



Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Gray, LEK;Ponsonby, AL;Collier, F;O'Hely, M;Sly, PD;Ranganathan, S;Tang, MLK;Carlin, JB;Saffery, R;Vuillermin, PJ;Burgner, D;Allen, KJ;Pezic, A

Title:

Deserters on the atopic march: Risk factors, immune profile and clinical outcomes of food sensitized-tolerant infants

Date:

2020-06-01

Citation:

Gray, L. E. K., Ponsonby, A. L., Collier, F., O'Hely, M., Sly, P. D., Ranganathan, S., Tang, M. L. K., Carlin, J. B., Saffery, R., Vuillermin, P. J., Burgner, D., Allen, K. J. & Pezic, A. (2020). Deserters on the atopic march: Risk factors, immune profile and clinical outcomes of food sensitized-tolerant infants. *Allergy European Journal of Allergy and Clinical Immunology*, 75 (6), pp.1404-1413. <https://doi.org/10.1111/all.14159>.

Persistent Link:

<https://hdl.handle.net/11343/275271>

1

2 DR. LAWRENCE EK GRAY (Orcid ID : 0000-0002-0773-4037)

3 DR. FIONA COLLIER (Orcid ID : 0000-0002-5438-480X)

4

5

6 Article type : Original Article: Food Allergy and Gastrointestinal Disease

7

8

9 **Deserters on the atopic march:**10 **Risk factors, immune profile and clinical outcomes of food sensitized-tolerant infants**

11

12 Lawrence E. K. Gray^{1,2}, Anne-Louise Ponsonby^{3,4}, Fiona Collier^{1,2,3}, Martin O’Hely^{1,3}, Peter D. Sly^{3,5},
13 Sarath Ranganathan^{3,4,6}, Mimi L. K. Tang^{3,4,6}, John B. Carlin^{3,4,6}, Richard Saffery^{3,4}, Peter J.
14 Vuillermin^{1,2,3} and the BIS Investigator Group[#].

15

16 [#]BIS Investigator Group: David Burgner, Katrina J. Allen, Angela Pezic

17

18 1. Deakin University, School of Medicine, Geelong, Victoria 3220, Australia

19 2. Barwon Health, Geelong, Victoria 3220, Australia

20 3. The Murdoch Children’s Research Institute, Parkville Victoria, 3052, Australia

21 4. The University of Melbourne, Parkville, Victoria 3052, Australia

22 5. University of Queensland, South Brisbane, Queensland 4101, Australia

23 6. The Royal Children’s Hospital, Parkville, Victoria 3052, Australia

24

25 Corresponding author:

Peter J Vuillermin

26

Child Health Research Unit Barwon Health

27

P.O. Box 281 Ryrie St, Geelong, Victoria 3220, Australia

28

Email: peter.vuillermin@deakin.edu.au

29

30 **Authorship statement**

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1111/ALL.14159](https://doi.org/10.1111/ALL.14159)

This article is protected by copyright. All rights reserved

31 All the authors have made substantial contributions to conception and design, or acquisition
32 of data, or analysis and interpretation of data; and been involved in drafting the manuscript
33 or revising it critically for important intellectual content; and given final approval of the
34 version to be published. Each author should have participated sufficiently in the work to
35 take public responsibility for appropriate portions of the content; and have agreed to be
36 accountable for all aspects of the work in ensuring that questions related to the accuracy or
37 integrity of any part of the work are appropriately investigated and resolved.

38

39 **Funding declaration**

40 This study was funded by the National Health and Medical Research Council of Australia
41 (1082307, 1147980, 11511322), the Australian Food Allergy Foundation, The Murdoch
42 Children's Research Institute, Barwon Health and Deakin University.

43

44 **Acknowledgements**

45 We would like to thank the study participants, as well as the entire BIS team which includes
46 interviewers, nurses, computer and laboratory technicians, clerical workers, research
47 scientists, volunteers, managers, and receptionists. We also thank the obstetric and
48 midwifery teams at Barwon Health and Saint John of God Hospital Geelong for their
49 assistance in recruitment and collection of biological specimens.

50

51 **Conflicts of interest**

52 None

53

54

55

56

57

58

59 **Abstract**

60 Background

61 Few studies have investigated the antecedents and outcomes of infants who demonstrate
62 IgE sensitization to foods that they clinically tolerate. Improved understanding of this
63 sensitized-tolerant phenotype may inform strategies for the prevention of food allergy.

64

65 Methods

66 In an Australian birth cohort (n=1074), assembled using an unselected antenatal sampling
67 frame, participants were categorised as non-sensitized (NS), sensitized-tolerant (ST) or food
68 allergic (FA) based on skin-prick testing and food challenge at 12 months of age.

69 Environmental exposures were recorded throughout. Cord blood regulatory T-cell
70 populations were measured at birth. Subsequent childhood allergic disease was assessed by
71 parent report, clinical examination and repeat skin-prick testing.

72

73 Results

74 The covariates of interest varied between NS(n=698), ST(n=27) and FA(n=61) groups as
75 follows, suggesting that across these measures the ST group was more similar to the NS
76 than the FA group: family history of eczema NS 44.6%, ST. 44.6%, FA 65.6%; pet ownership
77 at 12 months: NS 71.5%, ST 81.5%, FA 45.8%; eczema during the first 12 months: NS 19%, ST
78 32%, FA 64%; and aeroallergen sensitization at 4 years: NS 19.1%, ST 28.6%, FA 44.4%. At
79 birth a higher proportion of activated regulatory T cells was associated with ST (OR=2.89,
80 95%CI 1.03–8.16, $P=0.045$).

81

82 Conclusion

83 Food sensitized-tolerance in infancy appears to be associated with a similar pattern of
84 exposures, immunity and outcomes to non-sensitized infants. In addition, we found some
85 evidence that an elevated proportion of activated regulatory T cells at birth was specific to
86 the sensitized-tolerant infants, which may be relevant to suppression of clinical disease.

87

88

89

90

91

92

93
94
95
96
97
98
99
100
101
102
103
104
105
106
107
108
109
110
111
112
113
114
115
116
117
118
119
120
121
122
123
124

Author Manuscript

Graphical abstract

See attached file from Editor.

Highlights

See attached file from Editor

125
126
127
128
129
130
131
132
133
134
135
136
137
138
139
140
141
142
143
144
145
146
147
148
149
150
151
152
153
154
155
156

Key words

Allergy
Food allergy
Immune programming
Regulatory T-cell
Sensitized-tolerant

Abbreviations

BIS – Barwon Infant Study
DAG – directed acyclic graph
Ig – Immunoglobulin
PBS – phosphate buffered saline
SCORAD – scoring atopic dermatitis
SEIFA – Socio-Economic Indexes for Areas
SPT – skin prick testing
Treg – regulatory T-cell

157
158
159
160
161
162
163
164
165
166
167
168
169
170
171
172
173
174
175
176
177
178
179
180
181
182
183
184
185
186
187
188

Introduction

The “atopic march” of childhood allergic disease describes a putative causal pathway linking eczema in infancy to subsequent allergic sensitization to food, food allergy, hayfever, atopic wheeze and asthma.(1-5) Allergic sensitization to a food in infancy is common, occurring in up to 16% of infants at 12 months, but fewer than half the infants who are sensitized to foods are clinically allergic.(6) Many remain tolerant, able to ingest the food(s) without symptoms, despite having generated specific immunoglobulin E antibodies against the implicated food(s)(7, 8) and these infants are termed “sensitized-tolerant”.(9) Limited information exists on the protective factors and early immune profile that may prevent sensitized-tolerant infants from progression to food allergy.

Genetic risk factors for increased likelihood of allergic sensitization and food allergy include male sex and family history of allergic disease(10), but the genetic and demographic factors associated with sensitized-tolerance determined by food challenge remain unknown.

Evidence from studies of aeroallergen-mediated allergic disease indicates that progression from allergic sensitization to clinical expression of allergy may be influenced by environmental factors. For example, a study of children from urban vs. rural areas in Ethiopia found that rural children had an increased incidence of allergic sensitization to dust mite, but greatly decreased risk of wheeze and asthma compared to urban children.(11) This intriguing finding suggests a rural microbial and antigenic environment may promote the sensitized-tolerant phenotype, which is consistent with the notion of benign sensitization.(12)

Maternal exposure to environmental microbes influences antenatal immune programming, altering patterns of early immune response and associated clinical manifestations of allergic disease.(13-20) We, and others, have reported that infants who subsequently develop food allergy have a lower proportion of naïve regulatory T-cells (Treg) at birth(20-22) than non-allergic infants. However, few studies have examined Treg populations in relation to the sensitized-tolerant phenotype.(23-26)

189

190 Currently, there is limited knowledge of the risk factors, environmental exposures, immune
191 profile at birth and subsequent allergic disease outcomes of sensitized-tolerant infants.

192 Indeed, the existing evidence is based largely on studies performed in cohorts at high risk of
193 food allergy, with food allergy status defined by parent report rather than formal food
194 challenge.(27-30)

195

196 The aim of this study was to investigate, in a pre-birth cohort incorporating both skin-prick
197 testing and oral food challenge, the environmental factors, cord blood immune profile and
198 subsequent allergic disease outcomes of food sensitized-tolerant infants.

199

200

201

202

203

204

205

206

207 **Methods**

208 **Enrolment**

209 The Barwon Infant Study (BIS) is a birth cohort study (n=1074) conducted in south-eastern
210 Australia. Details of the study have been reported previously.(31) In brief, mothers were
211 recruited during pregnancy using an unselected antenatal sampling frame. The eligibility
212 criteria included: (i) residents of the defined geographical region in the Barwon area of
213 Victoria, (ii) less than 32-weeks gestation at the time of enrolment, and (iii) planning to give
214 birth at a local hospital. The final inception birth cohort constituted 1064 mothers and 1074
215 infants (10 sets of twins). Data were also collected on baseline characteristics of those
216 mothers who chose not to participate in the study. Ethics approval (10/24) for this study
217 was obtained from the Barwon Health Human Research Ethics committee.

218

219 **Determination of food sensitization phenotype at 12 months**

220 At the 12-month review, infants underwent a skin-prick test (SPT) to 10 food and
221 aeroallergens: cow's milk, egg, peanut, cashew, sesame, house dust mite, cat, dog, rye grass
222 and the fungus *Alternaria tenuis*, with a positive and negative control. A food allergen SPT
223 wheal size of at least 2mm greater than the negative control in the presence of a positive
224 histamine control was defined as food-sensitized. Food-sensitized infants and all
225 participants with food SPT wheals 1mm or greater than the negative control were offered
226 an in-hospital open food challenge. Food challenges were not performed on non-sensitized
227 participants. Participants with a positive oral food challenge were classified as food allergic.
228 In addition, those regularly ingesting the sensitized food at the time of SPT were defined as
229 sensitized-tolerant without formal challenge and included in the sensitized-tolerant group
230 for analysis. If, on clinical review, the participant had a clinical history and reaction
231 consistent with a diagnosis of IgE-mediated food allergy within 2 months either side of the
232 12-month review and a positive SPT, they were defined as food-allergic without proceeding
233 to food challenge and included in the food allergic group for analysis.

234

235 **Demographics and Risk Factors**

236 Birth record data and questionnaires administered during pregnancy were used to obtain
237 demographic information. Exposure to known or predicted risk factors for allergic disease
238 during the first 12 months was determined from clinical data and questionnaires completed
239 by parents at several timepoints during pregnancy and up to the 12-month review.

240

241 **Cord blood lymphocyte populations**

242 **Blood sampling and isolation of mononuclear cells**

243 Umbilical cord blood was collected at birth by syringe and immediately diluted in 10IU/mL
244 preservative-free sodium heparin (Pfizer) in 10ml of RPMI 1640 (Gibco, Life Technologies).
245 Mononuclear cells were isolated by density gradient centrifugation (Lymphoprep, Axis-
246 Shield), and $2-4 \times 10^4$ cells immediately used for flow cytometric measurement of Treg cells.

247 **Measurement of regulatory T-cell subsets by flow cytometry**

248 All blood samples were stained for flow cytometric analysis within 12 hours of collection.
249 Isotype controls were used to set up the instrument for positive gating, and, once
250 established, these settings were maintained throughout. Mononuclear cells were stained

251 with anti-CD4-PE, and anti-CD45RA-PECy5 and then washed in PBS and formalin fixed. After
252 overnight fixation, cells were permeabilized (0.5% Tween in PBS) and stained with anti-
253 FOXP3-Alexa Fluor488 followed by analysis on a 3-channel flow cytometer. Gating of naïve
254 Tregs (CD4⁺/FOXP3⁺/CD45RA⁺) and activated Tregs (CD4⁺/FOXP3⁺⁺/CD45RA⁻) was performed
255 as previously described (32) and reported as a proportion of the total CD4⁺ T-cell
256 population.

257 **Eczema status during the first 12 months**

258 Data on eczema were collected by questionnaires administered at 1, 3, 6, 9 and 12 months,
259 and clinical assessments conducted at 1, 6 and 12 months. Eczema was defined according to
260 the modified UK working party criteria.(33) The Scoring Atopic Dermatitis Scale (SCORAD)
261 was used to quantify eczema severity.(34, 35)

262

263 **Allergic sensitization at age 4 years**

264 Participants were assessed at 4-year review intended shortly after the 4th birthday (Mean
265 age 4.28 years, Standard deviation (SD) 0.35). At the 4-year review infants underwent SPT to
266 the same 10 food and aeroallergens using identical equipment and technique. Allergic
267 sensitization to an allergen at 4-years was defined as a wheal size of 3mm greater than the
268 negative control, in the presence of a positive control ≥ 3 mm and a negative saline control
269 ≤ 3 mm. Aeroallergen sensitization was defined as allergic sensitization to an aeroallergen
270 (any of house dust mite, cat, dog, rye grass or the fungus *Alternaria tenuis*).

271

272 **Wheeze, hayfever, atopic wheeze and doctor-diagnosed asthma at age 4 years**

273 At the 4-year review parents were asked if their child had wheezed or suffered from
274 hayfever symptoms in the past 12 months. Atopic wheeze was defined as allergic
275 sensitization to any allergen at age 4 years plus parent-reported wheeze in the preceding 12
276 months. At the 2 and 4-year reviews, parents were asked if a doctor had ever diagnosed
277 their child with asthma.

278

279 **Statistical analysis**

280 In order to avoid misclassification, analysis was restricted to those infants who could be
281 confidently classified as either non-sensitized (n=698), sensitized-tolerant (n=27) or food

282 allergic (n=61) (Table 1). To estimate odds ratios for the effect of cord blood Treg
283 populations, and infant eczema during the first 12 months, on infant food sensitization
284 phenotype, we fitted multinomial logistic regression models adjusted for relevant
285 covariates. These models differ from our previously reported analysis(21) by including a
286 third group, the sensitized-tolerant infants. Given the low baseline risk of sensitized
287 tolerance, the estimated odds ratios are interpretable as risk ratios (RR). To estimate risk
288 ratios for the effect of infant food sensitization phenotype on aeroallergic disease outcomes
289 to age 4 years we fitted logarithmic binomial regression models adjusted for relevant
290 covariates. Covariates included in analysis models were those known or potential risk
291 factors for allergic disease included in causal models represented by directed acyclic graphs
292 (DAGs)(Supplementary Figures 1-3). Data analysis used the statistical software Stata/SE
293 version 15.1 (Statacorp, TX, USA). See supplementary methods for further details of
294 methods.

295
296
297
298
299
300
301

302 **Results**

303 Table 1 lists demographic details for participants with food sensitization phenotype
304 determined in infancy (n=786). As previously reported(36), 845 infants were included in the
305 12-month review. Of these 93/845 (11.0%) were sensitized to one or more foods at 12
306 months. Following food challenge, 61/845 (7.2%) infants were food allergic and 27/845
307 (3.2%) were sensitized-tolerant. A further 59/845 (7.0%) were either: sensitized to
308 aeroallergens only (17/845), had a non IgE-mediated food allergy (1/845) or had
309 inconclusive results (41/845). These participants were excluded from the analysis in order to
310 specifically focus on the sensitized-tolerant and food allergic groups.

311
312
313

312 **Table 1.**

314 **Genetic and environmental exposures**

315 Environmental exposures did not appear to distinguish sensitized-tolerant from non-
316 sensitized individuals. By contrast, in comparison to food allergic infants, both non-
317 sensitized and sensitized-tolerant infants appeared to be less likely to have a family history
318 of eczema and asthma, and have higher rates of household pet ownership during gestation
319 and infancy (Table 1). A particularly strong difference was seen with respect to pet exposure
320 during infancy which was more frequent in sensitized-tolerant infants than food allergic
321 infants (RR 3.32, 95%CI 1.38-7.99, $P=0.007$). We did not find evidence that known risk
322 factors for increased allergic disease, such as male sex, no labour prior to delivery and
323 reduced household size, differed between groups.

324

325

326 **Cord blood regulatory T cells**

327 **Figure 1. (attached separately)**

328 **Lower cord blood naïve regulatory T-cells are associated with subsequent food allergy**

329 We previously reported that food allergic infants in our cohort had a lower proportion of
330 umbilical cord blood naïve Treg cells than non-sensitized participants (Odds ratio (OR) 0.63,
331 95% CI 0.44 – 0.90, $P= 0.010$). (20, 21) By contrast, there was no evidence of a difference in
332 the proportion of naïve Treg cells between the sensitized-tolerant and non-sensitized
333 groups (OR 0.89 95% CI 0.59 – 1.36, $P= 0.619$) (Figure1).

334 **Higher cord blood activated Tregs may be associated with subsequent sensitized- 335 tolerance**

336 In comparison to non-sensitized infants, there was some evidence that sensitized-tolerance
337 was associated with a higher proportion of umbilical cord blood activated Treg cells (OR =
338 2.89 95% CI 1.03 – 8.16, $P= 0.045$). There was however no evidence of a difference in the
339 proportion of activated Treg cells between the food allergic and non-sensitized groups (OR
340 0.72 95% CI 0.24 – 2.18, $P= 0.566$) (Figure 1).

341 **Eczema during the first 12 months**

342 As previously reported(36), at the 1-month review none of the participants reported
343 eczema. The cumulative prevalence of eczema up to the 3, 6, 9 and 12 month review was
344 9/763 (1.2%); 65/737 (8.8%); 126/685 (18.4%); and 162/701 (23.1%) respectively. Eczema
345 during infancy appeared to be strongly predictive of food allergy, and perhaps weakly
346 predictive of sensitized-tolerance (Table 2)(Supplementary figures 4 and 5)

347 **Table 2.**

348

349 **Aeroallergen sensitization and allergic disease to age 4 years**

350 We next investigated the relationship between food sensitization status at 12 months and
351 aeroallergen sensitization, hayfever, atopic wheeze, and doctor diagnosed asthma to age 4
352 years. SPT was performed in 546 participants at 4-year review and 156/546 (28.6%) children
353 were sensitized to aeroallergens. We did not find evidence that in comparison to non-
354 sensitized infants, sensitized-tolerance at 12 months were at increased risk of subsequent
355 aeroallergen sensitization, hayfever, or doctor-diagnosed asthma to age 4 years. However,
356 there was a weak indication that sensitized-tolerance might be associated with atopic
357 wheeze, with the 95% CI not excluding large effects. By contrast, in comparison to non-
358 sensitized infants, food allergy at 12 months strongly predicted subsequent aeroallergen
359 sensitization, hayfever, atopic wheeze at age 4 years and doctor-diagnosed asthma to age 4
360 years (Table 3). Further, food allergic infants were sensitized to a greater number of
361 aeroallergens at age 4 years (Supplementary Figure 6) and had a higher average
362 aeroallergen wheal size (Supplementary Figure 7) than either sensitized-tolerant infants or
363 non-sensitized infants.

364 **Table 3.**

365

366

367

368

369

370

371 **Discussion**

372 In this pre-birth cohort study, incorporating SPT and oral food challenge at 1 year, food
373 sensitized-tolerance during infancy appeared to appears to be associated with a similar

374 pattern of exposures and outcomes to non-sensitized infants. In addition, we found some
375 evidence that a regulatory immune profile at birth was associated with subsequent
376 sensitized-tolerance.

377

378 Genetic factors may influence the progression from food sensitization to clinically expressed
379 food allergy. A family history of allergic disease, in particular eczema and asthma, was
380 strongly associated with food allergy but not sensitized-tolerance. There is conflicting
381 evidence regarding associations between pet ownership and infant allergic disease.(37-39)
382 This is the first study to address the relationship between pet ownership and sensitized-
383 tolerance. Amongst sensitized infants, pet ownership at 12 months was strongly associated
384 with an increased incidence of sensitized-tolerance, and by contrast, a reduced incidence of
385 food allergy. This suggests that greater postnatal microbial exposure promotes a sensitized-
386 tolerant, rather than sensitized-allergic, phenotype; which is consistent with high levels of
387 sensitized-tolerance among children from rural versus urban Africa.(11) At 4 years of age
388 there was a very high proportion of dog ownership (422/559, 75.5%) but very low incidence
389 of dog sensitization (4/546, 0.7%). There was no evidence of concordance between dog
390 ownership and dog sensitization ($p = 0.474$). Cat ownership was less common than dog
391 ownership (204/559, 36.5%) and cat sensitization was more frequent than dog sensitization
392 (25/545, 4.6%). However, there was no evidence of concordance between cat ownership
393 and cat sensitization ($p = 0.281$). It is therefore unlikely that a relationship between dog/cat
394 ownership and dog/cat sensitisation is confounding the relationship between pet ownership
395 and aeroallergen sensitisation overall.

396

397

398 Associations between genetic and environmental factors, and food sensitization phenotype,
399 may reflect differences in immune function at birth. Previous studies have found
400 associations between a lower proportion of Tregs at birth and subsequent allergic
401 disease.(22, 24, 40). As far as we are aware, this is the first study to find this deficit is not
402 apparent among infants with sensitized-tolerance.

403

404 Interestingly, we found some evidence that sensitized-tolerant infants had an increased
405 proportion of umbilical cord blood activated Tregs in comparison to non-sensitized infants.

406 Both naïve and activated Tregs are equally suppressive but activated Tregs have a memory
407 (CD45RA^{neg}) phenotype(41) and are more proliferative(42). It has been recently reported
408 that sensitized-tolerant infants exhibit an increased capacity to produce and maintain
409 activated Tregs after oral food challenge.(26) There is mounting evidence regarding the
410 impact of the maternal microbial environment, microbiome and diet on foetal immune
411 development and Treg populations(13), although little is reported with specific reference to
412 activated Tregs. Our findings are consistent with an increase in activated Tregs during fetal
413 immune development in infants with subsequent sensitized-tolerance. Increased activated
414 Tregs are likely to provide sensitized-tolerant infants with a greater Treg response and
415 suppressive capacity, which may limit progression to food allergy.

416
417 Differences in the clinical expression of allergic disease among food sensitized-tolerant
418 versus food allergic infants were evident from early infancy and persisted throughout early
419 childhood. The association between eczema and food allergy, which was not apparent
420 between eczema and food sensitization, may reflect either causation or shared antecedent
421 factors. The dual-allergen-exposure model proposes that deficits in skin barrier function in
422 infancy are causally related to subsequent food allergy.(43) Alternatively, immune
423 phenotype in early infancy may underlie both eczema and food allergy. Infants with eczema
424 have been reported to have reduced Treg, with blunted responses to stimulation from
425 microbial components(23), however the relationship between the activated Treg cell
426 population and eczema has not been reported. It is plausible that an enhanced proliferative
427 response of activated Tregs reduces the risk of eczema and promotes sensitized-tolerance
428 by enabling more effective induction of tolerance following early allergen exposures. (42)
429 Antenatal exposure to allergen may augment this mechanism by promoting the production
430 of memory activated Tregs.(41)

431
432 Differences in the clinical expression of allergic disease by early food sensitization
433 phenotype persisted to age 4 years. In keeping with previous studies(44-46), food allergy
434 was strongly associated with subsequent aeroallergen sensitization, including the number of
435 aeroallergens sensitized and wheal size and was strongly associated with hayfever, atopic
436 wheeze and asthma. By contrast, sensitized-tolerant infants appeared to have a similar risk
437 of each of these outcomes to non-sensitized infants.

438

439 The strengths of this study include the longitudinal design, immune profiling at birth, and
440 determination of food allergy by formal food challenge. Food challenges provide robust
441 delineation of food sensitization phenotype in comparison to doctor diagnosis or parent
442 report which are often inaccurate (47, 48) but relied upon in previous studies.(27-30) A
443 potential limitation is the SPT wheal cut-offs chosen to define cases. In clinical practice food
444 sensitization at 12 months of age is defined as a SPT wheal size 3mm or greater than the
445 negative control(49), however recent studies have used a definition of 2mm or greater than
446 the negative control in infants.(50) This change in definition is supported by evidence that a
447 high proportion of 12 month old infants with a 2-3mm SPT response demonstrate clinically
448 apparent food allergy on formal challenge.(50) *A priori*, we therefore selected 2mm as an
449 appropriate definition of allergic sensitization at 12 months of age. Additionally, we adopted
450 a lower cut-off (1mm) to screen for infants who should undergo a formal food challenge in
451 order to optimize detection of clinically apparent food allergy in the cohort. We did not
452 have sufficient resources to conduct food challenges in the complete cohort at 12 months,
453 nor to conduct formal food challenges at 4 years. Another important limitation is the
454 relatively small number of sensitized-tolerant infants. There are substantial challenges
455 associated with performing food challenge in sufficiently large cohorts of infants to identify
456 enough children with sensitized-tolerance. Nonetheless, further delineation and
457 investigation of the sensitized-tolerant phenotype may well provide crucial insights.

458

459

460

461

462

463

464

465 **Conclusion**

466 Food sensitized-tolerance in infancy appears to be associated with a similar pattern of
467 exposures and outcomes to non-sensitized infants. In addition, an elevated proportion of
468 activated regulatory T cells at birth was specific to the sensitized-tolerant infants, and may

469 be relevant to suppression of clinical disease. Further understanding of the mechanisms
470 underlying the sensitized-tolerant phenotype may inform prevention of allergic disease.

471
472
473
474
475
476
477
478
479
480
481
482
483
484
485
486
487
488
489
490
491
492
493
494
495

Author Manuscript

References

496 1. Spergel JM, Paller AS. Atopic dermatitis and the atopic march. *The Journal of allergy*
497 *and clinical immunology*. 2003;112(6 Suppl):S118-27.

- 498 2. Burgess JA, Lowe AJ, Matheson MC, Varigos G, Abramson MJ, Dharmage SC. Does
499 eczema lead to asthma? The Journal of asthma : official journal of the Association for the
500 Care of Asthma. 2009;46(5):429-36.
- 501 3. Hill DA, Grundmeier RW, Ram G, Spergel JM. The epidemiologic characteristics of
502 healthcare provider-diagnosed eczema, asthma, allergic rhinitis, and food allergy in children:
503 a retrospective cohort study. BMC pediatrics. 2016;16:133.
- 504 4. Tsakok T, Marrs T, Mohsin M, Baron S, du Toit G, Till S, et al. Does atopic dermatitis
505 cause food allergy? A systematic review. Journal of Allergy and Clinical Immunology.
506 2016;137(4):1071-8.
- 507 5. Alduraywish SA, Standl M, Lodge CJ, Abramson MJ, Allen KJ, Erbas B, et al. Is there a
508 march from early food sensitization to later childhood allergic airway disease? Results from
509 two prospective birth cohort studies. Pediatric Allergy and Immunology. 2017;28(1):30-7.
- 510 6. Osborne NJ, Koplin JJ, Martin PE, Gurrin LC, Lowe AJ, Matheson MC, et al. Prevalence
511 of challenge-proven IgE-mediated food allergy using population-based sampling and
512 predetermined challenge criteria in infants. The Journal of allergy and clinical immunology.
513 2011;127(3):668-76.e1-2.
- 514 7. Sampson HA. Food allergy. Part 2: diagnosis and management. The Journal of allergy
515 and clinical immunology. 1999;103(6):981-9.
- 516 8. Sampson HA. Food allergy. Part 1: immunopathogenesis and clinical disorders. The
517 Journal of allergy and clinical immunology. 1999;103(5 Pt 1):717-28.
- 518 9. Nicolaou N, Poorafshar M, Murray C, Simpson A, Winell H, Kerry G, et al. Allergy or
519 tolerance in children sensitized to peanut: prevalence and differentiation using component-
520 resolved diagnostics. The Journal of allergy and clinical immunology. 2010;125(1):191-7.e1-
521 13.
- 522 10. Arshad SH, Tariq SM, Matthews S, Hakim E. Sensitization to common allergens and
523 its association with allergic disorders at age 4 years: a whole population birth cohort study.
524 Pediatrics. 2001;108(2):E33.
- 525 11. Yemaneberhan H, Bekele Z, Venn A, Lewis S, Parry E, Britton J. Prevalence of wheeze
526 and asthma and relation to atopy in urban and rural Ethiopia. The Lancet.
527 1997;350(9071):85-90.

- 528 12. Holt PG, Strickland D, Bosco A, Belgrave D, Hales B, Simpson A, et al. Distinguishing
529 benign from pathologic TH2 immunity in atopic children. *The Journal of allergy and clinical*
530 *immunology*. 2016;137(2):379-87.
- 531 13. Gray LE, O'Hely M, Ranganathan S, Sly PD, Vuillermin P. The Maternal Diet, Gut
532 Bacteria, and Bacterial Metabolites during Pregnancy Influence Offspring Asthma. *Frontiers*
533 *in immunology*. 2017;8:365.
- 534 14. Santner-Nanan B, Straubinger K, Hsu P, Parnell G, Tang B, Xu B, et al. Fetal-Maternal
535 Alignment of Regulatory T Cells Correlates with IL-10 and Bcl-2 Upregulation in Pregnancy.
536 *The Journal of Immunology*. 2013;191(1):145-53.
- 537 15. Gomez de Agüero M, Ganal-Vonarburg SC, Fuhrer T, Rupp S, Uchimura Y, Li H, et al.
538 The maternal microbiota drives early postnatal innate immune development. *Science (New*
539 *York, NY)*. 2016;351(6279):1296-302.
- 540 16. Schaub B, Liu J, Hoppler S, Schleich I, Huehn J, Olek S, et al. Maternal farm exposure
541 modulates neonatal immune mechanisms through regulatory T cells. *The Journal of allergy*
542 *and clinical immunology*. 2009;123(4):774-82.e5.
- 543 17. Lluís A, Depner M, Gaugler B, Saas P, Casaca VI, Raedler D, et al. Increased regulatory
544 T-cell numbers are associated with farm milk exposure and lower atopic sensitization and
545 asthma in childhood. *The Journal of allergy and clinical immunology*. 2014;133(2):551-9.
- 546 18. Pfefferle PI, Buchele G, Blumer N, Roponen M, Ege MJ, Krauss-Etschmann S, et al.
547 Cord blood cytokines are modulated by maternal farming activities and consumption of
548 farm dairy products during pregnancy: the PASTURE Study. *The Journal of allergy and clinical*
549 *immunology*. 2010;125(1):108-15.e1-3.
- 550 19. Douwes J, Cheng S, Travier N, Cohet C, Niesink A, McKenzie J, et al. Farm exposure in
551 utero may protect against asthma, hay fever and eczema. *European Respiratory Journal*.
552 2008;32(3):603-11.
- 553 20. Collier F, Ponsonby AL, O'Hely M, Tang MLK, Saffery R, Molloy J, et al. Naive
554 regulatory T cells in infancy: Associations with perinatal factors and development of food
555 allergy. *Allergy*. 2019.
- 556 21. Zhang Y, Collier F, Naselli G, Saffery R, Tang ML, Allen KJ, et al. Cord blood monocyte-
557 derived inflammatory cytokines suppress IL-2 and induce nonclassical "T(H)2-type" immunity
558 associated with development of food allergy. *Science translational medicine*.
559 2016;8(321):321ra8.

- 560 22. Smith M, Tourigny MR, Noakes P, Thornton CA, Tulic MK, Prescott SL. Children with
561 egg allergy have evidence of reduced neonatal CD4(+)CD25(+)CD127(lo/-) regulatory T cell
562 function. *The Journal of allergy and clinical immunology*. 2008;121(6):1460-6, 6 e1-7.
- 563 23. Ismail IH, Boyle RJ, Mah LJ, Licciardi PV, Tang ML. Reduced neonatal regulatory T cell
564 response to microbial stimuli associates with subsequent eczema in high-risk infants.
565 *Pediatric allergy and immunology : official publication of the European Society of Pediatric*
566 *Allergy and Immunology*. 2014;25(7):674-84.
- 567 24. Hinz D, Bauer M, Roder S, Olek S, Huehn J, Sack U, et al. Cord blood Tregs with stable
568 FOXP3 expression are influenced by prenatal environment and associated with atopic
569 dermatitis at the age of one year. *Allergy*. 2012;67(3):380-9.
- 570 25. Hawrylowicz CM, O'Garra A. Potential role of interleukin-10-secreting regulatory T
571 cells in allergy and asthma. *Nature reviews Immunology*. 2005;5(4):271-83.
- 572 26. Dang TD, Allen KJ, J. Martino D, Koplin JJ, Licciardi PV, Tang MLK. Food-allergic
573 infants have impaired regulatory T-cell responses following in vivo allergen exposure.
574 *Pediatric Allergy and Immunology*. 2016;27(1):35-43.
- 575 27. Schroeder A, Kumar R, Pongracic JA, Sullivan CL, Caruso DM, Costello J, et al. Food
576 allergy is associated with an increased risk of asthma. *Clinical and experimental allergy :*
577 *journal of the British Society for Allergy and Clinical Immunology*. 2009;39(2):261-70.
- 578 28. Sicherer SH, Wood RA, Vickery BP, Jones SM, Liu AH, Fleischer DM, et al. The natural
579 history of egg allergy in an observational cohort. *The Journal of allergy and clinical*
580 *immunology*. 2014;133(2):492-9.
- 581 29. Wood RA, Sicherer SH, Vickery BP, Jones SM, Liu AH, Fleischer DM, et al. The natural
582 history of milk allergy in an observational cohort. *The Journal of allergy and clinical*
583 *immunology*. 2013;131(3):805-12.
- 584 30. Alduraywish SA, Lodge CJ, Vicendese D, Lowe AJ, Erbas B, Matheson MC, et al.
585 Sensitization to milk, egg and peanut from birth to 18 years: A longitudinal study of a cohort
586 at risk of allergic disease. *Pediatric allergy and immunology : official publication of the*
587 *European Society of Pediatric Allergy and Immunology*. 2016;27(1):83-91.
- 588 31. Vuillermin P, Saffery R, Allen KJ, Carlin JB, Tang ML, Ranganathan S, et al. Cohort
589 Profile: The Barwon Infant Study. *International journal of epidemiology*. 2015;44(4):1148-
590 60.

- 591 32. Collier FM, Tang MLK, Martino D, Saffery R, Carlin J, Jachno K, et al. The ontogeny of
592 naïve and regulatory CD4(+) T-cell subsets during the first postnatal year: a cohort study.
593 *Clinical & translational immunology*. 2015;4(3):e34.
- 594 33. Williams HC, Burney PG, Pembroke AC, Hay RJ. The U.K. Working Party's Diagnostic
595 Criteria for Atopic Dermatitis. III. Independent hospital validation. *The British journal of*
596 *dermatology*. 1994;131(3):406-16.
- 597 34. Pucci N, Novembre E, Cammarata MG, Bernardini R, Monaco MG, Calogero C, et al.
598 Scoring atopic dermatitis in infants and young children: distinctive features of the SCORAD
599 index. *Allergy*. 2005;60(1):113-6.
- 600 35. Severity scoring of atopic dermatitis: the SCORAD index. Consensus Report of the
601 European Task Force on Atopic Dermatitis. *Dermatology (Basel, Switzerland)*.
602 1993;186(1):23-31.
- 603 36. Molloy J, Koplin JJ, Allen KJ, Tang ML, Collier F, Carlin JB, et al. Vitamin D insufficiency
604 in the first 6 months of infancy and challenge-proven IgE-mediated food allergy at 1 year of
605 age: a case-cohort study. *Allergy*. 2017.
- 606 37. Dick S, Friend A, Dynes K, AlKandari F, Doust E, Cowie H, et al. A systematic review of
607 associations between environmental exposures and development of asthma in children
608 aged up to 9 years. *BMJ open*. 2014;4(11):e006554.
- 609 38. Lodge CJ, Allen KJ, Lowe AJ, Hill DJ, Hosking CS, Abramson MJ, et al. Perinatal cat and
610 dog exposure and the risk of asthma and allergy in the urban environment: a systematic
611 review of longitudinal studies. *Clinical & developmental immunology*. 2012;2012:176484.
- 612 39. Lodrup Carlsen KC, Roll S, Carlsen KH, Mowinckel P, Wijga AH, Brunekreef B, et al.
613 Does pet ownership in infancy lead to asthma or allergy at school age? Pooled analysis of
614 individual participant data from 11 European birth cohorts. *PloS one*. 2012;7(8):e43214.
- 615 40. Hinz D, Bauer M, Roder S, Olek S, Huehn J, Sack U, et al. Cord blood Tregs with stable
616 FOXP3 expression are influenced by prenatal environment and associated with atopic
617 dermatitis at the age of one year. *Allergy*. 2012;67(3):380-9.
- 618 41. Miyara M, Yoshioka Y, Kitoh A, Shima T, Wing K, Niwa A, et al. Functional Delineation
619 and Differentiation Dynamics of Human CD4⁺ T Cells Expressing the FoxP3
620 Transcription Factor. *Immunity*. 2009;30(6):899-911.
- 621 42. Booth NJ, McQuaid AJ, Sobande T, Kissane S, Agius E, Jackson SE, et al. Different
622 proliferative potential and migratory characteristics of human CD4⁺ regulatory T cells that

623 express either CD45RA or CD45RO. Journal of immunology (Baltimore, Md : 1950).
624 2010;184(8):4317-26.

625 43. Lack G. Update on risk factors for food allergy. Journal of Allergy and Clinical
626 Immunology. 2012;129(5):1187-97.

627 44. Alduraywish SA, Lodge CJ, Campbell B, Allen KJ, Erbas B, Lowe AJ, et al. The march
628 from early life food sensitization to allergic disease: a systematic review and meta-analyses
629 of birth cohort studies. Allergy. 2016;71(1):77-89.

630 45. Tariq SM, Matthews SM, Hakim EA, Arshad SH. Egg allergy in infancy predicts
631 respiratory allergic disease by 4 years of age. Pediatric allergy and immunology : official
632 publication of the European Society of Pediatric Allergy and Immunology. 2000;11(3):162-7.

633 46. Vermeulen EM, Koplin JJ, Dharmage SC, Gurrin LC, Peters RL, McWilliam V, et al.
634 Food Allergy Is an Important Risk Factor for Childhood Asthma, Irrespective of Whether It
635 Resolves. The journal of allergy and clinical immunology In practice. 2018;6(4):1336-41.e3.

636 47. Thalayasingam M, Loo EXL, Tan MM, Bever HV, Shek LP-C. A review of oral food
637 challenges in children presenting to a single tertiary centre with perceived or true food
638 allergies. Singapore Medical Journal. 2015;56(11):622-5.

639 48. Bock SA, Atkins FM. Patterns of food hypersensitivity during sixteen years of double-
640 blind, placebo-controlled food challenges. The Journal of pediatrics. 1990;117(4):561-7.

641 49. Hill DJ, Heine RG, Hosking CS. The diagnostic value of skin prick testing in children
642 with food allergy. Pediatric allergy and immunology : official publication of the European
643 Society of Pediatric Allergy and Immunology. 2004;15(5):435-41.

644 50. Allen KJ, Koplin JJ, Ponsonby AL, Gurrin LC, Wake M, Vuillermin P, et al. Vitamin D
645 insufficiency is associated with challenge-proven food allergy in infants. The Journal of
646 allergy and clinical immunology. 2013;131(4):1109-16, 16.e1-6.

647
648
649
650
651
652
653
654

655
656
657
658
659
660
661
662
663
664

Table 1. Participant demographics for participants with known infant food sensitization phenotype

		Non-sensitized	Sensitized-tolerant	Food allergic	P-value for test of difference across all groups [#]
n	Total = 786	698	27	61	
Child sex	Male	346 (49.6%)	17 (63.0%)	34 (55.7%)	0.27
	Female	352 (50.4%)	10 (37.0%)	27 (44.3%)	
Plurality	Singleton	686 (98.3%)	27 (100.0%)	59 (96.7%)	0.53
	Twin	12 (1.7%)	0 (0.0%)	2 (3.3%)	
Maternal country of birth	Australia	627 (89.8%)	26 (96.3%)	56 (91.8%)	0.82
	Other	69 (9.9%)	1 (3.7%)	5 (8.2%)	
	Unknown	2 (0.3%)	0 (0.0%)	0 (0.0%)	
Paternal country of birth	Australia	605 (86.7%)	24 (88.9%)	49 (80.3%)	0.72
	Other	61 (8.7%)	2 (7.4%)	8 (13.1%)	
	Unknown	32 (4.6%)	1 (3.7%)	4 (6.6%)	
Family history of hayfever	Yes	438 (64.2%)	20 (76.9%)	47 (77.0%)	0.061
	No	244 (35.8%)	6 (23.1%)	14 (23.0%)	
Family history of eczema	Yes	303 (44.6%)	12 (44.4%)	40 (65.6%)	0.007
	No	376 (55.4%)	15 (55.6%)	21 (34.4%)	
Family history of asthma	Yes	331 (48.2%)	14 (51.9%)	43 (70.5%)	0.004
	No	356 (51.8%)	13 (48.1%)	18 (29.5%)	
Maternal age		31.93 (4.54)	31.64 (3.78)	32.21 (4.36)	0.83

at conception, mean (SD)					
Paternal age at conception, mean (SD)		33.91 (5.65)	33.20 (5.02)	34.35 (5.67)	0.62
Maternal highest education	Less than year 10	6 (0.9%)	0 (0.0%)	0 (0.0%)	0.85
	Year 10 or equivalent	35 (5.0%)	0 (0.0%)	2 (3.3%)	
	Year 12 or equivalent	93 (13.4%)	3 (11.1%)	12 (19.7%)	
	Trade certificate or Diploma	177 (25.5%)	7 (25.9%)	12 (19.7%)	
	Bachelor degree	249 (35.8%)	10 (37.0%)	23 (37.7%)	
	Postgraduate degree	135 (19.4%)	7 (25.9%)	12 (19.7%)	
	Paternal highest education	Less than year 10	16 (2.3%)	0 (0.0%)	
Year 10 or equivalent	44 (6.4%)	1 (4.0%)	5 (8.3%)		
Year 12 or equivalent	107 (15.6%)	2 (8.0%)	14 (23.3%)		
Trade certificate or Diploma	270 (39.5%)	11 (44.0%)	19 (31.7%)		
Bachelor degree	175 (25.6%)	6 (24.0%)	17 (28.3%)		
Postgraduate degree	72 (10.5%)	5 (20.0%)	5 (8.3%)		
SEIFA* disadvantage tertile	Low SEIFA (most disadvantaged)	219 (31.8%)	7 (26.9%)	18 (29.5%)	0.92
	Medium SEIFA	233 (33.8%)	8 (30.8%)	20 (32.8%)	
	High SEIFA (least	237 (34.4%)	11 (42.3%)	23 (37.7%)	

	disadvantaged)				
Household size during pregnancy	1 person	8 (1.1%)	0 (0.0%)	2 (3.3%)	0.21
	2 people	267 (38.4%)	13 (48.1%)	22 (36.1%)	
	3 people	240 (34.5%)	8 (29.6%)	28 (45.9%)	
	4 or more people	181 (26%)	6 (22.2%)	9 (14.8%)	
Sibling number at 12 months	No siblings	287 (41.1%)	13 (48.1%)	22 (36.1%)	0.50
	One sibling	244 (35.0%)	9 (33.3%)	28 (45.9%)	
	Two siblings	130 (18.6%)	3 (11.1%)	10 (16.4%)	
	Three or more siblings	37 (5.3%)	2 (7.4%)	1 (1.6%)	
Any maternal smoking during pregnancy	Any	84 (12.2%)	5 (18.5%)	9 (14.8%)	0.54
	None	607 (87.8%)	22 (81.5%)	52 (85.2%)	
Any maternal passive smoke exposure during pregnancy	Yes	77 (11.4%)	2 (7.4%)	6 (9.8%)	0.77
	No	601 (88.6%)	25 (92.6%)	55 (90.2%)	
Pet ownership during pregnancy	Yes	522 (75.0%)	21 (77.8%)	34 (55.7%)	0.004
	No	174 (25.0%)	6 (22.2%)	27 (44.3%)	
Pet ownership at 12 months	Yes	487 (71.5%)	22 (81.5%)	27 (45.8%)	<0.001
	No	194 (28.5%)	5 (18.5%)	32 (54.2%)	
Livestock exposure during pregnancy	Yes	56 (8.1%)	1 (3.7%)	1 (1.7%)	0.14
	No	635 (91.9%)	26 (96.3%)	59 (98.3%)	
Hospital type	Public hospital	477 (68.3%)	16 (59.3%)	41 (67.2%)	0.61
	Private hospital	221 (31.7%)	11 (40.7%)	20 (32.8%)	
Any labour prior to delivery	Yes	550 (78.9%)	24 (88.9%)	50 (82.0%)	0.40
	No	147 (21.1%)	3 (11.1%)	11 (18.0%)	
Birthweight, kg, mean		3.53 (0.53)8	3.72 (0.53)	3.51 (0.46)	0.34

	(SD)				
	Birthweight, Z-score (SD)	0.38 (0.91)	0.54 (0.97)	0.17 (0.88)	0.11
	Any breastfeeding				
	Yes	687 (98.4%)	27 (100.0%)	59 (96.7%)	0.48
	No	11 (1.6%)	0 (0.0%)	2 (3.3%)	

665

666 # P-values were calculated using Pearson's chi-squared test for binary or categorical
667 outcomes and a Kruskal-Wallis test for continuous outcomes.

668 * SEIFA - Socio-Economic Indexes for Areas. Lower score indicates greater relative socio-
669 economic disadvantage.

670

671

672

673

674

675

676

677

678

679

680

681

682

683

684

685

686

687

688

689

690 **Table 2. Eczema during infancy and subsequent food sensitization phenotype at 1 year in**
691 **comparison to non-sensitized infants**

Time point	Cumulative incidence of eczema			Adjusted risk ratio (aRR) † (95% Confidence interval)	
	NS	ST	FA	Sensitized-tolerant	Food allergic
3 months	4/678 (0.6%)	1/26 (3.9%)	4/59 (6.8%)	4.86 (0.49-48.11) p = 0.176	5.62 (0.83-38.00) p = 0.076
6 months	45/655 (6.9%)	3/26 (11.5%)	17/56 (30.3%)	1.58 (0.45-5.60) p = 0.478	4.18 (2.05-8.52) p < 0.001
9 months	96/610 (15.7%)	6/25 (24.0%)	24/50 (48.0%)	1.67 (0.62-4.47) p = 0.308	3.76 (1.97-7.16) p < 0.001
12 months	117/617 (19.0%)	8/25 (32.0%)	38/59 (64.4%)	2.05 (0.83-5.07) p = 0.119	5.81 (3.15-10.72) p < 0.001

692 †Adjusted for sex, family history of eczema, any siblings during pregnancy, pet ownership
693 and livestock exposure during pregnancy

694

695

696

697

698 **Table 3. Food sensitization phenotype at 1 year and subsequent aeroallergen sensitization**

699 **and disease in comparison to non-sensitized infants**

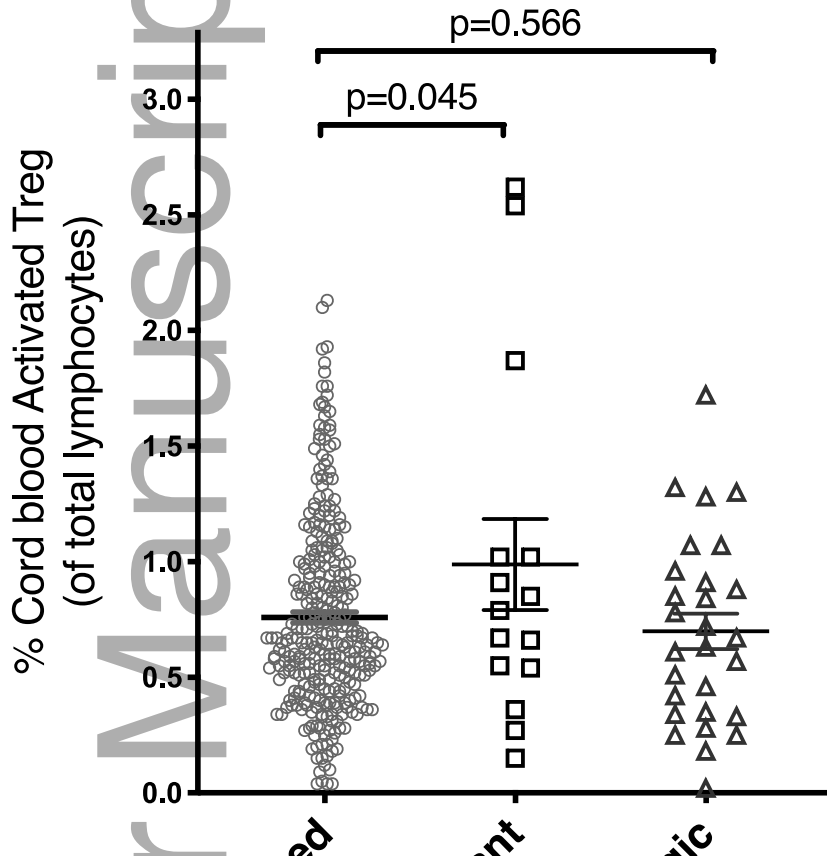
Allergic disease outcome	Incidence			Adjusted risk ratio (aRR) † (95% Confidence interval)	
	NS	ST	FA	Sensitized-tolerant	Food allergic
Aeroallergen sensitization at age 4 years	81/425 (19.1%)	4/14 (28.6%)	20/45 (44.4%)	1.39 (0.59-3.30) p = 0.449	3.84 (2.94-5.02) p < 0.001
Current	54/604	1/21	11/53	0.80	2.02

hayfever at age 4 years	(8.9%)	(4.8%)	(20.8%)	(0.21-3.02) p = 0.740	(1.17-3.50) p = 0.012
Current atopic wheeze at age 4 years	24/423 (5.7%)	2/14 (14.3%)	18/45 (40.0%)	2.95 (0.76-11.45) p = 0.117	5.97 (3.34-10.68) p < 0.001
Doctor-diagnosed asthma by age 2 years	36/551 (6.5%)	2/22 (9.1%)	8/45 (17.8%)	1.41 (0.35-5.59) p = 0.627	2.90 (1.42-5.86) p = 0.004
Doctor-diagnosed asthma by age 4 years	81/658 (12.3%)	3/24 (12.5%)	22/57 (38.6%)	0.92 (0.32-2.70) p = 0.883	2.80 (1.92-4.08) p < 0.001

700 † Adjusted for sex, family history of eczema, any siblings at 12 months and pet ownership at
701 12 months

702
703
704
705
706
707
708
709
710
711
712
713
714
715
716

717
718
719
720
721
722
723
724
725
726
727
728
729
730
731
732
733
734
735
736



737
738
739
740
741
742
743
744
745

Figure 1. Proportion of cord blood regulatory T-cells (naïve and activated) by food sensitization phenotype

Naïve and activated Treg proportions were determined as % of FOXP3^{low}CD45RA⁺ cells or FOXP3^{high}CD45RA⁻ cells respectively within the CD4⁺ cell fraction of cord blood mononuclear cells.

A. Reduced proportion of cord blood naïve Tregs in food allergic infants (n=28) compared to non-sensitized (n=313) (Odds ratio (OR) 0.63 per % change in proportion of naïve Treg, 95% confidence interval (CI) 0.44 – 0.90 , P= 0.010).(20, 21) B. Increased proportion of cord blood activated Tregs in the sensitized-tolerant group (n=15) compared to non-sensitized (n=313) (OR 2.89 per % change in proportion of activated Treg 95% CI 1.03 – 8.16, P= 0.045). Horizontal bar indicates mean. Whiskers indicate 95% CI of the mean.

