



# Genomic instability and genetic heterogeneity in aging: insights from clonal hematopoiesis (CHIP), monoclonal gammopathy (MGUS), and monoclonal B-cell lymphocytosis (MBL)

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**Abstract** Aging is a multifaceted process characterized by a gradual decline in physiological function and increased susceptibility to a range of chronic diseases. Among the molecular and cellular mechanisms driving aging, genomic instability is a fundamental hallmark, contributing to increased mutation load and genetic heterogeneity within cellular populations. This review explores the role of genomic instability and genetic heterogeneity in aging in

the hematopoietic system, with a particular focus on clonal hematopoiesis of indeterminate potential (CHIP), monoclonal gammopathy of undetermined significance (MGUS), and monoclonal B-cell lymphocytosis (MBL) as biomarkers. CHIP involves the clonal expansion of hematopoietic stem cells with somatic mutations. In contrast, MGUS is characterized by the presence of clonal plasma cells producing monoclonal immunoglobulins, while MBL is

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characterized by clonal proliferation of B cells. These conditions are prevalent in the aging population and serve as measurable indicators of underlying genomic instability. Studying these entities offers valuable insights into the mechanisms by which somatic mutations accumulate and drive clonal evolution in the hematopoietic system, providing a deeper understanding of how aging impacts cellular and tissue homeostasis. In summary, the hematopoietic system serves as a powerful model for investigating the interplay between genomic instability and aging. Incorporating age-related hematological conditions into aging research, alongside other biomarkers such as epigenetic clocks, can enhance the precision and predictive power of biological age assessments. These biomarkers provide a comprehensive view of the aging process, facilitating the early detection of age-related diseases and hopefully enabling personalized healthcare strategies.

**Keywords** Biological age · Biomarker · Aging · Clock · Mutation

## Introduction

Aging is a complex and multifaceted process characterized by a gradual decline in physiological function and increased vulnerability to diseases, ranging from cardiovascular diseases, neurodegenerative diseases,

and musculoskeletal diseases to cancers. Over the past two decades, significant advances in research have elucidated various molecular and cellular mechanisms that drive aging, collectively known as the hallmarks of aging [1–4]. These hallmarks provide a comprehensive framework for understanding the biological underpinnings of aging and include genomic instability, telomere attrition, epigenetic alterations, impaired cellular resilience and loss of proteostasis, deregulated nutrient sensing, mitochondrial dysfunction and oxidative stress, cellular senescence, stem cell exhaustion, and altered intercellular communication [4–6].

Among these hallmarks, genomic instability stands out as a fundamental mechanism contributing to aging [6–15]. Genomic instability encompasses a range of genetic alterations, including mutations, chromosomal aberrations, and DNA damage, which accumulate over time [10, 16–22]. This DNA damage accumulation results in increased mutation load and genetic heterogeneity (“somatic mosaicism” [23]) within dividing cellular populations, ultimately impairing tissue function and organismal health [24–39]. The accumulation of mutations with age varies significantly between different organs and cell types [25, 34–36, 40–46]. Somatic mosaicism predominantly affects tissues with high rates of cell turnover (e.g., skin, liver, and hematopoietic cells) [23]. The role of age-related genomic instability is especially important in stem cells [44]. Stem cells are responsible for the maintenance and regeneration of tissues throughout the life of an organism. As stem cells divide, they pass on accumulated genetic damage to their progeny, leading to clonal expansion of mutated cells. This can disrupt tissue homeostasis and contribute to the functional decline seen in aging tissues.

The hallmarks of aging are interconnected: age-related oxidative stress is a critical driver of DNA damage, while impaired DNA repair is a key component of the age-related decline in cellular resilience [4]. Many of these age-related changes are regulated at the level of the epigenome, where modifications in DNA methylation and histone modification patterns influence gene expression and cellular function. Cellular senescence, a state of permanent cell cycle arrest in response to stress, is closely linked to DNA damage. Increased cellular senescence with aging is causally associated with heightened DNA damage,

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contributing to the overall decline in tissue function and increased susceptibility to age-related diseases [18, 28, 29]. These interconnected processes underscore the complexity of aging and highlight the central role of genomic instability.

Understanding genomic heterogeneity in aging is crucial because it provides insights into the dynamic and adaptive nature of cellular populations in response to genomic stress. As cells acquire mutations, they diversify, leading to clonal expansions and shifts in tissue composition [47–56]. This heterogeneity not only reflects the underlying genomic instability but also influences the aging process and the development of age-related diseases.

Much of our understanding of genomic heterogeneity in aging stems from cross-sectional studies, which provide snapshots of genetic diversity at specific points in time across different age groups. However, studying genomic instability longitudinally, especially in humans, presents significant challenges. One primary obstacle is the invasive nature of obtaining repeated biopsies from the same individual over extended periods. While cross-sectional studies offer valuable insights, longitudinal studies are crucial for understanding the dynamics of genomic instability and its impact on aging at the individual level.

A particularly valuable organ system for studying the interplay between genomic instability and genetic heterogeneity in humans is the bone marrow [46, 57–59]. The bone marrow is a rich source of hematopoietic stem cells (HSCs) and progenitor cells, making it an ideal model for investigating how genetic changes accumulate and propagate within a highly dynamic cellular environment [23, 46]. For example, using single-cell whole-genome sequencing, researchers found that somatic mutations in B lymphocytes increase from less than 500 per cell in newborns to over 3000 per cell in centenarians, with mutational hotspots and B cell-specific mutation signatures [46]. In recent years, conditions such as clonal hematopoiesis of indeterminate potential (CHIP) [23], monoclonal gammopathy of undetermined significance (MGUS), and monoclonal B-cell lymphocytosis (MBL) [60, 61] have emerged as measurable consequences of genomic instability in the bone marrow. These clonal hematopoietic entities are characterized by the presence of nonmalignant clonal cell populations that arise from somatic mutations and mosaic

chromosomal alterations, and they are increasingly recognized as biomarkers of aging.

In this review, we will explore the mechanisms by which genomic instability contributes to aging, with a focus on the hematopoietic system. We will highlight the significance of CHIP, MGUS, and MBL as biomarkers of aging and discuss their potential to complement other aging biomarkers, such as epigenetic clocks. By understanding the role of genetic heterogeneity and genomic instability in human aging, we can gain valuable insights into the aging process and identify new avenues for therapeutic intervention.

### Genomic instability and aging

Genomic instability refers to the increased frequency of mutations within the genome, encompassing a broad spectrum of genetic alterations such as point mutations, insertions and deletions, chromosomal rearrangements, and aneuploidy. These alterations arise from intrinsic factors, including DNA replication errors and reactive oxygen species (ROS), as well as extrinsic factors, such as environmental toxins and radiation. The cellular machinery responsible for maintaining genomic integrity, including DNA repair pathways, the cell cycle checkpoint, and apoptosis, becomes less efficient with age. This decline in genomic maintenance allows genetic damage to accumulate, leading to increased genomic instability.

#### Role of mutation load in genetic heterogeneity

As organisms age, the accumulation of genetic mutations contributes to an increasing mutation load within cells [24–39]. This heightened mutation burden results in significant genetic heterogeneity among cellular populations [24–39]. In tissues with high cell turnover, such as the hematopoietic system, this heterogeneity is particularly pronounced. Each stem cell acquiring unique mutations leads to a mosaic of genetically distinct clones within the tissue. This clonal diversity can drive various cellular behaviors, with some clones potentially gaining a proliferative advantage or resistance to apoptosis. Consequently, this genetic heterogeneity is a direct outcome of the continuous accumulation of mutations, reflecting the underlying genomic instability.

## Impact of genomic instability on tissue function and aging

Genomic instability and the resultant genetic heterogeneity have profound implications for tissue function and overall aging [23, 41, 42, 62, 63]. As genetically diverse clones expand, they can disrupt the normal architecture and function of tissues. For instance, in the bone marrow, clonal expansions associated with conditions like CHIP and MGUS can impair hematopoiesis, leading to reduced production of healthy blood cells and increased risk of hematological malignancies. Additionally, these conditions are associated with an increased incidence of cardiovascular aging phenotypes, including a higher risk of thrombotic events and cardiovascular diseases [49, 50, 56, 64–70]. Additionally, genomic instability can induce cellular senescence, a state of irreversible growth arrest, contributing to the depletion of stem cell pools and impairing tissue regeneration [29]. The cumulative effect of these processes accelerates the functional decline of tissues, manifesting as the phenotypic hallmarks of aging and increasing susceptibility to age-related diseases. Understanding the impact of genomic instability is crucial for developing strategies to mitigate its effects and promote healthy aging.

## Hematopoietic system: a model for studying aging processes

The hematopoietic system is a complex network responsible for the production and regulation of blood cells. It encompasses the bone marrow, where hematopoietic stem cells (HSCs) reside and differentiate into various blood cell lineages, including red blood cells, white blood cells, and platelets. This system also includes peripheral blood, lymphoid tissues, and organs such as the spleen and thymus, which are essential for immune function and blood cell maturation. The hematopoietic system maintains homeostasis through a finely tuned balance of cell proliferation, differentiation, and apoptosis, ensuring a constant supply of functional blood cells throughout an individual's life.

## Relevance of the hematopoietic system in aging research

As individuals age, the functionality of the hematopoietic system declines, leading to reduced regenerative capacity, impaired immune responses, and increased susceptibility to diseases, including infections, anemia, and hematological cancers [71–74]. Age-related changes in the hematopoietic system include the exhaustion of HSCs, shifts in cell lineage potential, loss of reconstitution potential, and alterations in the bone marrow microenvironment. These changes provide a window into the broader impacts of aging on tissue function and organismal health, making the hematopoietic system an excellent model for studying the mechanisms of aging.

One significant application of studying the hematopoietic system in aging research is the use of blood cell analysis to determine biological age [75]. Biological age, which may differ from chronological age, reflects an individual's physiological state and susceptibility to age-related diseases [75–78]. Standard laboratory parameters derived from blood cell analysis, such as white blood cell counts and lymphocyte-to-monocyte ratios, have been incorporated into various “biological clocks” [75, 78]. These clocks are computational models that estimate biological age by integrating multiple hematological parameters with other laboratory parameters and/or physiological measurements [78]. Such measures provide a comprehensive assessment of an individual's health and biological aging, offering the potential for early detection of age-related decline and guiding interventions to promote healthy aging.

## Benefits of using the hematopoietic system to study genomic instability and genetic heterogeneity

The hematopoietic system offers numerous advantages for studying genomic instability and genetic heterogeneity [23]. Its high cellular turnover and the presence of long-lived HSCs make it highly susceptible to the accumulation of genetic mutations over time. These mutations contribute to clonal hematopoiesis, where specific cell clones expand due to their growth advantage, resulting in a diverse genetic landscape within the hematopoietic system.

One notable condition related to genomic instability in the hematopoietic system is CHIP. CHIP is

characterized by the presence of clonal blood cell populations derived from HSCs that harbor somatic mutations or mosaic chromosomal alterations [49–51, 70, 79–81]. These clonal populations can impact hematopoiesis and are associated with an increased risk of hematological malignancies and cardiovascular disease, making CHIP a valuable biomarker for aging research [49–51, 70, 79–81].

CHIP, as defined, is usually not associated with detectable peripheral blood count abnormalities. The progression of clonal myeloid cells may result in clonal cytopenias similar to those observed in myelodysplasia (MDS) or, rarely, polycythemia/thrombocytosis similar to those observed in myeloproliferative neoplasm (MPN). On the other hand, the progression of clonal lymphoid cells may result in monoclonal B-cell lymphocytosis (MBL) or, in rare instances, T-cell clones of uncertain significance (TCUS) [82]. Additionally, the proliferation of clonal B cells that mature to produce abnormal monoclonal immunoglobulins consequently leads to monoclonal gammopathy of unknown significance (MGUS) [83–87]. MGUS is considered a precursor to multiple myeloma, other plasma cell dyscrasias, and more rarely to other lymphoproliferative diseases and is more prevalent with advancing age [83–85]. Studying MGUS provides insights into how genomic instability and clonal expansions in B cells contribute to aging and disease.

The effects of receptor-diverse and homogeneous clonal hematopoiesis differ in their impact on immune function and disease risk. Receptor-diverse clones, which arise due to antigenic stimulation, tend to maintain a higher level of immune competence, preserving a range of T-cell or B-cell receptors. In contrast, homogeneous clonal hematopoiesis, where a single dominant clone expands, can reduce immune diversity and increase susceptibility to infections or immune dysregulation, as seen in conditions such as MGUS and MBL. This diversity or lack thereof may also influence the risk of progression to hematologic malignancies, with homogeneous clones potentially harboring a higher risk due to the lack of immune regulation and increased vulnerability to additional mutations.

Advances in single-cell sequencing and other high-throughput techniques have enhanced our ability to analyze the genetic and functional diversity of cells within the hematopoietic system [40, 46, 73]. These

technologies allow for detailed mapping of mutation patterns, clonal dynamics, and cellular behaviors, providing a comprehensive understanding of how genomic instability drives genetic heterogeneity and impacts aging [88–91].

### Clonal hematopoiesis of indeterminate potential (CHIP)

#### Definition and prevalence of CHIP

CHIP is characterized by the presence of an expanded clone of hematopoietic cells that harbor somatic mutations in genes commonly associated with hematologic malignancies in the absence of overt blood cancer or other hematologic abnormalities [23, 49, 50, 67, 70, 80, 81]. The present definition established the variant allele fraction (VAF) of  $\geq 2\%$  of somatic alterations of hematologic malignancy-associated genes, because the clinical consequence of variants below this threshold is less detectable [61]. CHIP is identified through genetic sequencing of blood cells, revealing specific mutations that confer a growth or survival advantage to particular cell clones [53, 54, 56, 92]. The prevalence of CHIP increases with age, affecting approximately 10–20% of individuals over the age of 70 [66, 80, 93]. Although it is asymptomatic and often detected incidentally, CHIP is a significant marker of clonal expansion within the hematopoietic system [23, 53, 54, 63, 93, 94].

#### Mechanisms leading to CHIP and its association with aging

The development of CHIP is driven by somatic mutations and mosaic chromosomal alterations affecting genes involved in hematopoietic cell regulation [23, 48, 49, 51, 53, 54, 56, 63, 64, 67, 80, 92, 95, 96]. These mutations occur in HSCs and confer a selective growth advantage preferentially to myeloid or lymphoid lineages, allowing the mutant clones to expand over time. Both myeloid and lymphoid driver gene mutations and mosaic chromosomal abnormalities can be detected, distinguishing between myeloid and lymphoid CHIP (M-CHIP and L-CHIP). Recent studies indicate that M-CHIP is more prevalent than L-CHIP [97]. Age-related declines in DNA repair mechanisms facilitate the accumulation of these

mutations, increased oxidative stress, and the overall aging of the hematopoietic stem cell pool. As a result, both M- and L-CHIP become more prevalent with advancing age, reflecting the broader phenomenon of genomic instability associated with the aging process. In M-CHIP, somatic mutations in three genes *DNMT3A*, *TET2*, and *ASXL1* (DTA) were implicated in approximately 80% of cases, all involved in epigenetic regulation. On the contrary, L-CHIP variants were more evenly spread across a larger number of genes [23, 97]. Several genes connected to L-CHIP are involved in DNA damage response, chromatin modifications, and maintaining genomic integrity, such as *ATM*, *KMT2D*, *KMT2C*, and *SPEN* [97].

#### Clinical implications of CHIP in the context of aging and disease

Although CHIP is believed to be asymptomatic, its presence has significant clinical implications [23, 49, 51, 52, 56, 67, 70, 93–95]. CHIP is a premalignant condition: individuals with CHIP have an increased risk of developing hematologic malignancies due to the potential for further genetic mutations that drive malignant transformation [51, 92, 98]. Myeloid and lymphoid CHIP variants clearly differentiate the type of malignancies: M-CHIP increases the likelihood of myeloid malignancies, while L-CHIP may lead to a higher incidence of lymphoid malignancies [97]. Additionally, preclinical and clinical studies indicate that the M-CHIP clonal events contribute to abnormal inflammatory and metabolic changes [23]. In humans, M-CHIP is associated with an elevated risk of cardiovascular disease, including atherosclerotic vascular disease leading to myocardial infarction [48–50, 56, 64, 67, 70, 79, 81] and stroke [66, 96], consequently adversely influencing the overall survival. The cardiovascular complications associated with M-CHIP may be explained by the loss of function related to the three main driver genes. However, the mechanisms underlying this association are not fully understood but may involve the pro-inflammatory effects of clonal hematopoietic cells, their migration to atherosclerotic lesions, and their contribution to both local and systemic inflammation [48–50, 56, 64, 67, 70, 79, 81]. On the contrary, L-CHIP is not associated with cardiovascular complications and increased mortality [97].

Beyond its role as a precursor to malignancy and cardiovascular disease, CHIP serves as a valuable biomarker for aging and age-related health risks [52]. Its detection can inform risk stratification and guide monitoring strategies for individuals at higher risk of adverse outcomes [52, 65, 99–103]. An important large-scale study by Nachun et al. demonstrated that CHIP is strongly associated with epigenetic age acceleration in multiple clocks [52]. Similar conclusions were reached by other studies as well [103]. CHIP carriers with mutations in multiple CHIP genes exhibit the most significant epigenetic age acceleration, which also associates with decreased telomere length [52]. The association of CHIP and epigenetic age acceleration identified a population at high risk of all-cause mortality (HR, 2.9) and coronary heart disease (HR, 3.24) [52]. Thus, the assessment of CHIP as a biomarker of age-related genomic instability offers potential avenues for early intervention and prevention of age-related diseases in high-risk populations.

#### Clonal cytopenia of unknown significance (CCUS)

##### Definition and prevalence of CCUS

Although the presence of CHIP mutations usually does not affect red cell distribution width (RDW), clonal cytopenia of undetermined significance (CCUS) [104–106] is characterized by the presence of M-CHIP along with one or more persistent cytopenias that cannot be explained by other hematologic or non-hematologic conditions and do not fulfill the diagnostic criteria for specific myeloid neoplasms. The definitions of cytopenias for CCUS, myelodysplastic syndromes (MDS), and MDS/myeloproliferative neoplasms (MDS/MPN) are harmonized and include the following: hemoglobin levels below 12–13 g/dL in females and males for anemia, an absolute neutrophil count below  $1.8 \times 10^9/L$  for leukopenia, and platelet counts below  $150 \times 10^9/L$  for thrombocytopenia. Anemia is the most prevalent form of cytopenia in older adults and serves as a key clinical and phenotypic marker of hematopoietic aging.

## Mechanisms leading to CCUS and its association with aging: clinical applications

Anemia in the elderly is most commonly attributed to nutrient deficiencies and chronic inflammation (ACI) [107, 108]. Despite extensive diagnostic resources, the cause of anemia remains unidentified in around one-third of the cases, regarded as unexplained anemia of the elderly (UAE) [109]. A large case–control study investigating the background of anemia revealed a higher prevalence of clonal hematopoiesis (M-CHIP) in cases with ACI and UAE compared to cases with nutrient deficiencies [110]. Although the occurrence of commonly detected DTA mutations did not differ between anemic and control cases, the anemia group exhibited a higher frequency of other mutations, including TP53 and SF3B1 [110]. Even a mild form of anemia in the elderly is correlated with significant morbidity and mortality [110].

## Monoclonal B-cell lymphocytosis (MBL)

### Definition, classification, and prevalence of MBL

MBL is characterized by the presence of a small monoclonal chronic lymphocytic leukemia/small lymphocytic lymphoma (CLL/SLL) phenotype B-cell clone in peripheral blood without diagnostic signs of B-lymphoproliferative disorder [60, 111–113]. In populations above the age of 40, the prevalence is estimated to be around 5–15% [114], and it rises to approximately 35% in individuals aged 70–79 years [111–113, 115].

MBL cases are categorized into low-count MBL ( $<0.05 \times 10^9/L$ ) and high-count MBL ( $0.05\text{--}0.5 \times 10^9/L$ ) based on the number of clonal lymphocytes, with high-count MBL comprising 5–15% of all MBL cases [116]. This subset has the potential to progress to chronic lymphocytic leukemia/small lymphocytic lymphoma (CLL/SLL) [116]. Immunophenotyping further divides MBL into three subtypes: CLL-type MBL (representing 75–85% of cases), atypical CLL-type MBL, and non-CLL-type MBL [117]. CLL-type MBL is the most common and shares many features with CLL, while the atypical and non-CLL types are less common and exhibit different immunophenotypic characteristics [117]. This classification aids in understanding the clinical

significance and potential progression of MBL to more serious conditions.

### Mechanisms leading to MBL

MBL emerges as the immune system is continuously stimulated throughout our lives, with genotypic changes occurring during each cell division. Certain genotypic alterations drive the expansion of B cells, leading to the propagation and formation of clones. These clones can retain their evolutionary advantages, continuing to expand even if the initial trigger stimuli are no longer present. Whole-genome sequencing of both low- and high-count MBL, as well as ultra-stable CLL (patients who do not show progression for over 10 years from initial diagnosis), revealed indistinguishable genomic profiles with low genomic complexity at both chromosomal and gene levels. The presence of somatic mutations in these cases indicated similar alterations, which likely occur before disease onset, probably at the hematopoietic stem cell level [118]. This suggests that MBL shares a common genetic background with CLL, including clinically relevant copy number alterations on a chromosomal level such as del13q, trisomy 12, del11q, and del17p [119], as well as gene mutations in TP53, NOTCH1, and SF3B1.

### Clinical implications of MBL in the context of aging and disease

MBL is an asymptomatic, premalignant stage of CLL with approximately a 1% annual risk of progression to CLL [120]. While individuals with low-count MBL have a 4.3 times higher risk of developing lymphoid malignancies compared to the general population, those with high-count MBL face a dramatically increased risk, estimated to be 74 times higher [116].

Monoclonality in MBL leads to a degree of immune impairment, increasing susceptibility to infections [115, 121]. This immune dysfunction is due to the clonal nature of the B cells, which affects the overall diversity and function of the immune system. Some studies have demonstrated decreased overall survival even in individuals with low-count MBL [122], although the literature is not entirely conclusive on this point, particularly regarding survival outcomes in low-count MBL cases [116].

## Monoclonal gammopathy of undetermined significance (MGUS)

Monoclonal gammopathy of undetermined significance (MGUS) is a condition characterized by the presence of an abnormal monoclonal protein (M protein) in the blood, produced by a clone of plasma cells [83, 87, 90]. Unlike multiple myeloma and other plasma cell dyscrasias, MGUS typically does not cause significant symptoms, organ damage, or a high level of M protein. MGUS is most commonly detected through blood tests conducted for unrelated reasons. It is a common condition, particularly among older adults, with a prevalence of over 5% in individuals 70 years of age or older and increasing to over 7.5% in those 85 years of age or older [83]. In the recent iStopMM study, the Icelandic population above the age of 40 years was systematically screened for the presence of MGUS, and 3.9% was found positive (5% in the > 50 years old population) [123, 124].

### Mechanisms leading to MGUS and its association with aging

MGUS is a premalignant condition that precedes multiple myeloma and carries an overall 1% annual risk of progression [125, 126]. The development of MGUS is believed to result from genetic and environmental factors that contribute to the clonal expansion of plasma cells producing monoclonal immunoglobulins. Targeted analyses have identified specific genetic abnormalities in MGUS, including IGH translocations, RB1 deletions, 1q gains, hyperdiploidy, and RAS gene mutations [125]. Somatic mutations in genes regulating cell growth, survival, and differentiation play a critical role in the pathogenesis of MGUS. In approximately half of the MGUS cases, hyperdiploidy is the founding genetic alteration. Recently, using an ingenious clonal mutation analysis approach, it was demonstrated that the first hyperdiploidy event may occur as early as the age of 20 years. As individuals age, the accumulation of age related as well as plasma cell-specific mutations and the decline in immune surveillance allow the abnormal plasma cell clones to expand, leading to the detection of M protein in the blood [90]. Age-related changes in the immune system, including immunosenescence and chronic inflammation, may create a

microenvironment conducive to the emergence and persistence of these clonal plasma cells [127–130].

### Clinical implications of MGUS in the context of aging and pathogenesis of age-related diseases

While MGUS is generally asymptomatic, its presence has important clinical implications, particularly in the context of aging [87, 90]. MGUS is considered a precursor condition to more serious plasma cell disorders, such as multiple multiple myeloma [90, 131]. The gross annual risk of progression from MGUS to multiple myeloma or related malignancies is approximately 1%, necessitating regular monitoring of individuals diagnosed with MGUS [90, 128, 131]. Patients with a “high-risk” MGUS may present with an M protein > 15 g/L, and/or abnormal serum free light chain ratio and/or a non IgG M protein. If all three clinical features are present, a more advanced MGUS situation is detected and the 20-year chance for myeloma progression is 58%, significantly more than the canonical overall 1% per year value [132]. Progressive or evolving MGUS may also be recharacterized as “early myeloma” as frequently myeloma-specific genetic events (such as DNA damage- or APOBEC-related mutations) could be identified in the isolated plasma cells.

Although CHIP and MGUS share comparable annual progression rates, MGUS involves the expansion of lineage-committed cells, whereas CHIP affects hematopoietic stem cells or less mature progenitor cells. Consequently, CHIP serves as a precursor state for a wider variety of hematologic neoplasms [80]. Early detection of progression can enable timely intervention and potentially improve clinical outcomes. In addition to its potential progression to malignancy, MGUS is associated with an increased risk of other health complications, including neuropathy [90, 133], nephropathy [134–138], skin changes [139–142], and occasionally even keratopathy [143–146]. It has also been shown that the thrombosis risk is increased in MGUS, but the risk is not related to the M protein level [68]. The exact mechanisms linking MGUS to these conditions are not fully understood but may involve the effects of the monoclonal protein on various tissues and organs, as well as the underlying genetic and immune changes associated with clonal plasma cell expansion [90]. Taken together, MGUS represents a significant

example of how genomic instability and clonal expansion manifest in the aging hematopoietic system. Thus, MGUS may serve as a valuable biomarker for aging and age-related diseases. Its detection can also provide insights into an individual's risk profile for developing hematologic malignancies and other complications.

### T-cell clones of uncertain significance (TCUS)

T-cell clones of uncertain significance (TCUS) represent the clonal expansion of T-cell large granular lymphocytes (T-LGL) and are considered a likely precursor state to T-cell large granular lymphocyte leukemia (LGLL) [147]. TCUS does not present symptoms typical of lymphoproliferative disease. Evaluation for TCUS is indicated when there is a persistent presence of at least  $0.5 \times 10^9/L$  of abnormal T lymphocytes, identified by flow cytometry as CD3+CD8+ or CD3+CD4+ with associated cytotoxic markers, for over 6 months, provided that other conditions, particularly viral infections, have been excluded [148]. Detection methods for TCUS include next-generation sequencing (NGS) and flow cytometry. Evidence suggests that the initial expansion of these T-cell clones is antigen-driven [149]. Genomic profiling may reveal mutations in STAT3 and STAT5B, which are predictive of malignant transformation [148]. Currently, the therapeutic approach for TCUS is a “watch and wait” strategy, similar to other premalignant conditions, as there is no effective treatment available for the condition at this time.

### Hematologic premalignant conditions as biomarkers of aging

Hematologic premalignant conditions are emerging as valuable biomarkers of aging due to their strong association with genomic instability. All these clonal conditions reflect underlying genetic alterations that accumulate with age, providing a measurable indication of genomic instability within the hematopoietic system [150, 151]. Importantly, in a recent study of 777 patients with a median age of 91 years (range 81–104), CHIP and MGUS prevalence were 17.5% and 9.5%, respectively, and the association between the two did not reach statistical significance ( $p=0.09$ )

[150, 152]. In another small-scale study, investigating bone marrow samples from 37 patients diagnosed with MGUS, a positive association between CHIP and MGUS was reported [153]. In this study, initial CHIP was frequent (27%) in MGUS patients, but this has to be confirmed in future larger-scale studies. The findings of the aforementioned study highlight potentially distinct pathways for CHIP and MGUS, while suggesting they share similar underlying mechanisms related to the aging process [150, 152]. The observed limited statistical association between CHIP and MGUS can be explained by the stochastic nature of mutations and clonal expansion. Since somatic mutations arise randomly, the occurrence and expansion of clonal populations in different cell types, such as hematopoietic stem cells in CHIP and plasma cells in MGUS, are not expected to have a direct one-to-one relationship. These clonal expansions are influenced by different biological processes and selective pressures within each cell population, leading to variability in their presence and extent. Therefore, while both conditions are indicative of age-related genomic instability, their independent occurrence reflects the complexity of clonal dynamics across different hematopoietic lineages. The detection of CHIP and MGUS in individuals can thus serve as a proxy for the broader genomic instability occurring throughout the body, highlighting their utility in aging research and underscoring the usefulness of incorporating both biomarkers into aging clocks to provide a more comprehensive assessment of biological aging.

In the realm of aging biomarkers, CHIP and MGUS offer unique insights compared to other widely studied measures such as epigenetic clocks. Epigenetic clocks estimate biological age based on specific patterns of DNA methylation, providing a quantifiable measure of age-related epigenetic changes. While epigenetic clocks are powerful tools for assessing biological age and predicting lifespan, CHIP and MGUS add an additional layer of information by directly indicating somatic mutations and clonal expansions. Unlike epigenetic changes, which are potentially reversible and influenced by environmental factors, the mutations leading to CHIP and MGUS are permanent and accumulate over time. This makes CHIP and MGUS particularly robust markers of cumulative genomic damage and clonal dynamics, complementing the temporal insights provided by epigenetic clocks.

The integration of CHIP and MGUS with existing biomarkers in aging studies holds significant potential for enhancing our understanding of the aging process and improving risk stratification for age-related diseases [50, 64, 66, 67, 81, 98, 154]. By combining the mutation-based insights from CHIP and MGUS with the information from epigenetic, proteomic, metabolomic, and/or lipidomic clocks, it may be possible to achieve a more comprehensive assessment of the biological age of an individual.

Furthermore, the inclusion of CHIP and MGUS in longitudinal aging studies can provide valuable data on the progression of genomic instability and its impact on health over time. This can facilitate the identification of early biomarkers of disease and the development of preventative measures. As research advances, the combined use of CHIP, MGUS, and other biomarkers will likely lead to a deeper understanding of the mechanisms driving aging and the discovery of novel therapeutic targets to promote healthy aging and longevity. The ability to detect and analyze these clonal events provides a valuable window into the dynamic processes underlying aging at the molecular and cellular levels.

One promising area of investigation is the development of anti-aging therapeutic interventions based on modulation of inflammation, which plays a central role in the progression of CHIP-related diseases [49, 155–158]. Anti-inflammatory therapies, such as IL-1 inhibitors like canakinumab, have demonstrated efficacy in reducing cardiovascular events in individuals with elevated inflammation driven by clonal hematopoiesis. In addition to inflammation modulation, targeted therapies that address the specific genetic mutations commonly associated with CHIP, including DNMT3A, TET2, and ASXL1, are being explored. These interventions aim to limit clonal expansion and its downstream effects. Cardiovascular risk reduction strategies, such as lipid-lowering therapies (e.g., statins) and antithrombotic agents, are also critical in addressing the heightened risk of thrombotic events seen in CHIP carriers. Together, these approaches hold promise for reducing both the hematological and cardiovascular risks associated with clonal expansions in aging populations.

## Conclusions

In summary, premalignant hematologic conditions are powerful biomarkers that offer unique and complementary insights into genomic instability and aging. Their incorporation into aging research, alongside other biomarkers such as epigenetic clocks, can enhance the precision and predictive power of biological age assessments. By providing a more comprehensive view of the aging process, CHIP and MGUS contribute to a deeper understanding of how genomic instability drives aging and age-related pathologies [159–169]. This knowledge can lead to more effective strategies for managing and mitigating the impacts of aging on human health. Ultimately, leveraging these biomarkers can improve early detection, risk stratification, and personalized interventions, promoting healthier aging and better quality of life for older adults.

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## Declarations

**Competing interests** Dr. Hajnalka Andrikovics, Dr. Andrea B. Maier, and Dr. Andrea Lehoczki serve as Associate Editor for GeroScience. Dr. Zoltan Ungvari serves as Editor-in-Chief for GeroScience.

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## References

1. Kaushik S, Tasset I, Arias E, Pampliega O, Wong E, Martinez-Vicente M, Cuervo AM. Autophagy and the hallmarks of aging. *Ageing Res Rev.* 2021;72:101468. <https://doi.org/10.1016/j.arr.2021.101468>.
2. Yousefzadeh MJ, Robbins PD, Huffman DM. Heterochronic parabiosis: a valuable tool to investigate cellular senescence and other hallmarks of aging. *Aging (Albany NY).* 2022;14:3325–8. <https://doi.org/10.18632/aging.204015>.
3. Wilson DM 3rd, Cookson MR, Van Den Bosch L, Zetterberg H, Holtzman DM, Dewachter I. Hallmarks of neurodegenerative diseases. *Cell.* 2023;186:693–714. <https://doi.org/10.1016/j.cell.2022.12.032>.
4. Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell.* 2013;153:1194–217. <https://doi.org/10.1016/j.cell.2013.05.039>.
5. Ungvari Z, Tarantini S, Sorond F, Merkely B, Csizsar A. Mechanisms of vascular aging, a geroscience perspective: JACC focus seminar. *J Am Coll Cardiol.* 2020;75:931–41. <https://doi.org/10.1016/j.jacc.2019.11.061>.
6. Ungvari Z, Tarantini S, Donato AJ, Galvan V, Csizsar A. Mechanisms of vascular aging. *Circ Res.* 2018;123:849–67. <https://doi.org/10.1161/CIRCRESAHA.118.311378>.
7. Sanchez-Roman I, Ferrando B, Holst CM, Mengel-From J, Rasmussen SH, Thinggaard M, Bohr VA, Christensen K, Stevnsner T. Molecular markers of DNA repair and brain metabolism correlate with cognition in centenarians. *Geroscience.* 2022;44:103–25. <https://doi.org/10.1007/s11357-021-00502-2>.
8. Shah AV, Bennett MR. DNA damage-dependent mechanisms of ageing and disease in the macro- and microvasculature. *Eur J Pharmacol.* 2017;816:116–28. <https://doi.org/10.1016/j.ejphar.2017.03.050>.
9. Barzilai A, Schumacher B, Shiloh Y. Genome instability: linking ageing and brain degeneration. *Mech Ageing Dev.* 2017;161:4–18. <https://doi.org/10.1016/j.mad.2016.03.011>.
10. Andriani GA, Vijg J, Montagna C. Mechanisms and consequences of aneuploidy and chromosome instability in the aging brain. *Mech Ageing Dev.* 2017;161:19–36. <https://doi.org/10.1016/j.mad.2016.03.007>.
11. Vermeij WP, Dolle ME, Reiling E, Jaarsma D, Payan-Gomez C, Bombardieri CR, Wu H, Roks AJ, Botter SM, van der Eerden BC, et al. Restricted diet delays accelerated ageing and genomic stress in DNA-repair-deficient mice. *Nature.* 2016;537:427–31. <https://doi.org/10.1038/nature19329>.
12. Cha HJ, Yim H. The accumulation of DNA repair defects is the molecular origin of carcinogenesis. *Tumour Biol.* 2013;34:3293–302. <https://doi.org/10.1007/s13277-013-1038-y>.
13. Durik M, Kavousi M, van der Pluijm I, Isaacs A, Cheng C, Verdonk K, Loot AE, Oeseburg H, Bhaggoe UM, Leijten F, et al. Nucleotide excision DNA repair is associated with age-related vascular dysfunction. *Circulation.* 2012;126:468–78. <https://doi.org/10.1161/CIRCULATIONAHA.112.104380>.
14. Rass U, Ahel I, West SC. Defective DNA repair and neurodegenerative disease. *Cell.* 2007;130:991–1004. <https://doi.org/10.1016/j.cell.2007.08.043>.
15. Hollander MC, Sheikh MS, Bulavin DV, Lundgren K, Augeri-Henmueller L, Shehee R, Molinaro TA, Kim KE, Tolosa E, Ashwell JD, et al. Genomic instability in Gadd45a-deficient mice. *Nat Genet.* 1999;23:176–84. <https://doi.org/10.1038/13802>.
16. Moskalev AA, Shaposhnikov MV, Plyusnina EN, Zhavoronkov A, Budovsky A, Yanai H, Fraifeld VE. The role of DNA damage and repair in aging through the prism of Koch-like criteria. *Ageing Res Rev.* 2013;12:661–84. <https://doi.org/10.1016/j.arr.2012.02.001>.
17. Seluanov A, Gladyshev VN, Vijg J, Gorbunova V. Mechanisms of cancer resistance in long-lived mammals. *Nat Rev Cancer.* 2018;18:433–41. <https://doi.org/10.1038/s41568-018-0004-9>.
18. Andriani GA, Almeida VP, Faggioli F, Mauro M, Tsai WL, Santambrogio L, Maslov A, Gadina M, Campisi J, Vijg J, Montagna C. Whole chromosome instability induces senescence and promotes SASP. *Sci Rep.* 2016;6:35218. <https://doi.org/10.1038/srep35218>.
19. Gorbunova V, Seluanov A, Zhang Z, Gladyshev VN, Vijg J. Comparative genetics of longevity and cancer: insights from long-lived rodents. *Nat Rev Genet.* 2014;15:531–40. <https://doi.org/10.1038/nrg3728>.
20. Campisi J, Vijg J. Does damage to DNA and other macromolecules play a role in aging? If so, how? *J Gerontol A Biol Sci Med Sci.* 2009;64:175–8. <https://doi.org/10.1093/gerona/gln065>.
21. Vijg J. Somatic mutations and aging: a re-evaluation. *Mutat Res.* 2000;447:117–35. [https://doi.org/10.1016/S0027-5107\(99\)00202-X](https://doi.org/10.1016/S0027-5107(99)00202-X).
22. Dolle ME, Giese H, Hopkins CL, Martus HJ, Hausdorff JM, Vijg J. Rapid accumulation of genome rearrangements in liver but not in brain of old mice. *Nat Genet.* 1997;17:431–4. <https://doi.org/10.1038/ng1297-431>.
23. Walsh K, Raghavachari N, Kerr C, Bick AG, Cummings SR, Druley T, Dunbar CE, Genovese G, Goodell MA, Jaiswal S, et al. Clonal hematopoiesis analyses in clinical, epidemiologic, and genetic aging studies to unravel underlying mechanisms of age-related dysfunction in humans. *Front Aging.* 2022;3:841796. <https://doi.org/10.3389/fragi.2022.841796>.
24. John L, Poos AM, Brobeil A, Schinke C, Huhn S, Prokoph N, Lutz R, Wagner B, Zangari M, Tirier SM, et al. Resolving the spatial architecture of myeloma and its microenvironment at the single-cell level. *Nat*

- Commun. 2023;14:5011. <https://doi.org/10.1038/s41467-023-40584-4>.
25. Busuttill RA, Garcia AM, Reddick RL, Dolle ME, Calder RB, Nelson JF, Vijg J. Intra-organ variation in age-related mutation accumulation in the mouse. *PLoS ONE*. 2007;2:e876. <https://doi.org/10.1371/journal.pone.0000876>.
  26. Busuttill R, Bahar R, Vijg J. Genome dynamics and transcriptional deregulation in aging. *Neuroscience*. 2007;145:1341–7. <https://doi.org/10.1016/j.neuroscience.2006.09.060>.
  27. Khrapko K, Kravtsov Y, de Grey AD, Vijg J, Schon EA. Does premature aging of the mtDNA mutator mouse prove that mtDNA mutations are involved in natural aging? *Aging Cell*. 2006;5:279–82. <https://doi.org/10.1111/j.1474-9726.2006.00209.x>.
  28. Vijg J. Impact of genome instability on transcription regulation of aging and senescence. *Mech Ageing Dev*. 2004;125:747–53. <https://doi.org/10.1016/j.mad.2004.07.004>.
  29. Busuttill RA, Dolle M, Campisi J, Vijg J. Genomic instability, aging, and cellular senescence. *Ann N Y Acad Sci*. 2004;1019:245–55. <https://doi.org/10.1196/annals.1297.041>.
  30. Vijg J, Dolle ME. Large genome rearrangements as a primary cause of aging. *Mech Ageing Dev*. 2002;123:907–15. [https://doi.org/10.1016/s0047-6374\(02\)00028-3](https://doi.org/10.1016/s0047-6374(02)00028-3).
  31. Nekhaeva E, Bodyak ND, Kravtsov Y, McGrath SB, Van Orsouw NJ, Pluzhnikov A, Wei JY, Vijg J, Khrapko K. Clonally expanded mtDNA point mutations are abundant in individual cells of human tissues. *Proc Natl Acad Sci U S A*. 2002;99:5521–6. <https://doi.org/10.1073/pnas.072670199>.
  32. Giese H, Snyder WK, van Oostrom C, van Steeg H, Dolle ME, Vijg J. Age-related mutation accumulation at a lacZ reporter locus in normal and tumor tissues of Trp53-deficient mice. *Mutat Res*. 2002;514:153–63. [https://doi.org/10.1016/s1383-5718\(01\)00329-1](https://doi.org/10.1016/s1383-5718(01)00329-1).
  33. Dolle ME, Vijg J. Genome dynamics in aging mice. *Genome Res*. 2002;12:1732–8. <https://doi.org/10.1101/gr.125502>.
  34. Dolle ME, Snyder WK, Dunson DB, Vijg J. Mutational fingerprints of aging. *Nucleic Acids Res*. 2002;30:545–9. <https://doi.org/10.1093/nar/30.2.545>.
  35. Martin SL, Hopkins CL, Naumer A, Dolle ME, Vijg J. Mutation frequency and type during ageing in mouse seminiferous tubules. *Mech Ageing Dev*. 2001;122:1321–31. [https://doi.org/10.1016/s0047-6374\(01\)00267-6](https://doi.org/10.1016/s0047-6374(01)00267-6).
  36. Dolle ME, Snyder WK, Gossen JA, Lohman PH, Vijg J. Distinct spectra of somatic mutations accumulated with age in mouse heart and small intestine. *Proc Natl Acad Sci U S A*. 2000;97:8403–8. <https://doi.org/10.1073/pnas.97.15.8403>.
  37. Dolle ME, Giese H, van Steeg H, Vijg J. Mutation accumulation in vivo and the importance of genome stability in aging and cancer. *Results Probl Cell Differ*. 2000;29:165–80. [https://doi.org/10.1007/978-3-540-48003-7\\_9](https://doi.org/10.1007/978-3-540-48003-7_9).
  38. Khrapko K, Bodyak N, Thilly WG, van Orsouw NJ, Zhang X, Collier HA, Perls TT, Upton M, Vijg J, Wei JY. Cell-by-cell scanning of whole mitochondrial genomes in aged human heart reveals a significant fraction of myocytes with clonally expanded deletions. *Nucleic Acids Res*. 1999;27:2434–41. <https://doi.org/10.1093/nar/27.11.2434>.
  39. Slagboom PE, Vijg J. Genetic instability and aging: theories, facts, and future perspectives. *Genome*. 1989;31:373–85. <https://doi.org/10.1139/g89-057>.
  40. Zhang L, Lee M, Maslov AY, Montagna C, Vijg J, Dong X. Analyzing somatic mutations by single-cell whole-genome sequencing. *Nat Protoc*. 2024;19:487–516. <https://doi.org/10.1038/s41596-023-00914-8>.
  41. Albert O, Sun S, Huttner A, Zhang Z, Suh Y, Campisi J, Vijg J, Montagna C. Chromosome instability and aneuploidy in the mammalian brain. *Chromosome Res*. 2023;31:32. <https://doi.org/10.1007/s10577-023-09740-w>.
  42. Ren P, Dong X, Vijg J. Age-related somatic mutation burden in human tissues. *Front Aging*. 2022;3:1018119. <https://doi.org/10.3389/fragi.2022.1018119>.
  43. Huang Z, Sun S, Lee M, Maslov AY, Shi M, Waldman S, Marsh A, Siddiqui T, Dong X, Pter Y, et al. Single-cell analysis of somatic mutations in human bronchial epithelial cells in relation to aging and smoking. *Nat Genet*. 2022;54:492–8. <https://doi.org/10.1038/s41588-022-01035-w>.
  44. Brazhnik K, Sun S, Alani O, Kinkhabwala M, Wolkoff AW, Maslov AY, Dong X, Vijg J. Single-cell analysis reveals different age-related somatic mutation profiles between stem and differentiated cells in human liver. *Sci Adv*. 2020;6:eaa2659. <https://doi.org/10.1126/sciadv.aax2659>.
  45. Lodato MA, Rodin RE, Bohrsen CL, Coulter ME, Barton AR, Kwon M, Sherman MA, Vitzthum CM, Luquette LJ, Yandava CN, et al. Aging and neurodegeneration are associated with increased mutations in single human neurons. *Science*. 2018;359:555–9. <https://doi.org/10.1126/science.aao4426>.
  46. Zhang L, Dong X, Lee M, Maslov AY, Wang T, Vijg J. Single-cell whole-genome sequencing reveals the functional landscape of somatic mutations in B lymphocytes across the human lifespan. *Proc Natl Acad Sci U S A*. 2019;116:9014–9. <https://doi.org/10.1073/pnas.1902510116>.
  47. Urban VS, Cegledi A, Mikala G. Multiple myeloma, a quintessential malignant disease of aging: a gero-science perspective on pathogenesis and treatment. *Geroscience*. 2023;45:727–46. <https://doi.org/10.1007/s11357-022-00698-x>.
  48. Polizio AH, Park E, Walsh K. Clonal hematopoiesis: connecting aging and inflammation in atherosclerosis. *Curr Atheroscler Rep*. 2023;25:105–11. <https://doi.org/10.1007/s11883-023-01083-5>.
  49. Gumuser ED, Schuermans A, Cho SMJ, Sporn ZA, Uddin MM, Paruchuri K, Nakao T, Yu Z, Haidermota S, Hornsby W, et al. Clonal hematopoiesis of indeterminate potential predicts adverse outcomes in patients with atherosclerotic cardiovascular disease. *J Am Coll Cardiol*. 2023;81:1996–2009. <https://doi.org/10.1016/j.jacc.2023.03.401>.
  50. Uddin MDM, Nguyen NQH, Yu B, Brody JA, Pampana A, Nakao T, Fornage M, Bressler J, Sotoodehnia N,

- Weinstock JS, et al. Clonal hematopoiesis of indeterminate potential, DNA methylation, and risk for coronary artery disease. *Nat Commun.* 2022;13:5350. <https://doi.org/10.1038/s41467-022-33093-3>.
51. Menendez-Gonzalez JB, Rodrigues NP. Exploring the associations between clonal hematopoiesis of indeterminate potential, myeloid malignancy, and atherosclerosis. *Methods Mol Biol.* 2022;2419:73–88. [https://doi.org/10.1007/978-1-0716-1924-7\\_5](https://doi.org/10.1007/978-1-0716-1924-7_5).
  52. Nachun D, Lu AT, Bick AG, Natarajan P, Weinstock J, Szeto MD, Kathiresan S, Abecasis G, Taylor KD, Guo X, et al. Clonal hematopoiesis associated with epigenetic aging and clinical outcomes. *Aging Cell.* 2021;20:e13366. <https://doi.org/10.1111/ace1.13366>.
  53. Jaiswal S. Clonal hematopoiesis and nonhematologic disorders. *Blood.* 2020;136:1606–14. <https://doi.org/10.1182/blood.2019000989>.
  54. Jaiswal S, Ebert BL. Clonal hematopoiesis in human aging and disease. *Science.* 2019;366. <https://doi.org/10.1126/science.aan4673>.
  55. Dulken BW, Buckley MT, Navarro Negredo P, Saligramma N, Cayrol R, Leeman DS, George BM, Boutet SC, Hebestreit K, Pluvinaige JV, et al. Single-cell analysis reveals T cell infiltration in old neurogenic niches. *Nature.* 2019;571:205–10. <https://doi.org/10.1038/s41586-019-1362-5>.
  56. Fuster JJ, Walsh K. Somatic mutations and clonal hematopoiesis: unexpected potential new drivers of age-related cardiovascular disease. *Circ Res.* 2018;122:523–32. <https://doi.org/10.1161/CIRCRESAHA.117.312115>.
  57. McKerrell T, Park N, Moreno T, Grove CS, Ponstingl H, Stephens J, Understanding Society Scientific Group, Crawley C, Craig J, Scott MA, et al. Leukemia-associated somatic mutations drive distinct patterns of age-related clonal hemopoiesis. *Cell Rep.* 2015;10:1239–1245. <https://doi.org/10.1016/j.celrep.2015.02.005>.
  58. Richardson C, Yan S, Vestal CG. Oxidative stress, bone marrow failure, and genome instability in hematopoietic stem cells. *Int J Mol Sci.* 2015;16:2366–85. <https://doi.org/10.3390/ijms16022366>.
  59. Ergen AV, Goodell MA. Mechanisms of hematopoietic stem cell aging. *Exp Gerontol.* 2010;45:286–90. <https://doi.org/10.1016/j.exger.2009.12.010>.
  60. Alaggio R, Amador C, Anagnostopoulos I, Attygalle AD, Araujo IBO, Berti E, Bhagat G, Borges AM, Boyer D, Calaminici M, et al. The 5th edition of the World Health Organization classification of haematolymphoid tumours: lymphoid neoplasms. *Leukemia.* 2022;36:1720–1748. <https://doi.org/10.1038/s41375-022-01620-2>.
  61. Khoury JD, Solary E, Abla O, Akkari Y, Alaggio R, Apperley JF, Bejar R, Berti E, Busque L, Chan JKC, et al. The 5th edition of the World Health Organization classification of haematolymphoid tumours: myeloid and histiocytic/dendritic neoplasms. *Leukemia.* 2022;36:1703–1719. <https://doi.org/10.1038/s41375-022-01613-1>.
  62. Vijg J, Austad SN. Biological restraints on indefinite survival. *Cold Spring Harb Perspect Med.* 2023;13. <https://doi.org/10.1101/cshperspect.a041200>.
  63. Ren P, Zhang J, Vijg J. Somatic mutations in aging and disease. *Geroscience.* 2024. <https://doi.org/10.1007/s11357-024-01113-3>.
  64. Zekavat SM, Viana-Huete V, Matesanz N, Jorshery SD, Zuriaga MA, Uddin MM, Trinder M, Paruchuri K, Zorita V, Ferrer-Perez A, et al. TP53-mediated clonal hematopoiesis confers increased risk for incident atherosclerotic disease. *Nat Cardiovasc Res.* 2023;2:144–58. <https://doi.org/10.1038/s44161-022-00206-6>.
  65. Bohme M, Desch S, Rosolowski M, Scholz M, Krohn K, Buttner P, Cross M, Kirchberg J, Rommel KP, Poss J, et al. Impact of clonal hematopoiesis in patients with cardiogenic shock complicating acute myocardial infarction. *J Am Coll Cardiol.* 2022;80:1545–56. <https://doi.org/10.1016/j.jacc.2022.08.740>.
  66. Bhattacharya R, Zekavat SM, Haessler J, Fornage M, Raffield L, Uddin MM, Bick AG, Niroula A, Yu B, Gibson C, et al. Clonal hematopoiesis is associated with higher risk of stroke. *Stroke.* 2022;53:788–97. <https://doi.org/10.1161/STROKEAHA.121.037388>.
  67. Jaiswal S, Natarajan P, Silver AJ, Gibson CJ, Bick AG, Shvartz E, McConkey M, Gupta N, Gabriel S, Ardisino D, et al. Clonal hematopoiesis and risk of atherosclerotic cardiovascular disease. *N Engl J Med.* 2017;377:111–21. <https://doi.org/10.1056/NEJMoa1701719>.
  68. Gkalea V, Fotiou D, Dimopoulos MA, Kastritis E. Monoclonal gammopathy of thrombotic significance. *Cancers (Basel).* 2023;15. <https://doi.org/10.3390/cancers15020480>.
  69. Soudet S, Jedraszak G, Evrard O, Marolleau JP, Garcon L, Pietri MAS. Is hematopoietic clonality of indetermined potential a risk factor for pulmonary embolism? *TH Open.* 2021;5:e338–42. <https://doi.org/10.1055/s-0041-1733856>.
  70. Khetarpal SA, Qamar A, Bick AG, Fuster JJ, Kathiresan S, Jaiswal S, Natarajan P. Clonal hematopoiesis of indeterminate potential reshapes age-related CVD: JACC review topic of the week. *J Am Coll Cardiol.* 2019;74:578–86. <https://doi.org/10.1016/j.jacc.2019.05.045>.
  71. Warren LA, Rossi DJ. Stem cells and aging in the hematopoietic system. *Mech Ageing Dev.* 2009;130:46–53. <https://doi.org/10.1016/j.mad.2008.03.010>.
  72. Snoeck HW. Aging of the hematopoietic system. *Curr Opin Hematol.* 2013;20:355–61. <https://doi.org/10.1097/MOH.0b013e3283623c77>.
  73. Jin X, Zhang R, Fu Y, Zhu Q, Hong L, Wu A, Wang H. Unveiling aging dynamics in the hematopoietic system insights from single-cell technologies. *Brief Funct Genomics.* 2024. <https://doi.org/10.1093/bfpgp/ela019>.
  74. Beerman I, Maloney WJ, Weissmann IL, Rossi DJ. Stem cells and the aging hematopoietic system. *Curr Opin Immunol.* 2010;22:500–6. <https://doi.org/10.1016/j.coi.2010.06.007>.
  75. Parker DC, Bartlett BN, Cohen HJ, Fillenbaum G, Huebner JL, Kraus VB, Pieper C, Belsky DW. Association of blood chemistry quantifications of biological aging with disability and mortality in older adults. *J Gerontol A Biol Sci Med Sci.* 2020;75:1671–9. <https://doi.org/10.1093/gerona/glz219>.
  76. Verschoor CP, Belsky DW, Ma J, Cohen AA, Griffith LE, Raina P. Comparing biological age estimates using domain-specific measures from the Canadian

- longitudinal study on aging. *J Gerontol A Biol Sci Med Sci.* 2021;76:187–94. <https://doi.org/10.1093/geronol/glaa151>.
77. Shapiro I, Belsky DW, Israel S, Youssim I, Friedlander Y, Hochner H. Familial aggregation of the aging process: biological age measured in young adult offspring as a predictor of parental mortality. *Geroscience.* 2023;45:901–13. <https://doi.org/10.1007/s11357-022-00687-0>.
  78. Kwon D, Belsky DW. A toolkit for quantification of biological age from blood chemistry and organ function test data: BioAge. *Geroscience.* 2021;43:2795–808. <https://doi.org/10.1007/s11357-021-00480-5>.
  79. Aviv A, Levy D. Hemothelium, clonal hematopoiesis of indeterminate potential, and atherosclerosis. *Circulation.* 2019;139:7–9. <https://doi.org/10.1161/CIRCULATIONAHA.118.038434>.
  80. Steensma DP, Bejar R, Jaiswal S, Lindsley RC, Sekeres MA, Hasserjian RP, Ebert BL. Clonal hematopoiesis of indeterminate potential and its distinction from myelodysplastic syndromes. *Blood.* 2015;126:9–16. <https://doi.org/10.1182/blood-2015-03-631747>.
  81. Libby P, Ebert BL. CHIP (clonal hematopoiesis of indeterminate potential): potent and newly recognized contributor to cardiovascular risk. *Circulation.* 2018;138:666–8. <https://doi.org/10.1161/CIRCULATIONAHA.118.034392>.
  82. Shi M, Olteanu H, Jevremovic D, He R, Viswanatha D, Corley H, Horna P. T-cell clones of uncertain significance are highly prevalent and show close resemblance to T-cell large granular lymphocytic leukemia. Implications *Lab Diagn Mod Pathol.* 2020;33:2046–57. <https://doi.org/10.1038/s41379-020-0568-2>.
  83. Kyle RA, Therneau TM, Rajkumar SV, Larson DR, Plevak MF, Offord JR, Dispenzieri A, Katzmann JA, Melton LJ 3rd. Prevalence of monoclonal gammopathy of undetermined significance. *N Engl J Med.* 2006;354:1362–9. <https://doi.org/10.1056/NEJMoa054494>.
  84. Sverrisdottir IS, Lund SH, Turesson I, Bjorkholm M, Goldin LR, Landgren O, Kristinsson SY. Parental longevity and survival among patients with multiple myeloma and monoclonal gammopathy of undetermined significance: a population-based study. *Br J Haematol.* 2019;186:37–44. <https://doi.org/10.1111/bjh.15883>.
  85. Park HK, Lee KR, Kim YJ, Cho HI, Eun Kim J, Woong Kim K, Jung Kim Y, Lee KW, Hyun Kim J, Bang SM, Lee JS. Prevalence of monoclonal gammopathy of undetermined significance in an elderly urban Korean population. *Am J Hematol.* 2011;86:752–5. <https://doi.org/10.1002/ajh.22095>.
  86. Hampel H, Schneider C, Hock C, Muller-Spann F, Ackenheil M. CNS demyelination in monoclonal gammopathy of undetermined significance (MGUS): possible cause of a dementia syndrome. *Eur Psychiatry.* 1996;11:46–9. [https://doi.org/10.1016/0924-9338\(96\)80458-5](https://doi.org/10.1016/0924-9338(96)80458-5).
  87. Guerard EJ, Tuchman SA. Monoclonal gammopathy of undetermined significance and multiple myeloma in older adults. *Clin Geriatr Med.* 2016;32:191–205. <https://doi.org/10.1016/j.cger.2015.08.012>.
  88. Varettoni M, Zibellini S, Defrancesco I, Ferretti VV, Rizzo E, Malcovati L, Galli A, Porta MGD, Boveri E, Arcaini L, et al. Pattern of somatic mutations in patients with Waldenstrom macroglobulinemia or IgM monoclonal gammopathy of undetermined significance. *Haematologica.* 2017;102:2077–85. <https://doi.org/10.3324/haematol.2017.172718>.
  89. Sahota SS, Leo R, Hamblin TJ, Stevenson FK. Ig VH gene mutational patterns indicate different tumor cell status in human myeloma and monoclonal gammopathy of undetermined significance. *Blood.* 1996;87:746–55.
  90. Kaur J, Valisekka SS, Hameed M, Bandi PS, Varma S, Onwughalu CJ, Ibrahim H, Mongia H. Monoclonal gammopathy of undetermined significance: a comprehensive review. *Clin Lymphoma Myeloma Leuk.* 2023;23:e195–212. <https://doi.org/10.1016/j.clml.2023.02.004>.
  91. Akkus E, Tuncali T, Akin HY, Aydin Y, Besisik SK, Gurkan E, Ratip S, Salihoglu A, Sargin D, Unal A, et al. Germline genetic variants in Turkish familial multiple myeloma/monoclonal gammopathy of undetermined significance cases. *Br J Haematol.* 2024;204:931–8. <https://doi.org/10.1111/bjh.19271>.
  92. Genovese G, Jaiswal S, Ebert BL, McCarroll SA. Clonal hematopoiesis and blood-cancer risk. *N Engl J Med.* 2015;372:1071–2. <https://doi.org/10.1056/NEJMc1500684>.
  93. Calvillo-Arguelles O, Jaiswal S, Shlush LI, Moslehi JJ, Schimmer A, Barac A, Thavendiranathan P. Connections between clonal hematopoiesis, cardiovascular disease, and cancer: a review. *JAMA Cardiol.* 2019;4:380–7. <https://doi.org/10.1001/jamacardio.2019.0302>.
  94. Luis TC, Wilkinson AC, Beerman I, Jaiswal S, Shlush LI. Biological implications of clonal hematopoiesis. *Exp Hematol.* 2019;77:1–5. <https://doi.org/10.1016/j.exphem.2019.08.004>.
  95. Cobo I, Tanaka T, Glass CK, Yeang C. Clonal hematopoiesis driven by DNMT3A and TET2 mutations: role in monocyte and macrophage biology and atherosclerotic cardiovascular disease. *Curr Opin Hematol.* 2022;29:1–7. <https://doi.org/10.1097/MOH.0000000000000688>.
  96. Jaiswal S, Fontanillas P, Flannick J, Manning A, Grauman PV, Mar BG, Lindsley RC, Mermel CH, Burt N, Chavez A, et al. Age-related clonal hematopoiesis associated with adverse outcomes. *N Engl J Med.* 2014;371:2488–98. <https://doi.org/10.1056/NEJMoa1408617>.
  97. Niroula A, Sekar A, Murakami MA, Trinder M, Agrawal M, Wong WJ, Bick AG, Uddin MM, Gibson CJ, Griffin GK, et al. Distinction of lymphoid and myeloid clonal hematopoiesis. *Nat Med.* 2021;27:1921–7. <https://doi.org/10.1038/s41591-021-01521-4>.
  98. Mitchell SR, Gopakumar J, Jaiswal S. Insights into clonal hematopoiesis and its relation to cancer risk. *Curr Opin Genet Dev.* 2021;66:63–9. <https://doi.org/10.1016/j.gde.2020.12.004>.
  99. Weeks LD, Ebert BL. Causes and consequences of clonal hematopoiesis. *Blood.* 2023;142:2235–46. <https://doi.org/10.1182/blood.2023022222>.
  100. Scolari FL, Brahmabhatt DH, Abelson S, Medeiros JF, Anker MS, Fung NL, Otsuki M, Calvillo-Arguelles O, Lawler PR, Ross HJ, et al. Clonal hematopoiesis confers an increased mortality risk in orthotopic heart transplant

- recipients. *Am J Transplant*. 2022;22:3078–86. <https://doi.org/10.1111/ajt.17172>.
101. Schenz J, Rump K, Siegler BH, Hemmerling I, Rahmel T, Thon JN, Nowak H, Fischer D, Hafner A, Tichy L, et al. Increased prevalence of clonal hematopoiesis of indeterminate potential in hospitalized patients with COVID-19. *Front Immunol*. 2022;13:968778. <https://doi.org/10.3389/fimmu.2022.968778>.
  102. Robertson NA, Latorre-Crespo E, Terradas-Terradas M, Lemos-Portela J, Purcell AC, Livesey BJ, Hillary RF, Murphy L, Fawkes A, MacGillivray L, et al. Longitudinal dynamics of clonal hematopoiesis identifies gene-specific fitness effects. *Nat Med*. 2022;28:1439–46. <https://doi.org/10.1038/s41591-022-01883-3>.
  103. Soerensen M, Tulstrup M, Hansen JW, Weischenfeldt J, Gronbaek K, Christensen K. Clonal hematopoiesis and epigenetic age acceleration in elderly Danish twins. *Hemasphere*. 2022;6:e768. <https://doi.org/10.1097/HS9.0000000000000768>.
  104. Valent P. ICUS, IDUS, CHIP and CCUS: diagnostic criteria, separation from MDS and clinical implications. *Pathobiology*. 2019;86:30–8. <https://doi.org/10.1159/000489042>.
  105. Steensma DP. The clinical challenge of idiopathic cytopenias of undetermined significance (ICUS) and clonal cytopenias of undetermined significance (CCUS). *Curr Hematol Malig Rep*. 2019;14:536–42. <https://doi.org/10.1007/s11899-019-00547-3>.
  106. Jain M, Tripathi A. ICUS/CCUS/CHIP: basics & beyond. *Expert Rev Hematol*. 2017;10:915–20. <https://doi.org/10.1080/17474086.2017.1371588>.
  107. Krishnamurthy S, Kumar B, Thangavelu S. Clinical and hematological evaluation of geriatric anemia. *J Family Med Prim Care*. 2022;11:3028–33. [https://doi.org/10.4103/jfmpc.jfmpc.2239\\_21](https://doi.org/10.4103/jfmpc.jfmpc.2239_21).
  108. Lanser L, Fuchs D, Kurz K, Weiss G. Physiology and inflammation driven pathophysiology of iron homeostasis-mechanistic insights into anemia of inflammation and its treatment. *Nutrients*. 2021;13. <https://doi.org/10.3390/nu13113732>.
  109. Eisenga MF, Stam SP, Bakker SJL. Redefining unexplained anemia in elderly. *JAMA Intern Med*. 2017;177:1394–5. <https://doi.org/10.1001/jamainternmed.2017.2958>.
  110. van Zeventer IA, de Graaf AO, Wouters H, van der Reijden BA, van der Klauw MM, de Witte T, Jonker MA, Malcovati L, Jansen JH, Huls G. Mutational spectrum and dynamics of clonal hematopoiesis in anemia of older individuals. *Blood*. 2020;135:1161–70. <https://doi.org/10.1182/blood.2019004362>.
  111. Tang C, Shen Y, Soosapilla A, Mulligan SP. Monoclonal B-cell lymphocytosis - a review of diagnostic criteria, biology, natural history, and clinical management. *Leuk Lymphoma*. 2022;63:2795–806. <https://doi.org/10.1080/10428194.2022.2092857>.
  112. Strati P, Shanafelt TD. Monoclonal B-cell lymphocytosis and early-stage chronic lymphocytic leukemia: diagnosis, natural history, and risk stratification. *Blood*. 2015;126:454–62. <https://doi.org/10.1182/blood-2015-02-585059>.
  113. Semenzato G, Ghobrial IM, Ghia P. Monoclonal B-cell lymphocytosis, monoclonal gammopathy of undetermined significance, and T-cell clones of uncertain significance: are these premalignant conditions sharing a common identity? *Lancet Haematol*. 2023;10:e549–56. [https://doi.org/10.1016/S2352-3026\(23\)00086-8](https://doi.org/10.1016/S2352-3026(23)00086-8).
  114. Nieto WG, Almeida J, Romero A, Teodosio C, Lopez A, Henriques AF, Sanchez ML, Jara-Acevedo M, Rasillo A, Gonzalez M, et al. Increased frequency (12%) of circulating chronic lymphocytic leukemia-like B-cell clones in healthy subjects using a highly sensitive multicolor flow cytometry approach. *Blood*. 2009;114:33–7. <https://doi.org/10.1182/blood-2009-01-197368>.
  115. Shanafelt TD, Kay NE, Parikh SA, Achenbach SJ, Lesnick CE, Hanson CA, Kleinstern G, Olson JE, Norman AD, Rabe KG, et al. Risk of serious infection among individuals with and without low count monoclonal B-cell lymphocytosis (MBL). *Leukemia*. 2021;35:239–44. <https://doi.org/10.1038/s41375-020-0799-8>.
  116. Slager SL, Parikh SA, Achenbach SJ, Norman AD, Rabe KG, Boddicker NJ, Olson JE, Kleinstern G, Lesnick CE, Call TG, et al. Progression and survival of MBL: a screening study of 10 139 individuals. *Blood*. 2022;140:1702–9. <https://doi.org/10.1182/blood.2022016279>.
  117. Dagklis A, Fazi C, Sala C, Cantarelli V, Scielzo C, Massacane R, Toniolo D, Caligaris-Cappio F, Stamatopoulos K, Ghia P. The immunoglobulin gene repertoire of low-count chronic lymphocytic leukemia (CLL)-like monoclonal B lymphocytosis is different from CLL: diagnostic implications for clinical monitoring. *Blood*. 2009;114:26–32. <https://doi.org/10.1182/blood-2008-09-176933>.
  118. Agathangelidis A, Ljungstrom V, Scarfo L, Fazi C, Gounari M, Pandzic T, Sutton LA, Stamatopoulos K, Tonon G, Rosenquist R, Ghia P. Highly similar genomic landscapes in monoclonal B-cell lymphocytosis and ultra-stable chronic lymphocytic leukemia with low frequency of driver mutations. *Haematologica*. 2018;103:865–73. <https://doi.org/10.3324/haematol.2017.177212>.
  119. Henriques A, Rodriguez-Caballero A, Nieto WG, Langerak AW, Criado I, Lecrevisse Q, Gonzalez M, Pais ML, Paiva A, Almeida J, Orfao A. Combined patterns of IGHV repertoire and cytogenetic/molecular alterations in monoclonal B lymphocytosis versus chronic lymphocytic leukemia. *PLoS ONE*. 2013;8:e67751. <https://doi.org/10.1371/journal.pone.0067751>.
  120. Rawstron AC, Bennett FL, O'Connor SJ, Kwok M, Fenton JA, Plummer M, de Tute R, Owen RG, Richards SJ, Jack AS, Hillmen P. Monoclonal B-cell lymphocytosis and chronic lymphocytic leukemia. *N Engl J Med*. 2008;359:575–83. <https://doi.org/10.1056/NEJMoa075290>.
  121. Moreira J, Rabe KG, Cerhan JR, Kay NE, Wilson JW, Call TG, Leis JF, Jelinek DF, Schwager SM, Bowen DA, et al. Infectious complications among individuals with clinical monoclonal B-cell lymphocytosis (MBL): a cohort study of newly diagnosed cases compared to controls. *Leukemia*. 2013;27:136–41. <https://doi.org/10.1038/leu.2012.187>.

122. Criado I, Rodriguez-Caballero A, Gutierrez ML, Pedreira CE, Alcoceba M, Nieto W, Teodosio C, Barcena P, Romero A, Fernandez-Navarro P, et al. Low-count monoclonal B-cell lymphocytosis persists after seven years of follow up and is associated with a poorer outcome. *Haematologica*. 2018;103:1198–208. <https://doi.org/10.3324/haematol.2017.183954>.
123. Sigurbergisdottir AY, Rognvaldsson S, Thorsteinsdottir S, Sverrisdottir I, Sigurethardottir GA, Vietharsson B, Onundarson PT, Agnarsson BA, Sigurethardottir M, Thornorsteinsdottir I, et al. Disease associations with monoclonal gammopathy of undetermined significance can only be evaluated using screened cohorts: results from the population-based iStopMM study. *Haematologica*. 2023;108:3392–8. <https://doi.org/10.3324/haematol.2023.283191>.
124. Rognvaldsson S, Love TJ, Thorsteinsdottir S, Reed ER, Oskarsson J, Petursdottir I, Sigurethardottir GA, Vietharsson B, Onundarson PT, Agnarsson BA, et al. Iceland screens, treats, or prevents multiple myeloma (iStopMM): a population-based screening study for monoclonal gammopathy of undetermined significance and randomized controlled trial of follow-up strategies. *Blood Cancer J*. 2021;11:94. <https://doi.org/10.1038/s41408-021-00480-w>.
125. Mikulasova A, Wardell CP, Murison A, Boyle EM, Jackson GH, Smetana J, Kufova Z, Pour L, Sandecka V, Almasi M, et al. The spectrum of somatic mutations in monoclonal gammopathy of undetermined significance indicates a less complex genomic landscape than that in multiple myeloma. *Haematologica*. 2017;102:1617–25. <https://doi.org/10.3324/haematol.2017.163766>.
126. Cowan A, Ferrari F, Freeman SS, Redd R, El-Khoury H, Perry J, Patel V, Kaur P, Barr H, Lee DJ, et al. Personalised progression prediction in patients with monoclonal gammopathy of undetermined significance or smouldering multiple myeloma (PANGEA): a retrospective, multicohort study. *Lancet Haematol*. 2023;10:e203–12. [https://doi.org/10.1016/S2352-3026\(22\)00386-6](https://doi.org/10.1016/S2352-3026(22)00386-6).
127. Plano F, Corsale AM, Gigliotta E, Camarda G, Vullo C, Di Simone M, Shekarkar Azgomi M, Speciale M, Carlisi M, Caccamo N, et al. Monoclonal gammopathies and the bone marrow microenvironment: from bench to bedside and then back again. *Hematol Rep*. 2023;15:23–49. <https://doi.org/10.3390/hematolrep15010004>.
128. Moscvin M, Evans B, Bianchi G. Dissecting molecular mechanisms of immune microenvironment dysfunction in multiple myeloma and precursor conditions. *J Cancer Metastasis Treat*. 2023;9. <https://doi.org/10.20517/2394-4722.2022.110>.
129. Garcia-Ortiz A, Rodriguez-Garcia Y, Encinas J, Maroto-Martin E, Castellano E, Teixido J, Martinez-Lopez J. The role of tumor microenvironment in multiple myeloma development and progression. *Cancers (Basel)*. 2021;13. <https://doi.org/10.3390/cancers13020217>.
130. Dutta AK, Alberge JB, Sklavenitis-Pistofidis R, Lightbody ED, Getz G, Ghobrial IM. Single-cell profiling of tumour evolution in multiple myeloma - opportunities for precision medicine. *Nat Rev Clin Oncol*. 2022;19:223–36. <https://doi.org/10.1038/s41571-021-00593-y>.
131. Barwick BG, Gupta VA, Vertino PM, Boise LH. Cell of origin and genetic alterations in the pathogenesis of multiple myeloma. *Front Immunol*. 2019;10:1121. <https://doi.org/10.3389/fimmu.2019.01121>.
132. Landgren O. Advances in MGUS diagnosis, risk stratification, and management: introducing myeloma-defining genomic events. *Hematol Am Soc Hematol Educ Program*. 2021;2021:662–72. <https://doi.org/10.1182/hematology.2021000303>.
133. Sobol U, Stiff P. Neurologic aspects of plasma cell disorders. *Handb Clin Neurol*. 2014;120:1083–99. <https://doi.org/10.1016/B978-0-7020-4087-0.00073-5>.
134. Yong ZH, Yu XJ, Liu JX, Zhou FD, Wang SX, Zhao MH. Kidney histopathologic spectrum and clinical indicators associated with MGRS. *Clin J Am Soc Nephrol*. 2022;17:527–34. <https://doi.org/10.2215/CJN.12890921>.
135. Inotani S, Horino T, Ishihara M, Ichii O, Matsumori A. Immunotactoid glomerulopathy associated with monoclonal gammopathy. *Lancet*. 2021;397:2081. [https://doi.org/10.1016/S0140-6736\(21\)00477-3](https://doi.org/10.1016/S0140-6736(21)00477-3).
136. Gozzetti A, Guarnieri A, Zamagni E, Zakharova E, Coriu D, Bittrich M, Pika T, Tovar N, Schutz N, Ciofini S, et al. Monoclonal gammopathy of renal significance (MGRS): real-world data on outcomes and prognostic factors. *Am J Hematol*. 2022;97:877–84. <https://doi.org/10.1002/ajh.26566>.
137. Ekladius A, Bhandari R, Javaid MM. Association of monoclonal gammopathy of undetermined significance and C3 glomerulopathy. *Intern Med J*. 2023;53:1712–5. <https://doi.org/10.1111/imj.16222>.
138. Alonso-Titos J, Martinez-Esteban MD, Lopez V, Leon M, Martin-Reyes G, Ruiz-Esteban P, Hernandez D. Monoclonal gammopathy of renal significance: early diagnosis is key. *Nefrologia (Engl Ed)*. 2021;41:502–13. <https://doi.org/10.1016/j.nefro.2021.11.008>.
139. Nickell AL, Corn M, Mannuru D, Hinze AM. Scleroderma in the setting of monoclonal gammopathy of unknown significance with progression to multiple myeloma: a case report. *Cureus*. 2023;15:e44968. <https://doi.org/10.7759/cureus.44968>.
140. Koutra E, Lusmoller E, Stadler R, Gutzmer R. Monoclonal gammopathy with cutaneous significance treated successfully with rituximab. *J Dtsch Dermatol Ges*. 2022;20:697–700. <https://doi.org/10.1111/ddg.14751>.
141. Huang H, Qian SX. Lichen myxedematosus associated with monoclonal gammopathy of undetermined significance: a case report and literature review. *Front Med (Lausanne)*. 2023;10:1118555. <https://doi.org/10.3389/fmed.2023.1118555>.
142. Claveau JS, Wetter DA, Kumar S. Cutaneous manifestations of monoclonal gammopathy. *Blood Cancer J*. 2022;12:58. <https://doi.org/10.1038/s41408-022-00661-1>.
143. Skalicka P, Dudakova L, Palos M, Huna LJ, Evans CJ, Mahelkova G, Meliska M, Stopka T, Tuft S, Liskova P. Paraproteinemic keratopathy associated with monoclonal gammopathy of undetermined significance (MGUS): clinical findings in twelve patients including recurrence after keratoplasty. *Acta Ophthalmol*. 2019;97:e987–92. <https://doi.org/10.1111/aos.14123>.

144. Milman T, Kao AA, Chu D, Gorski M, Steiner A, Simon CZ, Shih C, Aldave AJ, Eagle RC Jr, Jakobiec FA, Udell I. Paraproteinemic keratopathy: the expanding diversity of clinical and pathologic manifestations. *Ophthalmology*. 2015;122:1748–56. <https://doi.org/10.1016/j.ophtha.2015.05.029>.
145. Karakus S, Gottsch JD, Caturegli P, Eghrari AO. Monoclonal gammopathy of “ocular” significance. *Am J Ophthalmol Case Rep*. 2019;15:100471. <https://doi.org/10.1016/j.ajoc.2019.100471>.
146. Al Hariri M, Munder M, Lisch W, Schuster AK, Fehr EM, Jacobi B, Desuki A, Krefl A, Gericke A, Pfeiffer N, Wasielica-Poslednik J. Prevalence of corneal findings and their interrelation with hematological findings in monoclonal gammopathy. *PLoS ONE*. 2022;17:e0276048. <https://doi.org/10.1371/journal.pone.0276048>.
147. Calabretto G, Attardi E, Gurnari C, Semenzato G, Voso MT, Zambello R. LGL clonal expansion and unexplained cytopenia: two clues don't make an evidence. *Cancers (Basel)*. 2022;14. <https://doi.org/10.3390/cancers14215236>.
148. Semenzato G, Teramo A, Calabretto G, Gasparini VR, Zambello R. All that glitters is not LGL leukemia. *Leukemia*. 2022;36:2551–7. <https://doi.org/10.1038/s41375-022-01695-x>.
149. Garrido P, Ruiz-Cabello F, Barcena P, Sandberg Y, Canton J, Lima M, Balanzategui A, Gonzalez M, Lopez-Nevot MA, Langerak AW, et al. Monoclonal TCR-Vbeta13.1+/CD4+/NKa+/CD8-/T-LGL lymphocytosis: evidence for an antigen-driven chronic T-cell stimulation origin. *Blood*. 2007;109:4890–8. <https://doi.org/10.1182/blood-2006-05-022277>.
150. Da Via MC, Lionetti M, Matera A, Travaglino E, Lucca E, Riva E, Tettamanti M, Baldini L, Neri A, Della Porta MG, Bolli N. MGUS and chip: two faces, but not of the same medal. *Blood*. 2021;138(Supplement 1):3800. <https://doi.org/10.1182/blood-2021-147890>.
151. Popp HD, Flach J, Brendel S, Ruppenthal S, Kleiner H, Seifarth W, Schneider S, Schulze TJ, Weiss C, Wenz F, et al. Accumulation of DNA damage and alteration of the DNA damage response in monoclonal B-cell lymphocytosis and chronic lymphocytic leukemia. *Leuk Lymphoma*. 2019;60:795–804. <https://doi.org/10.1080/10428194.2018.1498494>.
152. Da Via MC, Lionetti M, Marella A, Matera A, Travaglino E, Signaroldi E, Galbusera AA, Lucca U, Mandelli S, Riva E, et al. MGUS and clonal hematopoiesis show unrelated clinical and biological trajectories in an older population cohort. *Blood Adv*. 2022;6:5702–6. <https://doi.org/10.1182/bloodadvances.2021006498>.
153. Quaney V, Kroger B, Sannareddy A, Khan U, Kalkan F, Collins RH, Madanat YF, Vusirikala M, Huang Y, Awan FT, et al. Abstract 5925: prevalence of clonal hematopoiesis in patients with monoclonal gammopathy of undetermined significance. *Cancer Res*. 2023;83(7\_Supplement):5925. <https://doi.org/10.1158/1538-7445.AM2023-5925>.
154. Miller PG, Fell GG, Foy BH, Scherer AK, Gibson CJ, Sperling AS, Burugula BB, Nakao T, Uddin MM, Warren H, et al. Clonal hematopoiesis of indeterminate potential and risk of death from COVID-19. *Blood*. 2022;140:1993–7. <https://doi.org/10.1182/blood.2022018052>.
155. Yalcinkaya M, Tall AR. Genetic and epigenetic regulation of inflammasomes: role in atherosclerosis. *Atherosclerosis*. 2024;396:118541. <https://doi.org/10.1016/j.atherosclerosis.2024.118541>.
156. Orkaby AR, Thomson A, MacFadyen J, Besdine R, Forman DE, Trivison TG, Ridker PM. Effect of canakinumab on frailty: a post hoc analysis of the CANTOS trial. *Aging Cell*. 2024;23:e14029. <https://doi.org/10.1111/accel.14029>.
157. Forman DE, Pignolo RJ. A pragmatic approach to introducing translational geroscience into the clinic: a paradigm based on the incremental progression of aging-related clinical research. *J Gerontol A Biol Sci Med Sci*. 2024;79. <https://doi.org/10.1093/gerona/glae062>.
158. Zuriaga MA, Yu Z, Matesanz N, Truong B, Ramos-Neble BL, Asensio-Lopez MC, Uddin MM, Nakao T, Niroula A, Zorita V, et al. Colchicine prevents accelerated atherosclerosis in TET2-mutant clonal haematopoiesis. *Eur Heart J*. 2024. <https://doi.org/10.1093/eurheartj/ehae546>.
159. Ungvari Z, Ungvari A, Bianchini G, Gyorffy B. Prognostic significance of a signature based on senescence-related genes in colorectal cancer. *Geroscience*. 2024;46:4495–504. <https://doi.org/10.1007/s11357-024-01164-6>.
160. Nyul-Toth A, Patai R, Csiszar A, Ungvari A, Gulej R, Mukli P, Yabluchanskiy A, Benyo Z, Sotonyi P, Prodan CI, et al. Linking peripheral atherosclerosis to blood-brain barrier disruption: elucidating its role as a manifestation of cerebral small vessel disease in vascular cognitive impairment. *Geroscience*. 2024. <https://doi.org/10.1007/s11357-024-01194-0>.
161. Gulej R, Nyul-Toth A, Csik B, Patai R, Petersen B, Negri S, Chandragiri SS, Shanmugarama S, Mukli P, Yabluchanskiy A, et al. Young blood-mediated cerebrovascular rejuvenation through heterochronic parabiosis: enhancing blood-brain barrier integrity and capillarization in the aged mouse brain. *Geroscience*. 2024;46:4415–42. <https://doi.org/10.1007/s11357-024-01154-8>.
162. Csiszar A, Ungvari A, Patai R, Gulej R, Yabluchanskiy A, Benyo Z, Kovacs I, Sotonyi P, Kirkpatrick AC, Prodan CI, et al. Atherosclerotic burden and cerebral small vessel disease: exploring the link through microvascular aging and cerebral microhemorrhages. *Geroscience*. 2024;46:5103–32. <https://doi.org/10.1007/s11357-024-01139-7>.
163. Magyar-Stang R, Pal H, Csanyi B, Gaal A, Mihaly Z, Czinege Z, Csipo T, Ungvari Z, Sotonyi P, Varga A, et al. Assessment of cerebral autoregulatory function and inter-hemispheric blood flow in older adults with internal carotid artery stenosis using transcranial Doppler sonography-based measurement of transient hyperemic response after carotid artery compression. *Geroscience*. 2023;45:3333–57. <https://doi.org/10.1007/s11357-023-00896-1>.
164. Gulej R, Nyul-Toth A, Ahire C, DeIFavero J, Balasubramanian P, Kiss T, Tarantini S, Benyo Z, Pacher P, Csik B, et al. Elimination of senescent cells by treatment with

- Navitoclax/ABT263 reverses whole brain irradiation-induced blood-brain barrier disruption in the mouse brain. *Geroscience*. 2023;45:2983–3002. <https://doi.org/10.1007/s11357-023-00870-x>.
165. Toth L, Czigler A, Hegedus E, Komaromy H, Amrein K, Czeiter E, Yabluchanskiy A, Koller A, Orsi G, Perlaki G, et al. Age-related decline in circulating IGF-1 associates with impaired neurovascular coupling responses in older adults. *Geroscience*. 2022;44:2771–83. <https://doi.org/10.1007/s11357-022-00623-2>.
166. Kiss T, Nyul-Toth A, Gulej R, Tarantini S, Csipo T, Mukli P, Ungvari A, Balasubramanian P, Yabluchanskiy A, Benyo Z, et al. Old blood from heterochronic parabionts accelerates vascular aging in young mice: transcriptomic signature of pathologic smooth muscle remodeling. *Geroscience*. 2022;44:953–81. <https://doi.org/10.1007/s11357-022-00519-1>.
167. Waigi EW, Pernomian L, Crockett AM, Costa TJ, Townsend P Jr, Webb RC, McQuail JA, McCarthy CG, Hollis F, Wenceslau CF. Vascular dysfunction occurs prior to the onset of amyloid pathology and Abeta plaque deposits colocalize with endothelial cells in the hippocampus of female APP<sup>swe</sup>/PSEN1<sup>dE9</sup> mice. *Geroscience*. 2024. <https://doi.org/10.1007/s11357-024-01213-0>.
168. van Dinther M, Voorter PHM, Zhang E, van Kuijk SMJ, Jansen JFA, van Oostenbrugge RJ, Backes WH, Staals J. The neurovascular unit and its correlation with cognitive performance in patients with cerebral small vessel disease: a canonical correlation analysis approach. *Geroscience*. 2024;46:5061–73. <https://doi.org/10.1007/s11357-024-01235-8>.
169. Sandor AD, Czinege Z, Szabo A, Losoncz E, Toth K, Mihaly Z, Sotonyi P, Merkely B, Szekely A. Cerebrovascular dysregulation and postoperative cognitive alterations after carotid endarterectomy. *Geroscience*. 2024. <https://doi.org/10.1007/s11357-024-01237-6>.

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