, Impara G, Galluppi E,
1783 Pizza M, Stefanelli P. 2010. Molecular analysis of two novel *Neisseria gonorrhoeae*
1784 virulent components: the macrophage infectivity potentiator and the outer
1785 membrane protein A. *New Microbiol* 33:167-70.

1786 203. Semchenko EA, Day CJ, Seib KL. 2020. The *Neisseria gonorrhoeae* vaccine candidate
1787 NHBA elicits antibodies that are bactericidal, opsonophagocytic and that reduce
1788 gonococcal adherence to epithelial cells. *Vaccines* 8:219.

1789 204. Masri HP, Cornelissen CN. 2002. Specific ligand binding attributable to individual
1790 epitopes of gonococcal transferrin binding protein A. *Infect Immun* 70:732-40.

1791 205. Price GA, Hobbs MM, Cornelissen CN. 2004. Immunogenicity of gonococcal
1792 transferrin binding proteins during natural infections. *Infect Immun* 72:277-83.

1793 206. Price GA, Masri HP, Hollander AM, Russell MW, Cornelissen CN. 2007. Gonococcal
1794 transferrin binding protein chimeras induce bactericidal and growth inhibitory
1795 antibodies in mice. *Vaccine* 25:7247-60.

1796 207. DeRocco AJ, Cornelissen CN. 2007. Identification of transferrin-binding domains in
1797 TbpB expressed by *Neisseria gonorrhoeae*. *Infect Immun* 75:3220-32.

1798 208. Ostberg KL, DeRocco AJ, Mistry SD, Dickinson MK, Cornelissen CN. 2013. Conserved
1799 regions of gonococcal TbpB are critical for surface exposure and transferrin iron
1800 utilization. *Infect Immun* 81:3442-50.

1801 209. Mickelsen PA, Blackman E, Sparling PF. 1982. Ability of *Neisseria gonorrhoeae*,
1802 *Neisseria meningitidis*, and commensal *Neisseria* species to obtain iron from
1803 lactoferrin. *Infect Immun* 35:915-20.

1804 210. Biswas GD, Anderson JE, Chen CJ, Cornelissen CN, Sparling PF. 1999. Identification
1805 and functional characterization of the *Neisseria gonorrhoeae* lbpB gene product.
1806 *Infect Immun* 67:455-9.

1807 211. Anderson JE, Hobbs MM, Biswas GD, Sparling PF. 2003. Opposing selective forces for
1808 expression of the gonococcal lactoferrin receptor. *Mol Microbiol* 48:1325-37.

1809 212. Pettersson A, Kortekaas J, Weynants VE, Voet P, Poolman JT, Bos MP, Tommassen J.
1810 2006. Vaccine potential of the *Neisseria meningitidis* lactoferrin-binding proteins
1811 LbpA and LbpB. *Vaccine* 24:3545-57.

1812 213. Adamiak P, Beddek AJ, Pajon R, Schryvers AB. 2012. Patterns of sequence variation
1813 within the *Neisseria meningitidis* lactoferrin binding proteins. *Biochem Cell Biol*
1814 90:339-50.

1815 214. Noinaj N, Cornelissen CN, Buchanan SK. 2013. Structural insight into the lactoferrin
1816 receptors from pathogenic *Neisseria*. *J Struct Biol* 184:83-92.

1817 215. Black JR, Dyer DW, Thompson MK, Sparling PF. 1986. Human immune response to
1818 iron-repressible outer membrane proteins of *Neisseria meningitidis*. *Infect Immun*
1819 54:710-3.

1820 216. Dyer DW, West EP, McKenna W, Thompson SA, Sparling PF. 1988. A pleiotropic iron-
1821 uptake mutant of *Neisseria meningitidis* lacks a 70-kilodalton iron-regulated protein.
1822 *Infect Immun* 56:977-83.

1823 217. Pettersson A, Kuipers B, Pelzer M, Verhagen E, Tiesjema RH, Tommassen J, Poolman
1824 JT. 1990. Monoclonal antibodies against the 70-kilodalton iron-regulated protein of
1825 *Neisseria meningitidis* are bactericidal and strain specific. *Infect Immun* 58:3036-41.

1826 218. van der Ley P, van der Biezen J, Suttmuller R, Hoogerhout P, Poolman JT. 1996.
1827 Sequence variability of FrpB, a major iron-regulated outer-membrane protein in the
1828 pathogenic neisseriae. *Microbiology (Reading)* 142 (Pt 11):3269-74.

1829 219. Carson SD, Klebba PE, Newton SM, Sparling PF. 1999. Ferric enterobactin binding and
1830 utilization by *Neisseria gonorrhoeae*. *J Bacteriol* 181:2895-901.

1831 220. Carson SD, Stone B, Beucher M, Fu J, Sparling PF. 2000. Phase variation of the
1832 gonococcal siderophore receptor FetA. *Mol Microbiol* 36:585-93.

1833 221. Stork M, Bos MP, Jongerius I, de Kok N, Schilders I, Weynants VE, Poolman JT,
1834 Tommassen J. 2010. An outer membrane receptor of *Neisseria meningitidis* involved
1835 in zinc acquisition with vaccine potential. *PLoS Pathog* 6:e1000969.

1836 222. Cornelissen CN, Hollander A. 2011. TonB-dependent transporters expressed by
1837 *Neisseria gonorrhoeae*. *Front Microbiol* 2:117.

1838 223. Maurakis S, Keller K, Maxwell CN, Pereira K, Chazin WJ, Criss AK, Cornelissen CN.
1839 2019. The novel interaction between *Neisseria gonorrhoeae* TdfJ and human S100A7
1840 allows gonococci to subvert host zinc restriction. *PLoS Pathog* 15:e1007937.

1841 224. Turner PC, Thomas CE, Stojiljkovic I, Elkins C, Kizel G, Ala'Aldeen DAA, Sparling PF.
1842 2001. Neisserial TonB-dependent outer-membrane proteins: detection, regulation
1843 and distribution of three putative candidates identified from the genome

1844 sequencesThe GenBank accession number for the sequence of tdfH from
 1845 meningococcal strain IR1074 reported in this paper is AF227418. Microbiology
 1846 147:1277-1290.

1847 225. Jean S, Juneau RA, Criss AK, Cornelissen CN. 2016. *Neisseria gonorrhoeae* evades
 1848 calprotectin-mediated nutritional immunity and survives neutrophil extracellular
 1849 traps by production of TdfH. *Infect Immun* 84:2982-94.

1850 226. Kammerman MT, Bera A, Wu R, Harrison SA, Maxwell CN, Lundquist K, Noinaj N,
 1851 Chazin WJ, Cornelissen CN. 2020. Molecular insight into TdfH-mediated zinc piracy
 1852 from human calprotectin by *Neisseria gonorrhoeae*. *mBio* 11:e00949-20.

1853 227. Clark VL, Knapp JS, Thompson S, Klimpel KW. 1988. Presence of antibodies to the
 1854 major anaerobically induced gonococcal outer membrane protein in sera from
 1855 patients with gonococcal infections. *Microb Pathog* 5:381-90.

1856 228. Boulanger MJ, Murphy ME. 2002. Crystal structure of the soluble domain of the
 1857 major anaerobically induced outer membrane protein (AniA) from pathogenic
 1858 *Neisseria*: a new class of copper-containing nitrite reductases. *J Mol Biol* 315:1111-
 1859 27.

1860 229. Ku SC, Schulz BL, Power PM, Jennings MP. 2009. The pilin O-glycosylation pathway of
 1861 pathogenic *Neisseria* is a general system that glycosylates AniA, an outer membrane
 1862 nitrite reductase. *Biochem Biophys Res Commun* 378:84-9.

1863 230. Falsetta ML, Steichen CT, McEwan AG, Cho C, Ketterer M, Shao J, Hunt J, Jennings
 1864 MP, Apicella MA. 2011. The composition and metabolic phenotype of *Neisseria*
 1865 *gonorrhoeae* biofilms. *Front Microbiol* 2:75.

1866 231. Smith H, Parsons NJ, Cole JA. 1995. Sialylation of *Neisseria* lipopolysaccharide: a
 1867 major influence on pathogenicity. *Microb Pathog* 19:365-77.

1868 232. Shell DM, Chiles L, Judd RC, Seal S, Rest RF. 2002. The *Neisseria* lipooligosaccharide-
1869 specific alpha-2,3-sialyltransferase is a surface-exposed outer membrane protein.
1870 Infect Immun 70:3744-51.

1871 233. Packiam M, Shell DM, Liu SV, Liu YB, McGee DJ, Srivastava R, Seal S, Rest RF. 2006.
1872 Differential expression and transcriptional analysis of the alpha-2,3-sialyltransferase
1873 gene in pathogenic *Neisseria* spp. Infect Immun 74:2637-50.

1874 234. Martin D, Cadieux N, Hamel J, Brodeur BR. 1997. Highly conserved *Neisseria*
1875 meningitidis surface protein confers protection against experimental infection. J Exp
1876 Med 185:1173-83.

1877 235. Gulati S, McQuillen DP, Sharon J, Rice PA. 1996. Experimental immunization with a
1878 monoclonal anti-idiotypic antibody that mimics the *Neisseria gonorrhoeae*
1879 lipooligosaccharide epitope 2C7. J Infect Dis 174:1238-48.

1880 236. Banerjee A, Wang R, Uljon SN, Rice PA, Gotschlich EC, Stein DC. 1998. Identification
1881 of the gene (*lgtG*) encoding the lipooligosaccharide beta chain synthesizing glucosyl
1882 transferase from *Neisseria gonorrhoeae*. Proc Natl Acad Sci USA 95:10872-7.

1883 237. Ngampasutadol J, Rice PA, Walsh MT, Gulati S. 2006. Characterization of a peptide
1884 vaccine candidate mimicking an oligosaccharide epitope of *Neisseria gonorrhoeae*
1885 and resultant immune responses and function. Vaccine 24:157-70.

1886 238. Gulati S, Agarwal S, Vasudhev S, Rice PA, Ram S. 2012. Properdin is critical for
1887 antibody-dependent bactericidal activity against *Neisseria gonorrhoeae* that recruit
1888 C4b-binding protein. J Immunol 188:3416-25.

1889 239. Chakraborti S, Lewis LA, Cox AD, St Michael F, Li J, Rice PA, Ram S. 2016. Phase-
1890 variable heptose I glycan extensions modulate efficacy of 2C7 vaccine antibody
1891 directed against *Neisseria gonorrhoeae* lipooligosaccharide. J Immunol 196:4576-86.

1892 240. Mulks MH, Knapp JS. 1987. Immunoglobulin A1 protease types of *Neisseria*
1893 gonorrhoeae and their relationship to auxotype and serovar. *Infect Immun* 55:931-
1894 936.

1895 241. Simpson DA, Hausinger RP, Mulks MH. 1988. Purification, characterization, and
1896 comparison of the immunoglobulin A1 proteases of *Neisseria gonorrhoeae*. *J*
1897 *Bacteriol* 170:1866-1873.

1898 242. Lomholt H, Kilian M. 1994. Antigenic relationships among immunoglobulin A1
1899 proteases from *Haemophilus*, *Neisseria*, and *Streptococcus* species. *Infect Immun*
1900 62:3178-83.

1901 243. Lin L, Ayala P, Larson J, Mulks M, Fukuda M, Carlsson SR, Enns C, So M. 1997. The
1902 *Neisseria* type 2 IgA1 protease cleaves LAMP1 and promotes survival of bacteria
1903 within epithelial cells. *Mol Microbiol* 24:1083-94.

1904 244. Lorenzen DR, D  x F, W  lk U, Tsirpouchtsidis A, Haas G, Meyer TF. 1999.
1905 Immunoglobulin A1 protease, an exoenzyme of pathogenic *Neisseriae*, is a potent
1906 inducer of proinflammatory cytokines. *J Exp Med* 190:1049-58.

1907 245. Karlinsky D, Prokopenko Y, Zinchenko A, Zhigis L, Kotelnikova O, Rumsh L, Smirnov I.
1908 2022. Highly similar sequences of mature IgA1 Proteases from *Neisseria meningitidis*,
1909 *Neisseria gonorrhoeae* and *Haemophilus influenzae*. *Pathogens* 11:734.

1910 246. Leuzzi R, Serino L, Scarselli M, Savino S, Fontana MR, Monaci E, Taddei A, Fischer G,
1911 Rappuoli R, Pizza M. 2005. Ng-MIP, a surface-exposed lipoprotein of *Neisseria*
1912 gonorrhoeae, has a peptidyl-prolyl cis/trans isomerase (PPIase) activity and is
1913 involved in persistence in macrophages. *Mol Microbiol* 58:669-81.

1914 247. Humbert MV, Christodoulides M. 2018. Immunization with recombinant truncated
1915 *Neisseria meningitidis*-Macrophage Infectivity Potentiator (rT-Nm-MIP) protein

1916 induces murine antibodies that are cross-reactive and bactericidal for *Neisseria*
 1917 gonorrhoeae. *Vaccine* 36:3926-3936.

1918 248. Christodoulides M. 2022. Update on the *Neisseria* macrophage infectivity
 1919 potentiator-like PPlase protein. *Front Cell Infect Microbiol* 12:861489.

1920 249. Delahay RM, Robertson BD, Balthazar JT, Shafer WM, Ison CA. 1997. Involvement of
 1921 the gonococcal MtrE protein in the resistance of *Neisseria gonorrhoeae* to toxic
 1922 hydrophobic agents. *Microbiology (Reading)* 143 (Pt 7):2127-2133.

1923 250. Lee EH, Shafer WM. 1999. The farAB-encoded efflux pump mediates resistance of
 1924 gonococci to long-chained antibacterial fatty acids. *Mol Microbiol* 33:839-45.

1925 251. Veal WL, Nicholas RA, Shafer WM. 2002. Overexpression of the MtrC-MtrD-MtrE
 1926 efflux pump due to an mtrR mutation is required for chromosomally mediated
 1927 penicillin resistance in *Neisseria gonorrhoeae*. *J Bacteriol* 184:5619-24.

1928 252. Baarda BI, Zielke RA, Nicholas RA, Sikora AE. 2018. PubMLST for antigen allele mining
 1929 to inform development of gonorrhea protein-based vaccines. *Front Microbiol*
 1930 9:2971.

1931 253. Piekarowicz A, Klyz A, Majchrzak M, Stein DC. 2016. Oral immunization of rabbits
 1932 with *S. enterica* Typhimurium expressing *Neisseria gonorrhoeae* filamentous phage
 1933 Phi6 induces bactericidal antibodies against *N. gonorrhoeae*. *Sci Rep* 6:22549.

1934 254. Klyz A, Piekarowicz A. 2018. Phage proteins are expressed on the surface of *Neisseria*
 1935 gonorrhoeae and are potential vaccine candidates. *PLoS One* 13:e0202437.

1936 255. Huang J, Zhang Q, Chen J, Zhang T, Chen Z, Chen Z, Yang J, Wang Y, Min Z, Huang M,
 1937 Min X. 2020. *Neisseria gonorrhoeae* NGO2105 is an autotransporter protein involved
 1938 in adhesion to human cervical epithelial cells and in vivo colonization. *Front*
 1939 *Microbiol* 11:1395.

1940 256. Thomas CE, Zhu W, Van Dam CN, Davis NL, Johnston RE, Sparling PF. 2006.
1941 Vaccination of mice with gonococcal TbpB expressed in vivo from Venezuelan equine
1942 encephalitis viral replicon particles. Infect Immun 74:1612-20.

1943

1944 **AUTHOR BIOGRAPHIES**

1945

1946 **Eloise Williams** is an Infectious Diseases Physician and Clinical Microbiologist at the
1947 Victorian Infectious Diseases Reference Laboratory, Melbourne, Australia, and is currently
1948 undertaking a Ph.D. through the Department of Infectious Diseases at the Peter Doherty
1949 Institute of Infection and Immunity at the University of Melbourne. She completed her
1950 medical studies (MBBS) at the University of Melbourne and a Masters of Public Health and
1951 Tropical Medicine at James Cook University. Her research interests include public health,
1952 sexually-transmitted infections and blood-borne viruses. A primary aim of her Ph.D. is to
1953 develop a *N. gonorrhoeae* oropharyngeal controlled-human infection model to further
1954 characterise the pathogenesis of oropharyngeal *N. gonorrhoeae* infection and accelerate
1955 the development of novel vaccines and therapeutics.

1956

1957 **Kate L. Seib** is a Professor of Microbiology at Griffith University, where she is a Group Leader
1958 and Associate Director of Research at the Institute for Glycomics. She completed a Ph.D in
1959 microbiology in 2004 at the University of Queensland and was a Postdoctoral Fellow and
1960 Project Leader at Novartis Vaccines, where she was part of the team working on the
1961 meningococcal B vaccine, 4CMenB. Her research focuses on *N. gonorrhoeae* pathogenesis
1962 and host immune response, with the aim of identifying therapeutic and preventative targets
1963 against *N. gonorrhoeae* infection. She has also led a number of studies modelling the impact

1964 of *N. gonorrhoeae* vaccines and is leading a multicentre randomised clinical trial evaluating
1965 the efficacy of the 4CMenB vaccine against *N. gonorrhoeae* infection.

1966

1967 **Christopher K. Fairley** is the Director of the Melbourne Sexual Health Centre and Professor
1968 of Public Health at Monash University. His principle research interests are the public health
1969 control of sexually transmitted infections and the effectiveness of clinical services. He has
1970 been described by the Lancet as a 'pioneer of sexually transmitted infection research'. His
1971 work has substantially contributed to the understanding of the epidemiology, microbiology
1972 and novel treatment and prevention approaches for *N. gonorrhoeae* infection. In particular
1973 his work has been pivotal in identifying the significant role of the oropharynx in gonorrhoea
1974 transmission.

1975

1976

1977 **Georgina L. Pollock** is a Post-Doctoral Researcher in the Department of Infectious Diseases
1978 at the Peter Doherty Institute for Infection and Immunity at the University of Melbourne.
1979 She completed a Ph.D investigating the molecular mechanisms used by pathogenic
1980 *Escherichia coli* to evade the immune responses in the gut at the University of Melbourne in
1981 2019, and is now part of a team that uses genomic approaches to investigate the dynamics
1982 of sexually transmitted infections. Her current research is focused on developing a *N.*
1983 *gonorrhoeae* oropharyngeal controlled human infection model.

1984

1985 **Jane S. Hocking** is a Professor of Epidemiology and implementation researcher at the
1986 Melbourne School of Population and Global Health at the University of Melbourne. She
1987 completed a Masters of Public Health and Ph.D at the University of Melbourne. Her research

1988 interests include the epidemiology and control of sexually transmitted infections, sexual
1989 health and the implementation and evaluation of primary care interventions. Her work has
1990 substantially contributed to the understanding of the significant role of the oropharynx in
1991 gonorrhoea transmission and novel approaches to gonorrhoea prevention and prevention.

1992

1993 **James S. McCarthy** is Director of the Victorian Infectious Diseases Service at the Royal
1994 Melbourne Hospital, and Professor of Medicine at the Doherty Institute. He received his
1995 medical degree from the University of Melbourne before undertaking clinical and research
1996 training in Australia, the UK, and the US at the University of Maryland and the Laboratory
1997 for Parasitic Diseases, National Institutes of Health, Bethesda, MD, before returning to
1998 Australia in 1997. His research has focussed on the diagnosis and treatment of parasitic
1999 diseases, with a major recent focus on the development and application of controlled
2000 human infection models of malaria and other pathogenic organisms, including *Neisseria*
2001 *gonorrhoeae*. This has enabled study of the host-pathogen interaction, development of
2002 diagnostic biomarkers and the evaluation of investigational drugs and vaccines

2003

2004 **Deborah A. Williamson** is a member of the Royal College of Physicians and a Fellow of the
2005 Royal College of Pathologists. She has had a number of roles in clinical and public health
2006 microbiology in Australia, including as Deputy Director of Microbiological Diagnostics Unit
2007 Public Health Laboratory, Director of Microbiology at Royal Melbourne Hospital and Director
2008 of the Victorian Infectious Diseases Reference Laboratory. Her research has focused on
2009 public health microbiology, particularly sexually-transmitted infections, microbial genomics
2010 and antimicrobial resistance. Over the past decade, Deborah has undertaken numerous

2011 studies of the epidemiology, microbiology and novel treatment and prevention strategies

2012 for *N. gonorrhoeae* infection.

TABLES

Table 1. Historical *Neisseria gonorrhoeae* vaccine trials in humans

Clinical trial design	Vaccine	Immunization schedule	Study population	Result	Reference
Randomized double-blind placebo-controlled trial	Inactivated whole-cell vaccine prepared from three strains of <i>N. gonorrhoeae</i>	1ml dose of intramuscular immunization 3 times at 1-week intervals	62 participants recruited from an indigenous population of Inuit in northern Canada (background yearly <i>N. gonorrhoeae</i> infection incidence of 25%)	Cumulative infection rate of 30% in immunized participants compared to 24% in placebo in the 12 month follow-up period following immunization (ns)	Greenberg et al 1974 (64)

Randomized double-blind placebo-controlled trial	Single-antigen pilus protein vaccine prepared from single strain of <i>N. gonorrhoeae</i>	0.1ml dose of intradermal immunization 2 times at 2-week interval	3250 US military personnel stationed in Korea (96% men; 39% with self-reported history of prior <i>N. gonorrhoeae</i> infection)	Cumulative infection rate 6.9% in immunized participants compared to 6.5% in placebo in 8-week follow-up period following immunization (ns)	Boslego et al 1991 (65)
Placebo-controlled human challenge trial	Outer membranes vaccine prepared from single strain of <i>N. gonorrhoeae</i>	Participants vaccinated (dosing schedule not available) then inoculated with homologous <i>N.</i>	63 male participants	Post-challenge infection rate 54% in immunized participants compared to 64% in placebo (ns)	Rice et al (66)

		<i>gonorrhoeae</i> strain per urethra 2-4 weeks later			
--	--	---	--	--	--

ns, not significant; US, United States.

Table 2. Potential *Neisseria gonorrhoeae* vaccine antigens discovered by traditional or reverse vaccinology approaches

Gene	Protein/ Antigen name	Function	Location	Conservation*	Immunogenicity	Data	Reference
Adherence and invasion of epithelial cells							
Phospholipase							
<i>pldA</i>	Outer membrane phospho- lipase A (OMPLA)	Phospholipid hydrolysis of endogenous phospholipids. Autolysin	Outer membrane	Highly conserved	Murine antibodies elicited by <i>N. meningitidis</i> homologue are not bactericidal or protective against infection	Preclinical	Senff et al 1976 (172); Cacciapuoti et al 1978 (173); Bos et al 2005 (174)
<i>PLD</i>	<i>Neisseria</i> <i>gonorrhoea</i> e phospho-	Regulator of gonococcal invasion of and survival within	Outer membrane	Highly conserved	Antibodies decrease adherence to and invasion of primary cervical cells	Preclinical	Edwards et al 2003 (92); Edwards and Apicella 2006

	lipase D (NgPLD)	cervical epithelia					(93); Edwards et al 2016 (175)
Pilin							
<i>pilE</i>	Major subunit of the type 4 pilus	Type 4 pilus fiber. Channel for pilus extrusion. Mediates adherence to epithelial cells	Outer membrane	Antigenically variable. Conserved at C terminus	Antibodies to pili block cell attachment but are directed at variable epitopes	Historical vaccine trial	Tramont et al 1981 (176); Siegel et al 1982 (177); Brinton et al 1982 (178); Schoolnik et al 1983 (179); Virji and Heckels 1984 (180); Rothbard et al 1985 (181); Tramont et al

							1985 (70); Boslego et al 1991 (65)
<i>pilC</i>	PilC	Type 4 pilus tip-associated adhesin. Plays key role in pilus biogenesis and adhesion	Outer membrane	Antigenically variable. Phase variable	No data	Preclinical	Backman et al 1998 (87); Morand et al 2001 (88)
<i>pilQ</i>	PilQ	Outer membrane channel for pilus extrusion.	Outer membrane	Antigenically variable. Conserved at C terminus	Antibodies elicited by <i>N. meningitidis</i> homologs are bactericidal	Preclinical	Drake and Koomey 1995 (89); Helm et al 2007 (90); Haghi et al 2012 (91)

		Essential role in pilus biogenesis					
Porin							
<i>porB</i>	Porin	Major outer membrane protein. Nutrient channel. Binds complement factors C4bp and Factor H to down- regulate	Outer membrane	Antigenically variable surface loops and conserved membrane- spanning regions	Antibodies are bactericidal, opsonophagocytotic and block gonococcal entry into epithelial cells	Preclinical	Hook et al 1984 (182); Heckels et al 1992 (183); Christodoulides et al 1993 (184); Ram et al 1998 (185); Ram et al 1998 (186); Ram et al 2001 (187); Edwards et al 2002 (188);

		complement activation at gonococcal surface. Suppresses neutrophil oxidative burst and neutrophil apoptosis					Zhu et al 2004 (130); Zhu et al 2005 (129); Kulewein et al 2006 (189); Garvin et al 2008 (190); Faulstich et al 2013 (191); McKnew et al 2013 (84); Yuen et al 2019 (86)
Other outer membrane proteins							

<i>opa</i>	Opacity proteins	Adherence and invasion of host cells Influence innate and adaptive immune responses by binding CEACAM receptors on T and B lymphocytes	Outer membrane	Antigenically variable. Phase variable	Antibodies are bactericidal	Preclinical and controlled human challenge studies	Virji et al 1993 (192); Plummer et al 1994 (193); Chen et al 1997 (194); de Jonge et al 2004 (143); Cole et al 2009 (195); Callaghan et al 2011 (196); Sadarangani et al 2011 (197);
------------	------------------	---	----------------	---	-----------------------------	--	--

<i>opcA</i>	OpcA	Adhesion and invasion of host epithelial and endothelial cells	Outer membrane	Antigenically variable	Antibodies elicited by <i>N. meningitidis</i> homologues are bactericidal	Preclinical	Zhu et al 2003 (198); Moore et al 2005 (199); Keiser et al 2010 (200);
<i>ompA</i>	Outer membrane protein A (OmpA)	Adhesion and invasion of host epithelial and endothelial cells	Outer membrane	Highly conserved	No data	Preclinical	Serino et al 2007 (201); Starnino et al 2010 (202)
<i>nhba</i>	<i>Neisseria</i> heparin binding	Involved in adherence to epithelial cells	Outer membrane	Highly conserved	Antibodies are bactericidal, opsono-phagocytotic and block	Preclinical	Marjuki et al 2019 (94); Semchenko et al

	antigen (NHBA)	and serum survival			gonococcal adherence to epithelial cells		2019 (153); Semchenko et al 2020 (96); Semchenko et al 2020 (203)
Nutrient acquisition and metabolism							
Iron metabolism							
<i>tbpA</i>	Transferrin- binding protein A (TbpA)	Essential receptor for iron uptake from transferrin	Outer membrane	Highly conserved	Antibodies are bactericidal	Preclinical and controlled human challenge studies	Cornelissen et al 1998 (97); Masri and Cornelissen 2002 (204); Price et al 2004 (205); Price et al 2005 (98); Price et al 2007 (206);

							Hobbs et al 2011 (51); Cash et al 2015 (81)
<i>tbpB</i>	Transferrin- binding protein B (TbpB)	Increases efficiency of iron uptake from transferrin	Outer membrane	Antigenically variable with conserved segments	Antibodies are bactericidal	Preclinical and controlled human challenge studies	Cornelissen et al 1998 (97); Price et al 2004 (205); Price et al 2005 (98); Price et al 2007 (206); de Rocco and Cornelissen 2007 (207); Hobbs et al 2011 (51); Ostberg et al

							2013 (208); Cash et al 2015 (81)
<i>lbpA</i>	Lactoferrin-binding protein A (LbpA)	Essential receptor for iron uptake from lactoferrin	Outer membrane	Highly conserved. Present in approximately half of isolates.	Antibodies elicited by <i>N. meningitidis</i> homologues are bactericidal but cross reactivity (in <i>N. meningitidis</i>) is limited	Preclinical and controlled human challenge studies	Mickelsen et al 1982 (209); Biswas et al 1999 (210); Anderson et al 2003 (211); Pettersson et al 2006 (212); Adamiak et al 2012 (213); Noinaj et al 2013 (214)

<i>lbpB</i>	Lactoferrin-binding protein B (LbpB)	Increases the efficiency of iron transport from lactoferrin	Outer membrane	Antigenically variable with conserved segments. Phase variable. Present in approximately half of isolates	Antibodies elicited by <i>N. meningitidis</i> homologues are bactericidal but cross reactivity (in <i>N. meningitidis</i>) is limited	Preclinical and controlled human challenge studies	Mickelsen et al 1982 (209); Biswas et al 1999 (210); Anderson et al 2003 (211); Pettersson et al 2006 (212); Adamiak et al 2012 (213); Noinaj et al 2013 (214)
<i>fetA</i>	Ferric entero-bactin	Involved in iron uptake through	Outer membrane	Antigenically variable. Phase variable	Antibodies elicited by <i>N. meningitidis</i> homologues are bactericidal but cross	Preclinical	Black et al 1986 (215); Dyer et al 1988 (216);

	transporter A (FetA)	scavenging siderophores from other bacteria via binding and transport of ferric enterobactin			reactivity (in <i>N. meningitidis</i>) is limited		Pettersson et al 1990 (217); Van der Ley et al 1996 (218); Carson et al 1999 (219); Carson et al 2000 (220)
<i>fetB</i>	Ferric entero- bactin transporter B (FetB)	Involved in iron uptake through scavenging siderophores from other bacteria via	Outer membrane	Antigenically variable	No data	Preclinical	Baarda et al (80)

		binding and transport of ferric enterobactin					
Zinc metabolism							
<i>tdfJ</i>	TonB- dependent family J (TdfJ)	Facilitates uptake of zinc via human protein S100A7	Outer membrane	Highly conserved	Antibodies elicited by <i>N. meningitidis</i> homologue are bactericidal	Preclinical	Stork et al 2010 (221); Cornelissen and Hollander 2011 (222); Maurakis 2019 (223)
<i>tdfH</i>	TonB- dependent family H (TdfH)	Facilitates uptake of zinc via human calprotectin	Outer membrane	Highly conserved	No data	Preclinical	Turner et al 2001 (224); Jean et al 2016 (225);

							Kammerman et al 2020 (226)
Anaerobic metabolism							
<i>aniA</i>	Anaerobically induced protein A (AniA)	Inducible nitrite reductase, required for anaerobic growth and biofilm formation	Outer membrane	Highly conserved	Antibodies block nitrite reductase activity	Preclinical	Clark et al 1988 (227); Boulanger et al 2002 (228); Ku et al 2008 (229); Falsetta et al 2009 (99); Falsetta et al 2011 (230); Shewell et al 2013 (102); Shewell et al 2017 (101)

Immune evasion							
<i>lst</i>	Alpha-2,3-sialy-transferase (Lst)	Sialylates the surface lipooligosaccharide to protect gonococci from complement-mediated killing and phagocytic killing by neutrophils. Incorporates	Cytoplasm (previously thought to be outer membrane)	Highly conserved	Antibodies partially inhibit sialyltransferase activity of <i>N. gonorrhoeae</i> however this is inhibited in the presence of exogenous 5'-cytidinemonophospho-N-acetylneuraminic acid (CMP-NANA) present in <i>N. gonorrhoeae</i> strains. KDO-specific monoclonal antibody 6E4 is opsonophagocytic.	Preclinical	Smith 1995 (231); Shell 2002 (232); Packiam 2006 (233); Wu & Jerse 2006 (104); Lewis et al 2015 (103); Jen et al 2021 (105)

		keto-deoxyoctanoate (KDO) as the terminal glycan on the LOS					
<i>nspA</i>	Neisserial surface protein A (NspA)	Subverts complement pathway activation by binding to complement inhibitor factor H	Outer membrane	Highly conserved	Antibodies are bactericidal and opsonophagocytic	Preclinical	Martin et al 1997 (234); Li et al 2011 (107); Lewis et al 2019 (106)
Intracellular survival							

<i>lgtG</i>	Lipooligo- saccharide (LOS) epitope 2C7	Inner glucose core of LOS. Promotes colonization and survival	Outer membrane	High antigenic conservation. Phase variable.	Antibodies are bactericidal and opsonophagocytotic	Preclinical	Gulati et al 1996 (108, 235); Banerjee et al 1998 (236); Ngampasutadol 2006 (237); Gulati et al 2012 (238), Gulati et al 2013 (110); Chakraborti et al 2016 (239) Gulati et al 2019 (109, 111)
<i>iga</i>	IgA1- specific	Promotes intracellular	Outer membrane	Highly conserved.	No data	Preclinical	Mulks and Knapp 1987

	protease (IgA1)	survival and release of inflammatory cytokines		Present in approximately 50% of isolates			(240); Simpson et al 1988 (241); Lomholt et al 1995 (242); Lin et al 1997 (243); Lorenzen et al 1999 (244); Karlinsky et al 2022 (245)
<i>mip</i>	Macro- phage Infectivity Potentiator (MIP) lipoprotein	Bacterial persistence within macrophages and protects <i>Neisseria</i>	Outer membrane	Highly conserved	Antibodies are bactericidal	Preclinical	Leuzzi et al 2005 (246) Humbert & Christodoulides 2018 (247);

		<i>gonorrhoeae</i> from bactericidal activity of immune effector cells					Christodoulides 2022 (248)
Oxidative stress and antimicrobial substance protection							
<i>msrA/B</i>	Methionine sulfoxide reductase (MsrA/B)	Protects from oxidative stress by reducing methionine sulfoxide to methionine	Outer membrane	Highly conserved	Antibodies are bactericidal, opsonophagocytic and functionally block the activity of MsrA/B by binding to its substrate, methionine sulfoxide	Preclinical	Jen et al 2019 (83)

<i>mtrE</i>	Multiple transferable resistance protein E (MtrE)	Surface-exposed channel of the MtrCDE and FarAB-MtrE efflux pumps that export antimicrobial substances	Outer membrane	Highly conserved. Expression upregulated in multi-drug resistant strains	Antibodies are bactericidal	Preclinical	Delahay et al 1997 (249); Lee & Schafer 1999 (250); Veal et al 2002 (251); Jerse & Deal 2013 (114); Wang et al 2018 (82); Handing et al 2018 (113); Baarda et al 2018 (252)
Other							

NgoΦ6	Filamentous bacteriophage proteins	Encodes proteins needed for progeny phage production	Outer membrane	Highly conserved	Antibodies are bactericidal and block adherence to cervical epithelial cells	Preclinical	Piekarowicz et al 2016 (253); Klyz & Piekarowicz 2018 (254)
Proteomic and bioinformatic vaccine antigen discovery							
<i>acp</i>	Adhesin complex protein (ACP)	Inhibition of host lysozyme activity, promotes host cell colonization	Outer membrane	Highly conserved	Antibodies are bactericidal and inhibit human lysozyme	Preclinical	Humbert et al 2017 (115); Almonacid-Mendoza et al 2018 (116)
<i>iga2</i>	IgA2 protease (AidA)	Putative adhesion and	Cell envelope	Antigenically variable	No data	Preclinical	El Rami et al 2019 (77); Huang et al

		penetration protein					2020 (255); Baarda et al 2021 (80)
<i>bamA</i>	Beta-barrel assembly machinery protein A (BamA)	Folds and inserts beta- barrel proteins into the outer membrane	Outer membrane	Highly conserved	Antibodies are bactericidal	Preclinical	Zielke et al 2016 (76); Baarda et al 2018 (252)
<i>bamE</i>	Beta-barrel assembly machinery protein E (BamE)	Contributes to outer membrane assembly and integrity	Outer membrane	Highly conserved	No data	Preclinical	El-Rami et al 2019 (77); Baarda et al 2021 (80)

<i>csgG</i>	Curl-specific gene G (CsgG)	Membrane protein	Outer membrane	Moderately conserved	No data	Preclinical	El Rami et al 2019(77); Baarda et al 2021 (80)
<i>lolB</i>	Lipoprotein outer membrane localization lipoprotein B (LoIB)	Putative role in lipoprotein trafficking to the outer membrane	Outer membrane	Moderately conserved	No data	Preclinical	El Rami et al 2019 (77); Baarda et al 2021 (80)
<i>lprI</i>	Lipoprotein I (LprI)	Putative lysozyme resistance protein	Cell envelope	Moderately conserved	No data	Preclinical	El Rami et al 2019 (77); Baarda et al 2021 (80)

<i>lptD</i>	Lipopoly-saccharide assembly protein D (LptD)	Lipopoly-saccharide assembly	Outer membrane	Moderately conserved	Antibodies are bactericidal	Preclinical	Zielke et al 2014 (76); Zielke et al 2016 (76)
<i>lptE</i>	Lipopoly-saccharide assembly protein E (LptE)	Putative role in lipopoly-saccharide assembly	Outer membrane	No data	No data	Preclinical	El-Rami et al 2019 (77)
<i>mafA</i>	Multiple adhesin family A (MafA)	Adhesin	Cell envelope	Antigenically variable	No data	Preclinical	El Rami et al 2019 (77); Baarda et al 2021 (80)

<i>metQ</i>	Methionine binding lipoprotein Q (MetQ)	Methionine transport adhesin involved in epithelial cell adherence and survival	Outer membrane	Highly conserved	Antibodies are bactericidal and block gonococcal adherence to human cervical epithelial cells	Preclinical	Zielke et al 2016 (76); Semchenko et al 2017 (118); El-Rami et al 2019 (77); Sikora et al 2020 (119);
NGO0416	-	Hypothetical protein with conserved domain similarity to N-terminal domain of LamB	Periplasm	Moderately conserved	Limited bactericidal antibodies	Preclinical	Zhu et al 2019 (79)

		carbohydrate-specific outer membrane porin					
NGO0425	-	Hypothetical protein	Cell envelope	Moderately conserved	No data	Preclinical	El Rami et al 2019 (77); Baarda et al 2021 (80)
NGO0690	-	Putative lipoprotein possibly involved in threonine biosynthesis	Periplasm/outer membrane	Moderately conserved	Antibodies are bactericidal	Preclinical	Zhu et al 2019 (79)

		and pilin antigenicity					
NGO0778	-	Membrane protein	Cell envelope	Highly conserved	No data	Preclinical	El Rami et al 2019 (77); Baarda et al 2021 (80)
NGO0948	-	Lipoprotein member of NlpB/DapX family	Periplasm/ outer membrane	Moderately conserved	Antibodies are bactericidal	Preclinical	Zhu et al 2019 (79)
NGO1043	-	Putative lipoprotein, possibly glycosylated and a	Periplasm/ outer membrane	Moderately conserved	Antibodies are bactericidal	Preclinical	Zhu et al 2019 (79)

		substrate for phospho- ethanolamine addition					
NGO1215	-	Putative protein with homology to a copper chaperone superfamily	Periplasm	Moderately conserved	Antibodies are bactericidal	Preclinical	Zhu et al 2019 (79)
NGO1251	-	Lipoprotein	Cell envelope	Highly conserved	No data	Preclinical	El Rami et al 2019 (77); Baarda et al 2021 (80)

NGO1701	-	Putative with homology to copper - binding protein of the DUF326 super family	Periplasm	Moderately conserved	Antibodies are bactericidal	Preclinical	Zhu et al 2019 (79)
NGO2054	-	Unknown	Outer membrane	Highly conserved	Antibodies are bactericidal	Preclinical	Zielke et al 2016 (76)
<i>ompU</i>	Outer membrane porin protein U (OmpU)	Putative iron uptake protein	Outer membrane	Moderately conserved	No data	Preclinical	El Rami et al 2019 (77); Baarda et al 2021 (80)

<i>sliC</i>	Surface-exposed lysozyme inhibitor of c-type lysozyme (SliC)	Inhibition of host lysozyme activity, promotes host colonization	Outer membrane	Highly conserved	No data	Preclinical	Zielke et al 2018 (117); Baarda et al 2021 (80)
<i>tamA</i>	Trans-location and assembly module A (TamA)	Translocation assembly	Outer membrane	Moderately conserved	Antibodies are bactericidal	Preclinical	Zielke et al 2016 (76)

*Conservation: Amino acid sequence conservation between *N. gonorrhoeae* strains: highly conserved $\geq 80\%$, moderately conserved $\geq 50\%$, antigenically variable $<50\%$.

Table 3. Contemporary *Neisseria gonorrhoeae* vaccines that have proceeded to preclinical studies in the experimental mouse model

Trial design	Vaccine	Immunization schedule	Immunogenicity	Attenuation of gonococcal infection	Reference
Estradiol-treated BALB/c mouse model inoculated vaginally with <i>N. gonorrhoeae</i> MS11 approximately 3 weeks after immunization	Gonococcal outer membrane preparation from <i>N. gonorrhoeae</i> strain MS11	IN or SC administration 3 times at 3 week intervals	Serum and vaginal antibodies induced by both IN and SC immunization. SBA to heterologous gonococcal strain induced by IN immunization.	Clearance of gonococcal colonization significantly faster in IN immunized compared to control mice.	Plante et al 2000 (139)

BALB/c mouse model immunized with putative vaccine	Gonococcal recombinant plasmid encoding PorB DNA (PorB DNA) from <i>N. gonorrhoeae</i> strain FA1090 prime vaccine followed by either PorB DNA, renatured recombinant PorB protein (rrPorB) plus Ribi R-700 adjuvant or PorB expressed from Venezuelan equine encephalitis virus	IM or epidermal gene gun bombardment administration with prime PorB DNA followed by boost with PorB DNA, rrPorB or PorB-VRPs 4- weeks later	Serum antibodies induced by both IM and epidermal gene gun bombardment, with Th1 response induced by IM administration and Th2 response induced by gene gun bombardment. Boosting with rrPorB and PorB VRPs significantly	No data	Zhu et al 2004 (130)
--	--	--	---	---------	-----------------------------

	replicon particles (PorB-VRPs)		increased PorB IgG and IgA antibodies. Serum OPA to homologous gonococcal strain, SBA not produced.		
BALB/c mouse model immunized with various combinations of putative vaccines	<i>N. gonorrhoeae</i> vaccines produced from strain FA1090, including renatured recombinant PorB protein (rrPorB) plus Ribi R-700 adjuvant or PorB expressed from Venezuelan equine	SC administration into dorsal area or hind footpad (rrPorB), SC administration into hind footpad (PorB VRP) or IN	Serum anti-PorB antibodies induced by all vaccines tested, with Th1 bias for PorB-VRP and rrPorB in footpad and Th2 bias when	No data.	Zhu et al 2005 (129)

	encephalitis virus replicon particles (PorB-VRPs) or outer membrane vesicle (OMV) vaccine	(OMV) 3-4 times, 3 weeks apart.	rrPorB given in dorsal area. IN OMV induced SBA whilst other vaccines did not.		
BALB/c mouse model immunized with various combinations of vaccine components	Various combinations of <i>N. gonorrhoeae</i> strain FA19 1) recombinant transferrin binding protein A (rTbpA) plus Ribi R-700 adjuvant, 2) recombinant transferrin binding protein B (rTbpB) and 3) cholera	IN or SC administration 3 times at 10 day intervals	Serum and vaginal antibodies induced by IN immunization for each Tbp antigen combined with Ctb. SBA induced by IN immunization.	No data	Price et al 2005 (98)

	toxin B subunit (Ctb), either as conjugates or admixed.				
BALB/c mouse model immunized with putative vaccine	<i>N. gonorrhoeae</i> vaccines produced from strain FA1090, including renatured recombinant TbpB (rrTbpB) and TbpB expressed from Venezuelan equine encephalitis virus replicon particles (TbpB-VRPs)	SC immunization at 0, 4, 7 and 10 weeks	Serum antibodies induced by both TbpB vaccines, with highest titers in mice immunized with rrTbpB. TbpB-VRP responses Th1- biased. Mucosal antibodies produced by both vaccines with	No data	Thomas et al 2006 (256)

			highest titers in mice immunized or boosted with rrTbpB. Bactericidal antibodies not produced		
Estradiol-treated BALB/c female mouse model inoculated with <i>N. gonorrhoeae</i> strain FA1090 approximately 2 weeks after immunization	OMV preparation from <i>N. gonorrhoeae</i> strain FA1090 combined with IL-12 microspheres	Intravaginal immunization 3 times at 1 week intervals	Intravaginal OMV/IL-12 microsphere vaccination induced serum and vaginal IgG and IgA antibodies against	Clearance of <i>N. gonorrhoeae</i> colonization significantly faster in mice immunized with OMV/IL-12 microsphere vaccine candidate compared to those	Liu et al 2017 (121)

			homologous and heterologous strains.	immunized with OMV or IL-12 microspheres alone	
Smith Webster (CFW) mouse model immunized with putative vaccine	Whole-cell formalin-inactivated microparticle vaccine from <i>N. gonorrhoeae</i> strain CDC-F62 loaded in dissolvable microneedles	Transdermal immunization applied for 20 minutes, 3 times, 2 weeks apart	Transdermal microparticle vaccination induced greater serum IgG antibodies than SC vaccination and induced elevated CD4+ and CD8+ responses comparable to SC vaccination.	No data	Gala et al 2018 (144)

BALB/c mouse model immunized with putative vaccine	<i>porB</i> gene DNA from <i>N. gonorrhoeae</i> strain WHO-A inserted into eukaryotic expression vector pVAX1 (pVAX1- <i>porB</i>) loaded in <i>S. enteritidis</i> ghosts (SE ghosts (pVAX1- <i>porB</i>))	PO immunization 3 times at 2 week intervals	SE bacterial ghosts (pVAX1- <i>porB</i>) vaccination induced greater serum IgG antibodies, CD4+ and CD8+ T cell responses than pVAX1- <i>porB</i> DNA vaccine alone. SBA induced.	No data	Jiao et al 2018 (132)
BALB/cAnNCr mouse model inoculated with <i>N. gonorrhoeae</i> F62 3 weeks after immunization	Meningococcal detoxified outer membrane vesicle (dOMV) vaccine	IP immunization 3 times at 3 week intervals	dOMV vaccine produced induced serum and vaginal	A significantly higher proportion of mice immunized with meningococcal dOMV	Beernik et al 2019 (138)

	prepared from meningococcal strains deleted for major outer membrane proteins (including PorA, PorB and RmpM) plus Alhydrogel adjuvant		antibodies. SBA not detected.	vaccines prepared from strains deleted for major outer membrane proteins were cleared of Ng compared to control.	
BALB/c mouse model immunized with putative vaccine	<i>nspA</i> gene DNA from <i>N. gonorrhoeae</i> strain WHO-A inserted into eukaryotic expression vector pVAX1 (pVAX1- <i>nspA</i>) either alone (SE ghosts pVAX1- <i>nspA</i>) or in combination with SE	PO immunization 3 times at 2 week intervals	Co-administered SE ghosts (pVAX1- <i>nspA</i>) and SE ghosts (pVAX1- <i>porB</i>) vaccination induced the highest level of anti- <i>nspA</i> and	No data	Jiao et al 2020 (133)

	ghosts (pVAX1-porB) vaccine described above		anti-porB serum IgG and the highest SBA titres		
BALB/c mouse model immunized inoculated with <i>N. gonorrhoeae</i> FA1090 approximately 3 weeks after immunization	Gonococcal recombinant MetQ protein combined with Titermax gold oil-in- water immersion adjuvant subcutaneous vaccine, then subsequently combined CpG 1826 adjuvant intranasal vaccine (rMetQ-CpG)	SC immunization, followed by 3 IN boosts on days 14, 24 and 35	Immunization with rMetQ-CpG induced the highest level of anti-MetQ IgG and IgA serum and vaginal antibodies, with a serum IgG1/IgG2a ratio suggestive of a Th1 response.	Clearance of <i>N.</i> <i>gonorrhoeae</i> colonization significantly faster and with a lower burden of infection in mice immunized with rMetQ- CpG vaccine candidate compared to those immunized with PBS or adjuvant alone.	Sikora et al 2020 (119)

CD-1 mouse model immunized with putative vaccine	Meningococcal native outer membrane vesicle (NOMV) vaccine prepared from meningococcal strain with genetically attenuated endotoxin and overexpressed factor H binding protein (FHbp) or inactivated gene encoding FHbp (NOMV-KO) or recombinant FHbp	IP immunization 2 times at 3-week intervals	Immunization with NOMV-FHbp and NOMV-KO induced gonococcal SBA	No data.	Matthias et al 2022 (137)
Estradiol-treated BALB/c mouse model inoculated	Gonococcal native outer membrane	IN or intravaginal immunization 2	IN and intravaginal	Female mice immunized with IN or	Liu et al 2023 (141)

with <i>N. gonorrhoeae</i> strain FA1090, FA19 or WHO strain F, L or W approximately 2 weeks after immunization; plus BALB/c male mouse model.	vesicle vaccine (NOMV) from strains FA1090; Gonococcal detergent-extracted OMV (dMV) from strain FA19 and double deletion mutant OMV (dm OMV) prepared from mutant <i>N. gonorrhoeae</i> strain MS11 in which genes for Rmp and LpxL1 were deleted to eliminate induction of blocking antibodies against Rmp and to	times at 2-week interval.	immunization of female mice with NOMV plus IL-12 ms induced comparable serum IgG, salivary IgA and vaginal IgG and IgA antigonococcal antibodies. IN immunization of male mice with NOMV plus IL-12 ms induced comparable serum	intravaginal NOMV plus IL-12 ms cleared gonococcal infection faster than mice immunized with control immunization. In addition, female mice cleared gonococcal infection with heterologous strains faster than mice immunized with control immunization. Gonococcal clearance was also accelerated in	
--	---	---------------------------	---	---	--

	<p>decrease LOS</p> <p>endotoxicity; all</p> <p>vaccines combined with</p> <p>IL-12 microspheres (ms)</p>		<p>IgG and saliva IgA</p> <p>antigonococcal</p> <p>antibodies to</p> <p>female mice. IFN-</p> <p>gamma</p> <p>production by</p> <p>CD4+ T cells from</p> <p>iliac lymph nodes</p> <p>was elevated after</p> <p>IN or intravaginal</p> <p>immunization</p> <p>with NOMV plus</p> <p>IL-12 ms.</p>	<p>mice immunized with</p> <p>deOMVs comparable to</p> <p>that seen for NOMV</p> <p>immunized mice;</p> <p>Gonococcal clearance</p> <p>was accelerated in mice</p> <p>immunized with OMV</p> <p>plus IL-12 ms vaccine</p> <p>produced from mutant</p> <p><i>N. gonorrhoeae</i> in</p> <p>which genes for Rmp</p> <p>and LpxL1 were deleted</p> <p>to eliminate induction</p> <p>of blocking antibodies</p> <p>against Rmp and to</p>	
--	---	--	--	--	--

				decrease LOS endotoxicity comparable to that seen for NOMV immunized mice	
Estradiol-treated BALB/c mouse model inoculated with <i>N. gonorrhoeae</i> strain FA1090	Gonococcal native outer membrane vesicle vaccine (NOMV) from Chilean gonococcal strain GC_08175680 (dmGC0817560 NOMV) and FA1090 (dmFA1090 NOMV) with <i>lpxL1</i> and <i>rmp</i> genes deleted to	Parenteral administration.	Immunization with dmGC_08175680 NOMV and dmFA1090 NOMV induced gonococcal specific serum and vaginal mucosal IgG and IgA	Immunization of mice with dmGC_08175680 OMV and dmFA1090 NOMV accelerated clearance of FA1090 from mice significantly faster than 4CMenB	MacLennan et al 2022 (140)

	<p>reduce reactogenicity,</p> <p>minimise production of</p> <p>potentially unprotective</p> <p>antibodies and increase</p> <p>NOMV yield; both</p> <p>vaccines formulated</p> <p>with aluminium</p> <p>hydroxide</p>		<p>antibodies and</p> <p>gonococcal-</p> <p>specific Th1/Th17</p> <p>CD4+ T cell</p> <p>responses</p>		
--	--	--	---	--	--

IM, intramuscular; IN, intranasal; IP, intraperitoneal; OPA: opsonophagocytic antibodies; PO, per oral; SBA: serum bactericidal antibodies; SC, subcutaneous

Table 4. Completed observational trials assessing vaccine effectiveness of meningococcal vaccines on gonorrhoea infection.

Clinical trial design	Vaccine	Immunization schedule	Study population	Result	Reference
Retrospective ecological study of <i>N. gonorrhoeae</i> infection incidence in Cuba before and after introduction of VA-MENGOC-BC vaccine	VA-MENGOC-BC <i>N. meningitidis</i> vaccine (OMV from <i>N. meningitidis</i> serogroup B strain CU385 plus serogroup C capsular polysaccharide from <i>N. meningitidis</i>	Intramuscular; single-dose given in mass vaccination program; 2-dose schedule, 2 months apart given in routine schedule.	Cuban national health registry data comprising annual incidence rates of <i>N. gonorrhoeae</i> infection, meningococcal disease and syphilis between 1970-2018, including period of vaccine efficacy trial of VA-MENGOC-BC, mass vaccination campaign	Decreased incidence of <i>N. gonorrhoeae</i> infection compared to other STIs observed between 1990-1993 after mass vaccination campaign, and between 2010-2018 compatible with possible impact of routine infant vaccination program.	Sotolongo et al 2007 (150); Perez et al 2009 (60); Azze et al 2019 (58); Reyes Diaz et al 2021 (59)

	serogroup C strain C11)		in age 3 months to 20 years from 1989-1990; and incorporation in routine vaccination schedule from 1991- 2018.		
Retrospective ecological study of <i>N. gonorrhoeae</i> infection incidence in Norway before and after introduction of MenBvac vaccine.	MenBvac <i>N.</i> <i>meningitidis</i> serogroup B vaccine (OMV from <i>N. meningitidis</i>	Intramuscular 2- dose schedule.	Norwegian national health registry data comprising incidence rates of gonorrhoea from 1993 onwards and data of vaccine efficacy trial in 13-15	Incidence rate ratio (IRRs) analysis (defined as number of new diagnoses of <i>N.</i> <i>gonorrhoeae</i> infection per 100,000 population for vaccinated cohort	Whelan et al 2016 (62)

	serogroup B strain H44/76)		<p>year-old students enrolled in secondary schools between 1988-1992.</p> <p>93,611 (63%) of the 148,589 children resident in Norway and born during 1973-1976 received MenBvac.</p> <p>Total 2,601 cases of <i>N. gonorrhoeae</i> infection reported during 1993-2008.</p>	<p>compared to pre-vaccination and post-vaccination cohorts) demonstrated reduced crude IRR for women aged 20-24 years in the vaccinated cohort (IRR 0.58, 95% CI 0.42-0.8) and reduced adjusted IRR for men aged 20-24 years in the vaccinated cohort (0.68, 95% CI 0.51-0.93) and post-vaccination cohort</p>	
--	-----------------------------------	--	---	---	--

				(0.51, 95% CI 0.33-0.78) between 1993-2008.	
Retrospective case-control study of 15–30-year-old sexual health clinic patients eligible to receive MeNZB vaccine in New Zealand. Cases are defined as confirmed laboratory detection of <i>N. gonorrhoeae</i> only from clinical specimen; and controls defined as confirmed laboratory	MeNZB <i>N. meningitidis</i> vaccine (OMV from <i>N. meningitidis</i> serogroup B strain NZ98/254)	Intramuscular, 3-dose schedule. Infants: age 6 weeks, 3 months and 5 months Children >6 months: 3 doses, 6 weeks apart	Sexual health clinic patients aged 15-30 years and eligible to receive MeNZB (mass vaccination program 2004-2006 age 6 weeks to 20 years and available in schools and primary care until 2008) and diagnosed with <i>N. gonorrhoeae</i> and/or <i>C. trachomatis</i> infection between Jan	Vaccinated individuals significantly less likely to be cases than controls (511 (41%) vs 6,424 (51%); adjusted OR 0.69 (95% CI 0.61-0.79; p<0.0001). Estimated vaccine effectiveness of MeNZB against <i>N. gonorrhoeae</i> infection adjusted for ethnicity, deprivation,	Petoussis-Harris et al 2017 (34)

detection of <i>C. trachomatis</i> only from clinical specimen			1, 2005 and Dec 31, 2016. 14,730 cases and controls for analysis: 1,241 incidences of <i>N.</i> <i>gonorrhoeae</i> infection; 12,487 incidences of <i>C.</i> trachomatis infection; and 1002 incidences of co-infection.	geographical area and sex 31% (95% CI 21-39; p<0.0001)	
Retrospective ecological study of <i>N. gonorrhoeae</i> infection incidence in Sanguenay-Lac-Saint-Jean	4CMenB <i>N.</i> <i>meningitidis</i> vaccine (OMV from	Intramuscular, 2- dose schedule.	Public health registry data comprising cases of <i>N.</i> <i>gonorrhoeae</i> infection	Decrease in the number of <i>N. gonorrhoeae</i> infections and incidence rate of among the	Longtin et al 2017 (61)

region of Quebec, Canada before and after introduction of 4CMenB vaccination program.	<i>N. meningitidis</i> serogroup B strain NZ98/254 plus three recombinant protein antigens)		notified between January 2006 and June, 2017 and vaccination uptake data for mass vaccination campaign (mass vaccination campaign of individuals aged 6 months to 20 years conducted May to December, 2014). Overall vaccine coverage was 82%. A total of 231	vaccinated cohort (age 14-20 years) observed during post-vaccination period, whereas it increased in unvaccinated cohort (age 21 years and older). Estimated vaccine impact: <i>N. gonorrhoeae</i> infection risk reduction of 59% (95% CI 22-84; p=0.1).	
--	--	--	--	---	--

			gonorrhoea cases were reported among persons aged 14 years and older between January, 2006 and June, 2017.		
Retrospective cohort study of individuals born 1984-1999 eligible for MeNZB vaccination 2004-2008 in New Zealand with primary outcome hospitalization for primary diagnosis of <i>N. gonorrhoeae</i> infection.	MeNZB <i>N. meningitidis</i> vaccine (OMV from <i>N. meningitidis</i> serogroup B strain NZ98/254)	Intramuscular vaccine, 3-dose schedule. Infants: age 6 weeks, 3 months and 5 months Children >6 months: 3 doses, 6 weeks apart.	Individuals born 1984-1999 and residing in New Zealand from 2004 until 2015 (mass vaccination program 2004-2006 age 6 weeks to 20 years and available in schools and primary care until	Vaccinated individuals were significantly less likely to be hospitalized due to <i>N. gonorrhoeae</i> infection after adjusting for gender, ethnicity and deprivation (HR 0.76, 95% CI 0.58-0.99) with estimated vaccine	Paynter et al 2019 (57)

			<p>2008) with data available through national registry on vaccination status, sex, ethnicity and deprivation.</p> <p>935,496 individuals included in the analysis. Overall vaccination coverage 59.2%. 261 cases of hospitalization attributable to <i>N. gonorrhoeae</i>.</p>	<p>effectiveness of 24% (95% CI 1-42%).</p>	
--	--	--	--	---	--

<p>Retrospective case-control study of 16-23 year-old individuals with <i>N. gonorrhoeae</i> or <i>Chlamydia trachomatis</i> infection in New York City and Philadelphia.</p> <p>Cases defined as confirmed laboratory detection of <i>N. gonorrhoeae</i> (NAAT or culture) but not <i>C. trachomatis</i>; and controls defined as confirmed laboratory detection of <i>C.</i></p>	<p>4CMenB</p> <p><i>N. meningitidis</i> vaccine (OMV from <i>N. meningitidis</i> serogroup B strain NZ98/254)</p>	<p>Intramuscular vaccine, 2-dose schedule minimum 30 days and maximum 180 days apart (single dose categorized as partial vaccination).</p>	<p>Individuals aged 16-23 years old with <i>C. trachomatis</i> and/or <i>N. gonorrhoeae</i> reported to STI surveillance systems of the New York City Department of Health and Mental Hygiene and the Philadelphia Department of Public Health, with data matched to vaccine registry data system to obtain number and</p>	<p>Vaccinated individuals were significantly less likely to be diagnosed with <i>N. gonorrhoeae</i> infection. Complete vaccination series unadjusted prevalence ratio (UPR) 0.64, 95% CI 0.51-0.79; $p<0.0001$ in bivariate analyses and adjusted prevalence ratio (APR) 0.60, 95% CI 0.47-0.77; $p<0.0001$ in multivariate analyses.</p>	<p>Abara et al 2022 (53)</p>
--	---	--	--	--	------------------------------

<p><i>trachomatis</i> only (NAAT or culture) but not <i>N. gonorrhoeae</i></p>			<p>dates of MenB-4C vaccine doses between Jan 1, 2016 and Dec 31, 2018.</p> <p>109,737 individuals with 167,706 reported STIs for analysis.</p> <p>124,876 <i>C. trachomatis</i> infections, 18,099 <i>N. gonorrhoeae</i> infections and 24,731 were gonococcal and chlamydia co-infections.</p>	<p>Partial vaccination series UPR 0.83, 95% CI 0.72-0.96, $p=0.0204$ in bivariate analyses and APR 0.74, 95% CI 0.63-0.88; $O=0.0012$.</p> <p>Estimated vaccine effectiveness for complete vaccination series 40% (95% CI 23-53) and partial vaccination series 26% (95% CI 12-37%).</p>	
--	--	--	--	--	--

			3,058 STIs occurred after complete vaccination series, 6,519 after partial vaccination series and 155,330 among vaccine-naïve individuals.		
Retrospective case-control study of adolescents and young adults with gonorrhoea or chlamydia infection in the state of South Australia, Australia.	4CMenB <i>N. meningitidis</i> vaccine (OMV from <i>N. meningitidis</i> serogroup B strain NZ98/254)	Intramuscular vaccine, 2-dose schedule, 8 weeks apart.	Individuals born between Feb 1, 1998 and Feb 1, 2005 that had <i>N. gonorrhoea</i> or <i>C. trachomatis</i> disease notification between Feb 1, 2019 and Jan	Estimated vaccine effectiveness using <i>C. trachomatis</i> infection as controls was 32.6% (95% CI 10.6-49.1) for individuals who received at least one	Wang et al 2022 (54)

<p>Cases defined as all gonorrhoea-positive cases who did or did not have <i>C. trachomatis</i> co-infection at the time of first episode <i>N. gonorrhoeae</i> infection.</p> <p>Controls defined as <i>C. trachomatis</i>-positive infections only.</p>			<p>31, 2021 (in 2019, a 2-dose vaccination schedule for 15-17 year-old school-based immunization programme was implemented and between 2019-2020, a catch-up programme was available for those aged 17-20 years).</p> <p>53,356 individuals received at least 1 dose of 4CMenB and</p>	<p>dose; and 32.7% (95% CI 8.3-50.6) for people who received two doses compared to those who were unvaccinated.</p>	
---	--	--	--	---	--

			46,083 received 2 doses. 512 patients with total 575 episodes of gonorrhoea and 3140 patients with 3847 episodes of chlamydia included in analysis.		
Retrospective matched cohort study of 15-30 year-olds who received 4CMenB (plus/minus MenACWY) or MenACWY only in Southern California, United States.	4CMenB <i>N. meningitidis</i> vaccine (OMV from <i>N. meningitidis</i> serogroup B strain NZ98/254);	4CMenB intramuscular vaccine, 2-dose schedule. Minimum 1 dose included in analysis.	Individuals aged 15-30 years old in Kaiser Permanente Southern California health records noted to be vaccinated with 4CMenB, matched in a	Incident gonorrhoea rates 2.0 (95% CI 1.3-2.8) per 1000-person years for 4CMenB recipients; 5.2 (95% CI 4.6-5.8) per 1000-person years for	Bruxvoort et al 2022 (55)

<p>The exposed group comprising recipients of 4CMenB were matched in a ratio of 1:4 to the unexposed group comprising recipients of MenACWY only by age, sex and year of index vaccination with study outcome positive gonorrhoea NAAT or culture or chlamydia NAAT (negative control).</p>	<p>MenACWY <i>N. meningitidis</i> vaccine (serogroup A, C, W, Y polysaccharide conjugate vaccine)</p>	<p>MenACWY intramuscular vaccine, 2-dose schedule. Minimum 1 dose included in analysis</p>	<p>ratio of 1:4 to recipients of MenACWY only by age, sex and year of index vaccination between Jan 1, 2016 and Dec 12, 2019.</p> <p>6,641 4CMenB recipients; matched to 26,471 MenACWY only recipients.</p>	<p>MenACWY only recipients. Incident chlamydia rates 12.4 (95% CI 10.7-14.4) per 1000-person years for 4CMenB recipients; 15.2 (95% CI 14.2-16.2) per 1000-person years for MenACWY only recipients.</p> <p>Hazard ratio (HR) for incident gonorrhoea in 4CMenB recipients compared to MenACWY</p>	
---	---	--	--	--	--

				only recipients 0.54 (95% CI 0.34-0.86) in multivariable analyses.	
Retrospective case-control study of ≥18 year old MSM living with HIV with gonorrhoea or syphilis, chlamydia or anal HPV in Milan, Italy. Cases defined as all gonorrhoea-positive cases by NAAT or culture; Controls were defined as	4CMenB <i>N. meningitidis</i> vaccine (OMV from <i>N. meningitidis</i> serogroup B strain NZ98/254)	Intramuscular vaccine, 2-dose schedule, 8 weeks apart.	≥18 year old MSM living with HIV diagnosed with gonorrhoea or syphilis, chlamydia or anal HPV included in the database of the Infectious Diseases Unit at the San Raffaele Scientific Institute, Milan, Italy	Estimated vaccine effectiveness was 42% (95% CI 6-64, p=0.027) and remained significant at 44% (95% CI 9-65, p=0.020) after adjustment in multivariable analysis.	Raccagni et al 2023 (56)

chlamydia positive by NAAT, syphilis positive by serology and HPV positive by anal NAAT to 28 HPV genotypes or following a diagnosis of condylomatosis.			between July, 2016 and February 2021. 349/1051 (33%) received 4CMenB vaccination. 103 cases and 948 controls analysed. Median follow up 3.8 years (2.1-4.3)		
---	--	--	---	--	--

HIV, human immunodeficiency virus; HPV, human papillomavirus; MSM, men who have sex with men; NAAT, nucleic acid amplification test;

OMV, outer membrane vesicle

Table 5. Randomised trials of vaccine effectiveness of meningococcal vaccines on gonorrhoea infection currently in design, recruitment or pre-publication phases.

Clinical trial design	Vaccine and immunization schedule	Study population	Recruitment strategy	Primary outcome	Reference
Phase III, double-blinded, randomised,	4CMenB; Intramuscular administration, 2	18-50 year-old men (cis and trans), transexual	730 participants enrolled and randomised	1. To measure whether 4CMenB changes the incidence of first	Seib et al (https://clinicaltrials.gov/study/NCT04415424)

<p>placebo-controlled, multi-centred trial evaluating the efficacy of 4CMenB in prevention of gonorrhoea infection (GoGoVax).</p>	<p>doses, 3 months apart OR placebo.</p>	<p>women and non-binary people who have sex with men; either HIV-negative and on PrEP or HIV-positive with HIV viral load <200 copies/ml and CD4</p>	<p>1:1. Recruitment for 12 months. After vaccination, all participants followed-up 3-monthly for 24 months.</p>	<p>episode <i>N. gonorrhoeae</i> infection.</p> <p>2. To compare overall incidence of all episodes of <i>N. gonorrhoeae</i> infection diagnosed during the study period between vaccine and placebo arms.</p>	
---	--	---	---	---	--

		count >350 cells/cmm.			
Phase II, randomised, observer- blind, placebo- controlled, multi-centre trial evaluating the efficacy of 4CMenB in prevention of urogenital/and	4CMenB; Intramuscular administration, 2 doses, 2 months apart OR placebo.	18-50 year- old healthy men and women	Approximately 2,200 participants and randomised 1:1. After vaccination, participants followed 3- monthly for 16 months.	To measure efficacy of 4CMenB in prevention of urogenital and/or anorectal infection.	Marazzo et al (https://clinicaltrials.gov/study/NCT04350138)

or anorectal gonorrhoea infection					
Single-site double- randomised controlled trial evaluating the efficacy of 4CMenB in prevention of gonorrhoea using a controlled	Initial vaccination phase: 2 doses of intramuscular 4CMenB OR quadrivalent influenza and tetanus/diphtheria vaccine; Post-challenge vaccination: crossover arm	18-35 year- old men without a history of 4CMenB vaccination	Approximately 120-140 participants enrolled and randomized 1:1.	Infectivity of <i>N.</i> <i>gonorrhoeae</i> inoculum defined as the proportion of participants with microbiological evidence of <i>N.</i> <i>gonorrhoeae</i> by culture or NAAT in urine or urethral swab culture on the	Duncan et al (https://clinicaltrials.gov/study/NCT05294588)

human experimental infection with <i>N. gonorrhoeae</i> strain FA1090	with receipt of either quadrivalent influenza and tetanus/diphtheria vaccines or 2 doses of 4Cmen B.			post-inoculation antibiotic treatment day in each study group.	
Single-site, parallel, double-blind, randomised, placebo- controlled trial evaluating the efficacy of	4CMenB; intramuscular administration, 2 doses, 1 month apart OR placebo	MSM aged 18 or above at risk of gonorrhoea infection (condomless sex with >1 man in last 6	150 participants.	Incidence of <i>N. gonorrhoeae</i> infection between control and intervention groups	Kwan T et al (https://clinicaltrials.gov/study/NCT05766904)

4CMenB in prevention of gonorrhoea infection		moths, history of STI, inclination to have condomless sex and other HIV PrEP-eligible criteria)			
Randomised, open-label, single-site trial evaluating the efficacy of	4CMenB; intramuscular administration, 2 doses, 3 months apart	18-50 year old gay and bisexual men that are currently	130 participants enrolled and randomised 1:1. Followed	Number of <i>N. gonorrhoeae</i> infections in participants over 2	Thng et al (147)

4CMenB in prevention of gonorrhoea infection (MenGo)		taking HIV PrEP or have been diagnosed with gonorrhoea in the past 3 months	3-monthly for 24 months.	years measured by NAAT	
--	--	---	--------------------------	------------------------	--

Footnotes: HIV, human immunodeficiency virus; MSM, men who have sex with men; NAAT, nucleic acid amplification test; PrEP, pre-exposure prophylaxis

