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Oral minoxidil bio-activation by hair follicle outer root sheath cell sulfotransferase enzymes predicts clinical efficacy in female pattern hair loss

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8 **Oral Minoxidil Bio-activation by Hair Follicle Outer Root Sheath Cell**  
9 **Sulfotransferase Enzymes Predicts Clinical Efficacy in Female Pattern Hair Loss**

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36 **Conflict of interest:**

37 Ramos PM: no conflict of interest

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39 Sinclair R: is the inventor of US patent 10226462 detection and treatment of excessive  
40 hair shedding

41 Miot HA: no conflict of interest

42 Topical minoxidil (TM) is the only FDA-approved medication for the treatment of female pattern  
43 hair loss (FPHL).<sup>1</sup> Minoxidil is a pro-drug. TM efficacy in FPHL requires bio-activation into  
44 minoxidil sulfate by sulfotransferase enzymes.<sup>2</sup> Sulfotransferases are xenobiotic metabolizing  
45 enzymes expressed in many tissues with the highest expression found in the human liver.<sup>3</sup> Hair  
46 follicle outer root sheath cells (ORS) also express sulfotransferase, and ORS sulfotransferase  
47 activity predicts TM therapeutic response.<sup>4-6</sup>

48 The clinical response to TM is variable. Several strategies have been previously explored to  
49 overcome lack of response to 5% TM secondary to low ORS sulfotransferases activity.<sup>7</sup> Use of  
50 higher concentrations of TM is effective in up to 60% of non-responders.<sup>8</sup>

51 We also recently conducted a head to head trial comparing 1mg daily oral minoxidil (OM) once  
52 daily to 5% topical once daily in the treatment of FPHL.<sup>9</sup> As OM is extensively metabolized by  
53 the liver, we sought to determine whether liver or ORS sulfotransferases are primarily  
54 responsible for OM bio-activation and efficacy in the treatment of FPHL. We also sought to  
55 develop a predictive biomarker for OM therapeutic response in FPHL.

56 Thirteen women with Sinclair's Stage II-IV FPHL received 1mg OM daily for 24 weeks. Hair  
57 counts were performed in a target (tattooed) area on the parietal scalp before and after  
58 treatment. Ten anagen hairs were plucked from the affected scalp, trimmed to a length of 1cm  
59 and immersed, bulb first, in 100 $\mu$ L of assay solution. Assay solution consisted of 50mM  
60 phosphate buffer (pH 8), 5mM potassium p-nitrophenyl sulfate, 20 $\mu$ M adenosine 3',5'-  
61 diphosphate, 100 $\mu$ M minoxidil, and 5mM MgCl<sub>2</sub>. Hair bulbs were then incubated for 24h at room  
62 temperature before the optical absorbance of the solution at 405nm was determined with a  
63 spectrophotometer (Shimadzu UV-1700, Kyoto, Japan) using a single scan and 1cm path  
64 length. Values less than 0.4 Absorbance Units (AU) are a validated marker of low follicular ST  
65 activity.<sup>4</sup>

66 The baseline hair count, percentile of hair increase and three cut-off parameters for OD are  
67 presented in figure 1. Women with low sulfotransferase activity had less hair re-growth (9.0+/-  
68 4.36) compared to women with a high sulfotransferase enzymatic activity (18.4+/-9.17) (p=0.06).  
69 A Receiver Operator Characteristics analysis (ROC) of the data (Graph 1) was conducted with a  
70 dichotomous categorization variable based on clinical response as defined by <13.7% increase  
71 in hair density. Using the previously reported response rate of OM in the treatment of FPHL of  
72 79.7%,<sup>10</sup> the optimal criteria for the ROC analysis is an OD>0.254 i.e., 100% of the subjects  
73 below 0.254 OD were non-responders. While the sample size of this study is small, the ROC  
74 analysis demonstrates that a lower follicular enzymatic activity threshold is required for bio-  
75 activation of OM compared to TM.

76 With respect to the relative the role of liver versus ORS sulfotransferase activity on OM bio-  
77 activation, our results show that ORS bioactivation predicts clinical response in the treatment of  
78 FPHL. While the dataset reached marginal significance (p=0.06) for an enzymatic activity cut-off  
79 of 0.4 OD, the ROC analysis demonstrated that the optimal cut-off for non-responders to OM in  
80 our study was 0.254 OD. This demonstrates that a lower follicular enzymatic activity threshold  
81 is required for bio-activation of OM compared to TM. This might be due to a contribution of liver  
82 and platelets on OM conversion and/or higher follicular accumulation of minoxidil. Using the

83 ROC analysis from this study as a “training dataset” for future investigations, we plan to  
84 elucidate the exact mechanism of OM bio-activation.

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89 **Figure 1.** Baseline hair density versus percentile of hair increase in this sample. Three cut-off points for  
90 Optical Density (OD) regarding sulfotransferase are represented.

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95 **Figure 2.** Receiver Operator Characteristics analysis (ROC) of the raw data with a dichotomous  
96 categorization variable based on response <13.7% hair re-growth.

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**Figure 1.** Baseline hair density versus percentile of hair increase in this sample. Three cut-off points for Optical Density (OD) regarding sulfotransferase are represented.



