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Title:

steps: Software for spatially and temporally explicit population simulations

Date:

2020-04-01

Citation:

Visintin, C., Briscoe, N. J., Woolley, S. N. C., Lentini, P. E., Tingley, R., Wintle, B. A. & Golding, N. (2020). steps: Software for spatially and temporally explicit population simulations. *Methods in Ecology and Evolution*, 11 (4), pp.596-603. <https://doi.org/10.1111/2041-210X.13354>.

Persistent Link:

<https://hdl.handle.net/11343/290256>

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8 Article type : Application

9 Editor : Dr Laura Graham

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steps: software for spatially- and temporally- explicit population simulations

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12 **Summary**

13 1. Species population dynamics are driven by spatial and temporal changes in the environment,
14 anthropogenic activities, and conservation management actions. Understanding how populations
15 will change in response to these drivers is fundamental to a wide range of ecological applications,
16 but there are few open-source software options accessible to researchers and managers that allow
17 them to predict these changes in a flexible and transparent way.

18
19 2. We introduce an open-source, multi-platform R package, *steps*, that models spatial changes in
20 species populations as a function of drivers of distribution and abundance, such as climate,
21 disturbance, landscape dynamics, and species ecological and physiological requirements.

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1111/2041-210X.13354](https://doi.org/10.1111/2041-210X.13354)

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22

23 3. To illustrate the functionality of *steps*, we model the population dynamics of the greater glider
24 (*Petauroides volans*), an arboreal Australian mammal. We demonstrate how *steps* can be used to
25 simulate population responses of the glider to forest dynamics and management with the types of
26 data commonly used in ecological analyses.

27

28 4. *steps* expands on the features found in existing software packages, can easily incorporate a
29 range of spatial layers (e.g., habitat suitability, vegetation dynamics and disturbances), facilitates
30 integrated and transparent analyses within a single platform, and produces interpretable outputs of
31 changes in species' populations through space and time. Further, *steps* offers both ready-to-use,
32 built-in functionality, as well as the ability for advanced users to define their own modules for custom
33 analyses. Thus, we anticipate that *steps* will be of significant value to environment and wildlife
34 managers and researchers from a broad range of disciplines.

35 **Keywords:** demography, ecological modelling, habitat, metapopulation, matrix models, population
36 dynamics, range shift, SDM

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37 **Introduction**

38 The need for spatial and temporal predictions of species' population dynamics in response to
39 environmental change and management actions has long been recognised (Akçakaya et al., 2004;
40 Keith et al., 2008), and population models remain central to ecological management and research
41 (Thuiller et al., 2013). Researchers and conservation managers need to be able to model the
42 cumulative and synergistic effects of multiple drivers of population dynamics, including the amount,
43 quality, and configuration of habitat (e.g. effects of climate and land-use change), stochastic
44 disturbances and catastrophes (e.g. fire), and spatially-varying influences on survival and fecundity
45 (e.g. disease, physiological constraints, predation).

46
47 Two well-established modelling approaches used to answer these questions are population viability
48 analysis (PVA) and correlative species distribution modelling (SDM). PVA requires knowledge of
49 species' population dynamics and estimates of vital rates, such as survival and fecundity, to
50 simulate population trajectories and quantify the likelihood of population persistence over a defined
51 time period (Boyce, 1992). Correlative SDM is a statistical modelling approach used to predict
52 species occurrence across a landscape using relevant spatial predictors and has been widely used
53 to predict potential impacts of environmental change and management interventions on biodiversity
54 (Guisan et al., 2013). Correlative SDM enables users to make spatial and temporal predictions of
55 species occurrence when these are the only data available. However, these models do not explicitly
56 account for dynamic population processes, such as dispersal, spatial and temporal variation in vital
57 rates, and density-dependence, and so may not provide reliable predictions of species persistence
58 (Fordham et al., 2012).

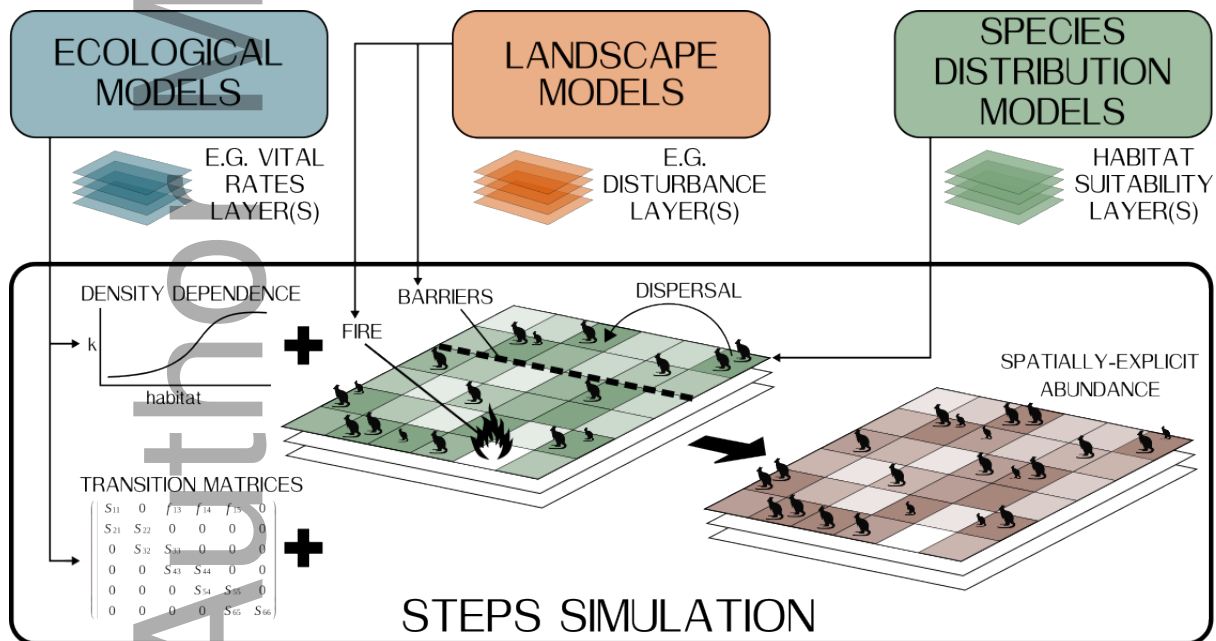
59
60 To address some of these limitations, spatially-explicit population models (SEPM) have been
61 developed, which combine spatially-explicit data with the population dynamics processes in PVA
62 (Fordham et al., 2013). Typically, SEPM start with spatial information on the availability and quality
63 of habitat patches through time – often derived from correlative SDM. This habitat suitability is
64 combined with information about initial population sizes, vital rates, and dispersal capacity, to
65 predict future population abundances via stochastic simulation (Fig. 1 and Akçakaya et al.
66 2004; Keith et al., 2008; Beeton et al., 2015).

67
68 Despite the potential of SEPM approaches to provide spatially- and temporally-explicit predictions of
69 population abundances, they remain relatively rare compared to correlative SDM (Briscoe et al.,
70 2019). A key barrier to more widespread use is the availability and accessibility of population
71 modelling software. As highlighted by Lurgi et al. (2015), common shortcomings of currently-
72 available population modelling software include limited flexibility for customisation, lack of
73 transparency and reproducibility, restrictions to specific computer operating systems, and advanced
74 computational skills required of users. There are several software options that are open-source and

75 multi-platform, however, these tend to be either highly customisable, but with complex set-up and
 76 coding requirements (e.g., *spaDES*), or more straightforward, but with limited modularity and
 77 documentation to assist with customisation (e.g., *demoniche* - Nenzén et al., 2012). Other freely
 78 available software with extensive built-in functionality may only run on single operating systems and
 79 have source code that is not available for users to scrutinise or customise (e.g. *RangeShifter* -
 80 Bocedi et al., 2014). One of the most recognised software packages for SEPM is *RAMAS Metapop*
 81 (Akçakaya, 1999), which has an intuitive interface and is well supported and tested, but has a
 82 license fee, only runs on Windows operating systems, and is not open-source. This prevents users
 83 from adding or integrating new modules that capture ecological or population processes that are not
 84 already integrated in the software. These limitations prevent more widespread adoption of
 85 comprehensive ecological models and inhibit synthesis studies that draw generalisations from the
 86 outcomes of a range of case studies and simulations.

87
 88 We present *steps* version 1.0.0, an *R* package that combines functionality from existing spatial
 89 population simulation software with high transparency, and a modular design that allows for future
 90 extensions by other researchers. Crucially, our software is open-source - written in the language of
 91 the widely used statistical software *R* (R Core Team, 2019) - and provides a zero-cost option for
 92 managers, consultants, citizen-scientists, and others across many different sectors.

93
 94



95
 96 **Figure 1:** *steps* is run on a grid-based architecture, which enables the easy
 97 integration of spatial products (grids) from other modelling software, including
 98 climate, landscape, physiological and disturbance information.

99

100 **Overview of software**

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101 While *steps* can be used to perform non-spatial PVA using information on initial population size
102 and a population growth function, its key aim is to enable spatio-temporal predictions of species
103 populations across a landscape. *steps* uses a regular grid to spatially represent a landscape and
104 define carrying capacities, initial abundances, and other miscellaneous spatial information that can
105 be used to define and modify population or landscape features (e.g. translocations or habitat
106 disturbances). Population dynamics within suitable habitat cells are then simulated based on an
107 age- or stage-structured transition matrix, with populations connected via dispersal (Fig. 1).

108
109 Because *steps* uses a regular grid to represent populations, it is straightforward to integrate different
110 ecological models and types of data representing landscapes, habitats and populations (Fig. 1).
111 Examples include defining habitat patches and/or carrying capacities using static or temporally-
112 variable spatial outputs from open-source software for landscape change (e.g. *LANDIS* - Mladenoff,
113 2004) or species distribution models (e.g. *dismo* – Hijmans et al., 2015); incorporating grids of
114 spatially- and temporally-explicit vital rates (e.g. biophysical models such as *NicheMapR* - Kearney
115 & Porter, 2019); and evaluating the relative benefits to species' populations arising from spatial
116 prioritisations of conservation actions (e.g. protected area designation or conservation
117 management) proposed using packages such as *Zonation* (Moilanen et al., 2009).

118
119 *steps* has been developed to run through the statistical software *R* on any operating system and
120 can easily operate in enhanced computational environments (i.e. high-performance server clusters).
121 *steps* architecture is modular and object-oriented (Fig. 2) to maximise flexibility and be
122 computationally-efficient (Appendix B).

123

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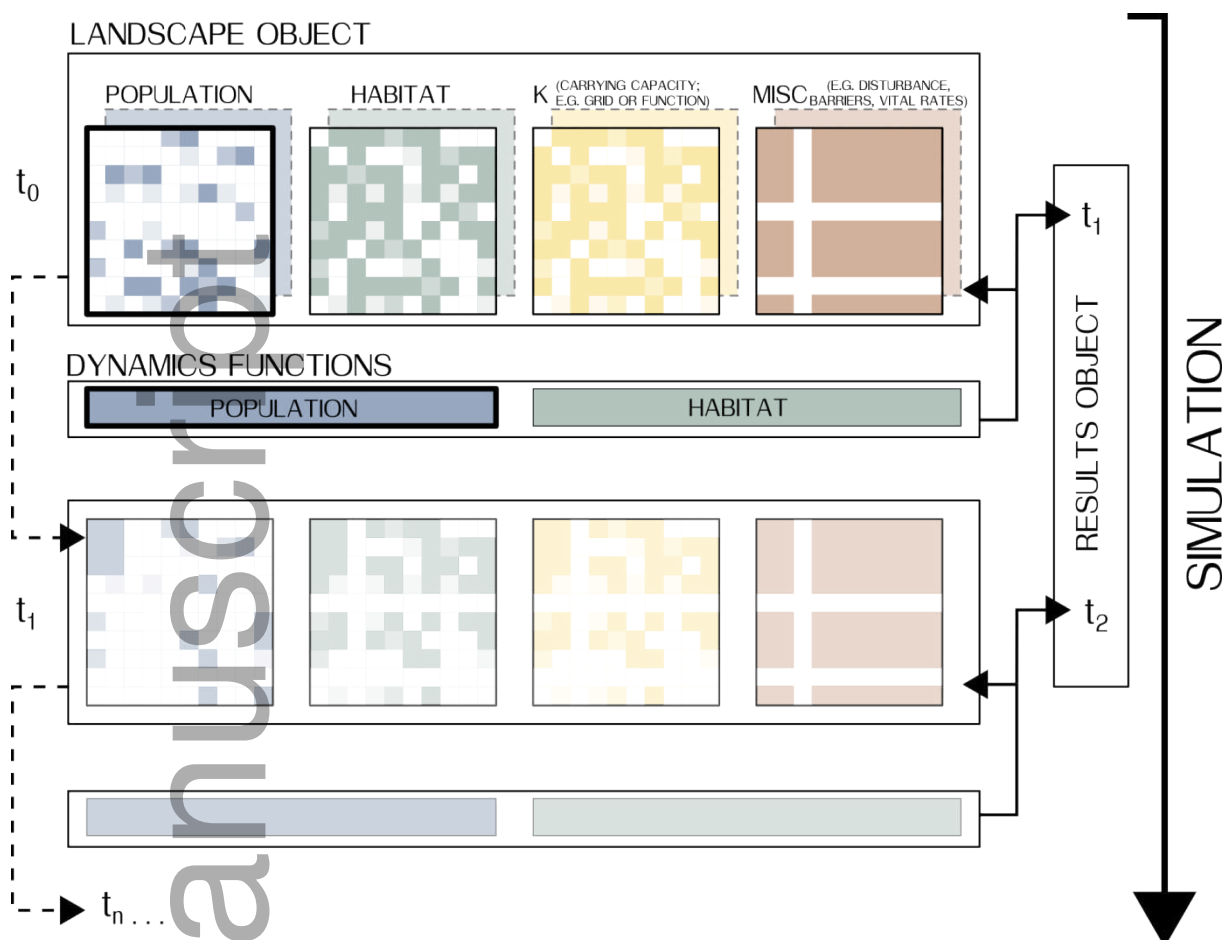


Figure 2: Landscape objects and dynamics functions are passed to a simulation. During the simulation, dynamics are applied to landscape objects at each time step/iteration (t_n) and stored in a results object (solid arrows). The landscape object is modified and reused at each iteration (dashed arrows). Only initial populations and a population growth function (bold boxes) are required to run a simulation - this is equivalent to a non-spatial population viability analysis.

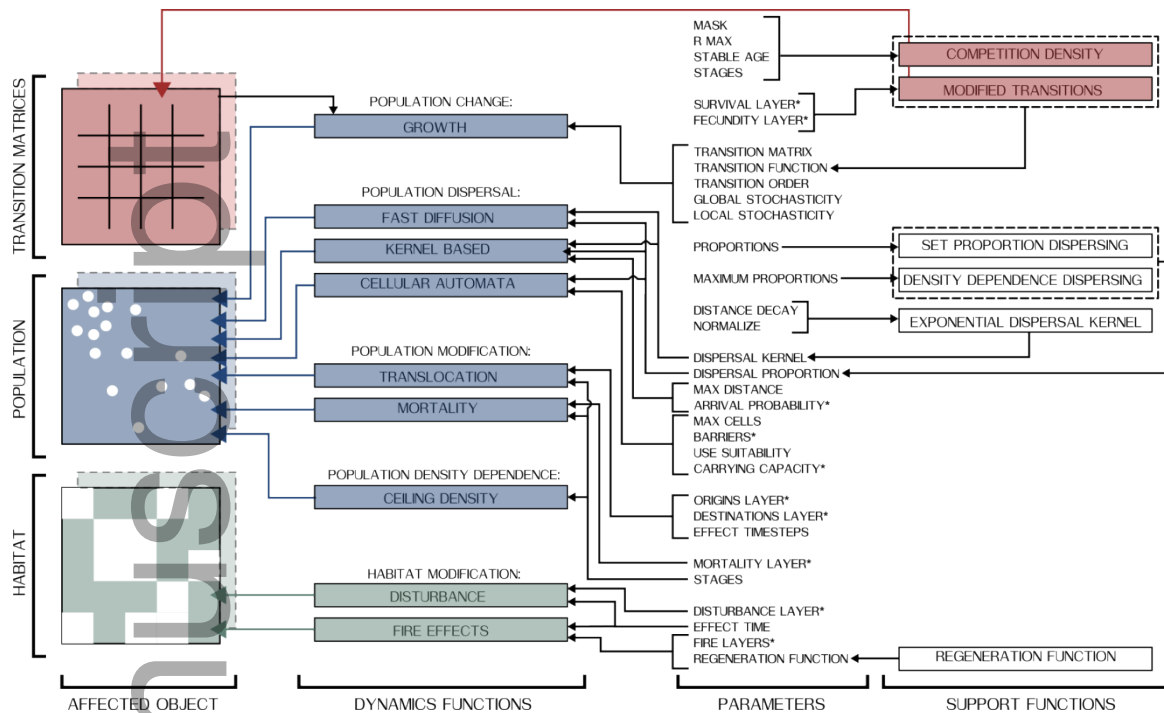
Software functionality

steps includes pre-defined functions to control changes in population growth, dispersal, density dependence, and modifications to populations and habitats (Fig. 3). We have included three types of dispersal function: a computationally-efficient diffusion kernel approach using a Fast Fourier Transform, a more flexible diffusion kernel approach in which dispersal can be constrained by habitat suitability or carrying capacity, and a cellular automata dispersal simulation that considers individual-based movements and accounts for landscape permeability. Both competition and ceiling density dependence functions are included to modify vital rates (e.g. Keith et al., 2008) and to cap population sizes (e.g. Zurell et al., 2012), respectively. Direct changes to populations based on management interventions, such as translocations, reintroductions, fertility control, or culling, can be simulated by calling functions to add or subtract cell populations at specified timesteps. Several plotting options are available, including the ability to plot spatial changes in populations over time as

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144 animations.

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Figure 3: Functionality of *steps*. Boxes in the “dynamics” and “support” categories indicate named inbuilt functions whilst boxes in the “affected object” category indicate landscape objects that are modified by functions during a simulation. Colours and coloured arrows indicate which functions operate on which objects. Black arrows represent information flow (i.e. objects/parameters supplied to functions). Parameters that require spatially-explicit inputs (i.e. grids) are marked with asterisks.

Users are not limited to performing simulations using pre-defined functions - the software accepts any custom function that can be implemented in *R* and we include a tutorial vignette in the software to assist users with creating their own functions. This balance of internal functions that cover many typical operations used in SEPM, and the flexibility to include other operations, offers unlimited potential for users to model and test unique scenarios.

Example

Here we demonstrate the functionality of our software by specifying a greater glider (*Petauroides volans*) SEPM within core habitat in South-Eastern Australia. Greater gliders are folivorous, arboreal mammals that rely on old growth forests for shelter and foraging. We used *steps* to simulate population trajectories over 50 years, accounting for key threats including forest fires. We stress that this example application is provided to demonstrate how to specify a SEPM using *steps*, and not to provide realistic predictions about the viability of the species under planned or proposed

168 management.

169
170 Our model landscape is a 250,000 hectare grid with a resolution of 500m x 500m (10,000 total
171 cells). We predicted habitat suitability in each timestep using a correlative species distribution model
172 fitted to occurrence data and dynamic climate and vegetation layers (Appendix A). We set initial
173 population sizes by randomly distributing approximately 4,000 gliders across suitable habitat
174 (likelihood of occurrence ≥ 0.5) in the landscape. Initial populations were comprised of three life-
175 stages - newborn (~29%), juvenile (~14%), and adult (~57%) - roughly based on stable age
176 distributions calculated from an initial age-based transition matrix (Table 1).

177
178 **Table 1:** A population transition matrix (Lefkovich matrix) representing post-
179 breeding survival and fecundity values for female greater gliders. The first row
180 indicates the expected number of newborn female gliders per timestep (one year),
181 per individual in each life stage that produce them, multiplied by their respective
182 survival. The second and third rows indicate the expected transition probabilities for
183 the three life stages. The juvenile and newborn life stages each span a year, so
184 there is no probability of an individual remaining in those life stages between
185 timesteps.

	Newborn	Juvenile	Adult
Newborn	0.000	0.425	0.425
Juvenile	0.500	0.000	0.000
Adult	0.000	0.850	0.850

186
187 We set up and ran the model using three functions: **landscape()**, **population_dynamics()**,
188 and **simulation()**. We first use **landscape()** to create an initial landscape object. This is
189 composed of our initial population raster stack, a habitat suitability raster stack obtained from a
190 species distribution model (Appendix A), and a carrying capacity object - in this case a user-defined
191 logistic k function that will create a raster of maximum population sizes at each timestep based on
192 the corresponding habitat suitability raster. We also include rasters for "fire", "development", and
193 "predation", which are used by the population and habitat dynamics functions to modify population
194 size, carrying capacity, and population parameters, such as survival. A link to code and data used to
195 create our inputs is provided in the "Data availability" section.

```

# carrying capacity as a logistic function of suitability
k_function <- function(landscape, timestep) {
  suit <- landscape$suitability[[timestep]]
  25 / (1 + exp(-(suit - 0.5) / 0.05))
}

glider_landscape <- landscape(population = glider_pop_500,
  suitability = glider_hab_suit_500,
  carrying_capacity = k_function,
  "fire" = glider_hab_fire_500,
  "development" = glider_hab_clear_500,
  "predation" = glider_owl_predation_500)

```

196

197 We use **population_dynamics()** to specify the functions to be executed on the landscape
 198 object at each timestep. We indicate that the population will grow according to a baseline transition
 199 matrix (Table 1), but with environmental stochasticity in each of the values. The transition matrices
 200 for each cell will be modified at each timestep by density-dependent competition between juveniles
 201 and adults, and by fires (affecting only survival).

202

203 We use cellular-automata dispersal and specify the maximum number of cells (five) across which
 204 individuals (juveniles only) move in each timestep to approximate a mean dispersal distance of one
 205 kilometre. We also use a function to set the proportion of individuals that disperse based on how
 206 close the population of a cell is to its carrying capacity. To simulate predation effects from owls, we
 207 use 'predation' rasters that define cells where predation reduces the population of juveniles by 50%.

```

glider_pop_dynamics_ls <- population_dynamics(
  change = growth(transition_matrix = glider_trans_mat,
    global_stochasticity = 0.005,
    transition_function = list(competition_density(stages = c(2, 3)),
      modified_transition(survival_layer = "fire")),
  dispersal = cellular_automata_dispersal(max_cells = c(0, 5, 0),
    dispersal_proportion = density_dependence_dispersing()),
  modification = mortality("predation", stages = 2),
  density_dependence = NULL
)

```

208

209 Lastly, we use **simulation()** to run population dynamics simulations across the landscape for the
 210 specified number of timesteps and replications, and return the results. We provide our landscape
 211 and population dynamics objects and specify ten replicates of fifty timesteps for the simulation.

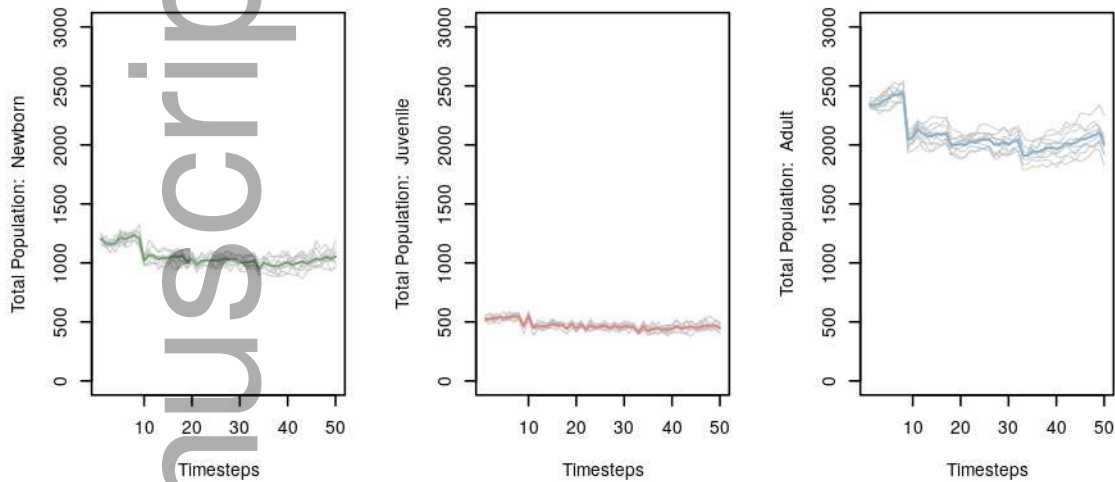
```

glider_baseline <- simulation(landscape = glider_landscape,
  population_dynamics = glider_pop_dynamics_ls,
  habitat_dynamics = NULL,
  timesteps = 50,
  replicates = 10)

```

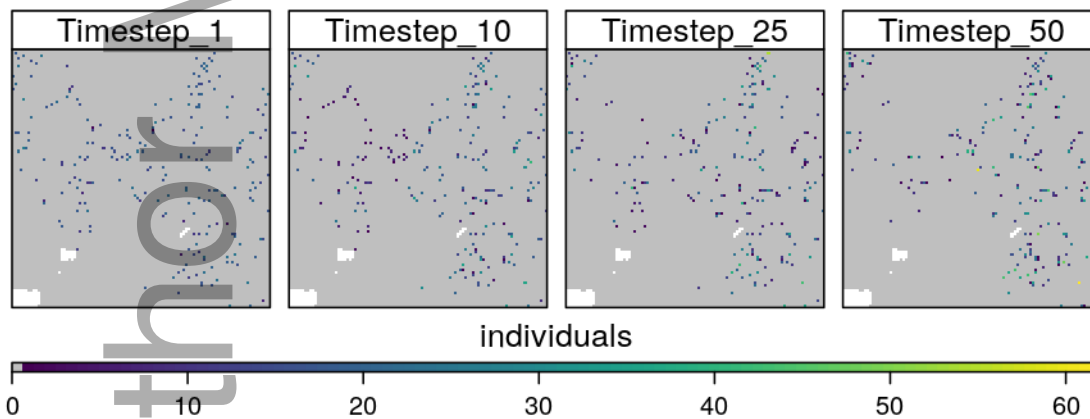
212

213 We illustrate two of the available options for plotting simulation results: trajectories of the total
214 population of each life stage across the landscape (Fig. 4) and spatial grids (Fig. 5). For the greater
215 glider, both plots indicate a sharp decline, due to a large fire occurring in year nine, followed by
216 population stabilisation.



217

218 **Figure 4:** Population trajectories over 50 years for each life stage. Grey lines
219 represent simulation replicates (ten total) whilst coloured lines are mean values.



220

221 **Figure 5:** Spatial representations of total population sizes in each 25ha grid cell for
222 years 1, 10, 25 and 50 of a single simulation replicate. Grey areas have zero
223 population and white areas indicate cells with missing values (e.g. water bodies).

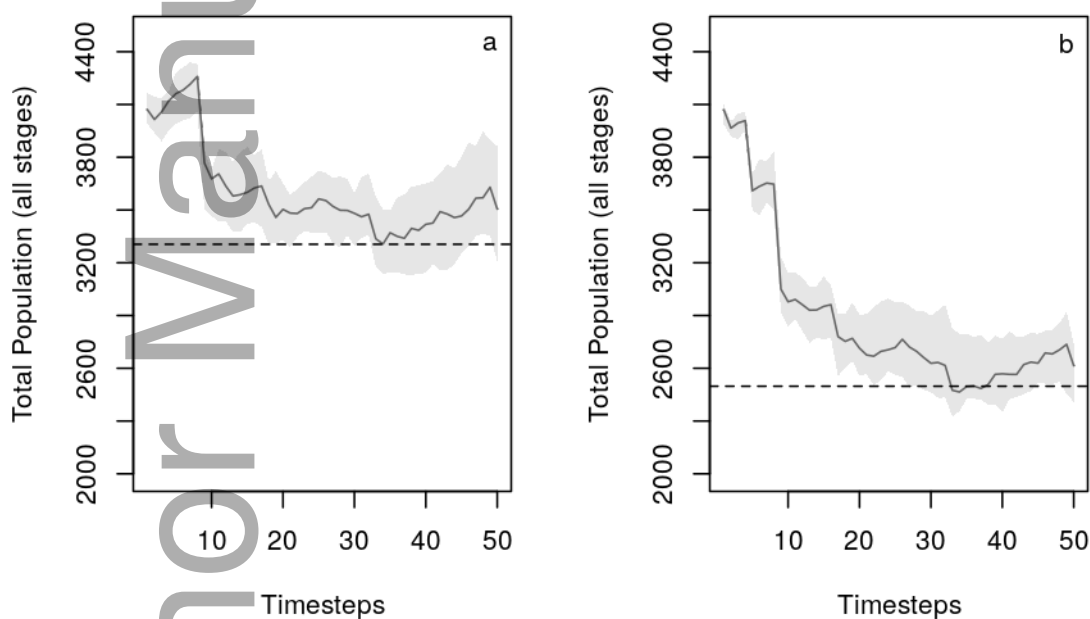
224

225 By adding functions or changing input data and parameters, we can test management scenarios for
226 the greater glider. As fire and logging were already incorporated into predicted habitat suitability, we
227 tested the effect of clearing approximately 43,000ha in the landscape for urban development to

228 assess population responses. This was done by adding a habitat dynamic function that applied
229 yearly disturbance layers to the habitat suitability rasters. We provided a raster stack representing
230 additive habitat clearing in each year containing zeros (cleared), and ones (no modification), which
231 was multiplied by the habitat suitability raster at each timestep.

```
glider_habmod <- simulation(landscape = glider_landscape,  
  population_dynamics = glider_pop_dynamics_ls,  
  habitat_dynamics = list(disturbance(disturbance_layers = "disturbance")),  
  timesteps = 50,  
  replicates = 10)
```

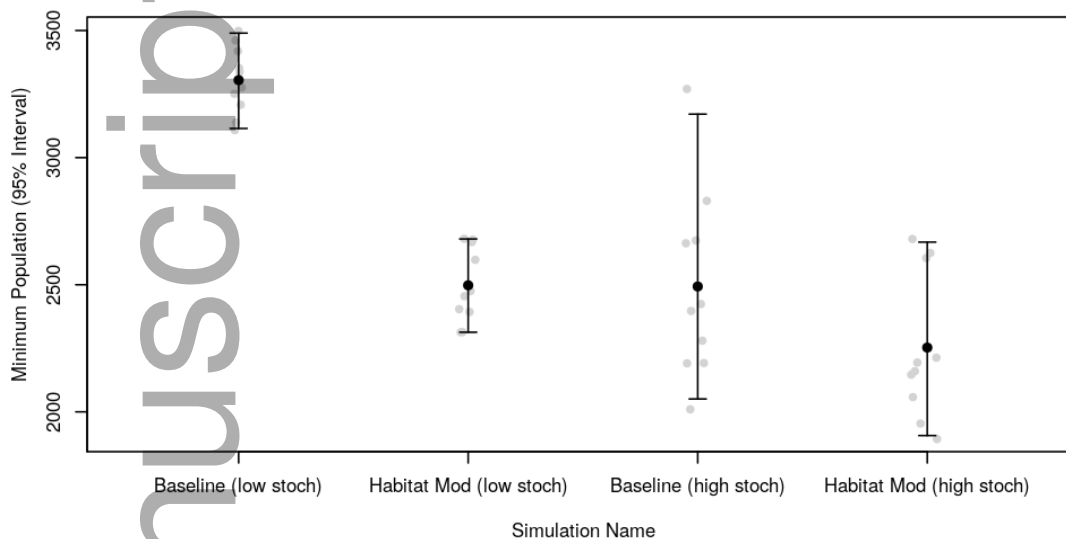
232
233 The simulation resulted in a corresponding change in the population trajectories of the species.
234 Whilst the population once remained relatively stable after the fire (Fig. 6a), it now continued to
235 decrease due to habitat modification (Fig. 6b).



236
237 **Figure 6:** Total population trajectory over the next 50 years with (a) no habitat
238 clearing for urban development and (b) habitat clearing for urban development
239 across the landscape. The grey area represents variability (95% interval) amongst
240 all simulation replicates (ten total) whilst the heavy black line represents mean
241 values. The dotted line is the mean of the minimum population estimates across all
242 replicates.

243
244 Simulation results can also be summarised across different management or environmental change
245 scenarios, or under varying model assumptions, using 'expected minimum abundance' (EMA) plots
246 (McCarthy & Thompson 2001). EMA is the mean of the minimum total population size across all

247 simulations. These plots allow the user to assess the sensitivity of predictions to model
248 assumptions, or to management or environmental change scenarios. In addition to simulating
249 disturbance, we tested each scenario with two different values of environmental stochasticity
250 (expressed as the standard deviation on survival and fecundity values) - a low value of 0.005 and a
251 ten-fold higher value of 0.05. Fig. 7 shows a comparison of all four simulations.



252
253 **Figure 7:** Expected minimum abundance (EMA) for different simulated scenarios,
254 with and without habitat modification, and at both high and low levels of
255 environmental stochasticity. The black dots represent expected minimum
256 abundance across all simulation replicates (each shown as a grey dot).

257 Discussion

258 We have developed open-source, flexible, and interoperable software that will enable widespread
259 and reproducible spatially-explicit simulations of species population dynamics. *steps* offers many
260 features currently available in commercially-licensed or platform-specific population simulators –
261 including dispersal, density-dependence, growth, and habitat dynamics – as well as the ability to
262 explicitly incorporate threats and management actions that can vary through space and time.

263
264 The grid-based approach adopted in *steps* differs from existing software such as *RAMAS Metapop*,
265 which represent landscapes using discrete habitat patches, which may differ in size and shape. This
266 has several advantages, for example, switching from small, isolated, defined patches to more
267 continuous landscapes is problematic when attempting to sensibly model dispersal using patch-
268 based approaches. Patch-based models also result in a loss of spatial information and, given the
269 increasing availability and quality of grid-based data, this may reduce model utility and flexibility.
270 However, grid-based model performance is dependent on the user selecting a suitable cell size for
271 analyses (see below). If motivated, users could simulate patch-based environments by aggregating
272 cell populations and setting all cells between patch-group cells to missing values. This may increase
273

274 computational efficiency since *steps* internally ignores all missing values.

275
276 Given that *steps* uses a regular grid to spatially organise a landscape, it is important for the user to
277 carefully consider the size of cells. For example, species attributes, such as home range size and
278 activity patterns, should influence the choice of grid cell resolution. In our example, we chose a grid
279 cell resolution of 500 m by 500 m (25ha). We could have chosen a cell size that is closer to the
280 1.5ha mean home range of greater gliders, as they tend to forage close to shelter; however, this
281 would only allow a maximum of two animals in each grid cell. These low carrying capacities make
282 the populations in each cell more sensitive to demographic stochasticity (although this can be
283 turned off globally), which may not be ecologically realistic in some circumstances. Where users are
284 unsure about the appropriate grid cell size to use, we recommend considering a cell size that sets
285 the maximum carrying capacity to more than a few individuals in highly suitable cells. Users may
286 need to experiment by toggling demographic stochasticity on and off to find a workable, and
287 ecologically reasonable, cell size.

288
289 Although we chose to simulate a management action that affected the amount and spatial
290 arrangement of available habitat, it is also possible to test impacts on population dynamics by
291 simulating processes that alter survival and/or fecundity (e.g. disease, heatwaves, drought),
292 including how these vary spatially and temporally. Our example analysed a species of conservation
293 concern, however, the modelling approach could equally be applied to management of
294 overabundant or pest species. *steps* provides a simple starting point, but a remarkable amount of
295 flexibility, enabling users to integrate models, data, and functions to produce robust and transparent
296 predictions about population change across landscapes.

297
298 **Future work**
299 We intend to develop an online repository for custom *steps* modules and advanced tutorials,
300 allowing users to share custom-written functions (e.g. management interventions or data extracting
301 utilities) for use with *steps*. Most importantly, a publicly-accessible repository will further support our
302 main motivation for developing the software - transparency, flexibility, and reproducibility.

303
304 **Acknowledgements**
305 We thank Will Morris and John Baumgartner for their contributions towards developing early
306 prototypes and modules of the software, Gerry Ryan for testing and providing feedback on the
307 software, Craig Nitschke for sharing the *LANDIS* model, and Barry Brook and one other anonymous
308 reviewer for providing insightful comments on the software and manuscript. This work was
309 supported by The University of Melbourne's Computational Biology Research Initiative, awarded to
310 N.J.B., P.E.L., R.T. and N.G., the National Environmental Science Program (NESP) Threatened
311 Species Recovery Hub, and Australian Research Council (ARC) Future Fellowship (FT100100819)

312 to B.A.W., R.T. and N.G. were partially funded by ARC DECRA fellowships (DE170100601 and
313 DE180100635). N.J.B. was supported by an ARC Discovery project (DP180101852), P.E.L. was
314 supported by an ARC Linkage project (LP160100439), and S.N.C.W. was supported by the Global
315 Ocean Biodiversity Initiative funded by the International Climate Initiative (IKI).

316

317 **Authors' contributions**

318 B.A.W., S.N.C.W. and N.J.B. conceptualised the modular SEPM freeware. N.G. devised the
319 software architecture. S.N.C.W. and B.A.W. developed early prototypes of the software. C.V.,
320 S.N.C.W. and N.G. led the advanced software development and testing. C.V. primarily wrote,
321 documented and published the software and led the writing of the manuscript. N.J.B. and B.A.W.
322 provided data for the example case study. N.J.B, P.E.L. and R.T. advised on the functionality of the
323 software and its applicability to management. All authors performed extended testing of the software
324 and contributed to the manuscript.

325

326 **Software accessibility**

327 The current development version of *steps* is hosted on Github at: [https://github.com/steps-](https://github.com/steps-dev/steps)
328 [dev/steps](https://github.com/steps-dev/steps). The latest stable version of *steps* can be installed from the comprehensive R archive
329 network (CRAN), within R, or directly at: <https://cran.r-project.org/web/packages/steps>.

330

331 **Data availability**

332 VISINTIN, CASEY; BRISCOE, NATALIE; Woolley, Skipton; LENTINI, PIA; Tingley, Reid; WINTLE,
333 BRENDAN; et al. (2020): Spatially- and temporally-explicit population simulator (*steps*) R package -
334 supporting data. University of Melbourne. Dataset. [https://melbourne.figshare.com/articles/Spatially-](https://melbourne.figshare.com/articles/Spatially-_and_temporally-explicit_population_simulator_steps_R_package_-_supporting_data/11608392)
335 [_and_temporally-explicit_population_simulator_steps_R_package_-_supporting_data/11608392](https://melbourne.figshare.com/articles/Spatially-_and_temporally-explicit_population_simulator_steps_R_package_-_supporting_data/11608392)

336 **References**

337 Akcakaya, H. R. (1999). RAMAS GIS. *Linking landscape data with population viability analysis.*
338 *Applied Biomathematics. Setauket, NY.*

339
340 Akcakaya, H. R., Radeloff, V. C., Mladenoff, D. J., & He, H. S. (2004). Integrating landscape and
341 metapopulation modeling approaches: viability of the sharp-tailed grouse in a dynamic landscape.
342 *Conservation Biology*, 18(2), 526-537.

343
344 Beeton, N. J., McMahon, C. R., Williamson, G. J., Potts, J., Bloomer, J., Bester, M. N., ... Johnson, C. N.
345 (2015). Using the spatial population abundance dynamics engine for conservation management.
346 *Methods in Ecology and Evolution*, 6(12), 1407-1416.

347
348 Briscoe, N. J., Elith, J., Salguero-Gómez, R., Lahoz-Monfort, J. J., Camac, J. S., Giljohann, K. M., ...
349 Guillera-Aroita, G. (2019), Forecasting species range dynamics with process-explicit models:
350 matching methods to applications. *Ecology Letters* doi:10.1111/ele.13348

351
352 Bocedi, G., Palmer, S. C., Pe'er, G., Heikkinen, R. K., Matsinos, Y. G., Watts, K., & Travis, J. M.
353 (2014). RangeShifter: a platform for modelling spatial eco-evolutionary dynamics and species'
354 responses to environmental changes. *Methods in Ecology and Evolution*, 5(4), 388-396.

355
356 Boyce, M. S. (1992). Population viability analysis. *Annual review of Ecology and Systematics*, 23(1), 481-
357 497.

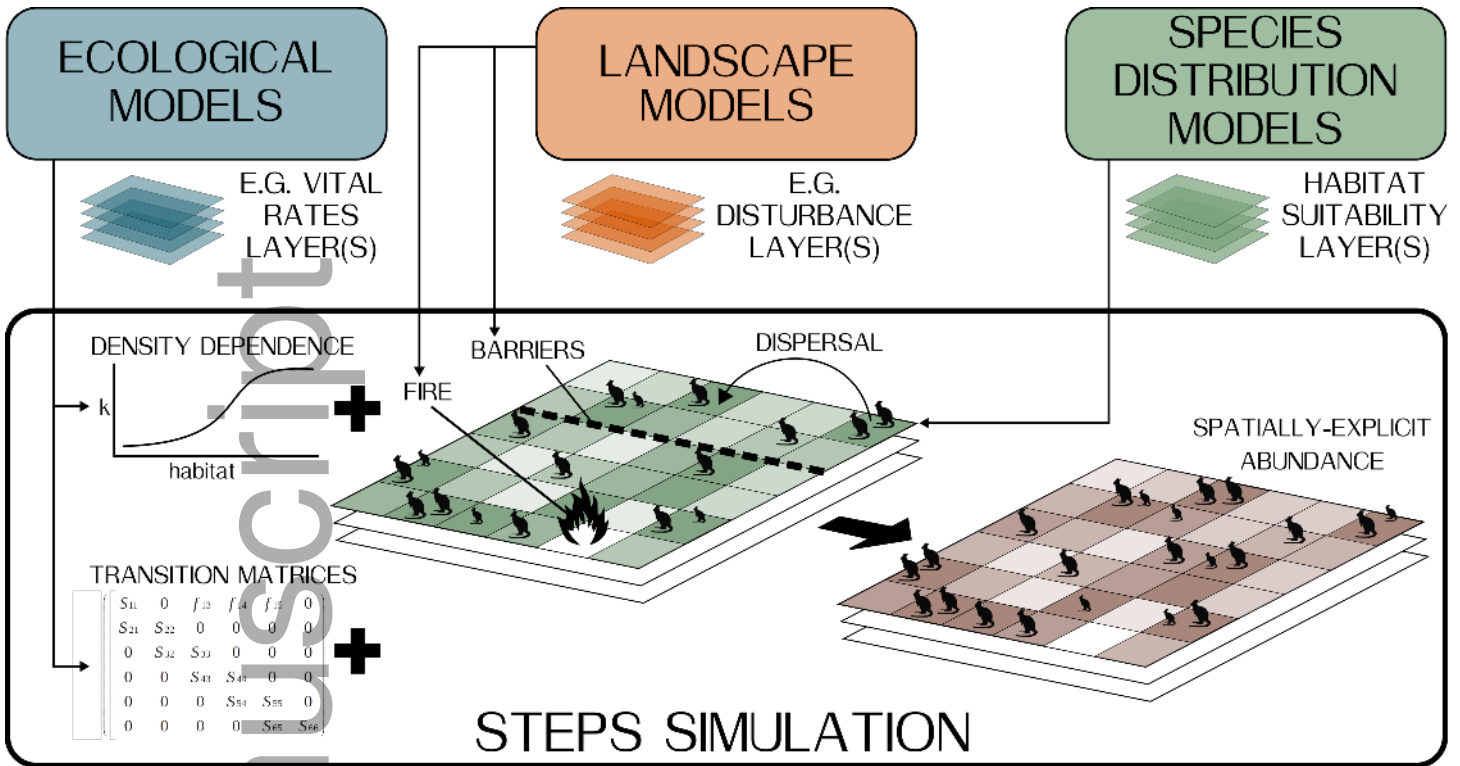
358
359 Fordham, D. A., Resit Akçakaya, H. , Araújo, M. B., Elith, J. , Keith, D. A., Pearson, R. , ...
360 Brook, B. W. (2012), Plant extinction risk under climate change: are forecast range shifts alone
361 a good indicator of species vulnerability to global warming? *Global Change Biology*, 18: 1357-
362 1371.

363
364 Fordham, D. A., Akçakaya, H. R., Araújo, M. B., Keith, D. A. and Brook, B. W. (2013), Tools for
365 integrating range change, extinction risk and climate change information into conservation
366 management. *Ecography*, 36: 956-964.

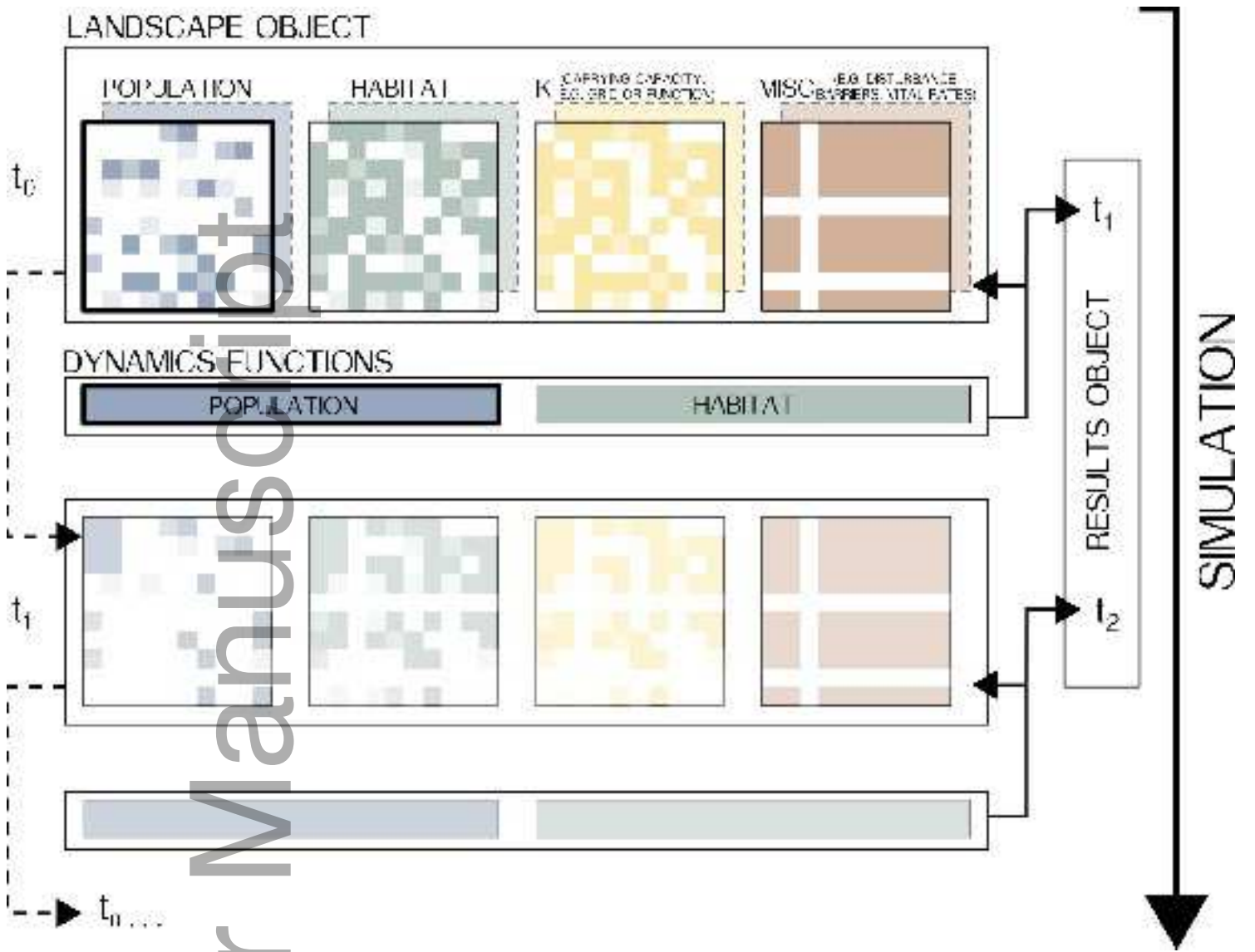
367
368 Guisan, A., Tingley, R., Baumgartner, J. B., Naujokaitis-Lewis, I., Sutcliffe, P. R., Tulloch, A. I., ...
369 Martin, T. G. (2013). Predicting species distributions for conservation decisions. *Ecology Letters*,
370 16(12), 1424-1435.

371
372 Hijmans, R.J., Phillips, S., Leathwick, J. and Elith, J. (2015) Dismo: Species Distribution Modeling.
373 R Package Version 1.1-4. <http://CRAN.R-project.org/package=dismo>

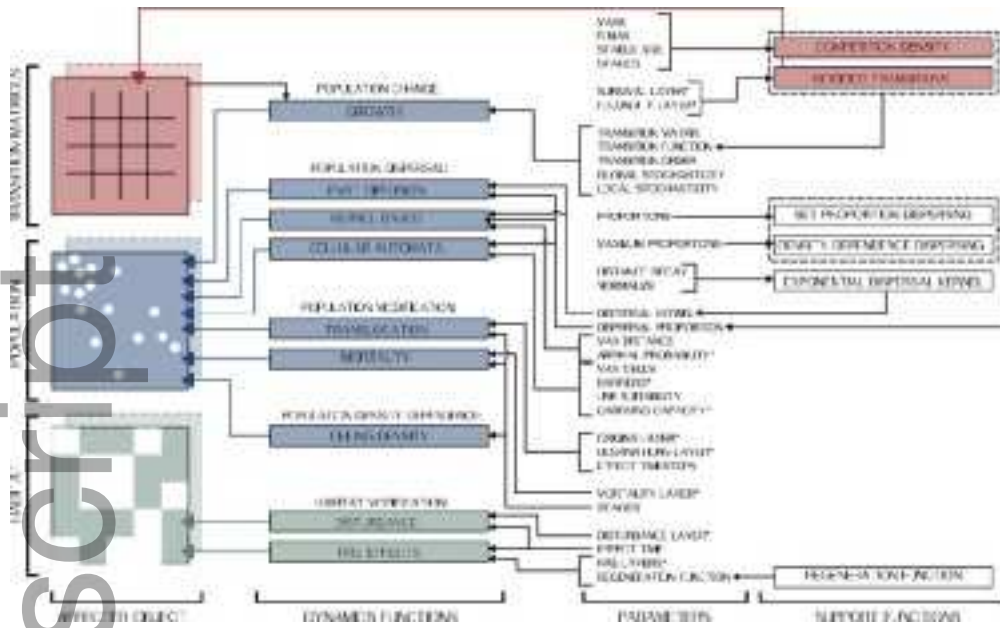
374
375 Kearney, M.R & Porter W.P (2019) NicheMapR - an R package for biophysical modelling: the
376 ectotherm and Dynamic Energy Budget models, *Ecography*, 10.1111/ecog.04680
377
378 Keith, D.A. et al. (2008). Predicting extinction risks under climate change: coupling stochastic
379 population models with dynamic bioclimatic habitat models. *Biology Letters*, 4, 560-563.
380
381 Lurgi, M., Brook, B. W., Saltre, F., & Fordham, D. A. (2015). Modelling range dynamics under global
382 change: which framework and why?. *Methods in Ecology and Evolution*, 6(3), 247-256.
383
384 McCarthy, M.A., and Thompson, C. (2001). Expected minimum population size as a measure of
385 threat. *Animal Conservation* 4:351-355.
386
387 Mladenoff, D. J. (2004). LANDIS and forest landscape models. *Ecological modelling*, 180(1), 7-19.
388
389 Moilanen, A., Kujala, H., & Leathwick, J. R. (2009). The Zonation framework and software for
390 conservation prioritization. *Spatial conservation prioritization*, 135, 196-210.
391
392 Nenzén, H. K., Swab, R. M., Keith, D. A. and Araújo, M. B. (2012). demoniche – an R-package for
393 simulating spatially-explicit population dynamics. *Ecography*, 35: 577-580.
394
395 R Core Team (2019). R: A language and environment for statistical computing. Vienna, Austria: R
396 Foundation for Statistical Computing. <https://www.R-project.org/>
397
398 Thuiller, W., Münkemüller, T., Lavergne, S., Moullot, D., Mouquet, N., Schiffrers, K., & Gravel, D.
399 (2013). A road map for integrating eco-evolutionary processes into biodiversity models. *Ecology*
400 *Letters*, 16, 94-105.
401
402 Wintle, B. A., Bekessy, S. A., Venier, L. A., Pearce, J. L., & Chisholm, R. A. (2005). Utility of
403 dynamic-landscape metapopulation models for sustainable forest management. *Conservation*
404 *Biology*, 19(6), 1930-1943.
405
406 Zurell, D., Grimm, V., Rossmannith, E., Zbinden, N., Zimmermann, N. E., & Schröder, B. (2012).
407 Uncertainty in predictions of range dynamics: black grouse climbing the Swiss Alps. *Ecography*,
408 35(7), 590-603.



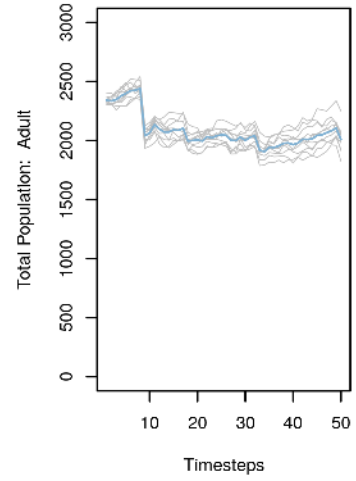
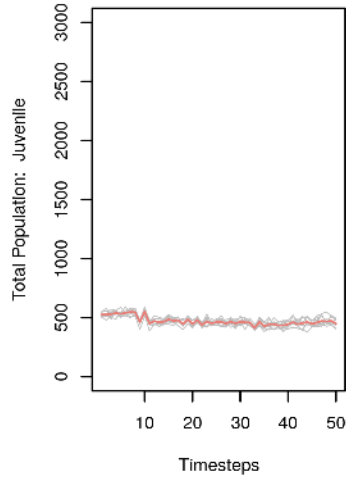
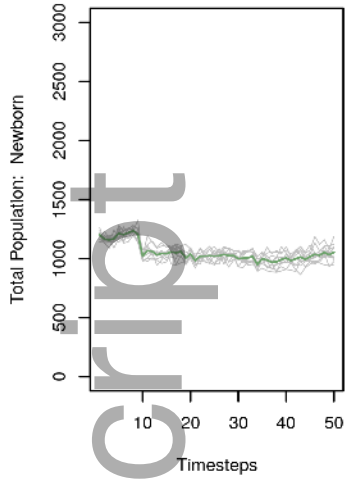
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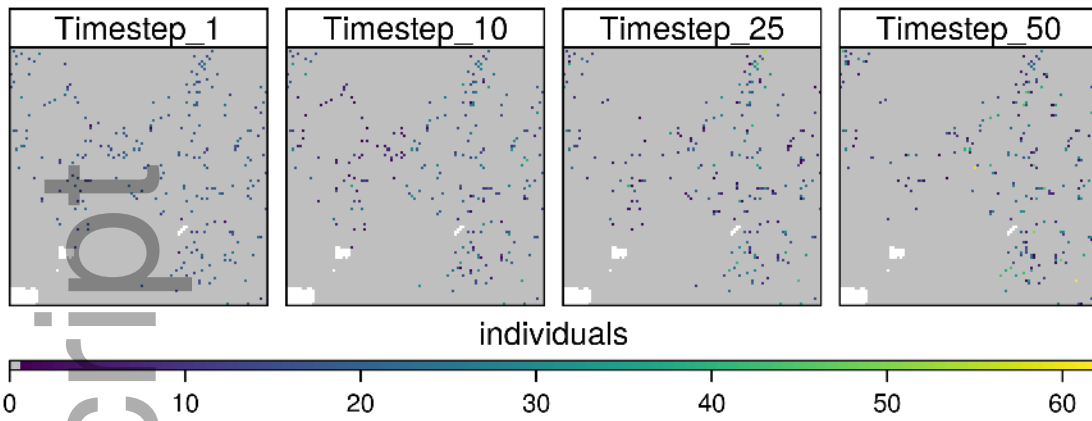


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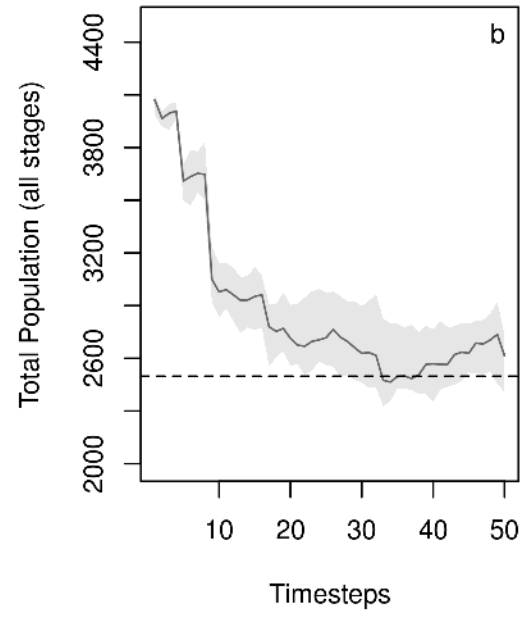
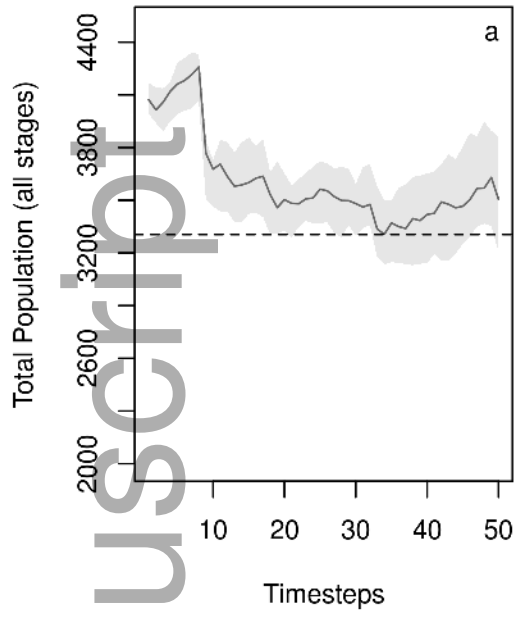


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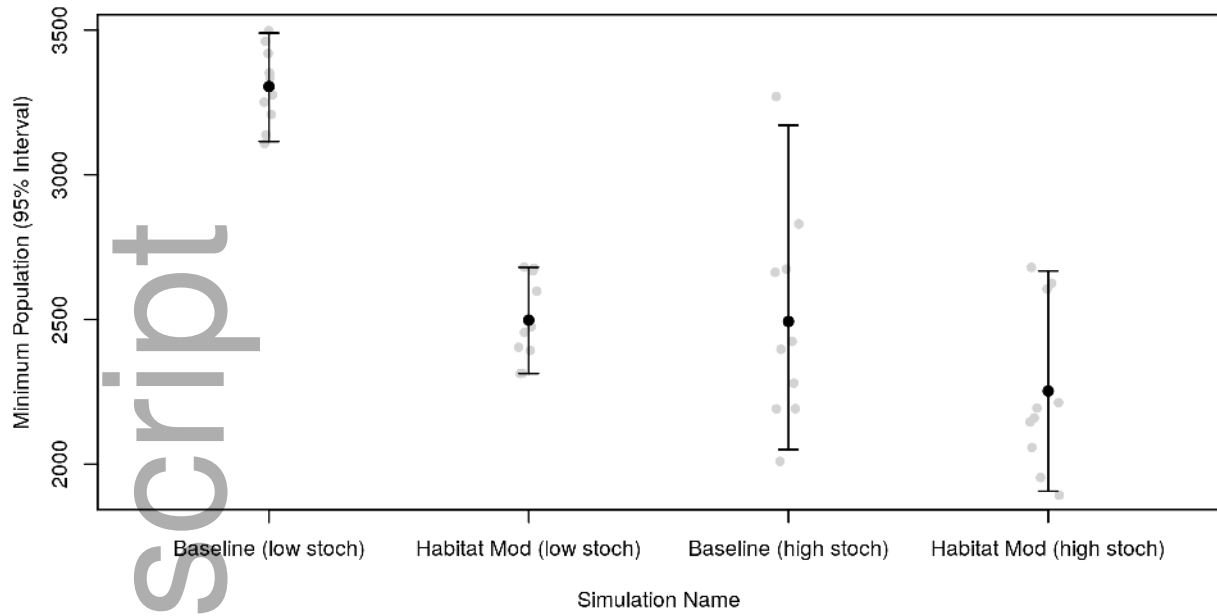
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