

1 **Ultrasonic Encapsulation – A Review**

2
3 Thomas S. H. Leong^{1,2}, Gregory J. O. Martin², Muthupandian Ashokkumar^{1*}

4
5 ¹ School of Chemistry, The University of Melbourne, Parkville, Victoria 3010, Australia

6 ² Department of Chemical & Biomolecular Engineering, The University of Melbourne,
7 Parkville, Victoria 3010, Australia

8 **Highlights**

- 9 • Ultrasonic preparation of encapsulated functional materials is reviewed.
- 10 • Mechanisms responsible for expression of functional properties are described.
- 11 • Promising applications for ultrasonically synthesized materials are identified.

12 Keywords: ultrasound, encapsulation, functional foods, bioproducts, emulsions,
13 sonoprocessing, responsive polymers

14
15 *corresponding author

16 17 18 **Abstract**

19 Encapsulation of materials in particles dispersed in water has many applications in nutritional
20 foods, imaging, energy production and therapeutic/diagnostic medicine. Ultrasonic technology
21 has been proven effective at creating encapsulating particles and droplets with specific physical
22 and functional properties. Examples include highly stable emulsions, functional polymeric
23 particles with environmental sensitivity, and microspheres for encapsulating drugs for targeted
24 delivery. This article provides an overview of the primary mechanisms arising from ultrasonics
25 responsible for the formation of these materials, highlighting examples that show promise
26 particularly in the development of food and bioproducts.

1. Introduction

Ultrasonics is a versatile technology with proven effectiveness to create a range of catalytic and functional materials that have applications across a multitude of fields including food [1], imaging [2], energy production [3] and therapeutic/diagnostic medicine [4]. The primary mechanism responsible for the creation of these materials is known as acoustic cavitation, which is the formation and collapse of bubbles influenced by ultrasound [5]. Ultrasound's versatility is owed in part to its broad active frequency region that can be tuned and applied specifically to control the intensity and number of cavitation events. These can be used to control aspects of materials such as particle size, surface roughness and structure.

Ultrasound can be used to promote the internalization of materials through a process known as encapsulation. The motivation for encapsulation is to protect, prolong or stabilize the internalised material from environmental deterioration and enables pharmaceuticals and/or nutrients to be delivered with enhanced efficacy in biological systems. These delivery systems take a number of different forms. A simple example found in foods is an emulsion [6]. The dispersion of an oil within water enables effective loading of oil soluble nutrients into aqueous food media, and is a useful strategy for preparing functional food products. Another example is the use of environmentally sensitive polymers to create core-shell structures that can be used to entrap materials such as drugs [7]. These polymer delivery agents respond to changes in pH, temperature or other external stimuli such that they release entrapped drug material only under specific conditions, thereby prolonging drug efficacy. These polymers can be synthetic or natural, such as proteins. Microspheres and microcapsules made from proteins have the advantage of being bio-compatible and bio-degradable, and have been extensively studied for pharmaceutical applications [8].

Whilst there are a number of reviews [4, 6, 7] covering the formation of different types of encapsulated materials, there has yet to be a review that brings together these different systems with details on how to effectively create them using ultrasonics. This review will provide a guide towards the application of ultrasound to promote encapsulation of materials, focusing on examples of relevance to the food, biomaterial and pharmaceutical industries.

2. Theory of applied ultrasound

This section provides an overview of the principles of ultrasound. A focus is made on the key physical and chemical effects of ultrasound in aqueous systems to provide background for the

subsequent discussion on the application of ultrasound-promoted encapsulation in aqueous systems.

2.1 Characteristics of ultrasound

Ultrasound is generally defined as sound at frequencies above 16 kHz. It is (generally) not audible when transmitted through the air. When sustained through a liquid medium (e.g. water), the ultrasonic pressure oscillations may cause in-phase expansion and contraction of the dissolved gas bubbles i.e. the bubble expands during the negative pressure cycle and contracts during the positive pressure cycle. This bubble oscillation is accompanied by diffusion of gas/vapour in and out of the bubble during the expansion and contraction respectively. The diffusion of gas in and out of the bubble is not equal [9] and under certain conditions, i.e. oscillation driven above a certain threshold pressure, the diffusion process can result in net accumulation of mass within the bubble over time. This results in net bubble growth and is known as rectified diffusion, a process unique to bubbles oscillating within a sound field. In a field containing multiple bubbles, the interaction of bubbles by collisions combining to form a larger bubble can also result in what is known as coalescence, and is another source of net bubble growth in an acoustic sound field [10].

Both rectified diffusion and coalescence cause bubble growth. Bubbles within a sound field will grow in size until they reach what is known as the bubble resonance size range, at which point they collapse. This formation, growth and collapse of a bubble due to the influence of ultrasound, is known as acoustic cavitation [5].

Of interest to material synthesis, the collapsing bubbles produce localised regions of extreme temperature and intense physical shearing. Bubbles driven at low ultrasonic frequency (~ 20-100 kHz), may collapse extremely violently, releasing sufficient energy to produce temperatures up to 10,000 K within the bubble core and pressures of several hundred atmospheres within a few hundred micron of the bubble collapse point [11]. This can lead to the formation of highly reactive radicals which can be used to promote chemical reactions. In water for example, hydrogen and hydroxyl radicals can be formed due to the splitting of the water molecule by pyrolysis. Both the physical shear and radical formation can be beneficially exploited to create materials with a range of desired functionality.

87 The ultrasound frequency regime ranges from 16 kHz to 500 MHz, although the frequency
 88 range most suitable for processing fluids is typically between 16-3000 kHz. When ultrasound
 89 is applied to fluids the cavitation effects are highly dependent on the frequency. The intensity
 90 of bubble collapse (i.e. amount of energy released) and the maximum bubble size prior to
 91 collapse (resonance size) are correlated and approximately inversely proportional to the applied
 92 frequency [12] (see Figure 1).

93 A simple relationship that can relate the resonance size of the bubble with the frequency is
 94 given by:

$$95 \qquad \qquad \qquad F \times R \approx 3 \qquad \qquad \qquad (1)$$

96 where F is the frequency in Hz and R is the bubble radius in m. Note that this equation gives
 97 only a very approximate theoretical resonance size and that there are other factors which may
 98 control the resonance size of the bubble [5, 13].

99 A more accurate version of (1) is the linear resonance radius which can be calculated using
 100 the following equation [13]:

101

$$102 \qquad \qquad \qquad R_r = \sqrt{\frac{3\gamma p_\infty}{\rho_L \omega^2}} \qquad \qquad \qquad (2)$$

103 where γ is the specific heat ratio of the gas inside the bubble, p_∞ is the ambient liquid
 104 pressure, ρ_L is the liquid density and ω is the angular frequency of ultrasound (all in SI
 105 units). In practice, the size for an active bubble is usually smaller than this radius due to the
 106 nonlinear nature of the bubble pulsation [14].

107 Ultrasound can be categorized into several different regions along the frequency spectrum. The
 108 *power ultrasound* region [12] spans the low frequency range between 16 – 100 kHz. It is
 109 characterized by large bubble resonance sizes followed by intense bubble collapse, often
 110 resulting in extremely strong physical effects including localized shear and high temperatures.
 111 This category of ultrasound delivers high energy density in the order of 10-1000 W/cm². Power
 112 ultrasound is often selected for material processing and to some extent in synthesis, owing to
 113 its strong physical shear and intense local temperature effects. Examples of processes in this

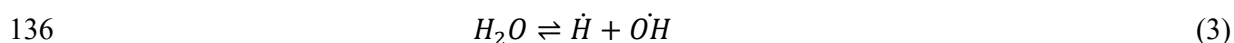
114 region include emulsification [15], homogenization [16], cell disruption [17] and
115 polymerization [18].

116 The region between 100 - 1000 kHz is usually labelled *intermediate ultrasound*. This region
117 results in only moderately intense bubble collapse, but importantly produces the most
118 'sonochemically active' bubble population that results in highly efficient radical production.
119 Koda et al. [19] and Mason et al. [20] have shown that peak radical production occurs
120 somewhere between 400 - 800 kHz, although it also depends on the power applied and the
121 physical and chemical properties of the fluid system. This intermediate ultrasound region is
122 selected when chemical modification is the primary goal.

123 Above 1000 kHz, the physical effects of bubble collapse become relatively benign due to small
124 bubble resonance size prior to collapse and a reduced proportion of bubbles undergoing
125 cavitation due to an increased cavitation threshold. The cavitation threshold refers to the
126 pressure (or size) above which bubble nucleation, a necessary precursor to cavitation, occurs.
127 However, despite a reduction in cavitation, radical formation is still possible at frequencies
128 around 1000 kHz [21] provided that sufficient energy intensity is employed. This regime,
129 typically labelled the *diagnostic* or *megasonic* region, is used if only gentle physical effects are
130 desired. It is particularly suitable for applications such as selective particle separation [22],
131 where the aim is to also preserve the natural integrity of the separated product.

132 **2.2 Radical formation and sonochemistry**

133 The concentrated energy released during bubble collapse can split solvent/solute molecules
134 that have diffused into the bubbles, to form radical species. For water, the following reaction
135 may take place [23]:



137 That is, water molecules can be split into highly reactive hydrogen and hydroxyl radicals.
138 Direct splitting of water as described above requires high temperatures resultant from high
139 intensity bubble collapse typically seen only in the power ultrasound region. Alternate reaction
140 pathways in water have been described by Yasui et al. [24] that allow for hydroxyl radical
141 production even at relatively low bubble temperatures that are typical of intermediate
142 ultrasound.

These radical species can be used to induce a whole range of redox reactions. As such, ultrasound can be used in organic synthesis reactions in which radicals are used to initiate and increase the reaction rates. Ultrasound has also been successfully used to increase yield by inducing modified reaction pathways that favor the formation of specific reaction products [4].

The radicals formed through cavitation can be used to initiate and accelerate the rates of free-radical polymerization and copolymerization [18], meaning that in some cases the use of ultrasound can obviate the need for a chemical initiator. Methyl methacrylate for example, can be polymerized by ultrasound without the use of an initiator [25], creating polymers with a molecular weight of up to 400,000 Da. Ultrasound-induced polymerization can however be complicated by the fact that the resulting polymers can also be simultaneously broken down by the intense shear forces resulting from the collapsing bubbles. These fragmented polymers may subsequently react to form side-products that may be quite different to conventional polymerization in the absence of acoustic cavitation.

Proteins are biological polymers that can be cross-linked to form larger networks and structures by ultrasound. These networks can be used to coat bubbles, droplets or other templates, forming rigid spheres that can be used for encapsulating materials for applications such as drug delivery [8]. The formation of protein microspheres by application of ultrasound results from a combination of shear-induced emulsification and radical formation [26, 27]. The high shear and temperature can partially unfold the proteins [28] which can then accumulate at the air or organic phase boundary and undergo cross-linking [27]. The protein cross-linking can be reversible, for instance through hydrophobic interactions and hydrogen bonding, or irreversible if covalent links are produced, for example disulphide bonds. In the latter case, this can potentially be facilitated by free radicals generated through ultrasonic cavitation [29].

Ultrasound can also be used to lower the temperature or pressure of some reactions, or reduce the requirement for solvents, which are expensive and often toxic. As such, ultrasonic synthesis techniques are often considered as ‘green chemistry’ alternatives for many applications [25].

2.3 Physical effects of ultrasound

Physical modification of materials by ultrasound can arise from the shear forces generated during bubble collapse that are associated with pressure shockwaves, liquid microjets and acoustic streaming. Acoustic streaming is the propagation of disturbances in the fluid caused

by the ultrasonically induced oscillation of gas bubbles. This results in localized shear forces in the immediate vicinity of the bubbles. Liquid microjets result from the asymmetric and extremely rapid collapse of bubbles during cavitation, leading to unidirectional expulsion of high velocity jets into the surrounding fluid. Naude and Ellis [30] first hypothesized that the observed pitting of solid surfaces and particle size reduction of colloids on exposure to ultrasound was in fact due to the formation of microjets during asymmetric bubble collapse. It has since been shown that microjets with velocities in the order of 100 m/s can be formed [31], and that these can create pitting and erosion of surfaces [25]. Microjetting is also the primary cause of ultrasound-induced bulk mixing. Symmetric bubble collapse generates shockwaves that propagate radially outwards from the collapse point into the surrounding fluid. These shockwaves can be used to increase the rates of mass transfer across interfacial boundaries, enhancing the efficiency of multi-phase reactions [25]. All of these physical effects are commonly utilized in laboratory ultrasonic baths to facilitate cleaning of surfaces and dissolution of solids.

Although collapsing bubbles may reach temperatures of many thousands of degrees Celsius, these extreme temperatures are confined to small areas at the core of the collapsing bubble and near the bubble surface [32]. In the bulk solution, the increase in temperature resultant from a single bubble collapse is small. Nevertheless, the dissipation of heat from many cavitation bubbles, vibrating transducer surfaces, shockwave propagation and acoustic streaming can all contribute to incremental increases to the surrounding bulk temperature. If required, temperature control (e.g. a cooling jacketed reactor) can be used to prevent undesired temperature effects, e.g. denaturation of proteins in foods.

The physical forces resultant from cavitation can increase particle interactions in suspensions [25]. Solid powders suspended in fluids, may experience an increase to their momentum in the vicinity of a cavitation bubble, which can cause them to collide together with greater force than under quiescent conditions. Inorganic solids can be fractured and disrupted upon collision, leading to a reduction to their average particle size [25]. The minimum size achievable is dependent on characteristics of the solid, solvent and cavitation intensity. The lower limit is reached when the momentum of the particles become too small to create further impacts to cause particle fragmentation.

Ultrasound can be used to form encapsulating particles over a broad range of sizes, from around 100 nm to 20 μ m in diameter. The particle size can be controlled to a large extent by selecting

appropriate sonication conditions such as power intensity, frequency and reactor configuration. The duration of processing [33], the type of reaction vessel [15] and sonifier used [34] are also variables that control the size of the particles that are formed. Importantly, in addition to being able to target a desired average particle size, ultrasound is able to produce particles with a narrow size distribution. For example, it has been shown that the use of a flow-through horn system could generate lysozyme coated nanospheres of very narrow size distribution ranging between 550-650 nm, compared with a larger 3 mm ultrasound horn that resulted in formation of particles with a broader range between 850-1200 nm [34]. Zhou et al. [35] also used high frequency ultrasound as a post-sonication technique to further narrow down the size distribution of ultrasonically-generated microspheres. By using 213 kHz ultrasound, lysozyme microspheres with a distribution of initially 0.5-4 μm were narrowed to 0.5-2 μm due to selective breakage of the larger microspheres by the ultrasound.

3. Applications of ultrasonic encapsulation

Ultrasonics can be used to promote specific functionality in different materials. For example, in foods comprising emulsions, the shelf-stability and physical appearance are dependent on the droplet size of the dispersed phase. Ultrasonics can be used to disperse different organic/oil phases into various aqueous phases in a controlled manner, to create emulsified products that are very shelf-stable and attractive in appearance [6]. Ultrasonics can also be used to promote the formation of polymer systems that are responsive to specific environmental conditions, such as pH and temperature. These polymer systems are useful for controlling drug release in biological systems. A combination of emulsification and polymerization can be promoted by ultrasound to form protein cross linkages, resulting in the formation of protein microspheres. These entities can be made biocompatible and biodegradable to enable their use as drug-delivery vehicles. This section will describe the effect of ultrasound on important functional properties of food emulsions (3.1), polymer particles for controlled drug release (3.2), and protein microspheres (3.3).

3.1 Functional food emulsions

Droplet size and polydispersity are key attributes that govern the functionality and stability of emulsions. The intense shear forces generated during ultrasonic cavitation can be used to create emulsions with very small and relatively uniformly sized droplets [36]. Two mechanisms are responsible for the emulsification effect of ultrasound. First, the application of the sound field

produces interfacial waves, which become unstable resulting in the dispersion of the oil phase into the continuous water phase as mid- to large-sized droplets. Secondly, the physical effects resultant from cavitation break up these initially formed droplets of dispersed oil into droplets of sub-micron size [25].

Ultrasonics is particularly useful for the production of, for example, food emulsions. In food emulsions, the size of the emulsified droplets influences its visual appearance, mouth-feel and shelf-life stability among other things [6]. Whereas large sized emulsion droplets are characterized by a ‘milky’ opaque appearance, emulsions with emulsion droplet size (EDS) smaller than ~ 100 nm, can appear translucent and almost clear [15] due to the reduction in light scattering by the smaller droplets.

While emulsions are inherently thermodynamically unstable, when the droplets are smaller than ~ 100 nm they become kinetically stable [37]. At these sizes, the Brownian motion of the droplets overcomes the natural buoyancy force of the droplets to rise and cream. The instability is dependent instead on colloidal forces such as Ostwald Ripening [38] and droplet-droplet collisions that leads to coalescence and eventual phase separation. These are usually very slow processes, such that nano-sized emulsions are shelf-stable for many months [39].

The formation of nano-sized droplets requires the application of strong shear forces to break apart the liquid droplets. This is typically achieved using low frequency ultrasound in the *power ultrasound* region (20-100 kHz) delivered at high energy intensity > 10 W/cm². The emulsified droplets then need to be stabilized by a surfactant in the system, to prevent spontaneous phase separation by coalescence [15]. Midsonic and megasonic frequencies > 400 kHz are usually ineffective at forming emulsions, due to insufficient physical shear forces arising from the collapse of smaller resonance size bubbles at these frequencies.

Higher frequency ultrasound (midsonic to megasonic) has however been reported useful at forming nano-size emulsion droplets when applied following low frequency ultrasound through a process known as *tandem acoustic emulsification* [39, 40]. Oleic acid/water nano-emulsions were prepared by Kamogawa et al. [40] using this technique, while Nakabayashi et al. [39] also reported the production of transparent emulsions of ethylenedioxythiophene (EDOT) monomer formed by sequential emulsification at 20 kHz, 1.6 MHz and 2.4 MHz ultrasound. The nano-emulsions formed by Nakabayashi et al. were stable even in the absence of additional surfactant for 1 to 2 years. These nanoemulsions are not only stable and attractive

in appearance, but can be used to conduct direct electropolymerization (see Section 3.2) in the absence of additional surfactant [41].

It was proposed that the small droplets achieved in tandem acoustic emulsification upon application of higher frequency ultrasound was not due to destructive shear forces such as microjets and shockwaves prominent at low frequency ultrasound. Instead, it is due to the enhanced acceleration of solvent and the emulsion droplets caused by acoustic radiation forces and acoustic streaming [40] such that they collide together and break apart into smaller droplets. These acceleration forces become stronger with increased frequency, and the sequence in which the different frequencies of ultrasound are applied is noted to be important, with reversal of the order (i.e. high frequency followed by low frequency) resulting in ineffective emulsification.

Oil-in-water and water-in-oil emulsions can both be produced successfully using the tandem emulsification technique. Although the application of tandem acoustic emulsification has yet to be reported for food applications, it would be an attractive avenue for creating nano-sized surfactant-free emulsions.

For foods, the EDS plays a significant role in the sensory characteristics [42]. In general, emulsions containing smaller droplets have a higher viscosity [42] which are purported to provide improved sensory properties such as ‘creamier’ mouthfeel [6] in a range of products such as cheese [43] and creams [44].

Emulsions can be used to load hydrophobic or amphiphilic materials with biological functionality or nutritional benefit into an aqueous fluid. In the case of amphiphilic materials, it may be desirable to maximize the surface area of the droplets by reducing the emulsion droplet size. Smaller droplets are also better at retaining a larger amount of volatile material within the oil phase of an emulsion during spray drying for the production of encapsulated microparticles [45]. This is because the smaller emulsion droplets are less likely to be broken apart by the atomizer within the spray dryer [45]. Spray dried encapsulating microparticles can be used to create products that are able to mitigate the release of undesirable odors or smells e.g. fish oil powders.

Another way to encapsulate materials in emulsions is to create what is known as a double emulsion. Double emulsions are emulsions entrapped within emulsions. Their capacity to encapsulate aqueous components within oil droplets makes them promising delivery vehicles

for bioactives, for flavour masking and for fat reduction in foods [46]. The entrapped inner phase is protected from degradation by environmental factors in the external phase, and release of inner material can be delayed until it enters the digestive system, thereby masking potentially undesirable flavours. Fat reduction can be achieved without compromising the sensory properties of the fat phase by displacing fat without reducing the apparent volume fraction of the fat droplets.

There is significant commercial interest, with a large number of examples having been developed for the production of flavour-enhanced and reduced-fat salad dressings [47], and also reduced-fat cheese [48-50]. Instability is a potential issue for using double emulsions in food applications. Rapid phase separation can arise due to the relatively large droplets (typically greater than 20 μm) [51] that are formed at the low shear rates which are required to avoid release of the entrapped material. This issue may be resolved to an extent by use of ultrasonication as reported by Tang et al. [52, 53]. Ultrasonication was successfully used to form double emulsions of sub-micron size range for the purpose of aspirin encapsulation, achieving both high stability (1 month prolonged storage) and entrapment yield (up to 99% encapsulation) [53].

The use of ultrasonics has been compared with most conventional and state of the art emulsification techniques. Some of the more common methods applied in industrial emulsification are rotor-stator systems [54] and high pressure homogenization [55]. In addition to conventional high pressure homogenization, a modified technology known as the MicrofluidizerTM involves impinging two pressurized streams against each other. The MicrofluidizerTM (MF) has been shown to be highly effective at nano-emulsion preparation [56-58]. MF has relatively high energy efficiency for producing emulsions with very small and narrowly distributed EDS [57] and is commonly used in the pharmaceutical industry to make nano-emulsions. Madhi Jafari et al. [57] have compared emulsion preparation using US at matched specific energies with MF, and found comparable performance. It was found that when using matched 20 kJ/kg energy input, particle size reduction by MF achieved mean volume-weighted particle size of 0.83 μm compared with 1.02 μm for ultrasonication at 20 kHz.

Emulsification by ultrasonication and MF occurs via common causal mechanisms including cavitation and shear [58]. Although MF has been noted to be superior in size reduction and generating emulsions with more narrow size distributions, ultrasonication is deemed to be

significantly easier to operate, clean and maintain [57]. With extended duration of processing, ultrasonication has been shown to be able to achieve comparably small emulsion droplets to MF [58]. Leong et al. [15]. demonstrated the capability of ultrasound to produce emulsions with comparable particle size to microfluidization, provided that the energy density and surfactant system was optimized.

Typically, the formation of emulsions requires a large amount of surfactant to cover the newly formed surfaces and hence stabilize the dispersed droplets. The tandem-emulsification technique reported by Nakabayashi et al. [39] and Kamogawa et al. [40] is exceptional in that ultrasound can produce nano-sized emulsions in the absence of surfactant, although the technique is yet to be proven for a wide range of oils. Recently, Shanmugam et al. [59] have shown that ultrasonic emulsification can be used to create stable food-based emulsions of flax seed oil directly in skim-milk without the requirement for additional surfactants. The native milk proteins are partially denatured (less than 1%) by the ultrasound and allowing them to effectively coat the formed oil droplets, stabilizing the emulsion for at least 7 days. Emulsification could not be achieved in the absence of ultrasound even when using matched applied energies in a rotor-stator system, suggesting the importance of acoustic cavitation to the stabilization process.

3.2 *Formation of polymeric particles for controlled drug release*

In the treatment of certain diseases, drugs must be delivered at rates corresponding to the physiological needs of the patient. In conventional drug delivery, the concentration of the drug within the patient's blood stream rises, peaks then declines. Each drug has a different concentration above which it becomes toxic and below which it is rendered ineffective. Controlled drug release is desirable in treating certain illnesses, as it enables maintenance of a drug within a desired therapeutic range with a single dose that is responsive to the needs of the patient.

Polymer-based materials that are sensitive to environmental factors such as temperature, pH and ionic strength, have potential to be used as responsive drug delivery vehicles [7]. If the structure of the polymer can be externally regulated (e.g. by magnetic, ultrasonic, thermal and electric stimulation) or self-regulated (i.e. by changing environmental conditions), it is possible to release the entrapped drug in a controlled manner. Ultrasonics has been investigated as a tool to assist in synthesizing such polymers with a range of functionality. For a more detailed

review of ultrasonically enhanced synthesis of polymers, readers are invited to read the review by Price [18].

As described in Section 2, acoustic cavitation leads to both chemical and physical phenomena that can be controlled to create polymers with improved rates of reaction and more defined characteristics such as molecular weight. Radical polymerization is one of the most studied [18] sonochemically-enhanced polymerization processes. The radicals formed during acoustic cavitation can be used to initiate the polymerization process in place of conventional initiators [60, 61]. A particular system in which this has been successful is in vinyl monomers such as methyl methacrylate [62]. Another notable advantage of generating radicals using ultrasound is the ability to perform the reaction at reduced temperatures (i.e. between -10 to 60 °C compared to between 50 to 100 °C for more conventional radical polymerization reactions of PMMA).

The physical effects of ultrasound can be used to control various properties of the resulting polymer. The intensity of the ultrasound applied, which influences the strength and number of acoustic cavitation bubbles, is one variable that can be modulated to control the yield of polymer produced as well as the final molecular weight of the resulting polymer [63]. A larger number of collapsing bubbles creates more radicals, which can increase the frequency of polymer initiation events. Simultaneously, the shearing forces resultant from the collapse of bubbles can break apart some of the long polymer chains that are formed, effectively reducing the molecular weight of the final polymer. The intensity of collapse, and the duration over which sonication is applied, can control the molecular weight of the polymers [63]. The ability to influence the polymerization process by manipulating ultrasound variables provides scope to transform conventional polymerization processes to form new materials. It should be noted though that sonication has little to no effect on the actual propagation reaction of the polymer formation. The main effects are largely confined to the initiation process and subsequent chain breakage of the formed polymer chains.

Ultrasound can promote emulsion polymerization to form latex particles of approximately equal size to the emulsion droplets [64]. The latex particles formed by ultrasound are typically smaller than those formed by conventional emulsion polymerization, resulting in an increased surface area [65]. Further, the use of ultrasound removes the need for chemical initiators or co-stabilizers, reduces the required reaction temperature, increases the rate of polymerization and results in higher monomer conversion and potentially higher molecular weights. As a number

of these advantages reduce chemical and energy consumption, ultrasonic emulsion polymerisation can be considered a 'green' alternative to conventional polymerization reactions. A proposed mechanism for ultrasonically-promoted emulsion polymerization is presented in Figure 2.

Ultrasonic emulsion polymerization has also been used to create latex coated magnetic nanoparticles using a simple, one-step method [66]. The particles exhibited colloidal stability for up to 12 months with no observed deterioration, and strong magnetic properties. The suspensions behaved as conventional magnetic fluids in their response to a magnetic field.

Temperature responsive polymers of poly(N-isopropylacrylamide) and poly(N-vinylcaprolactam) have been prepared using ultrasound, and their swelling behavior in the presence of different concentrations of surfactant (SDS) studied [67]. The potential for these types of polymers to be used as drug delivery vehicles was demonstrated by their ability to entrap rhodamine B dye. The structure of these polymers is temperature dependent. At 20 °C, the polymer chain is an expanded coil that is soluble in water resulting in clear aqueous solutions. When heated to 32 °C, the chains collapse to a globular structure which decreases the solvation properties causing the polymer solution to become turbid. The release kinetics of the entrapped rhodamine B was consequently found to be dependent on the temperature, with higher release rates occurring at 40 °C compared with 20 °C [67]. The release of dye was found to follow Fickian diffusion kinetics, with the diffusion coefficients being 4×10^{-12} and $3.6 \times 10^{-11} \text{ m}^2/\text{s}$ at 20 °C and 40 °C respectively i.e. an order of magnitude increase in release rate. There was an apparent limit to the maximum amount of dye release (approximately 62 %), which was attributed to the concentration gradient of the dye within the polymer and bulk solution approaching zero.

3.3 Formation of protein-coated microspheres

Ultrasonics have been used to prepare protein microspheres, which have a wide range of potential biomedical applications including acting as echo contrast agents for sonography and magnetic resonance imaging, and as vehicles for drug delivery [4]. These microspheres (typically several μm in diameter) consist of a protein shell surrounding a core which can be either a gas or a liquid. The solid shell is a barrier to permeation between the interior phase and the aqueous exterior, conferring long term storage stability to the protein microspheres.

Protein microspheres are formed by a combination of two acoustic phenomena: emulsification and cavitation. In the emulsification stage bubbles or liquid droplets are created, which acts as a 'template' for the protein shell to form around. The radicals produced by acoustic cavitation lead to the formation of superoxide species which promote the formation of intermolecular disulphide crosslinks between the proteins covering the interface. It has been shown that emulsification alone (via vortex mixing) was not sufficient to produce long-lived microbubbles, indicating that chemical cross-linking arising from ultrasonic cavitation was required to produce stable protein microbubbles [26].

One of the first commercially available protein microspheres were albumin-coated microbubbles, marketed under the name Albunex® and Optison™ [8]. These microspheres have been used primarily as contrast agents for ultrasound imaging, with the air-filled core providing enhanced signal response.

Protein microspheres can be used as drug delivery vehicles with therapeutic agents either loaded on the surface of air-filled protein microspheres, or if liquid filled, entrapped within the liquid phase of the protein microsphere. Of importance for drug delivery, they are inherently biodegradable and likely to be more biocompatible than microsphere made from synthetic polymers. They can also be functionalized with ligands (e.g. antibodies, peptides or vitamins) to target specific entities within the body.

To achieve further functionality, proteins may be used that infer useful biological properties to the formed microspheres. Cavalieri et al. [27] first reported the formation of lysozyme protein microspheres which were stable for several months using a sonochemical approach. Lysozyme, derived from hen egg white, has natural anti-microbial properties and the microspheres formed from lysozyme were found to retain some of the enzymatic functionality and anti-microbial activity of the native protein. This work by Cavalieri et al. [27] confirmed the need to release free thiol groups via partial protein denaturation in order to initiate crosslinking required to stabilize protein microspheres (Figure 3). Alternatively, the microbubbles can be used as a carrier for antibiotics. Avivi et al. [68] encapsulated tetracycline into bovine serum albumin-coated microspheres using a sonochemical approach, and found that up to 65% encapsulation efficiency could be achieved. Importantly, it was found that the majority of the encapsulated tetracycline, approximately 97%, was loaded within the core of the bubble and not simply adsorbed to the surface of the protein microspheres. Avivi et al. [68] confirmed that the

antimicrobial activity of the entrapped tetracycline when released by gentle heating was identical to equivalent amounts of free tetracycline when applied to different strains of bacteria.

Zhou et al. [69] used the same approach to create liquid-encapsulating lysozyme microspheres loaded with various oils (sunflower oil, tetradecane, dodecane and perfluorohexane). Liquid-filled microspheres can theoretically be loaded with significantly larger quantities of oil-soluble drugs, than air-filled bubbles where the active drug component needs to be functionalized on the surface of the bubble. The type of liquid encapsulated in the microspheres was found to influence the physical properties (i.e. size, polydispersity, and shell wall strength) of the formed microspheres.

The approach used to synthesize lysozyme microspheres can be extended to the synthesis of protein-mimicking polymer-coated microspheres. Cavalieri et al. [29] first reported a one-step sonochemical process to synthesise microspheres made from synthetic thiolated polymers of polymethacrylic acid. Important physical properties of the formed microspheres could be controlled by adjusting the thiol content in the macromolecules. The size, surface roughness, and shell thickness were all found to increase with increasing number of thiol groups in the monomer backbone. Recent work has further demonstrated the versatility of this method with the fabrication of new types of microspheres, including chitosan/titanium dioxide hybrids [70]. These hybrid microspheres have composite properties including high mechanical strength and antibacterial activity.

The size of the active sonochemical region delivering the ultrasound has been shown to effect the size distribution of formed microspheres, offering a means of controlling size [34]. In this study, a novel flow-through sonication horn with a very small active sonochemical region created smaller and more monodisperse microspheres than larger diameter horns with larger sonochemical regions.

In addition to promoting the formation of protein microspheres, ultrasound is a potential tool for targeted drug release. Ultrasound has been shown to break apart chitosan/titanium oxide hybrid nanospheres, releasing the entrapped contents [70]. This ability could potentially be used to induce rupture of protein microspheres to increase the localized delivery of a drug to specific parts of the body.

4 Industry application

Ultrasonics has been successfully used to generate a range of functional food and biomaterials in the laboratory. Industrial uptake of ultrasonics is not currently widespread, but is gaining considerable traction. A number of potential issues identified in early studies are gradually being debunked or resolved. Some of these issues will be discussed in the following section.

4.1 *Generation of particulate metal contaminants*

The strong physical phenomena generated by cavitation are capable of affecting not just the product, but also the transducer and reactor surfaces. The potential for release of metallic particles into the product that may be too small to remove has raised some health concerns. Recently however, Mawson et al. [71] assessed the production of metal particulates from ultrasonic transducers and found no evidence for the formation of harmful nanoparticles (<80 nm). In their study, no nano-particulate material was observed even after prolonged exposure (up to 7.5 hours) to high intensity ultrasound (20 kHz and 174 J/mL). However, most food or drug related applications involving ultrasound only require a few second of ultrasonic processing [72], meaning the risk of contamination of sensitive products such as foods and pharmaceuticals is minimal. Contamination-free reactors [73] have also been developed, and these can be used for the production of high-valued products that require the utmost purity.

4.2 *Degradation of functional properties*

The formation of radicals by ultrasonic cavitation can be beneficial in promoting and enhancing sonochemical reactions, but they can also potentially degrade redox sensitive components. This is a particular concern for foods and bioproducts, where the flavor and nutritional properties must be taken into account. Vitamins, fats, and other lipids are particularly susceptible to reactions induced by oxidative radicals. Fatty acid oxidation and lipolysis can significantly modify the flavor profile of the food [74] and produce off-flavors associated with ‘burnt rubber’, ‘grass’ or ‘rancid fruit’. Destruction of vitamins and anti-oxidants in foods may reduce the nutritional value or induce undesirable color changes.

However, these problems can be mitigated by selecting operation at lower frequency ultrasound and employing shorter exposure times [72]. As can be observed in Figure 4, radical formation at 20 kHz has been shown to be minimal relative to mid and high frequency ultrasound, and at short sonication times essentially negligible and unlikely to result in any significant change to

the functionality of biomaterials. For instance, for a solution of 5 % protein, the radical concentration resultant from sonication at 20 kHz for 5 min would typically be approximately less than 10 ppm (i.e. 10 moles of radical per million moles of protein). A recent study performed by Juliano et al. [75] showed that by limiting the duration of sonication (i.e. low specific energy), off-flavor volatiles from oxidation of milk fat could be kept below detectable sensory thresholds, even when operating with mid (400 kHz) or high (1 MHz) frequency ultrasound. The reason is because many natural food products such as milk contain natural antioxidants, which mitigate detrimental changes to the product.

4.3 Augmentation or replacement of current industrial techniques

Ultrasound can be used to replace or complement conventional techniques such as emulsification and polymerization that are used to create encapsulating particles in aqueous systems. Acoustic cavitation can provide efficient high-shear processing and radical polymerization as described in sections 3.1 and 3.2. However, ultrasound cannot replace industrial techniques currently used to create dried encapsulating particles, for example, spray drying, freeze drying, extrusion coating, fluidized bed coating, and coacervation [76]. Although studies have employed ultrasonic atomization as a technique to generate microcapsules at lab scale [77], spray drying is the most feasible method on an industrial scale, particularly for foods [76, 78]. Spray drying is highly energy efficient and effective technique by which internalized materials can be stabilized during storage. Emulsions can be formed as a precursor to encapsulate lipids and oil-soluble material during spray drying. Ultrasound can play a complementary role in aiding the formation of stable small-sized emulsions which may have beneficial outcomes within the spray dried product [45].

Hydrodynamic cavitation [79, 80], imparts similar cavitation-borne mechanisms to materials as ultrasonic cavitation, with the advantage of higher throughput due to its more conventional unit design. In some situations, hydrodynamic cavitation can be more efficient due to its ability to generate cavitation over a large volume/region [79].

4.4 Towards scale-up

Ultrasonic technology is yet to be widely implemented at an industrial scale, not due to uncertainty of its efficacy, but to challenges in scaling up. For a comprehensive review of the design considerations for efficient scale up of sonochemical reactors readers are directed to

Gogate et al. [81]. A key issue for scale-up of sonoprocessing that is worth highlighting is the strong attenuation in effectiveness with distance from the ultrasound source. This complicates scale-up as the effective volume is confined to the active sonochemical regions close to the transducers which, in some cases, can be quite small and narrow. One strategy is to use flow-through cells, where liquid passes through a narrow region close to the transducer to ensure all elements of fluid are subjected to ultrasound. Alternatively, flow chambers incorporating multiple transducer horns can be effective at providing uniform delivery of ultrasound to large volumes of material. Many commercial flow-through sonication products are now available, and can be tailored for a range of applications.

The current cost of the technology, although not prohibitive, is still often higher than conventional alternatives. However, the ability of ultrasound to produce unique, high-value products with improved functionality while reducing chemical and energy consumption in some applications, may compensate for the extra cost. It is envisioned that the continual development of the technology will lead to gradual industrial uptake of ultrasonics and eventually its mainstream use for the production of valuable functional materials.

5. Conclusion

The ultrasonic synthesis of functional food and bio-materials has a bright future with many yet-to-be realized commercial opportunities. Many of the issues identified in early studies are being overcome, paving the way for ultrasonic synthesis of the next generation of drug delivery agents, functional biomaterials and food products.

Acknowledgments

The authors acknowledge funding from the Australian Government through the ARC Dairy Innovation Hub.

References

- [1] F. Chemat, M.K. Khan, Applications of ultrasound in food technology: processing, preservation and extraction, *Ultrason. Sonochem.*, 18 (2011) 813-835.
- [2] B.B. Goldberg, J.-B. Liu, F. Forsberg, Ultrasound contrast agents: A review, *Ultrasound in Medicine & Biology*, 20 (1994) 319-333.
- [3] B.G. Pollet, The use of ultrasound for the fabrication of fuel cell materials, *international journal of hydrogen energy*, 35 (2010) 11986-12004.

- [4] K.S. Suslick, G.J. Price, Applications of ultrasound to materials chemistry, *Annual Review of Materials Science*, 29 (1999) 295-326.
- [5] T.G. Leighton, *The Acoustic Bubble*, Academic Press, San Diego, 1994.
- [6] S. Kentish, T. Wooster, M. Ashokkumar, S. Balachandran, R. Mawson, L. Simons, The use of ultrasonics for nanoemulsion preparation, *Innov. Food Sci. Emerg.*, 9 (2008) 170-175.
- [7] J. Kost, R. Langer, Responsive polymeric delivery systems, *Advanced Drug Delivery Reviews*, 6 (1991) 19-50.
- [8] K. Ferrara, R. Pollard, M. Borden, Ultrasound microbubble contrast agents: Fundamentals and application to gene and drug delivery, *Ann Rev Bio Eng*, 9 (2007) 415-447.
- [9] A.I. Eller, Growth of Bubbles by Rectified Diffusion, *The Journal of the Acoustical Society of America*, 46 (1969) 1246-1250.
- [10] M. Ashokkumar, J. Lee, S. Kentish, F. Grieser, Bubbles in an acoustic field: an overview, *Ultrason. Sonochem.*, 14 (2007) 470-475.
- [11] E.B. Flint, K.S. Suslick, The temperature of cavitation, *Science*, 253 (1991) 1397-1399.
- [12] T. Leong, M. Ashokkumar, S. Kentish, The fundamentals of power ultrasound—a review, *Acoust Aust*, 39 (2011) 54-63.
- [13] F.R. Young, *Cavitation*, McGraw-Hill, London, 1989.
- [14] K. Yasui, T. Tuziuti, J. Lee, T. Kozuka, A. Towata, Y. Iida, The range of ambient radius for an active bubble in sonoluminescence and sonochemical reactions, *J. Chem. Phys.*, 128 (2008) 184705-184712.
- [15] T.S.H. Leong, T.J. Wooster, S.E. Kentish, M. Ashokkumar, Minimising oil droplet size using ultrasonic emulsification, *Ultrason. Sonochem.*, 16 (2009) 721-727.
- [16] M. Villamiel, P. de Jong, Influence of high-intensity ultrasound and heat treatment in continuous flow on fat, proteins, and native enzymes of milk, *J Agr Food Chem*, 48 (2000) 472-478.
- [17] G. Cravotto, L. Boffa, S. Mantegna, P. Perego, M. Avogadro, P. Cintas, Improved extraction of vegetable oils under high-intensity ultrasound and/or microwaves, *Ultrason. Sonochem.*, 15 (2008) 898-902.
- [18] G.J. Price, Ultrasonically enhanced polymer synthesis, *Ultrason. Sonochem.*, 3 (1996) S229-S238.
- [19] S. Koda, T. Kimura, T. Kondo, H. Mitome, A standard method to calibrate sonochemical efficiency of an individual reaction system, *Ultrason. Sonochem.*, 10 (2003) 149-156.
- [20] T.J. Mason, A.J. Cobley, J.E. Graves, D. Morgan, New evidence for the inverse dependence of mechanical and chemical effects on the frequency of ultrasound, *Ultrason. Sonochem.*, 18 (2011) 226-230.
- [21] L. Johansson, T. Singh, T. Leong, R. Mawson, S. McArthur, R. Manasseh, P. Juliano, Cavitation and non-cavitation regime for large-scale ultrasonic standing wave particle separation systems—In situ gentle cavitation threshold determination and free radical related oxidation, *Ultrason. Sonochem.*, 28 (2016) 346-356.
- [22] T. Leong, L. Johansson, P. Juliano, S.L. McArthur, R. Manasseh, Ultrasonic separation of particulate fluids in small and large scale systems: a review, *Ind. Eng. Chem. Res.*, 52 (2013) 16555-16576.
- [23] A. Weissler, Formation of Hydrogen Peroxide by Ultrasonic Waves: Free Radicals, *J. Am. Chem. Soc.*, 81 (1959) 1077-1081.
- [24] K. Yasui, T. Tuziuti, M. Sivakumar, Y. Iida, Theoretical study of single-bubble sonochemistry, *J Chem Phys*, 122 (2005) 224706.
- [25] L. Thompson, L. Doraiswamy, Sonochemistry: science and engineering, *Ind. Eng. Chem. Res.*, 38 (1999) 1215-1249.
- [26] M.W. Grinstaff, K.S. Suslick, Air-filled proteinaceous microbubbles: synthesis of an echo-contrast agent, *P Natl Acad Sci USA*, 88 (1991) 7708-7710.

- [27] F. Cavalieri, M. Ashokkumar, F. Grieser, F. Caruso, Ultrasonic synthesis of stable, functional lysozyme microbubbles, *Langmuir*, 24 (2008) 10078-10083.
- [28] A. Shanmugam, J. Chandrapala, M. Ashokkumar, The effect of ultrasound on the physical and functional properties of skim milk, *Innov. Food Sci. Emerg.*, 16 (2012) 251-258.
- [29] F. Cavalieri, M. Zhou, F. Caruso, M. Ashokkumar, One-pot ultrasonic synthesis of multifunctional microbubbles and microcapsules using synthetic thiolated macromolecules, *Chemical Communications*, 47 (2011) 4096-4098.
- [30] C.F. Naudé, A.T. Ellis, On the mechanism of cavitation damage by nonhemispherical cavities collapsing in contact with a solid boundary, *Journal of Fluids Engineering*, 83 (1961) 648-656.
- [31] K.S. Suslick, *Sonochemistry, science*, 247 (1990) 1439-1445.
- [32] D.J. Flannigan, K.S. Suslick, Plasma formation and temperature measurement during single-bubble cavitation, *Nature*, 434 (2005) 52-55.
- [33] T. Feczko, J. Tóth, G. Dósa, J. Gyenis, Influence of process conditions on the mean size of PLGA nanoparticles, *Chem. Eng. Process. Process Intensification*, 50 (2011) 846-853.
- [34] M. Zhou, F. Cavalieri, F. Caruso, M. Ashokkumar, Confinement of acoustic cavitation for the synthesis of protein-shelled nanobubbles for diagnostics and nucleic acid delivery, *ACS Macro Letters*, 1 (2012) 853-856.
- [35] M. Zhou, F. Cavalieri, M. Ashokkumar, Modification of the size distribution of lysozyme microbubbles using a post-sonication technique, *Instrumentation Science & Technology*, 40 (2012) 51-60.
- [36] B. Abismaïl, J.P. Canselier, A.M. Wilhelm, H. Delmas, C. Gourdon, Emulsification by ultrasound: drop size distribution and stability, *Ultrason. Sonochem.*, 6 (1999) 75-83.
- [37] S.M. Jafari, Y. He, B. Bhandari, Production of sub-micron emulsions by ultrasound and microfluidization techniques, *J Food Eng.*, 82 (2007) 478-488.
- [38] C. Solans, P. Izquierdo, J. Nolla, N. Azemar, M. Garcia-Celma, Nano-emulsions, *Current Opinion in Colloid & Interface Science*, 10 (2005) 102-110.
- [39] K. Nakabayashi, F. Amemiya, T. Fuchigami, K. Machida, S. Takeda, K. Tamamitsu, M. Atobe, Highly clear and transparent nanoemulsion preparation under surfactant-free conditions using tandem acoustic emulsification, *Chem. Commun.*, 47 (2011) 5765-5767.
- [40] K. Kamogawa, G. Okudaira, M. Matsumoto, T. Sakai, H. Sakai, M. Abe, Preparation of oleic acid/water emulsions in surfactant-free condition by sequential processing using midsonic-megasonic waves, *Langmuir*, 20 (2004) 2043-2047.
- [41] R. Asami, M. Atobe, T. Fuchigami, Electropolymerization of an immiscible monomer in aqueous electrolytes using acoustic emulsification, *J. Am. Chem. Soc.*, 127 (2005) 13160-13161.
- [42] D. Kilcast, S. Clegg, Sensory perception of creaminess and its relationship with food structure, *Food Quality and Preference*, 13 (2002) 609-623.
- [43] H. Goudédranche, J. Fauquant, J.-L. Maubois, Fractionation of globular milk fat by membrane microfiltration, *Le lait*, 80 (2000) 93-98.
- [44] M. Akhtar, J. Stenzel, B.S. Murray, E. Dickinson, Factors affecting the perception of creaminess of oil-in-water emulsions, *Food Hydrocolloid*, 19 (2005) 521-526.
- [45] A. Soottitantawat, H. Yoshii, T. Furuta, M. Ohkawara, P. Linko, Microencapsulation by spray drying: influence of emulsion size on the retention of volatile compounds, *JOURNAL OF FOOD SCIENCE-CHICAGO-*, 68 (2003) 2256-2262.
- [46] G. Muschiolik, Multiple emulsions for food use, *Current Opinion in Colloid & Interface Science*, 12 (2007) 213-220.
- [47] A.G. Gaonkar, Stable multiple emulsions comprising interfacial gelatinous layer, flavor-encapsulating multiple emulsions and low/no-fat food products comprising the same, in, *Google Patents*, 1994.

- [48] C. Lobato-Calleros, A. Sosa-Pérez, J. Rodríguez-Tafoya, O. Sandoval-Castilla, C. Pérez-Alonso, E. Vernon-Carter, Structural and textural characteristics of reduced-fat cheese-like products made from W 1/O/W 2 emulsions and skim milk, *LWT-Food Science and Technology*, 41 (2008) 1847-1856.
- [49] C. Lobato-Calleros, J. Reyes-Hernández, C. Beristain, Y. Hornelas-Urbe, J. Sánchez-García, E. Vernon-Carter, Microstructure and texture of white fresh cheese made with canola oil and whey protein concentrate in partial or total replacement of milk fat, *Food Res Int*, 40 (2007) 529-537.
- [50] C. Lobato-Calleros, E. Rodríguez, O. Sandoval-Castilla, E. Vernon-Carter, J. Alvarez-Ramirez, Reduced-fat white fresh cheese-like products obtained from W 1/O/W 2 multiple emulsions: Viscoelastic and high-resolution image analyses, *Food Res Int*, 39 (2006) 678-685.
- [51] H. Lamba, K. Sathish, L. Sabikhi, Double Emulsions: Emerging Delivery System for Plant Bioactives, *Food Bioprocess Tech.*, 8 (2015) 709-728.
- [52] S.Y. Tang, M. Sivakumar, B. Nashiru, Impact of osmotic pressure and gelling in the generation of highly stable single core water-in-oil-in-water (W/O/W) nano multiple emulsions of aspirin assisted by two-stage ultrasonic cavitation emulsification, *Colloid Surface B*, 102 (2013) 653-658.
- [53] S.Y. Tang, M. Sivakumar, Design and evaluation of aspirin-loaded water-in-oil-in-water submicron multiple emulsions generated using two-stage ultrasonic cavitation emulsification technique, *Asia-Pacific Journal of Chemical Engineering*, 7 (2012) S145-S156.
- [54] Y.F. Maa, C. Hsu, Liquid-liquid emulsification by rotor/stator homogenization, *Journal of Controlled Release*, 38 (1996) 219-228.
- [55] S. Schultz, G. Wagner, K. Urban, J. Ulrich, High-pressure homogenization as a process for emulsion formation, *Chemical Engineering and Technology*, 27 (2004) 361-368.
- [56] C. Qian, D.J. McClements, Formation of nanoemulsions stabilized by model food-grade emulsifiers using high-pressure homogenization: Factors affecting particle size, *Food Hydrocolloids*, 25 (2011) 1000-1008.
- [57] S. Mahdi Jafari, Y. He, B. Bhandari, Nano-emulsion production by sonication and microfluidization—a comparison, *International Journal of Food Properties*, 9 (2006) 475-485.
- [58] Y.F. Maa, C.C. Hsu, Performance of sonication and microfluidization for liquid-liquid emulsification, *Pharmaceutical Development and Technology*, 4 (1999) 233-240.
- [59] A. Shanmugam, M. Ashokkumar, Ultrasonic preparation of stable flax seed oil emulsions in dairy systems—Physicochemical characterization, *Food Hydrocolloid*, 39 (2014) 151-162.
- [60] P. Kruus, M. O'Neill, D. Robertson, Ultrasonic initiation of polymerization, *Ultrasonics*, 28 (1990) 304-309.
- [61] P. Kruus, T. Patraboy, Initiation of polymerization with ultrasound in methyl methacrylate, *J Phys Chem*, 89 (1985) 3379-3384.
- [62] G.J. Price, D.J. Norris, P.J. West, Polymerization of methyl methacrylate initiated by ultrasound, *Macromolecules*, 25 (1992) 6447-6454.
- [63] G. Price, P. West, P. Smith, Control of polymer structure using power ultrasound, *Ultrason. Sonochem.*, 1 (1994) S51-S57.
- [64] B.M. Teo, S.W. Prescott, M. Ashokkumar, F. Grieser, Ultrasound initiated miniemulsion polymerization of methacrylate monomers, *Ultrason. Sonochem.*, 15 (2008) 89-94.
- [65] S.H. Sonawane, B.M. Teo, A. Brothie, F. Grieser, M. Ashokkumar, Sonochemical synthesis of ZnO encapsulated functional nanolatex and its anticorrosive performance, *Ind. Eng. Chem. Res.*, 49 (2010) 2200-2205.

- [66] B.M. Teo, F. Chen, T.A. Hatton, F. Grieser, M. Ashokkumar, Novel one-pot synthesis of magnetite latex nanoparticles by ultrasound irradiation, *Langmuir*, 25 (2009) 2593-2595.
- [67] B.M. Teo, S.W. Prescott, G.J. Price, F. Grieser, M. Ashokkumar, Synthesis of temperature responsive poly (N-isopropylacrylamide) using ultrasound irradiation, *J Phys Chem B*, 114 (2010) 3178-3184.
- [68] Avivi, Y. Nitzan, R. Dror, A. Gedanken, An easy sonochemical route for the encapsulation of tetracycline in bovine serum albumin microspheres, *J. Am. Chem. Soc.*, 125 (2003) 15712-15713.
- [69] M. Zhou, T.S.H. Leong, S. Melino, F. Cavalieri, S. Kentish, M. Ashokkumar, Sonochemical synthesis of liquid-encapsulated lysozyme microspheres, *Ultrason. Sonochem.*, 17 (2010) 333-337.
- [70] M. Zhou, B. Babgi, S. Gupta, F. Cavalieri, Y. Alghamdi, M. Aksu, M. Ashokkumar, Ultrasonic fabrication of TiO₂/chitosan hybrid nanoporous microspheres with antimicrobial properties, *RSC Advances*, 5 (2015) 20265-20269.
- [71] R. Mawson, M. Rout, G. Ripoll, P. Swiergon, T. Singh, K. Knoerzer, P. Juliano, Production of particulates from transducer erosion: Implications on food safety, *Ultrason. Sonochem.*, (2014).
- [72] M. Ashokkumar, D. Sunartio, S. Kentish, R. Mawson, L. Simons, K. Vilkuh, C. Versteeg, Modification of food ingredients by ultrasound to improve functionality: A preliminary study on a model system, *Innov. Food Sci. Emerg.*, 9 (2008) 155-160.
- [73] S. Freitas, G. Hielscher, H.P. Merkle, B. Gander, Continuous contact-and contamination-free ultrasonic emulsification—a useful tool for pharmaceutical development and production, *Ultrason. Sonochem.*, 13 (2006) 76-85.
- [74] F. Martinez, A. Davidson, J. Anderson, S. Nakai, I. Desai, A. Radcliffe, Effects of ultrasonic homogenization of human milk on lipolysis, IgA, IgG, lactoferrin and bacterial content, *Nutr Res*, 12 (1992) 561-568.
- [75] P. Juliano, A.E. Torkamani, T. Leong, V. Kolb, P. Watkins, S. Ajlouni, T.K. Singh, Lipid oxidation volatiles absent in milk after selected ultrasound processing, *Ultrason. Sonochem.*, 21 (2014) 2165-2175.
- [76] K.G.H. Desai, H. Jin Park, Recent developments in microencapsulation of food ingredients, *Drying technology*, 23 (2005) 1361-1394.
- [77] Y. Yeo, K. Park, A new microencapsulation method using an ultrasonic atomizer based on interfacial solvent exchange, *Journal of controlled release*, 100 (2004) 379-388.
- [78] A. Gharsallaoui, G. Roudaut, O. Chambin, A. Voilley, R. Saurel, Applications of spray-drying in microencapsulation of food ingredients: An overview, *Food Res Int*, 40 (2007) 1107-1121.
- [79] P.R. Gogate, A.B. Pandit, A review and assessment of hydrodynamic cavitation as a technology for the future, *Ultrason. Sonochem.*, 12 (2005) 21-27.
- [80] V. Moholkar, P.S. Kumar, A. Pandit, Hydrodynamic cavitation for sonochemical effects, *Ultrason. Sonochem.*, 6 (1999) 53-65.
- [81] P.R. Gogate, V.S. Sutkar, A.B. Pandit, Sonochemical reactors: important design and scale up considerations with a special emphasis on heterogeneous systems, *Chem. Eng. J.*, 166 (2011) 1066-1082.

Figures

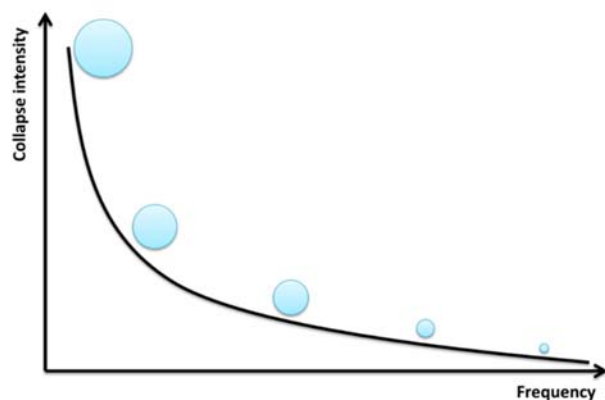


Figure 1: Schematic representation of the relationship between ultrasonic frequency applied and the relative intensity and size of the collapsing bubbles. Not drawn to scale.

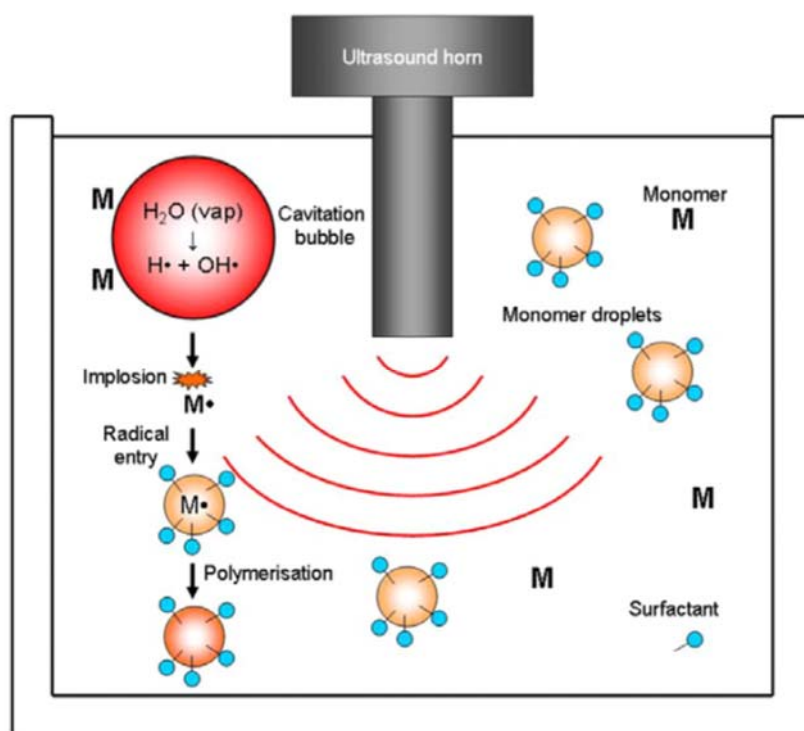


Figure 2: A schematic diagram of a proposed emulsion polymerization process. Reprinted from Teo et al. [64], Copyright 2008, with permission from Elsevier.

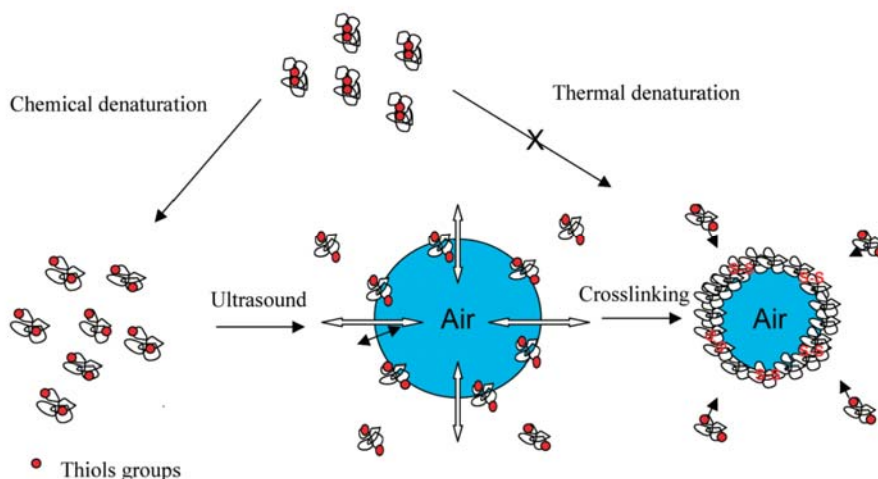


Figure 3: Mechanism proposed for the formation of lysozyme protein microspheres. Reprinted with permission from Cavalieri et al [27]. Copyright 2008 American Chemical Society.

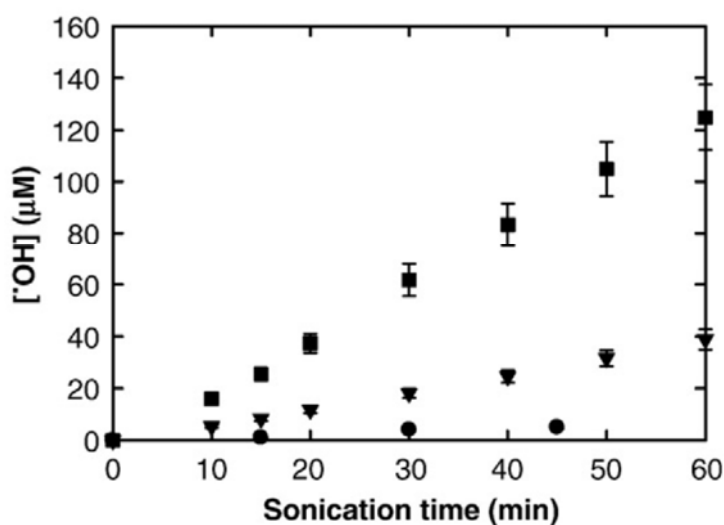


Figure 4: OH radical yield generated in water upon sonication at different acoustic frequencies with matched power 0.9 W cm^{-2} . Adapted from Ashokkumar et al. [72], Copyright 2008, with permission from Elsevier.