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RESEARCH ARTICLE

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# Enhanced RAD21 cohesin expression confers poor prognosis in BRCA2 and BRCAX, but not BRCA1 familial breast cancers

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## Abstract

**Introduction:** The *RAD21* gene encodes a key component of the cohesin complex, which is essential for chromosome segregation, and together with BRCA1 and BRCA2, for high-fidelity DNA repair by homologous recombination. Although its expression correlates with early relapse and treatment resistance in sporadic breast cancers, it is unclear whether familial breast cancers behave in a similar manner.

**Methods:** We performed an immunohistochemical analysis of RAD21 expression in a cohort of 94 familial breast cancers (28 BRCA1, 27 BRCA2, and 39 BRCAX) and correlated these data with genotype and clinicopathologic parameters, including survival. In these cancers, we also correlated RAD21 expression with genomic expression profiling and gene copy-number changes and miRNAs predicted to target RAD21.

**Results:** No significant differences in nuclear RAD21 expression were observed between BRCA1 (12 (43%) of 28), BRCA2 (12 (44%) of 27), and BRCAX cancers (12 (33%) of 39 ( $p = 0.598$ )). No correlation was found between RAD21 expression and grade, size, or lymph node, ER, or HER2 status (all  $P > 0.05$ ). As for sporadic breast cancers, RAD21 expression correlated with shorter survival in grade 3 ( $P = 0.009$ ) and but not in grade 1 ( $P = 0.065$ ) or 2 cancers ( $P = 0.090$ ). Expression of RAD21 correlated with poorer survival in patients treated with chemotherapy ( $P = 0.036$ ) but not with hormonal therapy ( $P = 0.881$ ). RAD21 expression correlated with shorter survival in BRCA2 ( $P = 0.006$ ) and BRCAX ( $P = 0.008$ ), but not BRCA1 cancers ( $P = 0.713$ ). Changes in *RAD21* mRNA were reflected by genomic changes in DNA copy number ( $P < 0.001$ ) and by RAD21 protein expression, as assessed with immunohistochemistry ( $P = 0.047$ ). High *RAD21* expression was associated with genomic instability, as assessed by the total number of base pairs affected by genomic change ( $P = 0.048$ ). Of 15 miRNAs predicted to target RAD21, mir-299-5p inversely correlated with RAD21 expression ( $P = 0.002$ ).

**Conclusions:** Potential use of RAD21 as a predictive and prognostic marker in familial breast cancers is hence feasible and may therefore take into account the patient's BRCA1/2 mutation status.

## Introduction

It is estimated that 5% to 10% of all breast cancers are attributable to inherited mutations, of which the two most important and highly penetrant are BRCA1 and BRCA2 [1]. Studies have demonstrated key differences

in spontaneous BRCA-associated tumors [2,3]. BRCA1 cancers are more likely to show a basal phenotype, with 80% to 90% of BRCA1 cancers being negative for ER and HER2 and positive for basal cytokeratins [4,5]. BRCA1 cancers also have characteristic gene-expression and genomic profiles, and appear to be sensitive to DNA damage by cisplatin. Although reports suggest that lobular carcinomas may be more frequent in BRCA2 carriers, no specific molecular phenotype has been described for BRCA2-associated tumors, which

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usually show a ductal, no-special-type morphology and ER positivity [6].

RAD21 is a component of the multiprotein complex cohesin, which is involved in maintaining alignment and cohesion of replicated “sister” chromatids. RAD21, together with SMC1, SMC3, and STAG1/2, forms a tripartite ring, which according to the “ring model,” promotes sister chromatid cohesion (SCC) by encircling sister chromatids [7]. SCC not only is vital for correct chromosome segregation during mitosis and meiosis, but also plays an important role in the repair of double-stranded DNA breaks (DSBs). By promoting sister-chromatid alignment, cohesin allows BRCA1/2-mediated homologous recombination to occur between sister chromatids [8]. In addition to its role in maintaining sister-chromatid cohesion, evidence now suggests cohesins are involved in promoting and inhibiting gene transcription. In MCF-7 cell lines, genes cobound by cohesin and ER are preferentially regulated by estrogen [9]. Cohesin may also act as a negative regulator of gene expression, by physically blocking enhancer/promoter interaction [10].

Of the four proteins that compose the core cohesin complex, RAD21 has emerged as a key marker of tumor behavior. A meta-analysis of gene-expression data from clinical cancer specimens showed that increased *RAD21* expression was a feature of poorly differentiated breast, ovarian, bladder, and lung cancers [11]. Gene-expression profiling of 31 breast cancer patients with supraclavicular lymph node metastasis revealed *RAD21* as one of six genes that were differentially expressed between good- and poor-outcome groups [12]. Our previous study on sporadic breast cancers showed that RAD21 overexpression correlated with early relapse in high-grade breast cancers regardless of intrinsic tumor subtype [13]. We, and others, also showed that *RAD21* knockdown confers *in vitro* resistance to DNA-damaging chemotherapeutic agents, which recapitulated the findings in our cohort of sporadic breast cancers [13,14].

Although RAD21 overexpression correlates with early relapse and treatment resistance in sporadic cancers, it is unclear whether familial breast cancers behave in a similar manner. This may be of particular relevance, as RAD21, BRCA1, and BRCA2 are all involved in DNA repair through homologous recombination; hence RAD21 overexpression in the absence of either BRCA1 or -2 may not necessarily confer the same predictive and prognostic implications as in sporadic cancers with intact BRCA1/2. We therefore performed an immunohistochemical analysis of RAD21 expression in a cohort of familial breast cancers. We also postulated that enhanced RAD21 expression may be associated with changes in both DNA copy number and reduced expression of microRNAs (miRNAs), and therefore correlated

RAD21 expression with genomic changes and miRNAs predicted to target RAD21.

## Materials and methods

### Patients

Breast cancer specimens were collected from a previously characterized cohort of 139 female patients from the kConFab family breast cancer registry (Table 1) [15]. Classification of BRCA1, BRCA2, and BRCAX status was performed as described previously [15]. The flow of patients according to the REMARK guidelines (Additional file 1 Table S1) [16]. Of the 139 cases, 18 cases were excluded because of the lack of tissue available for array construction, and a further 27 cases were excluded because of the absence of tumor on the array stained for RAD21. The final cohort was composed of 94 cases, which included 28 BRCA1, 27 BRCA2, and 39 BRCAX cases. This study has ethics committee approval (Peter MacCallum Cancer Centre, 09/36). Consent to participate in the study and consent to publish were obtained by kConFab in accordance with its family-enrollment and data-collection guidelines [17]. Patients were followed up for a median period of 64.0 months (range, 0.4 to 299.0 months). During this time, 38 patients relapsed, and 33 died of breast cancer (deaths unrelated to breast cancer were censored). Relapse-free survival was defined as the time to first reappearance of tumor at any site after definitive treatment, whereas breast cancer-specific survival was defined as time from primary surgical excision to breast cancer-related death. All patients with HER2-positive tumors were diagnosed prior to 2000, and did not receive trastuzumab therapy.

### Immunohistochemistry

Tumor-tissue microarrays (1-mm cores), with a twofold redundancy, were prepared from archival formalin-fixed, paraffin-embedded tissue blocks. RAD21 staining was performed as previously described by using a rabbit polyclonal anti-RAD21 antibody (1:200; Abcam, Cambridge, UK) [13]. Nuclear RAD21 expression was assessed for intensity (0, no staining; 1, weak; 2, moderate; 3, strong) and the percentage of positive cells (0, 0; 1, < 10%; 2, 10% to 50%; 3, 51% to 80%; 4, > 80% positive cells). The scores for intensity and percentage were added, and a cut-off of 6, the median, was used to define two approximately equal sized groups of patients (with low and high RAD21 expression) for subsequent statistical analyses [13].

HER2 chromogenic *in situ* hybridization (CISH) and immunoperoxidase staining for ER $\alpha$ , PgR, HER2, CK5/6, and EGFR were performed for all tumors. By using stratification of intrinsic phenotypes based on Nielsen *et al.* [18], we placed tumors into luminal (ER positive, HER2 negative, cytokeratin (CK) 5/6 negative or positive),

**Table 1 Clinical and tumor characteristics**

	BRCA1 <i>n</i> (%)	BRCA2 <i>n</i> (%)	BRCAX <i>n</i> (%)	All familial <i>n</i> (%)
<b>Age</b>				
≤ 50 years	25 (89%)	15 (56%)	24 (61%)	64 (68%)
> 50 years	3 (11%)	12 (44%)	15 (39%)	30 (32%)
<b>Tumor size</b>				
< 20 mm	20 (71%)	13 (52%)	16 (47%)	49 (56%)
> 20 mm	8 (29%)	12 (48%)	18 (53%)	38 (44%)
Unknown	0	2	5	7
<b>Nodal status</b>				
Negative	26 (93%)	19 (70%)	25 (64%)	70 (74%)
Positive	2 (7%)	8 (30%)	14 (36%)	24 (26%)
Unknown	0	0	0	0
<b>Grade</b>				
I	0	1 (4%)	2 (6%)	3 (4%)
II	2 (9%)	10 (44%)	8 (24%)	20 (25%)
III	21 (93%)	12 (52%)	23 (70%)	56 (71%)
Unknown	5	4	6	15
<b>ERα</b>				
Negative	23 (85%)	5 (21%)	11 (32%)	39 (46%)
Positive	4 (15%)	19 (79%)	23 (68%)	46 (54%)
Unknown	1	3	5	9
<b>PgR</b>				
Negative	23 (85%)	9 (38%)	16 (47%)	48 (56%)
Positive	4 (15%)	15 (62%)	18 (53%)	37 (44%)
Unknown	1	3	5	9
<b>HER2 status</b>				
Negative	26 (100%)	22 (100%)	28 (85%)	76 (94%)
Positive	0	0	5 (15%)	5 (6%)
Unknown	2	5	6	13
<b>Endocrine therapy</b>				
Not given	22 (96%)	16 (76%)	21 (64%)	59 (77%)
Given	1 (4%)	5 (24%)	12 (36%)	18 (23%)
Unknown	5	6	6	17
<b>Chemotherapy</b>				
Not given	8 (30%)	15 (60%)	13 (39%)	16 (42%)
Given	19 (70%)	10 (40%)	20 (61%)	49 (58%)
Unknown	1	2	6	9

*n* = 94.

basal (HER2 and ER negative; CK5/6 positive), HER2 (HER2 positive, ER and CK5/6 negative or positive), and null/negative (HER2, ER, and CK5/6 negative). For HER2, EGFR, and CK5/6, the cut-offs were derived from Neilsen *et al.* [18]. An Allred score of > 2/8 was considered positive for ERα [19].

#### Gene-expression and copy-number analysis of *RAD21*

Gene-expression and copy-number variation (CNV) data from a cohort of familial breast cancers were derived from a previous study, by using the Illumina Human-6 BeadArray (Illumina, San Diego, CA, USA) and the CNV370 SNP array (Illumina), respectively [20]. In this study, the familial tumors were classified into one of the breast tumor subtypes: basal-like, luminal A, luminal B, HER2-positive, and normal-like. These data were used to determine the expression of *RAD21* in 75 (19 BRCA1, 30 BRCA2, 25 BRCAX, one unknown) familial breast cancers. Of these 75 cases, 34 had matching CNV data available (11 BRCA1, nine BRCA2, and 14 BRCAX), and 18 had matching protein-expression data as assessed by immunohistochemistry (seven BRCA1, eight BRCA2, three BRCAX). Copy number was determined by using SNP-CGH data for 34 tumors. All data were imported and visualized in Beadstudio v3.2. The logR ratio was used to perform frequency plots of genomic gain or loss by using CGH explorer [21]. R was used to perform SOMATICS [22] to identify regions containing genomic aberrations. The copy number of *RAD21* in each tumor was inferred from the average logR value of 22 single-nucleotide polymorphisms (SNPs), which were within the *RAD21* intron (*n* = 2) or in sequences flanking the gene (*n* = 20). All these data are available on GEO (Accession Number GSE19177).

#### microRNA and gene-expression data mining

Matching gene-expression and miRNA profiles of 215 sporadic breast cancer specimens were obtained by mining a microarray dataset described by Buffa *et al.* [23], in GEO (Accession number GSE22220), samples BCmicroRNA 1 to 110). Normalized log2 signal intensities were obtained after background subtraction and quantile normalization, as previously described by Buffa *et al.* [23]. This signal intensity for *RAD21* was used for survival analysis, as outlined subsequently.

#### microRNA microarray

miRNA expression was assessed for 11 BRCA1 basal breast cancers and 13 normal breast specimens via microarrays. For each sample, 250 ng of total RNA was labeled and hybridized on Human v2 MicroRNA Expression BeadChips (Illumina). The BeadChips were scanned with the Illumina iScan Reader [24]. Data were imported into GenomeStudio (Illumina), from which raw data with background subtraction were exported to the PARTEK Genomics Suite (St. Louis, MO, USA) for further analysis. Raw probe intensities were shifted, such that the minimum probe intensity for each sample was equal to 1. All values were transformed by taking logs (base 2), followed by quantile normalization [25,26].

Probe mapping for Illumina MicroRNA Expression v2 BeadChips was based on miRBase v.12.0 [27].

### Statistical analysis

Correlations were evaluated by using the Mann-Whitney  $U$  or  $\chi^2$  tests where appropriate. Kaplan-Meier survival curves were calculated for breast cancer-specific death and were compared with the log rank test. The Cox proportional hazard regression model was used to identify independent prognostic factors for breast cancer-specific survival. Analyses were performed with SPSS 16.0 software (SPSS Inc., 233 South Wacker Drive, Chicago, IL, USA). A two-tailed  $P$  value test was used in all analyses, and a  $P$  value of less than 0.05 was considered statistically significant.

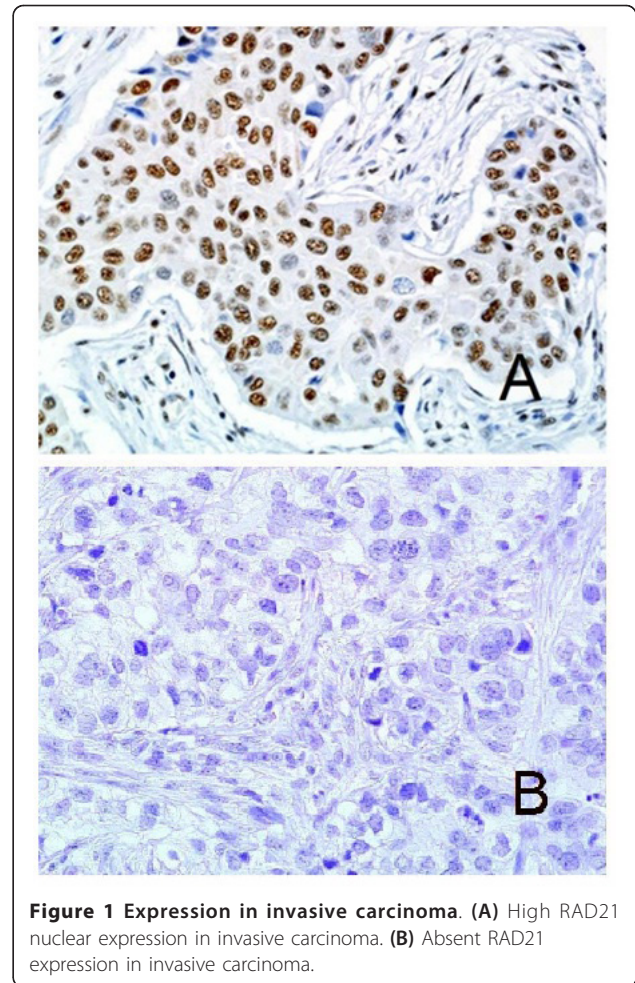
### Results

#### RAD21 protein expression in familial breast cancers, their relation with intrinsic subtypes and clinicopathologic parameters

The median age of the cohort was 45.6 years. No difference was found in breast cancer-specific survival when stratified by age, by using a cut-off of 40 years ( $P = 0.442$ ). The tumors in our cohort showed either absent, low, or high RAD21 expression (as defined earlier; Figure 1). With direct comparison of individual expression scores or a median cut-off score, no significant differences in nuclear RAD21 expression were observed between BRCA1 (12 (43%) of 28), BRCA2 (12 (44%) of 27), and BRCAX cancers (12 (33%) of 39) ( $P = 0.598$ ) (Additional file 2 Table S2a). Similarly, no differences in Rad21 expression were seen within the intrinsic breast cancer subtypes (luminal, 17 (42%) of 41; basal, 14 (44%) of 32; HER2, two (50%) of four; and null, one (20%) of five;  $P = 0.768$ ) (Additional file 2 Table S2b). No correlation was seen between RAD21 expression and tumor grade, size, lymph node status, or ER or HER2 status (all  $P > 0.05$ , Table 2).

#### RAD21 expression and survival in familial breast cancers

A significant correlation was noted between high-RAD21 expression and shorter relapse-free survival ( $P = 0.038$ ) and breast cancer-specific survival ( $P = 0.001$ , Figure 2A) across the entire familial cancer group. Correlation with breast cancer-specific survival was confirmed in a multivariate analysis (including ER, HER2, tumor grade, size, lymph-node status), with RAD21 as a continuous score out of seven (homologous recombination (HR) = 1.66;  $P = 0.003$ ; 95% CI, 1.19 to 2.32; Table 3). Similar to our previous findings in sporadic breast cancers [13], high RAD21 expression correlated with poorer relapse-free survival ( $P = 0.008$ ) and breast cancer-specific survival for grade 3 familial



**Figure 1 Expression in invasive carcinoma. (A)** High RAD21 nuclear expression in invasive carcinoma. **(B)** Absent RAD21 expression in invasive carcinoma.

breast cancers ( $P = 0.009$ , Figure 2B) [13], but not for grade 1 and 2 cancers ( $P = 0.065$  and  $0.090$ , respectively).

Correlation of RAD21 expression with breast cancer-specific survival was further assessed for BRCA1, BRCA2, and BRCAX cancers. Interestingly, high RAD21 expression correlated with poorer survival in BRCA2 ( $P = 0.006$ ) (Figure 2C) and BRCAX cancers ( $P = 0.008$ ) (Figure 2D), but not in BRCA1 cancers ( $P = 0.71$ ) (Figure 2E). When the intrinsic subtypes were individually analyzed, high RAD21 expression correlated with worse survival in luminal breast cancers ( $P = 0.010$ ) (Figure 2F). A similar divergence of the survival curves was observed in basal cancers, although this was not statistically significant ( $P = 0.075$ ) (Figure 2G). Insufficient numbers of HER2 and null-type cancers were available for survival analyses to be performed ( $n = 2$  and  $3$ , respectively).

High RAD21 expression correlated with shorter survival for patients treated with adjuvant chemotherapy ( $P = 0.036$ ) (Figure 2H). Expression of RAD21 did not

**Table 2 Correlation of RAD21 expression with clinicopathologic features (n = 94)**

	RAD21 negative	RAD21 positive	P value
<b>Age</b>			
≤ 50 years	40 (70%)	24 (65%)	0.589
> 50 years	17 (30%)	13 (35%)	
<b>Tumor size</b>			
< 20 mm	29 (55%)	20 (59%)	0.706
> 20 mm	24 (45%)	14 (41%)	
<b>Nodal status</b>			
Negative	42 (74%)	28 (76%)	0.829
Positive	15 (26%)	9 (24%)	
<b>Grade</b>			
I	2 (5%)	1 (3%)	0.128
II	7 (16%)	13 (36%)	
III	34 (79%)	22 (61%)	
<b>ERα</b>			
Negative	22 (44%)	17 (49%)	0.677
Positive	28 (56%)	18 (51%)	
<b>HER2 status</b>			
Negative	46 (96%)	30 (91%)	0.366
Positive	2 (4%)	3 (9%)	
<b>Relapse</b>			
Negative	44 (79%)	18 (51%)	0.007
Positive	12 (21%)	17 (49%)	
<b>BCSS</b>			
Negative	45 (80%)	19 (54%)	0.006
Positive	11 (20%)	16 (46%)	

BCSS, Breast cancer-specific survival.

correlate with survival in patients receiving hormonal therapy ( $p = 0.88$ ).

#### Validation of RAD21 expression as a prognostic marker in the cohort of sporadic cancers from Buffa *et al*

The correlation of *RAD21* gene expression to 10-year relapse-free survival was explored in a validation cohort of 215 breast cancer patients, with tumors previously characterized on the Illumina Human RefSeq-8 microarray, by Buffa *et al.* [23]. Kaplan-Meier curves were charted after stratifying the cohort into two groups, with high *RAD21* expression being defined at the 66<sup>th</sup> percentile (that is, top third of tumors). The same analysis was repeated with the cut-off set at the 50<sup>th</sup> percentile. High *RAD21* expression correlated with poorer survival at both cut-offs ( $P = 0.007$  and  $P = 0.024$ , respectively; Figure 3). At a cut-off at the 66<sup>th</sup> percentile, *RAD21* was an independent indicator of 10-year relapse-free survival in a multivariate analysis including ER status, lymph node status, grade, size, and age (HR = 1.62;  $P = 0.046$ ; 95% CI, 1.08 to 2.61).

#### Gene expression correlates with relative copy number and protein expression of RAD21 in familial breast cancers

The cohort of familial tumors was previously analyzed on the basis of gene-expression and copy-number analysis [20]. No significant difference was found in *RAD21* gene expression between BRCA1, BRCA2, and BRCA3 cancers ( $P = 0.170$ , Kruskal-Wallis test). Similarly, no significant difference was noted in *RAD21* copy number among the BRCA subtypes ( $P = 0.141$ ). Similar to our findings in sporadic breast cancers, no difference was seen in *RAD21* copy number between basal-like and luminal cancers ( $P = 0.749$ ). The 34 tumors with both gene-expression and copy-number data showed a significant correlation between *RAD21* expression and estimated copy number ( $r = 0.619$ ;  $P < 0.001$ ) (Figure 4). Five (15%) of the 34 tumors showed a copy-number gain (copy number of 3). Of these five tumors with copy number gain, there were four basal-like tumors and one luminal tumor.

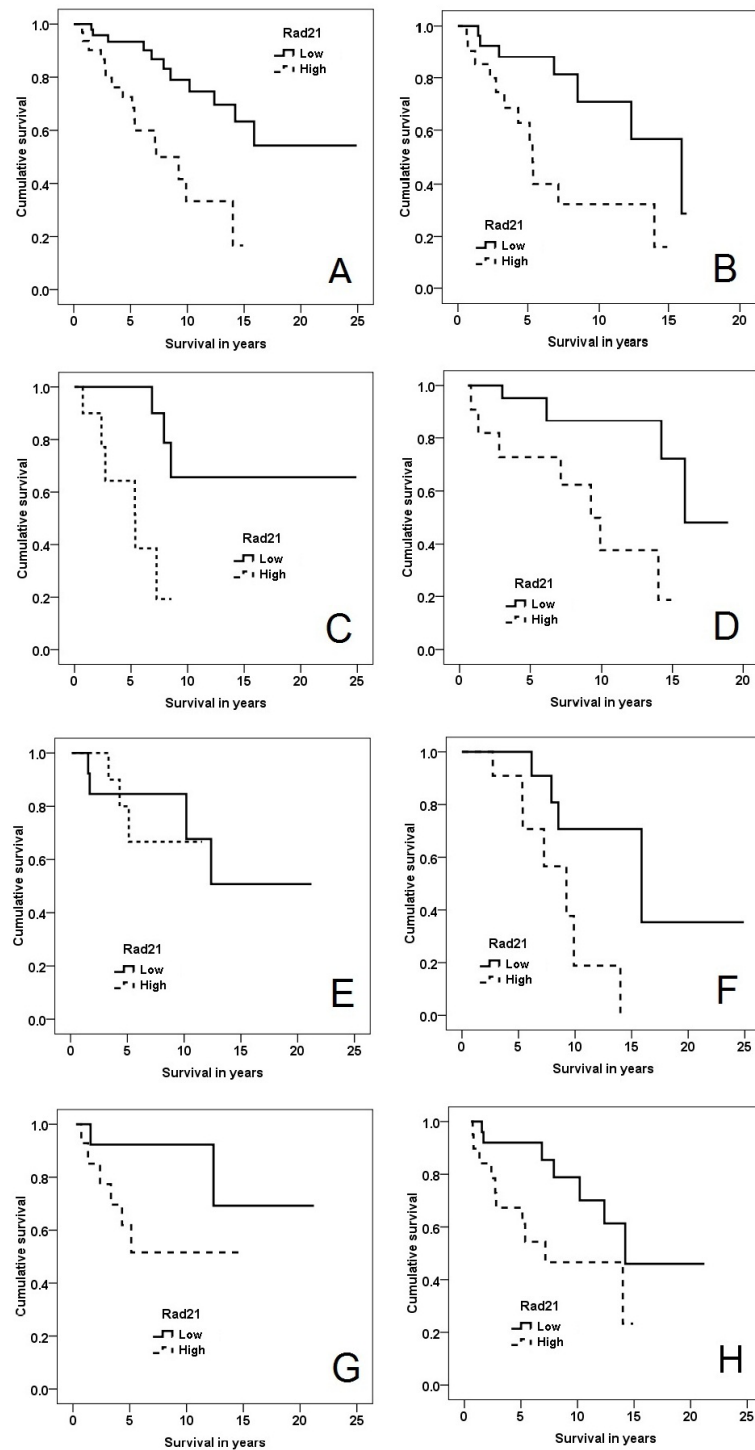
For 18 tumors, *RAD21* gene expression was matched with protein expression, as assessed by immunohistochemistry. This showed a significant correlation between *RAD21* gene expression and *RAD21* protein expression (of seven) ( $r = 0.475$ ;  $P = 0.047$ ) (Figure 5).

#### RAD21 expression is linked with genomic instability in familial breast cancers

Thirty-four tumors were analyzed for genomic change by using SNP-CGH profiling, as previously described by Waddell *et al.* [20]. SOMATICS [22] was used to identify copy-number change and copy-neutral loss of heterozygosity. The total number of chromosomal aberrations and the total number of base pairs affected by genomic change were compared between high *RAD21* (top third;  $n = 11$ ) and low *RAD21* (bottom third;  $n = 11$ ) expressing tumors, as assessed by gene-expression analysis. Tumors with high *RAD21* expression had a higher number of base pairs affected by genomic change (mean =  $1.92 \times 10^9$ ), compared with tumors with low *RAD21* expression (mean =  $1.26 \times 10^9$ ;  $P = 0.048$ ; Figure 6). Although no significant difference was found in the total number of chromosomal aberrations between high and low *RAD21*-expressing tumors ( $P = 0.660$ ), the difference in the number of base pairs affected by genomic change suggests that *RAD21* expression is linked with a higher level of genomic instability.

#### mir-299-5p is predicted to target RAD21 and inversely correlates with RAD21 expression in sporadic breast cancers

A search of potential miRNAs that may target *RAD21* was performed on MicroCosm Targets, version 5 [28].



**Figure 2 Kaplan-Meier curves, breast cancer-specific overall survival stratified by Rad21 expression. (A)** All familial cancers ( $P = 0.001$ ). **(B)** Grade 3 familial cancers ( $P = 0.009$ ). **(C)** BRCA2 cancers ( $P = 0.006$ ). **(D)** BRCA1 cancers ( $P = 0.008$ ). **(E)** Familial luminal cancers ( $P = 0.010$ ). **(F)** Familial basal cancers ( $P = 0.075$ ). **(G)** Familial cancers treated with adjuvant chemotherapy ( $P = 0.036$ ).

**Table 3 Cox regression model: factors influencing breast cancer-specific survival in familial breast cancers**

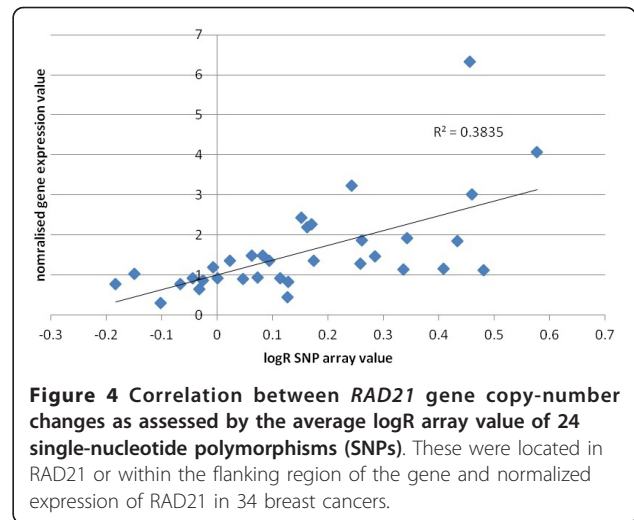
	<i>P</i> value	Hazard ratio	95% Confidence interval
<b>RAD21 (score out of 7)</b>	0.003	1.66	1.19-2.32
<b>ER</b>	0.088	0.33	0.09-1.18
<b>HER2</b>	0.088	2.24	0.51-9.97
<b>Grade</b>	0.030	5.66	1.18-27.22
<b>Size</b>	0.691	0.99	0.97-1.02
<b>Lymph-node status</b>	0.003	6.05	1.82-20.15

The correlation between *RAD21* gene expression and the expression of 15 miRNAs predicted to target *RAD21* was interrogated by mining matching gene-expression and miRNA-array data from the cohort of Buffa *et al.* (108 tumors from miRNA arrays 1 to 110) [23]. Of the 15 miRNAs examined, mir-299-5p inversely correlated with *RAD21* expression (Pearson  $r = -0.294$ ;  $P = 0.002$ ).

### Discussion

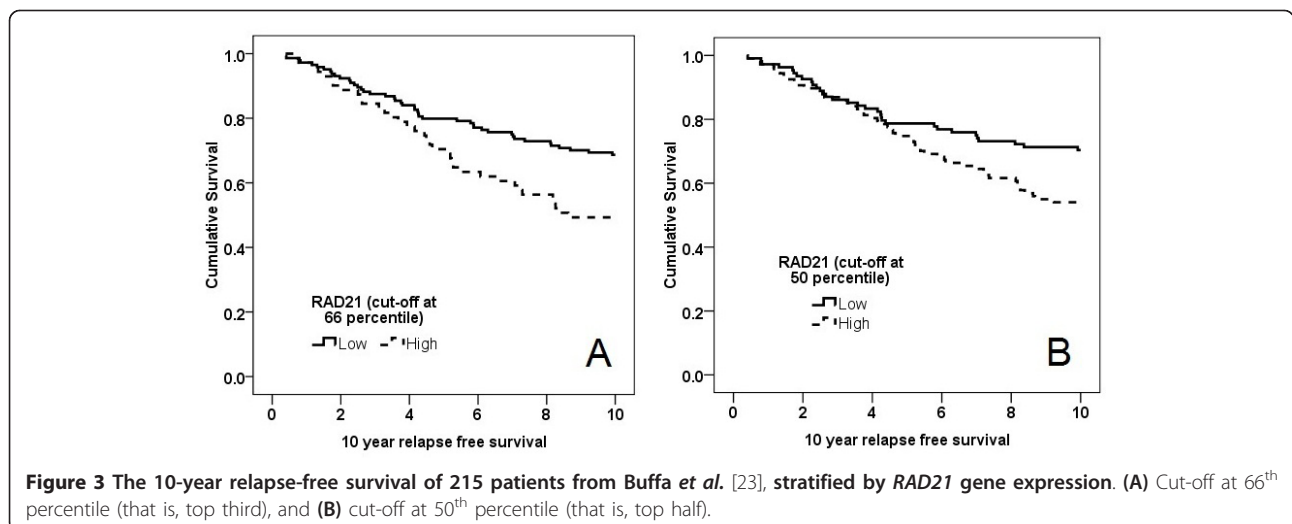
Although *RAD21* expression is associated with a poor prognosis and treatment resistance in sporadic breast cancers, the role of *RAD21* in familial breast cancers is unclear, as *RAD21* expression in the absence of either functional *BRCA1* or 2 may not necessarily confer the same predictive and prognostic implications as in sporadic cancers with intact *BRCA1/2*. We therefore performed the first analysis of *RAD21* in a cohort of fully characterized familial breast cancers and investigated potential mechanisms that may mediate *RAD21* levels, including DNA copy number and reduced expression of targeting microRNA.

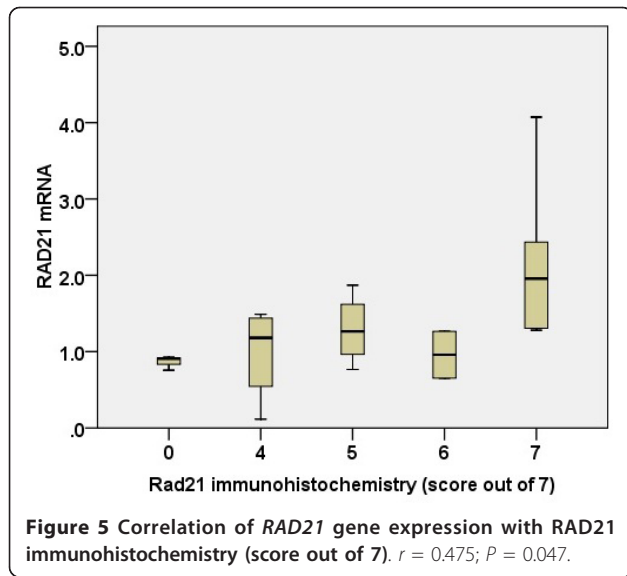
Our findings in a cohort of familial breast cancers recapitulated our previous findings in sporadic breast



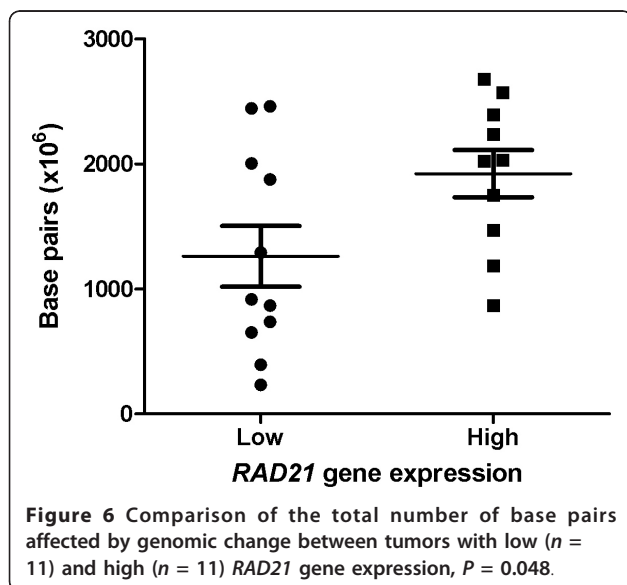
cancers, with enhanced expression of *RAD21* occurring in a subset of tumors regardless of grade, size, or intrinsic subtype [13]. Similarly, *RAD21* expression conferred a poor prognosis in grade 3, but not in grade 1 or 2 cancers. The previously demonstrated correlation of *RAD21* with survival was also confirmed in the cohort of Buffa *et al.* of sporadic cancers assessed with gene-expression arrays and in our cohort of familial breast cancers.

We found that *RAD21* expression does not correlate with *BRCA* status. However, its expression associates with a poor prognosis in *BRCA2* and *BRCAX* cancers. This finding is consistent with our previous findings in sporadic cancers [13]. Interestingly, *RAD21* was not associated with a poor prognosis in *BRCA1* cancers. How *RAD21* expression influences the poor prognosis in *BRCA2* and *BRCAX* but not in *BRCA1* patients remains to be determined. This finding is likely to be





explained by the final roles of RAD21 and BRCA1/2 in homologous recombination (HR). Like BRCA1 and BRCA2, cohesins are important regulators of genomic stability. Cohesins facilitate error-free repair of double-stranded breaks (DSBs) in DNA by HR, possibly by promoting the alignment and cohesion of sister chromatids [8]. Defects in HR result in DSB repair through the alternative, error-prone, nonhomologous end-joining and single strand-annealing pathways; this may lead to deletions, translocations, and chromosomal instability [29]. HR is particularly important in the repair of DNA damage caused by chemotherapeutic agents and radiotherapy. Although both BRCA1 and BRCA2 participate in HR, evidence from genetic studies suggests that



BRCA1 functions upstream of BRCA2 [30]. Hence, in the setting of BRCA1 cancers in which DSB repair has already been compromised by deleterious BRCA1 mutations, overexpression of RAD21 may have no effect on tumor behavior. Furthermore, removal of the cohesin complex after DNA repair requires proteolytic cleavage of RAD21 by caspase 3 [31]. Activation and translocation of caspase 3 into the nucleus requires functioning wild-type BRCA1 and is inhibited by mutated BRCA1 [32]. Failure to cleave RAD21 due to BRCA1-dependent caspase 3 inhibition may explain loss of HR repair in BRCA1 but not in BRCA2 cancers.

Another possible explanation relates to the emerging role of cohesin as a key regulator of gene transcriptions [33,34]. RAD21 is implicated in mediating estrogen-regulated transcription through ER $\alpha$  in MCF7 breast cancer cells [9]. Hence RAD21 expression may influence a subset of gene expression through interplay with ER $\alpha$ , and thereby contribute to the poor prognosis in BRCA2 and BRCX patients. This notion is consistent with ER status in our cohort, in which 85% of BRCA1 tumors are ER $\alpha$  negative. Although RAD21 may be implicated in ER-regulated gene expression, functional genomic screens performed on estrogen-dependent cell lines have so far yielded conflicting results regarding the role of RAD21 in tamoxifen resistance. Silencing of *RAD21* via sh-RNA suggests that RAD21 expression is associated with sensitivity to tamoxifen [35], whereas transduction by retroviral cDNA suggests that RAD21 expression is associated with resistance [36]. Neither of these findings appears to be reflected by our clinical cohorts, as Rad21 expression did not correlate with survival in either sporadic ( $P = 0.231$ ) or familial ( $P = 0.881$ ) breast cancers treated with hormone therapy.

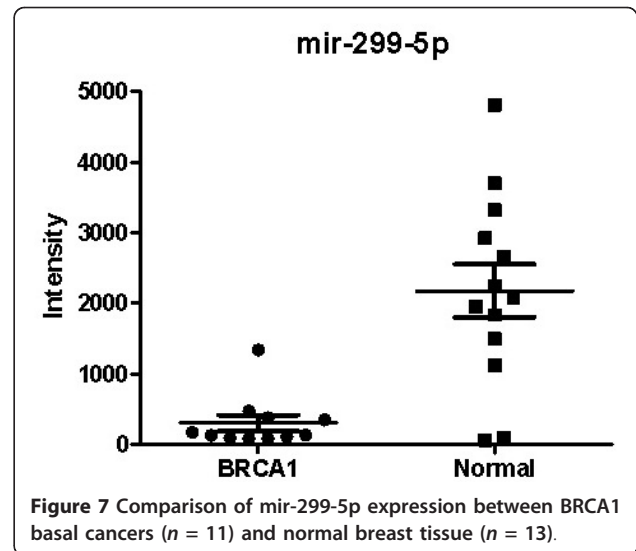
Our previous study demonstrated that RAD21 expression is associated with chemotherapy resistance in cell lines and in a cohort of sporadic cancers [13]. Similarly, in our cohort of familial cancers receiving adjuvant chemotherapy, RAD21 expression correlated with a poorer prognosis. This is despite our finding that RAD21 does not correlate with survival in BRCA1 cancers and the higher use of chemotherapy in BRCA1 cancers. This further supports our argument that BRCA1 cancers behave differently from other cancers in response to RAD21 overexpression.

Although the function of RAD21 has been extensively investigated in the literature, relatively little is known regarding the regulation of its expression. In keeping with our previous findings in sporadic breast cancers [13], *RAD21* gene expression correlated with copy number in our cohort of familial breast cancers. This is further supported by the positive correlation between *RAD21* gene expression and protein expression, as assessed by immunohistochemistry in matched tumor

samples. Increased *RAD21* expression is associated with increased numbers of base pairs being affected by genomic change, which suggests that increased expression is linked to genomic instability. The effect of genomic changes on *RAD21* expression appears to be independent of *BRCA* status and intrinsic tumor subtype. *RAD21* upregulation may be an early event that occurs before tumor invasion. This is supported by the presence of increased *RAD21* expression in 46% of the DCIS cases we previously examined [13]. In addition, interrogation of the cohort of Schuetz *et al.* [37] showed no difference in *RAD21* expression between 14 micro-dissected matching invasive and *in situ* ductal carcinoma samples ( $P = 0.154$ ).

In addition to genomic changes, we postulated that other mechanisms are likely to control *RAD21* expression, such as through targeting microRNAs. Of the 15 miRNAs predicted to target *RAD21*, *mir-299-5p* expression significantly inversely correlated with *RAD21* expression. Reduced circulating *mir-299-5p* was previously demonstrated in the serum of breast cancer patients [38], compared with normal controls. Silencing of *mir-299-5p* has also been implicated in the pathogenesis of oral squamous cell carcinomas [39]. Limited data derived from breast cancer cell lines showed that *mir-299-5p* knockdown is associated with upregulation of osteopontin [40], a glycoprotein involved in invasion, metastasis, and resistance to radiotherapy and chemotherapeutic agents [41,42]. Aberrant regulation of *RAD21* by *mir-299-5p* is also supported by a comparison of 11 *BRCA1* basal and 13 normal breast samples, in which a sevenfold reduction in *mir-299-5p* was observed between *BRCA1* cancers and normal breast tissue ( $P = 0.0002$ ; Figure 7). Nevertheless, although *mir-299-5p* is clearly reduced in breast cancer, this appears appear to play a relatively minor role in regulating *RAD21* expression ( $r^2 = 0.0864$ ;  $P = 0.002$ ) compared with genomic changes in copy number ( $r^2 = 0.286$ ;  $P < 0.001$ ).

In summary, our findings show that *RAD21* expression is associated with a poorer prognosis in *BRCA2* and *BRCAX*, but not in *BRCA1* cancers. Because increased *RAD21* expression may confer resistance to DNA-damaging agents, alternative treatment strategies may be useful in *RAD21*-positive *BRCA2* and *BRCAX* cancers. This may include dosage intensification and the use of chemotherapeutic agents with alternative modes of action. The efficacy of poly-ADP ribose polymerase (PARP)-inhibitor therapy is dependent on defective HR repair [43,44]. In view of the role of *RAD21* in increasing HR activity [33], tumor *RAD21* status may be particularly relevant in patients being considered for PARP-inhibitor therapy.



**Figure 7** Comparison of *mir-299-5p* expression between *BRCA1* basal cancers ( $n = 11$ ) and normal breast tissue ( $n = 13$ ).

## Conclusions

*RAD21* expression in familial cancers is reflected by expression changes in copy-number expression. It is also inversely correlated with *mir-299-5p*, the expression of which is suppressed in breast cancers. Similar to our earlier findings in sporadic breast cancers, increased *RAD21* expression in *BRCA2* and *BRCAX* cancers confers a poor prognosis and resistance to DNA-damaging chemotherapeutic agents. This association does not apply to *BRCA1* cancers, in which repair by HR between sister chromatids may be compromised by *BRCA1* deficiency. *RAD21* is thus a potential *BRCA1/2* mutation status-dependent predictive and prognostic marker in familial breast cancers.

## Additional material

**Additional file 1: Table S1.** Flow of familial breast cancer patients through the study, according to REMARK criteria.

**Additional file 2: Table S2.** Nuclear *RAD21* expression in familial breast cancers (score out of 7).

## Abbreviations

*BRCA1*: breast cancer 1, early onset; *BRCA2*: breast cancer 2, early onset; CGH: chromogenic hybridization; CISH: chromogenic *in situ* hybridization; CNV: copy-number variation; DCIS: ductal carcinoma *in situ*; DSB: double-stranded break; ER: estrogen receptor; HER2: human epidermal growth receptor 2; HR: homologous recombination; miRNA: microRNA; PARP: poly-ADP ribose polymerase; SNP: single-nucleotide polymorphism.

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#### Authors' contributions

MY contributed to the conception and design of the study, analyzed the data, and drafted and revised the manuscript. HX contributed to the conception and design of the study, performed the immunohistochemistry, and revised the manuscript. NW performed the CGH and mRNA array experiments. KS performed the miRNA experiments. IH contributed to the statistical analysis and design of the miRNA array experiments. kConFab contributed to the provision of study materials and the collection of clinicopathologic and follow-up survival data. MJM contributed to the conception and design of the study and the analysis, drafting, and revision of the manuscript. SBF contributed to the conception and design of the study, provision of study materials, analysis, drafting, and revision of the manuscript. All authors read and approved the final manuscript.

#### Competing interests

The authors declare that they have no competing interests.

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#### References

- Palacios J, Honrado E, Osorio A, Cazorla A, Sarrío D, Barroso A, Rodríguez S, Cigudosa JC, Diez O, Alonso C, Sanchez L, Rivas C, Benitez J: **Immunohistochemical characteristics defined by tissue microarray of hereditary breast cancer not attributable to BRCA1 or BRCA2 mutations: differences from breast carcinomas arising in BRCA1 and BRCA2 mutation carriers.** *Clin Cancer Res* 2003, **9**:3606-3614.
- Lakhani SR, Gusterson BA, Jacquemier J, Sloane JP, Anderson TJ, van der Vijver MJ, Venter D, Freeman A, Antoniou A, McGuffog L, Smyth E, Steel CM, Haites N, Scott RJ, Goldgar D, Neuhausen S, Daly PA, Ormiston W, McManus R, Scherneck S, Ponder BA, Futreal PA, Peto J, Stoppa-Lyonnet D, Bignon YJ, Stratton MR: **The pathology of familial breast cancer: histological features of cancers in families not attributable to mutations in BRCA1 or BRCA2.** *Clin Cancer Res* 2000, **6**:782-789.
- Palacios J, Honrado E, Osorio A, Cazorla A, Sarrío D, Barroso A, Rodríguez S, Cigudosa JC, Diez O, Alonso C, Lerma E, Dopazo J, Rivas C, Benitez J: **Phenotypic characterization of BRCA1 and BRCA2 tumors based in a tissue microarray study with 37 immunohistochemical markers.** *Breast Cancer Res Treat* 2005, **90**:5-14.
- Foulkes WD, Stefansson IM, Chappuis PO, Begin LR, Goffin JR, Wong N, Trudel M, Akslén LA: **Germline BRCA1 mutations and a basal epithelial phenotype in breast cancer.** *J Natl Cancer Inst* 2003, **95**:1482-1485.
- Lakhani SR, Reis-Filho JS, Fulford L, Penault-Llorca F, van der Vijver M, Parry S, Bishop T, Benitez J, Rivas C, Bignon YJ, Chang-Claude J, Hamann U, Cornelisse CJ, Devilee P, Beckmann MW, Nestle-Kramling C, Daly PA, Haites N, Varley J, Laloo S, Radice P, Scherneck S, Sobol H, Jacquemier J, Wagner T, Peto J, Stratton MR, McGuffog L, Easton DF: **Prediction of BRCA1 status in patients with breast cancer using estrogen receptor and basal phenotype.** *Clin Cancer Res* 2005, **11**:5175-5180.
- Armes JE, Egan AJ, Southey MC, Dite GS, McCredie MR, Giles GG, Hopper JL, Venter DJ: **The histologic phenotypes of breast carcinoma occurring before age 40 years in women with and without BRCA1 or BRCA2 germline mutations: a population-based study.** *Cancer* 1998, **83**:2335-2345.
- Nasmyth K, Haering CH: **Cohesin: its roles and mechanisms.** *Annu Rev Genet* 2009, **43**:525-558.
- Peters JM, Tedeschi A, Schmitz J: **The cohesin complex and its roles in chromosome biology.** *Genes Dev* 2008, **22**:3089-3114.
- Schmidt D, Schwalie PC, Ross-Innes CS, Hurtado A, Brown GD, Carroll JS, Flicek P, Odom DT: **A CTCF-independent role for cohesin in tissue-specific transcription.** *Genome Res* 2010, **20**:578-588.
- Rollins RA, Korom M, Aulner N, Martens A, Dorsett D: **Drosophila nipped-B protein supports sister chromatid cohesion and opposes the stromalin/Scc3 cohesion factor to facilitate long-range activation of the cut gene.** *Mol Cell Biol* 2004, **24**:3100-3111.
- Rhodes DR, Yu J, Shanker K, Deshpande N, Varambally R, Ghosh D, Barrette T, Pandey A, Chinnaiyan AM: **Large-scale meta-analysis of cancer microarray data identifies common transcriptional profiles of neoplastic transformation and progression.** *Proc Natl Acad Sci USA* 2004, **101**:9309-9314.
- Oishi Y, Nagasaki K, Miyata S, Matsuura M, Nishimura S, Akiyama F, Iwai T, Miki Y: **Functional pathway characterized by gene expression analysis of supraclavicular lymph node metastasis-positive breast cancer.** *J Hum Genet* 2007, **52**:271-279.
- Xu X, Yan M, Natrajan R, Yan Y, Swagemakers S, Tomaszewski JM, Verschoor S, Millar EKA, van der Spek P, Reis-Filho J, Ramsay RG, O'Toole SA, McNeil CM, Sutherland RL, Mackay MJ, Fox SB: **Enhanced Rad21 cohesin expression confers poor prognosis and resistance to chemotherapy in high grade luminal, basal and HER2 breast cancers.** *Breast Cancer Res* 2011, **32**:13 R9.
- Atienza JM, Roth RB, Rosette C, Smylie KJ, Kammerer S, Rehbock J, Ekblom J, Denissenko MF: **Suppression of RAD21 gene expression decreases cell growth and enhances cytotoxicity of etoposide and bleomycin in human breast cancer cells.** *Mol Cancer Ther* 2005, **4**:361-368.
- Yan M, Rayoo M, Takano EA, Fox SB: **Nuclear and cytoplasmic expressions of ERbeta1 and ERbeta2 are predictive of response to therapy and alters prognosis in familial breast cancers.** *Breast Cancer Res Treat* 2011, **126**:395-405.
- McShane LM, Altman DG, Sauerbrei W, Taube SE, Gion M, Clark GM: **Reporting recommendations for tumor marker prognostic studies.** *J Clin Oncol* 2005, **23**:9067-9072.
- Kathleen Cuningham Foundation Consortium for Research into Familial Breast Cancer, Enrolment of Families and Data Collection. [http://www.kconfab.org/Index.shtml].
- Nielsen TO, Hsu FD, Jensen K, Cheang M, Karaca G, Hu Z, Hernandez-Boussard T, Livasy C, Cowan D, Dressler L, Akslén LA, Ragaz J, Gown AM, Gilks CB, van de Rijn M, Perou CM: **Immunohistochemical and clinical characterization of the basal-like subtype of invasive breast carcinoma.** *Clin Cancer Res* 2004, **10**:5367-5374.
- Leake R, Barnes D, Pinder S, Ellis I, Anderson L, Anderson T, Adamson R, Rhodes T, Miller K, Walker R: **Immunohistochemical detection of steroid receptors in breast cancer: a working protocol; UK Receptor Group, UK NEQAS, The Scottish Breast Cancer Pathology Group, and The Receptor and Biomarker Study Group of the EORTC.** *J Clin Pathol* 2000, **53**:634-635.
- Waddell N, Arnold J, Cocciardi S, da Silva L, Marsh A, Riley J, Johnstone CN, Orloff M, Assie G, Eng C, Reid L, Keith P, Yan M, Fox S, Devilee P, Godwin AK, Hogervorst FB, Couch F, Grimmond S, Flanagan JM, Khanna K, Simpson PT, Lakhani SR, Chenevix-Trench G: **Subtypes of familial breast tumours revealed by expression and copy number profiling.** *Breast Cancer Res Treat* 2010, **123**:661-677.
- Lingjaerde OC, Baumbusch LO, Liestol K, Glad IK, Borresen-Dale AL: **CGH-Explorer: a program for analysis of array-CGH data.** *Bioinformatics* 2005, **21**:821-822.
- Assie G, LaFramboise T, Platzer P, Bertherat J, Stratakis CA, Eng C: **SNP arrays in heterogeneous tissue: highly accurate collection of both germline and somatic genetic information from unpaired single tumor samples.** *Am J Hum Genet* 2008, **82**:903-915.
- Buffa FM, Camps C, Winchester L, Snell CE, Gee HE, Sheldon H, Taylor M, Harris AL, Ragoussis J: **microRNA associated progression pathways and potential therapeutic targets identified by integrated mRNA and**

- microRNA expression profiling in breast cancer. *Cancer Res* 2011, **71**:5635-5645.
24. Chen J, Lozach J, Garcia EW, Barnes B, Luo S, Mikoulitch I, Zhou L, Schroth G, Fan JB: **Highly sensitive and specific microRNA expression profiling using BeadArray technology.** *Nucleic Acids Res* 2008, **36**:e87.
  25. Pradervand S, Weber J, Thomas J, Bueno M, Wirapati P, Lefort K, Dotto GP, Harshman K: **Impact of normalization on miRNA microarray expression profiling.** *RNA (New York)* 2009, **15**:493-501.
  26. Pradervand S, Weber J, Lemoine F, Consales F, Paillusson A, Dupasquier M, Thomas J, Richter H, Kaessmann H, Beaudoin E, Hagenbuchle O, Harshman K: **Concordance among digital gene expression, microarrays, and qPCR when measuring differential expression of microRNAs.** *BioTechniques* 2010, **48**:219-222.
  27. Griffiths-Jones S, Saini HK, van Dongen S, Enright AJ: **miRBase: tools for microRNA genomics.** *Nucleic Acids Res* 2008, **36**:D154-158.
  28. **Microcosm Targets.** [<http://www.ebi.ac.uk/enright-srv/microcosm/cgi-bin/targets/v5/search.pl>].
  29. Tirkkonen M, Kainu T, Loman N, Johannsson OT, Olsson H, Barkardottir RB, Kallioniemi OP, Borg A: **Somatic genetic alterations in BRCA2-associated and sporadic male breast cancer.** *Genes Chromosomes Cancer* 1999, **24**:56-61.
  30. Wang W: **Emergence of a DNA-damage response network consisting of Fanconi anaemia and BRCA proteins.** *Nat Rev Genet* 2007, **8**:735-748.
  31. Chen F, Kamradt M, Mulcahy M, Byun Y, Xu H, McKay MJ, Cryns VL: **Caspase proteolysis of the cohesin component RAD21 promotes apoptosis.** *J Biol Chem* 2002, **277**:16775-16781.
  32. Martin SA, Ouchi T: **BRCA1 phosphorylation regulates caspase-3 activation in UV-induced apoptosis.** *Cancer Res* 2005, **65**:10657-10662.
  33. Xu H, Tomaszewski JM, McKay MJ: **Can corruption of chromosome cohesin create a conduit to cancer?** *Nat Rev Cancer* 2011, **11**:199-210.
  34. Dorsett D: **Cohesin: genomic insights into controlling gene transcription and development.** *Curr Opin Genet Dev* 2011, **21**:199-206.
  35. Mendes-Pereira AM, Sims D, Dexter T, Fenwick K, Assiotis I, Kozarewa I, Mitsopoulos C, Hakas J, Zvelebil M, Lord CJ, Ashworth A: **Genome-wide functional screen identifies a compendium of genes affecting sensitivity to tamoxifen.** *Proc Natl Acad Sci USA* 2012, **109**:2730-2735.
  36. van Agthoven T, Sieuwerts AM, Meijer D, Meijer-van Gelder ME, van Agthoven TL, Sarwari R, Sleijfer S, Foekens JA, Dorsers LC: **Selective recruitment of breast cancer anti-estrogen resistance genes and relevance for breast cancer progression and tamoxifen therapy response.** *Endocr Relat Cancer* 2010, **17**:215-230.
  37. Schuetz CS, Bonin M, Clare SE, Nieselt K, Sotlar K, Walter M, Fehm T, Solomayer E, Riess O, Wallwiener D, Kurek R, Neubauer HJ: **Progression-specific genes identified by expression profiling of matched ductal carcinomas in situ and invasive breast tumors, combining laser capture microdissection and oligonucleotide microarray analysis.** *Cancer Res* 2006, **66**:5278-5286.
  38. van Schooneveld E, Wouters M, Van der Auwera I, Peeters D, Huget P, van Dam PA, Vermeulen PB, Van Laere SJ, Dirix LY: **Detection of circulating miRNAs in serum from patients with breast cancer and their association with the presence of metastatic disease.** *J Clin Oncol* 2011, **29**:abstr 10506.
  39. Kozaki K, Imoto I, Mogi S, Omura K, Inazawa J: **Exploration of tumor-suppressive microRNAs silenced by DNA hypermethylation in oral cancer.** *Cancer Res* 2008, **68**:2094-2105.
  40. Shevde LA, Metge BJ, Mitra A, Xi Y, Ju J, King JA, Samant RS: **Spheroid-forming subpopulation of breast cancer cells demonstrates vasculogenic mimicry via hsa-miR-299-5p regulated de novo expression of osteopontin.** *J Cell Mol Med* 2010, **14**:1693-1706.
  41. Hähnel A, Wichmann H, Kappler M, Kotszsch M, Vordermark D, Taubert H, Bache M: **Effects of osteopontin inhibition on radiosensitivity of MDA-MB-231 breast cancer cells.** *Radiat Oncol* 2012, **5**:82.
  42. Pang H, Cai L, Yang Y, Chen X, Sui G, Zhao C: **Knockdown of osteopontin chemosensitizes MDA-MB-231 cells to cyclophosphamide by enhancing apoptosis through activating p38 MAPK pathway.** *Cancer Biother Radiopharm* 2011, **26**:165-173.
  43. Alli E, Sharma VB, Sunderesakumar P, Ford JM: **Defective repair of oxidative DNA damage in triple-negative breast cancer confers sensitivity to inhibition of poly(ADP-ribose) polymerase.** *Cancer Res* 2009, **69**:3589-3596.
  44. Ashworth A: **A synthetic lethal therapeutic approach: poly(ADP) ribose polymerase inhibitors for the treatment of cancers deficient in DNA double-strand break repair.** *J Clin Oncol* 2008, **26**:3785-3790.

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