



Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Man, K;Kallies, A;Vasanthakumar, A

Title:

Resident and migratory adipose immune cells control systemic metabolism and thermogenesis

Date:

2022-03-01

Citation:

Man, K., Kallies, A. & Vasanthakumar, A. (2022). Resident and migratory adipose immune cells control systemic metabolism and thermogenesis. *Cellular and Molecular Immunology*, 19 (3), pp.421-431. <https://doi.org/10.1038/s41423-021-00804-7>.

Persistent Link:

<https://hdl.handle.net/11343/302736>

License:

CC BY

REVIEW ARTICLE OPEN



Resident and migratory adipose immune cells control systemic metabolism and thermogenesis

Kevin Man^{1,2}, Axel Kallies^{1,2} and Ajithkumar Vasanthakumar^{1,2,3}

© The Author(s) 2021

Glucose is a vital source of energy for all mammals. The balance between glucose uptake, metabolism and storage determines the energy status of an individual, and perturbations in this balance can lead to metabolic diseases. The maintenance of organismal glucose metabolism is a complex process that involves multiple tissues, including adipose tissue, which is an endocrine and energy storage organ that is critical for the regulation of systemic metabolism. Adipose tissue consists of an array of different cell types, including specialized adipocytes and stromal and endothelial cells. In addition, adipose tissue harbors a wide range of immune cells that play vital roles in adipose tissue homeostasis and function. These cells contribute to the regulation of systemic metabolism by modulating the inflammatory tone of adipose tissue, which is directly linked to insulin sensitivity and signaling. Furthermore, these cells affect the control of thermogenesis. While lean adipose tissue is rich in type 2 and anti-inflammatory cytokines such as IL-10, obesity tips the balance in favor of a proinflammatory milieu, leading to the development of insulin resistance and the dysregulation of systemic metabolism. Notably, anti-inflammatory immune cells, including regulatory T cells and innate lymphocytes, protect against insulin resistance and have the characteristics of tissue-resident cells, while proinflammatory immune cells are recruited from the circulation to obese adipose tissue. Here, we review the key findings that have shaped our understanding of how immune cells regulate adipose tissue homeostasis to control organismal metabolism.

Keywords: adipose tissue; immune cells; metabolism*Cellular & Molecular Immunology* (2022) 19:421–431; <https://doi.org/10.1038/s41423-021-00804-7>

INTRODUCTION

While the primary function of the immune system is to protect against invading pathogens, several landmark studies have uncovered noncanonical functions of immune cells that extend beyond immune surveillance [1, 2]. Indeed, immune cells regulate several vital physiological processes, including tissue regeneration and repair, as well as organismal glucose metabolism [2, 3]. Adipose tissue is an endocrine and energy storage organ that is critical for the regulation of systemic metabolism. Impaired adipose tissue function is therefore closely linked to obesity and type 2 diabetes (T2D), both of which are both major public health problems in the developed and developing world. In addition, obesity is tightly linked to increased cancer incidence and impaired immune responses to infectious diseases [4, 5]. Lean adipose tissue secretes a variety of soluble mediators, including hormones, cytokines and chemokines, which regulate neuronal and metabolic circuits that control satiety, food intake, metabolite storage and catabolism [6, 7]. Notably, adipose tissue contains a diverse array of immune cells that directly impact its function. Thus, adipose tissue integrates organismal energy homeostasis with the immune system.

Adipose tissue can be found in many distinct anatomical locations and accordingly is heterogeneous in its composition and function. Visceral adipose tissue (VAT), for example, is located inside the

abdominal cavity around the inner organs and plays a particularly important role in metabolism. Thus, impaired VAT function is tightly linked to metabolic disease [7, 8]. In contrast, subcutaneous adipose tissue (SCAT), which is found under the dermal layer of the skin, is particularly important for thermal regulation [9]. Indeed, all endothermic animals, including mammals and birds, use heat generated during cellular metabolism to maintain a stable core body temperature (homeothermy), which is critical for survival and allows for adaptation to diverse environmental climates [10, 11]. Most of the adipose tissue in adults consists of white adipose tissue (WAT), which is mainly an energy store. In contrast, brown adipose tissue (BAT), which is morphologically and transcriptionally distinct from WAT, has higher mitochondrial density and expression of mitochondrial uncoupling proteins with specialized functions in heat production (thermogenesis) [12]. Immune cells in adipose tissue also regulate thermogenesis by promoting beiging, which is a process in which WAT upregulates thermogenic transcriptional programs and mitochondrial uncoupling proteins to morphologically resemble BAT. Beige adipose tissue and BAT drive increased energy expenditure during cold exposure to maintain homeothermy, which is a major energy utilization program in endothermic mammals [12].

Adipose tissue is a multicellular organ composed of adipocytes, endothelial cells, mesenchymal stromal cells (MSCs) and immune

¹Peter Doherty Institute for Infection and Immunity, Melbourne, VIC, Australia. ²Department of Microbiology and Immunology, University of Melbourne, Melbourne, VIC, Australia.

³Olivia Newton-John Cancer Research Institute, Melbourne, VIC, Australia. ✉email: kkman@unimelb.edu.au; axel.kallies@unimelb.edu.au; ajith.vasanthakumar@onjcri.org.au

Received: 9 September 2021 Accepted: 28 October 2021

Published online: 26 November 2021

cells [13]. Immune cells enriched in lean adipose tissue are largely anti-inflammatory and promote normal metabolic homeostasis. Many of these cells seed the tissue early in life and become permanently resident in adipose tissue [14]. During aging or the development of obesity, however, proinflammatory immune cells are progressively recruited to adipose tissue, which drives the development of insulin resistance [15]. These changes are in part driven by alterations in the secretome of adipose tissue, favoring the infiltration of immune cells that amplify adipose inflammation. For example, immune cell-derived proinflammatory cytokines such as TNF block insulin signaling by inactivating insulin receptor substrate (IRS), leading to insulin resistance and exacerbating blood glucose levels due to the downregulation of glucose transporters on adipocytes [16]. Thus, during chronic alterations in energy balance, the distribution and composition of adipose-resident immune cell populations is dramatically altered, which directly influences adipose tissue function. Here, we review the ongoing immunological characteristics of adipose tissue in healthy and obese contexts. Specifically, we discuss the differentiation and homeostatic requirements of immune cells and the multidirectional communication between immune cells, adipocytes, endothelial cells and stromal cells that regulate adipose tissue function and systemic metabolism. We propose a model in which adipose tissue homeostasis and function are regulated by an intricate and highly dynamic network of resident and recruited immune cells. Understanding the functional consequences of the interactions between immune and parenchymal cells in adipose tissue will further the development of new therapeutic strategies to treat obesity and diseases linked to obesity, including metabolic diseases and cancer.

TISSUE-RESIDENT IMMUNE CELLS PRESERVE INSULIN SENSITIVITY

WAT deposits, both visceral and subcutaneous, primarily function as sites of nutrient storage and lipid mobilization. These tissues also perform endocrine functions by secreting adipokines (leptin, adiponectin, resistin), lipokines (palmitoleic acid) and chemokines (CCL2) that are involved in modulating local and systemic immunity and metabolic homeostasis [2, 17–19]. While adipocytes themselves secrete many of these mediators, immune cells and MSCs produce multiple cytokines, such as interleukin-4 (IL-4), IL-5, TNF, IFN γ and IL-33, in response to the energy state of adipose tissue, which critically determines the metabolic fitness of organisms [20–23].

Myeloid cells are the most abundant adipose tissue-resident immune cells. Tissue-resident macrophages derive from the yolk sac and are the first immune cells to seed adipose tissue, where they undergo local expansion [24, 25]. In lean adipose tissue, macrophages display an alternatively activated M2 phenotype (CD206+CD301+CD11c $^{-}$) and promote immune suppression. Under homeostatic conditions, M2 macrophages maintain adipocyte turnover by clearing dead adipocytes and debris through phagocytosis and lysosomal activation and by restraining the differentiation of adipocyte progenitors [26]. IL-4 produced by eosinophils preserves the alternative activation status of these macrophages, which in turn produce the anti-inflammatory cytokine IL-10 and the IL-1 decoy receptor to inhibit IL-1 β signaling [22]. Adipocyte-derived adiponectin, which is abundant in lean adipose tissue, also polarizes macrophages to an M2 phenotype [27]. Several transcription factors are implicated in the differentiation of M2 macrophages. In addition to IL-4 and IL-13, the induced transcription factors STAT6, PPAR γ , PPAR δ , KLF4 and IRF4 are key drivers of M2 polarization [23, 28–31].

The role of conventional dendritic cells (cDCs) in adipose tissue homeostasis is still controversial [32]. This is partly due to difficulties in separating these cells from macrophages and monocytes. In most tissues, cDCs can be identified by high

expression of CD11c and MHCII, but activated macrophages and monocytes in adipose tissue can also express prototypic DC markers [32, 33]. A recent study utilizing Zbtb46 reporter mice, in which cDCs can be distinguished from other myeloid cells with confidence, showed that both type 1 and type 2 conventional dendritic cells (cDC1s and cDC2s) play an anti-inflammatory role in lean adipose tissue, by producing IL-10 [34]. Notably, the anti-inflammatory function of cDC1s in adipose tissue requires the Wnt/ β -catenin pathway (*Ctnnb1*) for IL-10 production, while cDC2s upregulate PPAR γ to maintain a tolerogenic anti-inflammatory state in adipose tissue [34]. Thus, both tissue-resident macrophages and cDCs contribute to the maintenance of adipose tissue homeostasis.

Adipose tissue is also rich in innate lymphoid cells (ILCs), which are a major source of type 2 cytokines. Group 2 innate lymphoid cells (ILC2s) dominate lean adipose tissue and play an important role in preserving the T $_H$ 2 milieu by regulating the recruitment of eosinophils, which in turn maintain M2 macrophages [35]. ILC2s depend on IL-33 and are important producers of the type 2 cytokines IL-5 and IL-13, which are thought to contribute to adipose tissue health [2, 35–37]. In addition, invariant natural killer T (NKT) cells, which produce IL-4, IL-13 and IL-10, are enriched in adipose tissue [38, 39]. Compared to splenic NKT cells, adipose NKT cells have a distinct phenotype and express the transcription factors Nfil3, T-bet and GATA3, while being negative for PLZF [39]. In a lean state, NKT cells preserve the M2 phenotype of macrophages by producing IL-10 in an Nfil3-dependent manner and facilitate the expansion of regulatory T cells by producing IL-2 [39]. Similar to those of NKT cells, lineage commitment and the differentiation of ILC2s also depend upon Nfil3, GATA3, T-bet and Id2, whereas PPAR γ is critical for IL-33-dependent activation and functional licensing [40–42]. Notably, both ILC2s and NKT cells are bona fide adipose tissue-resident populations, as demonstrated by parabiosis experiments [36, 39].

Regulatory T (Treg) cells are the major anti-inflammatory adaptive immune cell subset enriched in lean adipose tissue. Tregs are specialized CD4 $^{+}$ T cells with suppressive and tissue-regulatory functions. In adipose tissue, these cells restrain inflammation to prevent the development of insulin resistance. Treg cells that reside in adipose tissue, in particular the VAT, display a unique phenotype as well as distinct transcriptional and cytokine requirements compared to their lymphoid tissue counterparts. These cells express high amounts of the adipocyte transcription factor PPAR γ , which is essential for their development and maintenance [43, 44]. Indeed, the loss of PPAR γ specifically in Tregs results in the specific loss of Tregs in VAT [44]. Unlike their lymphoid tissue counterparts, adipose Treg cells specifically require the cytokine IL-33 for survival and expansion [45]. The distribution, phenotype and homeostatic requirements of Treg cells in subcutaneous adipose tissue, however, are distinct from their counterparts in VAT [36]. The importance of Treg cells in controlling adipose inflammation and insulin resistance has been demonstrated by multiple studies. Systemic ablation of Treg cells using *Foxp3^{DTR}* mice or specific ablation within the adipose tissue using *Pparg^{fl/fl}Foxp3^{Cre}* mice led to the development of insulin resistance during diet-induced obesity [43, 44]. We have shown that treatment of diet-induced or genetically obese mice with recombinant IL-33 could expand Treg cells in adipose tissue, restrain inflammation and revert glucose intolerance [46]. Expanding Treg cells systemically in obese mice using an IL-2 antibody complex also mitigated adipose inflammation and insulin resistance [43]. Consistent with the central role of Treg cells, pioglitazone, a PPAR γ agonist that is used as an antidiabetic drug, was shown to exert its effects at least in part by activating PPAR γ in adipose Treg cells [44]. Treg cells in adipose tissue express the enzyme hydroxyprostaglandin dehydrogenase (HPGD), which converts prostaglandin E $_2$ (PGE $_2$) into 15-keto PGE $_2$, and Treg cell-specific loss of HPGD exacerbates adipose

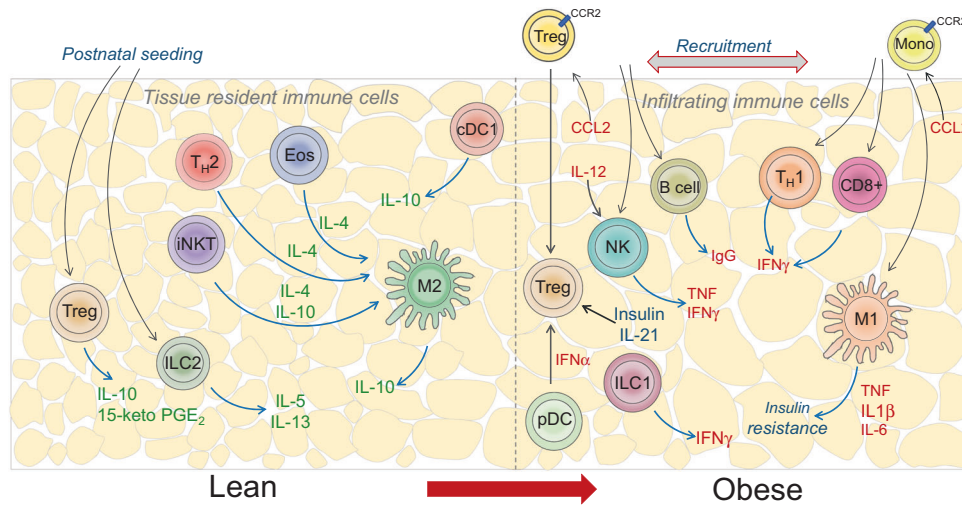


Fig. 1 Immune spectrum in lean and obese adipose tissue. Lean adipose tissue is enriched in anti-inflammatory immune cells and immune cells that contribute to a T_H2 -dominant environment, including Treg cells, eosinophils, T_H2 cells, M2 macrophages, ILC2s, NKT cells and $\gamma\delta$ T cells, and are critical in preserving insulin sensitivity. Importantly, most of these immune cells migrate to adipose tissue early in life, where they establish tissue residency. Obesity displaces these immune cells by facilitating the recruitment of inflammatory immune cells such as M1 macrophages, T_H1 cells, CD8+ T cells, NK cells, ILC1s and B cells. This effect is mediated at least in part by hypoxic and ER stress signals, as well as chemokines such as MCP-1/CCL2

inflammation and insulin resistance in response to diet-induced obesity [47].

Treg cells seed adipose tissue within the first weeks of life [45], increase in number during maturation, and decline during later stages of life [43, 48]. However, even in adult mice, Tregs are continuously recruited to adipose tissue [36]. Indeed, adipose Treg cells are known to arise from peripheral Treg cells that express low levels of PPAR γ . After migrating to adipose tissue, these cells acquire the cardinal features of adipose Treg cells, including expression of the IL-33 receptor ST2 and the terminal differentiation marker KLRG1 [49, 50]. It is currently unclear whether Treg cells recruited during adulthood differ developmentally or functionally from Treg cells that seed adipose tissue during postnatal development. However, during all stages of development, T cell receptor (TCR) signaling is a vital requirement for adipose Treg cell differentiation and maintenance. This effect is mediated through the transcription factors BATF and IRF4, which are induced by TCR signaling, and activate PPAR γ expression and IL-33 responsiveness by inducing the expression of ST2 [46]. Similarly, the transcription factor Blimp1, which is downstream of IRF4, preserves the transcriptional signature of Treg cells in adipose tissue by directly regulating the expression of ST2, PPAR γ and IL-10 [36]. Recently, it was shown that insulin directly regulates the differentiation and function of adipose tissue Treg cells by inducing Hif1 α -dependent PPAR γ expression [51, 52]. Overall, lean adipose tissue is enriched in anti-inflammatory immune cells that are seeded early in life, display hallmarks of tissue residency and play a critical role in maintaining adipose tissue homeostasis and function (Fig. 1).

ADIPOSE TISSUE NICHES FOR IMMUNE CELLS

The notion that adipose tissue is enriched for an array of tissue-resident anti-inflammatory cell types suggests that this tissue contains specialized anatomical niches that promote the survival of these cells. One of the key mediators of such a niche may be IL-33, which plays a critical role in adipose tissue homeostasis by regulating the expansion and activity of ILC2s and Treg cells [2, 45, 46]. We and others have shown that PDGFR α +Pdpn+ MSCs are the main sources of IL-33 in adipose tissue [20, 36, 53, 54] (Fig. 2). Therefore, MSCs facilitate the accumulation of anti-inflammatory lymphocytes and directly contribute to sustaining

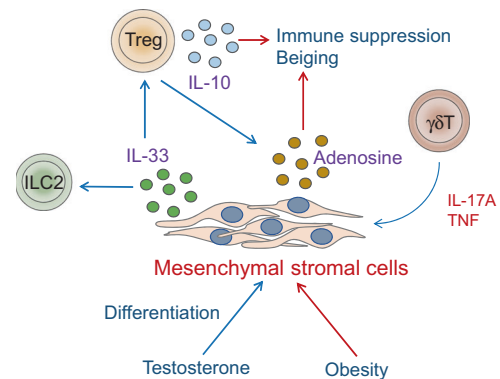


Fig. 2 Immune-stromal cell crosstalk. PDGFR α + mesenchymal stromal cells are the primary sources of IL-33 in adipose tissue. The cytokines TNF and IL-17A produced by $\gamma\delta$ T cells activate IL-33 production in MSCs. Treg cells and ILC2s rely on MSC-derived IL-33 for differentiation, maintenance, survival and activation. MSCs also express CD73, which catalyzes the production of adenosine, which coordinates with Treg cells to promote immune suppression in adipose tissue. Male sex hormones facilitate the differentiation of IL-33+CD73+MSCs, and obesity impairs this population

the T_H2 phenotype of immune cells and homeostasis in adipose tissue [36, 53]. Notably, we found that IL-33+ MSCs develop in a sex hormone-dependent manner, and the male sex hormone testosterone is critical for their differentiation [36]. Male mice show an enrichment in IL-33+ MSCs, and male but not female adipose tissue is specifically enriched in IL-33-dependent Treg cells [36]. On the other hand, ILC2s, which also rely on IL-33, did not show sexual dimorphism in their adipose distribution [36], indicating that other factors contribute to the sex-specific distribution and phenotype of Treg cells. Similarly, sex differences were not observed in Treg cells in SCAT, indicating unique sex hormone-mediated processes in VAT [36]. TNF and IL-17A production by PLZF-expressing $\gamma\delta$ T cells was shown to be important for supporting IL-33 expression in MSCs [54] and therefore indirectly promoted immune suppression in adipose tissue (Fig. 2). Although this is just one example of the immune-stromal cell crosstalk that maintains tissue homeostasis and

residency, it is likely that other factors contribute to the anti-inflammatory state and preserve the health and function of lean adipose tissue. Consistent with this idea, IL-33+ MSCs also exhibit high expression of the immunoregulatory molecule CD73 [36], an ectonucleotidase that converts AMP to adenosine [55], suggesting that MSCs play additional immunomodulatory roles distinct from IL-33-dependent regulation of immune cells (Fig. 2). CD73 is also highly expressed on a subset of VAT-resident Treg cells, thus contributing to adenosine production and beige fat biogenesis [52]. Interestingly, insulin signaling was shown to drive the transition of CD73^{hi}ST2^{lo} to CD73^{lo}ST2^{hi} Treg cells by inducing PPAR γ expression [52], suggesting that VAT contains at least two subsets of Treg cells with distinct functions.

Notably, immune cells in adipose tissue niches have coopted aspects of the transcriptional machinery of adipocytes for their development and function. Indeed, key factors that are known to regulate adipogenesis and adipocyte function, including PPAR γ and the Wnt signaling pathway [56], have been used by adipose-resident immune cells to promote various anti-inflammatory functions. These mechanisms may also confer immune cells with tissue tropic functions, which is evident from the critical role of PPAR γ and IL-33 in adipose resident Treg cells. Thus, these adipose tissue-specific transcriptional networks, which are utilized by different cell types, may act as molecular links between adipocytes, MSCs and immune cells to enable tissue residency and communicate the physiological state of adipose tissue. For example, PPAR γ is a molecule that controls lipid metabolism in adipocytes, MSCs and in immune cells [20, 28, 44]. Indeed, crosstalk with MSCs could be one of the mechanisms by which adipose immune cell residency and homeostasis are maintained.

ADIPOSE IMMUNE INFILTRATION DRIVES INSULIN RESISTANCE

Excess energy intake and low caloric output lead to obesity, a physiological condition marked by adipocyte hyperplasia (increased numbers) and hypertrophy (increased size). The growth and expansion of adipocytes results in hypoxia, the upregulation of oxidative and membrane/ER stress pathways and adipocyte death [57, 58]. Signals generated from stressed and dying adipocytes inhibit insulin action by initiating inhibitory serine phosphorylation of IRS proteins via JNK, MyD88 and IKK- β [59]. Furthermore, hypertrophic adipocytes upregulate chemokines, including CCL2, CCL5 and CCL8, leading to the recruitment of monocytes to adipose tissue, where these cells differentiate in situ into inflammatory M1 macrophages, which are a predominant source of TNF, IL-1 β , IL-6 and IL-18 [24]. Inflammatory signals are then amplified locally and systemically, leading to the recruitment of other immune cells, including NK cells, ILC1s, B cells and CD8+ T cells [60–64], eventually replacing and inhibiting the function of resident M2 macrophages, Treg cells, NKT cells and ILC2s [2, 38, 43]. Obesity also results in dysregulated endocrine function. For example, obese adipose tissue exhibits reduced secretion of adiponectin, which mediates insulin sensitivity and is one of the most abundant adipokines secreted by adipocytes [65]. In contrast, the levels of the satiety hormone leptin are increased in obesity to counteract food intake [66, 67]. Notably, adipokines also have a direct impact on immune cell differentiation and activation. Leptin, for example, regulates multiple immune cells, including macrophages, NK cells and Treg cells [68]. Obesity therefore changes the local and systemic cytokine and adipokine environment and has profound implications on systemic metabolism, inflammation and immunity.

Macrophages are thought to be key mediators of adipose tissue inflammation and metabolic disease [69]. In obese adipose tissue, M1 macrophages are clustered around dead adipocytes and form crown-like structures, unlike resident M2 macrophages, which are interspersed between adipocytes and in the vasculature [70]. TNF production by macrophages recruited from the periphery is

central to insulin resistance [21]. TNF inhibits glucose uptake by adipocytes by downregulating the expression of glucose transporters (slc2a4) [16] and reduce insulin signaling by inducing inhibitory serine phosphorylation of insulin receptor tyrosine kinase proteins [71]. Analogous to TNF, IL-6 can also inhibit insulin signaling by promoting serine phosphorylation of IRS proteins [72]. During obesity, recruited macrophages phagocytose adipocytes with high lipid levels to become lipid-laden macrophages [73]. Intracellular lipids are known to activate inflammatory pathways in adipose tissue macrophages [74], although precisely how M1 macrophages are activated has been a subject of controversy. Saturated fatty acid (SFA) signaling and TLR receptors play important roles in inflammatory gene activation in macrophages [75], while the activation of inflammasome (NLRP3)- and caspase-1-dependent pathways are important for mature IL-1 β and IL-18 secretion by adipose macrophages [75]. Given the role of TLR4 in sensing not only LPS but also saturated fatty acids, it is widely believed that the SFA-TLR4 axis is involved in macrophage activation [76–78]. A recent paper, however, showed that SFAs do not activate macrophages via TLR4 but instead induce JNK signaling to reprogram macrophage metabolism during inflammation [79]. Accordingly, stress-induced JNK signaling is critical for the differentiation of M1 macrophages during obesity, and specific ablation of JNK in macrophages protected mice from diet-induced insulin resistance [80, 81].

While the M1 and M2 nomenclature provides a useful framework for the study of tissue macrophages, it is insufficient to describe the inflammatory status of adipose macrophages during obesity, as these cells often express markers of both M1 and M2 macrophages [14]. M1 macrophages are often identified by the expression of CD11c, and the ablation of CD11c+ cells (using CD11c-DTR mice) had a positive effect on ameliorating diet-induced insulin resistance [32]. However, CD11c is also expressed by conventional dendritic cells and monocytes, making it difficult to interpret some of these studies. While CD11c+ macrophages can be distinguished from dendritic cells by the expression of CD64 and MerTK, further investigations are required to delineate the functions of adipose macrophages and DCs. A recent study exploring macrophage heterogeneity using single-cell RNA-seq revealed a distinct inflammatory population that was CD9+Ly6C–, expressed genes related to lipid metabolism and was distributed in the obese adipose tissue of both mice and humans [82]. However, the precise function of this population has yet to be determined. Notably, inflammatory macrophages also contribute to the maintenance of adipose tissue homeostasis. During lipolysis and fasting-induced weight loss, inflammatory macrophages can phagocytose nonesterified fatty acids that are liberated during lipolysis and from dead adipocytes to prevent lipotoxicity [83]. Furthermore, blocking IL-6 trans-signaling prevented the accumulation of M1 macrophages but did not improve insulin tolerance [84]. Thus, the precise role of macrophages is likely to be multifaceted, and the signals that control their recruitment and function in adipose tissue remain a topic of great interest. Overall, further work is required to determine the developmental origin and function of this heterogeneous adipose myeloid population and to molecularly characterize distinct cell types that contribute to adipose tissue homeostasis and function.

Another important source of TNF during obesity, besides from macrophages, are NK cells, which are limited in distribution to epididymal VAT depots [85]. Obesity drives the upregulation of the NK cell activating receptor NCR1 on adipocytes. This, in turn, triggers IFN γ production by NK cells and facilitates the differentiation of inflammatory macrophages that promote insulin resistance [63]. Similarly, IFN γ produced by ILC1s reinforces inflammatory macrophage polarization [86] while simultaneously counteracting IL-33-mediated activation of ILC2s [37]. Accordingly, mice lacking *Nfil3*, which is critical for NK cell differentiation, or genetic loss of

NCR1 or IFN γ resulted in improved insulin sensitivity [63], while treating mice with IL-15 to expand NK cells led to insulin resistance [85].

Although multiple lines of evidence suggest that macrophages and NK cells are critical in the initiation of adipose tissue inflammation and insulin resistance, many other cell types have been implicated in these processes. For example, B cells have been shown to expand in obese adipose tissue and promote the activation of M1 macrophages, as well as CD4 $^+$ and CD8 $^+$ T cells. B cell deficiency protected mice from the development of insulin resistance, whereas the transfer of pathogenic IgG from obese mice into B cell-deficient mice induced inflammation and insulin resistance [62]. Similarly, T cells play central roles in adipose tissue inflammation. For example, adipose infiltration of CD8 $^+$ effector T cells precedes macrophage infiltration, suggesting that CD8 $^+$ T cells initiate obesity-driven adipose inflammation [60]. Furthermore, CD4 $^+$ T cells, particularly T $_H$ 1 cells, have been shown to play a proinflammatory role during obesity [87]. In line with this conclusion, deficiency of the transcription factor T-bet, which regulates the differentiation of T $_H$ 1 cells and many other immune cell types [88], results in improved insulin sensitivity [87]. This effect appears to be intrinsic to CD4 $^+$ T cells, as adoptive transfer of wild-type CD4 $^+$ T cells promoted insulin resistance in *Rag* $^{-/-}$ mice fed a high-fat diet, whereas the transfer of T-bet-deficient CD4 $^+$ T cells failed to initiate inflammation [87]. In support of a key role of T cells in metabolic disease, treating genetically obese *ob/ob* mice with anti-CD3 antibody minimized the expansion of T $_H$ 1 cells and mitigated adipose inflammation and insulin resistance [61]. Collectively, multiple types of immune cells from the innate and adaptive arms of the immune system infiltrate adipose tissue during obesity to participate in the inflammatory cascade that culminates in insulin resistance and metabolic dysfunction (Fig. 1). In contrast, tissue-resident immune cell populations found in adipose tissue during homeostasis are critical for maintaining adipocyte homeostasis and function.

IMPAIRED IMMUNOSUPPRESSION MECHANISMS IN OBESITY

Obesity-induced adipose inflammation and the influx of newly recruited inflammatory immune cells impair immunosuppressive mechanisms and further amplify inflammation and insulin resistance. Obesity has been shown to result in a loss of Treg cells and ILC2s specifically in VAT [2, 43]. Although the signals that lead to obesity-mediated Treg cell loss are not fully understood, excessive weight gain was found to drive the downregulation of *Pparg* and other adipose Treg cell-specific genes [48]. Obesity is also associated with the loss of IL-33-expressing MSCs, which affects not only Treg cells but also ILC2s [20, 36]. Finally, obesity results in increased IFN α production by plasmacytoid dendritic cells, which was shown to be toxic to Treg cells [89]. Hyperinsulinemia also directly impairs Treg cell population expansion and function, and accordingly, ablation of the insulin receptor specifically in Treg cells led to the expansion of adipose Treg cells, reduced adipose tissue inflammation and restored metabolic health [51]. Finally, IL-21 has been shown to be increased in adipose tissue during obesity, and IL-21 deficiency leads to the expansion of adipose tissue Treg cells and preserves insulin sensitivity [90]. Similar to Treg cells, ILC2s are impaired through the loss of IL-33-expressing MSCs during obesity [53]. Furthermore, IFN γ impairs the activation of ILC2s in adipose tissue during aging and obesity [37]. This, in turn, may also affect Treg cells, which are thought to interact with ILC2s through ICOS [37].

Interestingly, our recent work revealed an inflammation-driven pathway that recruits Treg cells to adipose tissue. Adipose tissue-derived CCL2 attracts Treg cells to adipose tissue [36], exploiting the same molecular pathway utilized by monocytes to infiltrate adipose tissue [36, 91]. Thus, the inflammation-mediated loss of Treg cells is counteracted by simultaneous de novo recruitment of

peripheral Treg cells. Overall, however, Treg cell influx and expansion decline during the late stages of obesity and with physiological aging, allowing for the expansion of proinflammatory immune cells, exacerbating adipose tissue inflammation [43]. Importantly, the decline in adipose Treg cells and ILC2s during obesity is conserved across mice and humans [2, 43], suggesting that immune cell homeostasis is mediated by evolutionarily conserved mechanisms. In summary, obesity not only facilitates the infiltration of inflammatory immune cells into adipose tissue but also disables the protective mechanisms required to maintain insulin sensitivity and glucose homeostasis.

IMMUNE CONTROL OF THERMOGENESIS

In addition to energy storage and the regulation of systemic metabolism, adipose tissue also plays a critical role in thermogenesis, which is impacted by immune cells. Mammals have specialized heat-generating adipose tissue deposits, including brown and beige adipose tissue, which have high mitochondrial density and expression of mitochondrial uncoupling protein 1 (UCP1), a transmembrane protein that creates a proton channel in the mitochondrial inner membrane to allow the translocation of protons and the dissipation of the electrochemical gradient, leading to the uncoupling of oxidative phosphorylation from the synthesis of ATP and the generation of heat as a byproduct [92]. Upon environmental cold exposure or activation of the sympathetic nervous system via beta-3 adrenergic stimulation, inguinal or subcutaneous WAT deposits can also engage in adaptive thermogenesis by upregulating the expression of UCP1. UCP1 $^+$ cells are known as 'beige' adipocytes and are transcriptionally distinct from white or brown adipocytes [93, 94]. These cells are derived from Myf5 $^+$ PDGFR α^+ precursor cells [95, 96] or by the direct conversion or transdifferentiation of existing white adipocytes [97, 98]. However, despite its central role in thermogenesis, UCP1 is not essential because *Ucp1* $^{-/-}$ mice show no defects in adaptation to long-term cold exposure [99]. UCP1-independent mechanisms of thermogenesis occur predominantly in the form of futile metabolic cycling processes, during which tandem inverse reactions occur simultaneously, and the only net effect is the hydrolysis of ATP and dissipation of energy in the form of heat [100–103].

In a lean state, adipose tissue-resident immune cells participate in the regulation of adaptive thermogenesis predominantly via the secretion of cytokines that influence the differentiation and function of mesenchymal stromal cells and adipocyte precursor cells or by controlling the differentiation and phenotype of other adipose-resident immune cells, indirectly impacting adipocyte precursors. M2 macrophages support the terminal differentiation of PDGFR α^+ stromal cells to beige adipocytes upon cold exposure [104, 105] (Fig. 3). In a related circuit, eosinophil-derived IL-4, together with ILC2-derived IL-13, stimulates the proliferation and differentiation of PDGFR α^+ stromal cells to the beige adipocyte lineage [104, 106]. Accordingly, mice lacking eosinophils, IL-4 and IL-13, or IL-4Ra, or mice with a macrophage-specific deletion of *IL-4ra*, exhibit deficiencies in beige adipocyte formation, cold-induced thermogenesis and decreased energy expenditure [106] (Fig. 3). IL-33-dependent ILC2s are also necessary for sustaining the proliferation and commitment of PDGFR α^+ adipocyte precursor cells to the beige lineage [106]. Additionally, IL-33 induces the expression of methionine-enkephalin peptides in ILC2s, which induces beige fat biogenesis via an unknown mechanism [107]. Recently, $\gamma\delta$ TCR T cells were shown to be important for regulating thermogenesis by directing the innervation of BAT and by increasing the expression of tyrosine hydroxylase through IL-17F- and adipocyte IL-17Rc-dependent signaling [108] (Fig. 3). Finally, activated NKT cells contribute to WAT beiging by increasing the expression of FGF21, a hormone involved in stimulating adipocyte glucose uptake [109] (Fig. 3).

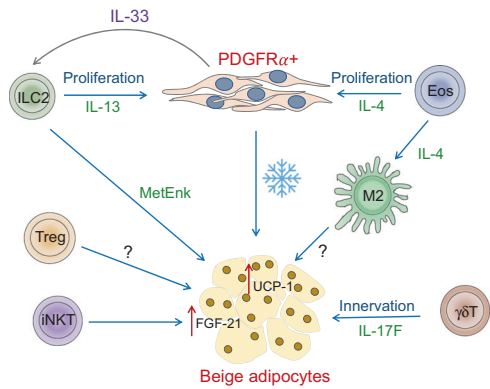


Fig. 3 Immune regulation of thermogenesis. Upon cold exposure, M2 macrophages promote the differentiation of beige adipocytes from PDGFR α + stromal cells. This process is mediated by IL-4 produced by eosinophils. IL-4 and IL-13 derived from eosinophils and ILC2s also promote the proliferation of PDGFR α + cells to facilitate beiging. While the mechanism by which M2 macrophages promote beiging is unclear, ILC2s and NKT cells promote the upregulation of UCP-1 and FGF-21 in adipose tissue, respectively. Innervation mediated by $\gamma\delta$ T cell-derived IL-17F also promotes beiging and thermogenesis. The role of Treg cells in thermogenesis is controversial

Adipokines produced by adipocytes can also induce thermogenesis and increase energy expenditure. For example, adiponectin is increased in subcutaneous WAT by chronic cold exposure and promotes beige adipocyte formation and BAT UCP1 expression via its effects on macrophage polarization, which increases M2 macrophage polarization in WAT without affecting type 2 cytokine expression or ILC2 abundance [110]. BAT also contains resident innate and adaptive immune cells, although their tissue tropic functions are not well understood. For example, BAT harbours transcriptionally distinct Treg cells that regulate the inflammatory tone of the tissue, although Treg depletion seems to have only minor effects on BAT thermogenic gene programs and thermogenesis [111, 112]. It remains to be determined whether BAT-resident macrophages and ILC2s also participate in regulating thermogenesis in a similar manner to their counterparts in WAT.

Energy balance is a critical regulator of the extent of adaptive thermogenesis in adipose tissue. Inflammatory pathways activated by obesity inhibit adaptive thermogenic programs. For example, IL-1 β and TNF signaling in adipocytes inhibits UCP-1 expression and cold-induced thermogenesis [113, 114]. In a similar manner, I κ B kinase- ϵ (IKK ϵ) $^{-/-}$, IRF3- and TGF- β -dependent inflammatory pathways inhibit adipose tissue thermogenesis and energy expenditure during obesity, and blocking these pathways stimulates adipose tissue thermogenesis [115, 116]. Interestingly, exercise and energy restriction regimens (caloric restriction or intermittent fasting) promote beige adipocyte formation and energy expenditure through immune cell-derived cytokines. Specifically, the positive effect of exercise and fasting on thermogenesis requires IL-4 production by eosinophils and alternative activation of macrophages in WAT [104] and signaling in an IL-4R α - and STAT6-dependent manner [105]. Exercise was also shown to increase the production of meteorin-like hormone in muscle and adipose tissue, which stimulated the differentiation of M2 macrophages, thereby indirectly increasing beige differentiation and thermogenic capacity [105]. Intermittent fasting, while acting on the IL-4/IL-13 signaling cascade, also influences microbiota composition and enhances hepatic production of FGF21 to increase beiging in a PPAR α -dependent manner [117]. Whether exercise and caloric restriction regimens directly influence adipocyte thermogenesis and the mechanisms remain to be elucidated.

While fasting and caloric restriction are associated with increases in health span and longevity [118], physiological aging induces a decline in the thermogenic capacity of adipose tissue, which is accompanied by increased accumulation of inflammatory B cells, $\alpha\beta$ -TCR T cells and M1 macrophages with senescence-associated gene signatures and pathways associated with catecholamine catabolism to suppress lipolysis [119–121]. With the increase in inflammatory senescent immune cells in adipose tissue, there is an accompanying decline in M2 macrophages and ILC2s, which are important for preserving thermogenic capacity, as detailed above. ILC2s become dysfunctional during aging, and accordingly, IL-33-mediated expansion of aged ILC2s failed to promote thermogenesis in aged mice. Accordingly, the transfer of ILC2s from young mice restored thermogenic capacity in aged mice during cold exposure [122].

Similar to insulin signaling, the thermogenic capacity of adipose tissue also depends upon the activity of various anti-inflammatory resident immune cells, which are impaired by inflammatory signaling cascades that are upregulated during obesity and decline in function with physiological aging. Overall, adipose tissue immune cells have indispensable roles in regulating beige and brown thermogenic adipocyte differentiation, thermogenic capacity and systemic energy expenditure by regulating the sympathetic innervation of adipose tissue and the differentiation of thermogenic PDGFR α + adipocyte precursor cells.

CONTROVERSIES AND OPEN QUESTIONS

Adipose tissue contains a multitude of different immune and stromal cells and is impacted by factors such as diet, age and sex. Most studies, however, focus on the roles of only one or a few different cell types under one set of conditions. Thus, it is not surprising that controversies have arisen about the relative impacts of certain cell types on adipose tissue functions. Conflicting results are most likely due to differences in experimental design. Sex in particular plays a critical role, and male mice are far more susceptible to the development of insulin resistance and obesity than female mice [36, 123]. This difference has been attributed to adipose tissue-intrinsic differences [124] and differences in the abundance and phenotype of Tregs and stromal cells in VAT [36]. Another important factor is age. NKT cells are particularly abundant in adipose tissue in mice between 8 and 16 weeks of age [43], while Treg cells accumulate until 6–8 months of age before declining [43]. Similarly, the length of dietary interventions or the microbiota [125], the use of nonlittermate controls or differences in the experimental readouts [126] have been shown to play critical roles in the outcomes of studies of adipose tissue function.

Even understanding the role of individual cell populations is not without challenges. NKT cells, for example, can exert both pro- and anti-inflammatory effects, and given their abundance in lean VAT in mice and humans [38, 127], several studies have examined their role in obesity-induced inflammation with partially contradictory results [125, 128–133]. This discrepancy may be due to differences in the genetic models used to deplete NKT cells (*Cd1d* $^{-/-}$ or *Ja18* $^{-/-}$ mice), which lack different types of NKT cells [134]. Critically, the ablation of CD1d specifically on adipocytes resulted in inflammation and insulin resistance, indicating an adipocyte-intrinsic role for CD1d [135]. Single-cell sequencing technologies to understand the heterogeneity of different NKT cell subsets and the use of cell type-specific knockout models are important to specifically target NKT cells during early and established obesity to fully understand their role in metabolic regulation.

Similar to NKT cells, the role of Treg cells is also somewhat controversial in the context of adipose tissue biology. Although these cells are widely accepted as mediators that suppress inflammation, including that in adipose tissue [136], they have

also been implicated in exacerbating adipose inflammation [137, 138]. The examination of mice with Treg cell-specific ablation of PPAR γ suggested a negative role of adipose tissue Treg cells during aging [137]. Similarly, two studies suggested that Treg cells dampen adipose tissue beiging in a Blimp1- and IL-10-dependent manner [137, 139]. In our own experiments, the loss of Blimp1 in Treg cells resulted in the depletion of adipose tissue-resident Treg cells and led to impaired systemic glucose homeostasis [36]. One must consider alternative explanations for the observed phenotypes of mice with Treg cell-specific deletion of Blimp1 or IL-10. Deletion of both molecules is known to affect Treg cells in many tissues, including in the gastrointestinal tract [140], thus resulting in tissue inflammation and weight loss and indirectly contributing to increased glucose metabolism. Similarly, it is possible that Treg cells that are impaired by aging or the loss of critical regulatory molecules, such as PPAR γ , lose expression of their lineage-defining transcription factor Foxp3 and acquire an inflammatory phenotype, subsequently contributing to adipose inflammation. Fate mapping mouse models would help in understanding the abundance of ex-Treg cells and their contribution to inflammatory status in different mouse models. Finally, it is currently unclear whether regulatory circuits that are active in murine Treg cells also contribute to Treg cell differentiation and function in human adipose tissue. For example, our data suggested that IL-33 signaling plays a role in mouse and human adipose tissue Treg cells [46], while others have failed to identify ST2+ Treg cells in human adipose tissue [141]. Given the paucity of Treg cells in female and obese adipose tissue [36, 43, 46], future studies need to take into account the sex and adiposity of human subjects. In-depth characterization of immune cells embedded in human adipose tissue will help to further the understanding of evolutionarily conserved immune-mediated mechanisms. Single-cell genomic technologies may further illuminate the complexity of immune cells in human adipose tissue and variations associated with sex, age and adiposity. Finally, the role of Treg cells in thermogenesis is controversial and requires further investigation. While one study has shown that the loss of Treg cell function (Blimp1 or IL10 deletion) improves thermogenesis [138], another study demonstrated that adoptive transfer of Treg cells promoted the beiging of subcutaneous adipose tissue [142].

There is a common idea that lean adipose tissue is T_H2-biased, and obesity promotes T_H1-mediated inflammation. Indeed, several types of immune cells in lean adipose tissue produce T_H2 cytokines, such as IL-4, IL-13, IL-5 and IL-10 [2, 38, 44]. MSCs also contribute to T_H2 inflammation in lean adipose tissue by producing IL-33 [20, 36, 53]. Consistent with the T_H2 profile, immune cells such as ILC2s and Treg cells also express the T_H2 transcription factor Gata3 in lean adipose tissue [2, 36]. While systemic loss of T-bet protected against the development of insulin resistance [87], the ablation of IFN γ , the major T_H1 cytokine, only modestly improved insulin sensitivity in obese mice [143]. However, there have been no systematic studies that have examined the role of T_H1 and T_H2 cytokines in female mice. Given the pronounced differences in susceptibility to metabolic diseases [36, 144] and in VAT Treg cell phenotype and function between males and females [36], revisiting the T_H1/T_H2 model in adipose tissue health and disease is urgently needed.

Notably, adaptive thermogenesis influences immune recruitment and composition in adipose and peripheral tissues, including the liver and gastrointestinal tract [145–147]. Modulating thermogenic programs, such as by increasing housing temperature to the thermoneutral zone, can alter phenotypes driven by obesity-induced inflammation, atherosclerosis, bacterial sepsis, nonalcoholic fatty liver disease, colitis and cancer [145, 147, 148] to those more consistently observed in human physiology. Normal vivarium conditions impose significant thermal stress on experimental animals, which can obscure experimental results and represent an additional obstacle for predictive modeling of human

diseases and therapies, as humans spend most of their lives under thermoneutral conditions [149]. Thermogenesis and ambient housing temperature, therefore, should be experimental variables that are carefully considered when carrying out metabolic and immunological studies.

Finally, it must be acknowledged that there are certain limitations in regard to animal models and their use in obesity and metabolic studies. For example, there are differences in pancreatic islet architecture in mice compared with humans [150]. Furthermore, genome-wide association studies showed that human obesity is polygenic in nature and associated with over 100 candidate genes, making monogenetic mouse models of obesity less amenable to therapeutic translation [151]. Therefore, the validation of findings based on murine adipose tissue is required in both healthy and obese humans of both sexes to fully understand the role of the adipose immune system in regulating metabolism and obesity-related chronic inflammatory diseases. Overall, many questions remain, and further work is required to untangle the complicated relationships between adipose tissue function and the immune system.

CONCLUDING REMARKS

Over the last two decades, several landmark studies have uncovered the remarkable impact of the immune system on systemic metabolism, as outlined in this review. The quest to understand the precise role of each immune cell subset in protecting adipose tissue homeostasis or contributing to adipose inflammation and associated pathology has revealed cellular networks involving immune cells, adipocytes and MSCs. It has become apparent that most immune cells that are anti-inflammatory and contribute to the preservation of systemic glucose metabolism populate adipose tissue early in life, expand locally and are not frequently replenished by circulating immune cells. The adaptation to adipose tissue microenvironments and tissue-derived signals, as well as the use of molecular regulators, such as IL-33 and PPAR γ , are common in many of these adipose tissue-resident immune cells. Stromal cells may play an important yet poorly defined role in mediating the development, maintenance, and intercellular communication of adipose tissue immune cells.

Reestablishing immunoregulatory mechanisms in adipose tissue could be a therapeutic approach to treat metabolic inflammation and insulin resistance. Additionally, there is great interest in understanding the cellular and molecular pathways that increase and sustain beige and brown adipocyte thermogenesis as a mechanism to regulate energy expenditure and metabolism in humans, and there are potential implications for the management of obesity and diabetes [152]. Overall, understanding the population dynamics of immune cells and their functions in adipose tissue will aid in the design of novel therapeutic interventions that dampen adipose inflammation to restore insulin sensitivity and glucose homeostasis.

REFERENCES

1. Theret M, Mounier R, Rossi F. The origins and non-canonical functions of macrophages in development and regeneration. *Development*. 2019;146:dev156000. <https://doi.org/10.1242/dev.156000>.
2. Brestoff JR, Kim BS, Saenz SA, Stine RR, Monticelli LA, Sonnenberg GF, et al. Group 2 innate lymphoid cells promote beiging of white adipose tissue and limit obesity. *Nature*. 2015;519:242–6. <https://doi.org/10.1038/nature14115>.
3. Davies LC, Jenkins SJ, Allen JE, Taylor PR. Tissue-resident macrophages. *Nat Immunol*. 2013;14:986–95. <https://doi.org/10.1038/ni.2705>.
4. Stone TW, McPherson M, Gail Darlington L. Obesity and cancer: existing and new hypotheses for a causal connection. *EBioMedicine*. 2018;30:14–28. <https://doi.org/10.1016/j.ebiom.2018.02.022>.
5. Rebello CJ, Kirwan JP, Greenway FL. Obesity, the most common comorbidity in SARS-CoV-2: is leptin the link? *Int J Obes*. 2020;44:1810–7. <https://doi.org/10.1038/s41366-020-0640-5>.

6. Nawrocki AR, Scherer PE. The delicate balance between fat and muscle: adipokines in metabolic disease and musculoskeletal inflammation. *Curr Opin Pharm.* 2004;4:281–9. <https://doi.org/10.1016/j.coph.2004.03.003>.
7. Ouchi N, Parker JL, Lugus JJ, Walsh K. Adipokines in inflammation and metabolic disease. *Nat Rev Immunol.* 2011;11:85–97. <https://doi.org/10.1038/nri2921>.
8. West-Eberhard MJ. Nutrition, the visceral immune system, and the evolutionary origins of pathogenic obesity. *Proc Natl Acad Sci USA.* 2019;116:723–31. <https://doi.org/10.1073/pnas.1809046116>.
9. van der Lans AA, Hoeks J, Brans B, Vijgen GH, Visser MG, Vosselman MJ, et al. Cold acclimation recruits human brown fat and increases nonshivering thermogenesis. *J Clin Invest.* 2013;123:3395–403. <https://doi.org/10.1172/JCI68993>.
10. Harms M, Seale P. Brown and beige fat: development, function and therapeutic potential. *Nat Med.* 2013;19:1252–63. <https://doi.org/10.1038/nm.3361>.
11. Man K, Kutayin VI, Chawla A. Tissue immunometabolism: development, physiology, and pathobiology. *Cell Metab.* 2017;25:11–26. <https://doi.org/10.1016/j.cmet.2016.08.016>.
12. Rosell M, Kaforou M, Frontini A, Okolo A, Chan YW, Nikolopoulou E, et al. Brown and white adipose tissues: intrinsic differences in gene expression and response to cold exposure in mice. *Am J Physiol Endocrinol Metab.* 2014;306:E945–964. <https://doi.org/10.1152/ajpendo.00473.2013>.
13. Reilly SM, Sattler AR. Adapting to obesity with adipose tissue inflammation. *Nat Rev Endocrinol.* 2017;13:633–43. <https://doi.org/10.1038/nrendo.2017.90>.
14. Russo L, Lumeng CN. Properties and functions of adipose tissue macrophages in obesity. *Immunology.* 2018;155:407–17. <https://doi.org/10.1111/imm.13002>.
15. Sam S, Mazzone T. Adipose tissue changes in obesity and the impact on metabolic function. *Transl Res.* 2014;164:284–92. <https://doi.org/10.1016/j.trsl.2014.05.008>.
16. Lumeng CN, Deyoung SM, Sattler AR. Macrophages block insulin action in adipocytes by altering expression of signaling and glucose transport proteins. *Am J Physiol Endocrinol Metab.* 2007;292:E166–174. <https://doi.org/10.1152/ajpendo.00284.2006>.
17. Schwartz DR, Lazar MA. Human resistin: found in translation from mouse to man. *Trends Endocrinol Metab.* 2011;22:259–65. <https://doi.org/10.1016/j.tem.2011.03.005>.
18. Rosen ED, Spiegelman BM. What we talk about when we talk about fat. *Cell.* 2014;156:20–44. <https://doi.org/10.1016/j.cell.2013.12.012>.
19. Yamauchi T, Kamon J, Waki H, Terachi Y, Kubota N, Hara K, et al. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipodystrophy and obesity. *Nat Med.* 2001;7:941–6. <https://doi.org/10.1038/90984>.
20. Spallanzani RG, Zemmour D, Xiao T, Jayewickreme T, Li C, Bryce PJ et al. Distinct immunocyte-promoting and adipocyte-generating stromal components coordinate adipose tissue immune and metabolic tenors. *Sci Immunol.* 2019;4. <https://doi.org/10.1126/sciimmunol.aaw3658>.
21. Uysal KT, Wiesbrock SM, Marino MW, Hotamisligil GS. Protection from obesity-induced insulin resistance in mice lacking TNF- α function. *Nature.* 1997;389:610–4. <https://doi.org/10.1038/39335>.
22. Wu D, Molofsky AB, Liang HE, Ricardo-Gonzalez RR, Jouihan HA, Bando JK, et al. Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. *Science.* 2011;332:243–7. <https://doi.org/10.1126/science.1201475>.
23. Kang K, Reilly SM, Karabacak V, Gangl MR, Fitzgerald K, Hatano B, et al. Adipocyte-derived Th2 cytokines and myeloid PPAR δ regulate macrophage polarization and insulin sensitivity. *Cell Metab.* 2008;7:485–95. <https://doi.org/10.1016/j.cmet.2008.04.002>.
24. Hassnain Waqas SF, Noble A, Hoang AC, Ampem G, Popp M, Strauß S, et al. Adipose tissue macrophages develop from bone marrow-independent progenitors in *Xenopus laevis* and mouse. *J Leukoc Biol.* 2017;102:845–55. <https://doi.org/10.1189/jlb.1A0317-082RR>.
25. Schulz C, Gomez Perdiguero E, Chorro L, Szabo-Rogers H, Cagnard N, Kierdorf K, et al. A lineage of myeloid cells independent of Myb and hematopoietic stem cells. *Science.* 2012;336:86–90. <https://doi.org/10.1126/science.1219179>.
26. Nawaz A, Aminuddin A, Kado T, Takikawa A, Yamamoto S, Tsuneyama K, et al. CD206(+) M2-like macrophages regulate systemic glucose metabolism by inhibiting proliferation of adipocyte progenitors. *Nat Commun.* 2017;8:286. <https://doi.org/10.1038/s41467-017-00231-1>.
27. Ohashi K, Parker JL, Ouchi N, Higuchi A, Vita JA, Gokke N, et al. Adiponectin promotes macrophage polarization toward an anti-inflammatory phenotype. *J Biol Chem.* 2010;285:6153–60. <https://doi.org/10.1074/jbc.M109.088708>.
28. Odegaard JI, Ricardo-Gonzalez RR, Goforth MH, Morel CR, Subramanian V, Mukundan L, et al. Macrophage-specific PPAR γ controls alternative activation and improves insulin resistance. *Nature.* 2007;447:1116–20. <https://doi.org/10.1038/nature05894>.
29. Liao X, Sharma N, Kapadia F, Zhou G, Lu Y, Hong H, et al. Kruppel-like factor 4 regulates macrophage polarization. *J Clin Invest.* 2011;121:2736–49. <https://doi.org/10.1172/JCI45444>.
30. Eguchi J, Kong X, Tenta M, Wang X, Kang S, Rosen ED. Interferon regulatory factor 4 regulates obesity-induced inflammation through regulation of adipose tissue macrophage polarization. *Diabetes.* 2013;62:3394–403. <https://doi.org/10.2337/db12-1327>.
31. Sica A, Mantovani A. Macrophage plasticity and polarization: in vivo veritas. *J Clin Invest.* 2012;122:787–95. <https://doi.org/10.1172/JCI59643>.
32. Patsouris D, Li PP, Thapar D, Chapman J, Olefsky JM, Neels JG. Ablation of CD11c-positive cells normalizes insulin sensitivity in obese insulin resistant animals. *Cell Metab.* 2008;8:301–9. <https://doi.org/10.1016/j.cmet.2008.08.015>.
33. Stefanovic-Racic M, Yang X, Turner MS, Mantell BS, Stolz DB, Sumpter TL, et al. Dendritic cells promote macrophage infiltration and comprise a substantial proportion of obesity-associated increases in CD11c+ cells in adipose tissue and liver. *Diabetes.* 2012;61:2330–9. <https://doi.org/10.2337/db11-1523>.
34. MacDougall CE, Wood EG, Loschko J, Scaglioni V, Cassidy FC, Robinson ME, et al. Visceral adipose tissue immune homeostasis is regulated by the crosstalk between adipocytes and dendritic cell subsets. *Cell Metab.* 2018;27:588–601 e584. <https://doi.org/10.1016/j.cmet.2018.02.007>.
35. Molofsky AB, Nussbaum JC, Liang HE, Van Dyken SJ, Cheng LE, Mohapatra A, et al. Innate lymphoid type 2 cells sustain visceral adipose tissue eosinophils and alternatively activated macrophages. *J Exp Med.* 2013;210:535–49. <https://doi.org/10.1084/jem.20121964>.
36. Vasanthakumar A, Chisanga D, Blume J, Gloury R, Britt K, Henstridge DC, et al. Sex-specific adipose tissue imprinting of regulatory T cells. *Nature.* 2020;579:581–5. <https://doi.org/10.1038/s41586-020-2040-3>.
37. Molofsky AB, Van Gool F, Liang HE, Van Dyken SJ, Nussbaum JC, Lee J, et al. Interleukin-33 and Interferon-gamma Counter-Regulate Group 2 Innate Lymphoid Cell Activation during Immune Perturbation. *Immunity.* 2015;43:161–74. <https://doi.org/10.1016/j.immuni.2015.05.019>.
38. Lynch L, Nowak M, Varghese B, Clark J, Hogan AE, Toxavidis V, et al. Adipose tissue invariant NKT cells protect against diet-induced obesity and metabolic disorder through regulatory cytokine production. *Immunity.* 2012;37:574–87. <https://doi.org/10.1016/j.immuni.2012.06.016>.
39. Lynch L, Michelet X, Zhang S, Brennan PJ, Busslinger M, Lester C, et al. Regulatory iNKT cells lack expression of the transcription factor PLZF and control the homeostasis of T(reg) cells and macrophages in adipose tissue. *Nat Immunol.* 2015;16:85–95. <https://doi.org/10.1038/ni.3047>.
40. Fali T, Aychek T, Ferhat M, Jouzeau JY, Busslinger M, Moulin D, et al. Metabolic regulation by PPAR γ is required for IL-33-mediated activation of ILC2s in lung and adipose tissue. *Mucosal Immunol.* 2021;14:585–93. <https://doi.org/10.1038/s41385-020-00351-w>.
41. Seillet C, Rankin LC, Groom JR, Mielke LA, Tellier J, Chopin M, et al. Nfil3 is required for the development of all innate lymphoid cell subsets. *J Exp Med.* 2014;211:1733–40. <https://doi.org/10.1084/jem.20140145>.
42. Hoyler T, Klose CS, Souabni A, Turqueti-Neves A, Pfeifer D, Rawlins EL, et al. The transcription factor GATA-3 controls cell fate and maintenance of type 2 innate lymphoid cells. *Immunity.* 2012;37:634–48. <https://doi.org/10.1016/j.immuni.2012.06.020>.
43. Feuerer M, Herrero L, Cipolletta D, Naaz A, Wong J, Nayer A, et al. Lean, but not obese, fat is enriched for a unique population of regulatory T cells that affect metabolic parameters. *Nat Med.* 2009;15:930–9. <https://doi.org/10.1038/nm.2002>.
44. Cipolletta D, Feuerer M, Li A, Kamei N, Lee J, Shoelson SE, et al. PPAR- γ is a major driver of the accumulation and phenotype of adipose tissue Treg cells. *Nature.* 2012;486:549–53. <https://doi.org/10.1038/nature11132>.
45. Kolodin D, van Panhuys N, Li C, Magnuson AM, Cipolletta D, Miller CM, et al. Antigen- and cytokine-driven accumulation of regulatory T cells in visceral adipose tissue of lean mice. *Cell Metab.* 2015;21:543–57. <https://doi.org/10.1016/j.cmet.2015.03.005>.
46. Vasanthakumar A, Moro K, Xin A, Liao Y, Gloury R, Kawamoto S, et al. The transcriptional regulators IRF4, BATF and IL-33 orchestrate development and maintenance of adipose tissue-resident regulatory T cells. *Nat Immunol.* 2015;16:276–85. <https://doi.org/10.1038/ni.3085>.
47. Schmidleithner L, Thabet Y, Schönfeld E, Köhne M, Sommer D, Abdullah Z, et al. Enzymatic activity of HPGD in Treg cells suppresses tconv cells to maintain adipose tissue homeostasis and prevent metabolic dysfunction. *Immunity.* 2019;50:1232–48. <https://doi.org/10.1016/j.immuni.2019.03.014>. e1214.
48. Cipolletta D, Cohen P, Spiegelman BM, Benoist C, Mathis D. Appearance and disappearance of the mRNA signature characteristic of Treg cells in visceral adipose tissue: age, diet, and PPAR γ effects. *Proc Natl Acad Sci USA.* 2015;112:482–7. <https://doi.org/10.1073/pnas.1423486112>.
49. Li C, DiSpirito JR, Zemmour D, Spallanzani RG, Kuswanto W, Benoist C, et al. TCR transgenic mice reveal stepwise, multi-site acquisition of the distinctive fat-Treg phenotype. *Cell.* 2018;174:285–99. <https://doi.org/10.1016/j.cell.2018.05.004>. e212.
50. Delacher M, Imbusch CD, Hotz-Wagenblatt A, Mallm JP, Bauer K, Simon M, et al. Precursors for nonlymphoid-tissue treg cells reside in secondary lymphoid organs and are programmed by the transcription factor BATF. *Immunity.* 2020;52:295–312. <https://doi.org/10.1016/j.immuni.2019.12.002>. e211.

51. Wu D, Wong CK, Han JM, Orban PC, Huang Q, Gillies J, et al. T reg-specific insulin receptor deletion prevents diet-induced and age-associated metabolic syndrome. *J Exp Med*. 2020;217. <https://doi.org/10.1084/jem.20191542>.
52. Li Y, Lu Y, Lin SH, Li N, Han Y, Huang Q, et al. Insulin signaling establishes a developmental trajectory of adipose regulatory T cells. *Nat Immunol*. 2021;22:1175–85. <https://doi.org/10.1038/s41590-021-01010-3>.
53. Rana B, Jou E, Barlow JL, Rodriguez-Rodriguez N, Walker JA, Knox C, et al. A stromal cell niche sustains ILC2-mediated type-2 conditioning in adipose tissue. *J Exp Med*. 2019;216:1999–2009. <https://doi.org/10.1084/jem.20190689>.
54. Kohlgruber AC, Gal-Oz ST, LaMarche NM, Shimazaki M, Duquette D, Koay HF, et al. gammadelta T cells producing interleukin-17A regulate adipose regulatory T cell homeostasis and thermogenesis. *Nat Immunol*. 2018;19:464–74. <https://doi.org/10.1038/s41590-018-0094-2>.
55. Roh M, Wainwright DA, Wu JD, Wan Y, Zhang B. Targeting CD73 to augment cancer immunotherapy. *Curr Opin Pharm*. 2020;53:66–76. <https://doi.org/10.1016/j.coph.2020.07.001>.
56. Bagchi DP, MacDougall OA. Wnt signaling: from mesenchymal cell fate to lipogenesis and other mature adipocyte functions. *Diabetes*. 2021;70:1419–30. <https://doi.org/10.2337/dbi20-0015>.
57. Hosogai N, Fukuhara A, Oshima K, Miyata Y, Tanaka S, Segawa K, et al. Adipose tissue hypoxia in obesity and its impact on adipocytokine dysregulation. *Diabetes*. 2007;56:901–11. <https://doi.org/10.2337/db06-0911>.
58. Strissel KJ, Stancheva Z, Miyoshi H, Perfield JW, DeFuria J, Jick Z, et al. Adipocyte death, adipose tissue remodeling, and obesity complications. *Diabetes*. 2007;56:2910–8. <https://doi.org/10.2337/db07-0767>.
59. Hotamisligil GS. Endoplasmic reticulum stress and the inflammatory basis of metabolic disease. *Cell*. 2010;140:900–17. <https://doi.org/10.1016/j.cell.2010.02.034>.
60. Nishimura S, Manabe I, Nagasaki M, Eto K, Yamashita H, Ohsugi M, et al. CD8+ effector T cells contribute to macrophage recruitment and adipose tissue inflammation in obesity. *Nat Med*. 2009;15:914–20. <https://doi.org/10.1038/nm.1964>.
61. Winer S, Chan Y, Paltser G, Truong D, Tsui H, Bahrami J, et al. Normalization of obesity-associated insulin resistance through immunotherapy. *Nat Med*. 2009;15:921–9. <https://doi.org/10.1038/nm.2001>.
62. Winer DA, Winer S, Shen L, Wadia PP, Yantha J, Paltser G, et al. B cells promote insulin resistance through modulation of T cells and production of pathogenic IgG antibodies. *Nat Med*. 2011;17:610–7. <https://doi.org/10.1038/nm.2353>.
63. Wensveen FM, Jelenčić V, Valentić S, Šestan M, Wensveen TT, Theurich S, et al. NK cells link obesity-induced adipose stress to inflammation and insulin resistance. *Nat Immunol*. 2015;16:376–85. <https://doi.org/10.1038/ni.3120>.
64. Wang H, Shen L, Sun X, Liu F, Feng W, Jiang C, et al. Adipose group 1 innate lymphoid cells promote adipose tissue fibrosis and diabetes in obesity. *Nat Commun*. 2019;10:3254. <https://doi.org/10.1038/s41467-019-11270-1>.
65. Kadowaki T, Yamauchi T, Kubota N, Hara K, Ueki K, Tobe K. Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. *J Clin Invest*. 2006;116:1784–92. <https://doi.org/10.1172/JCI29126>.
66. Rosenbaum M, Leibel RL. 20 years of leptin: role of leptin in energy homeostasis in humans. *J Endocrinol*. 2014;223:F83–96. <https://doi.org/10.1530/JOE-14-0358>.
67. Lonnqvist F, Arner P, Nordfors L, Schalling M. Overexpression of the obese (ob) gene in adipose tissue of human obese subjects. *Nat Med*. 1995;1:950–3. <https://doi.org/10.1038/nm0995-950>.
68. de Candia P, Praticchizzo F, Garavelli S, Alviggi C, La Cava A, Matarese G, et al. The pleiotropic roles of leptin in metabolism, immunity, and cancer. *J Exp Med*. 2021;218. <https://doi.org/10.1084/jem.20191593>.
69. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest*. 2003;112:1796–808. <https://doi.org/10.1172/JCI19246>.
70. Zheng C, Yang Q, Cao J, Xie N, Liu K, Shou P, et al. Local proliferation initiates macrophage accumulation in adipose tissue during obesity. *Cell Death Dis*. 2016;7:e2167. <https://doi.org/10.1038/cddis.2016.54>.
71. Hotamisligil GS, Budavari A, Murray D, Spiegelman BM. Reduced tyrosine kinase activity of the insulin receptor in obesity-diabetes. Central role of tumor necrosis factor- α . *J Clin Invest*. 1994;94:1543–9. <https://doi.org/10.1172/JCI117495>.
72. Senn JJ, Klopper PJ, Nowak IA, Mooney RA. Interleukin-6 induces cellular insulin resistance in hepatocytes. *Diabetes*. 2002;51:3391–9. <https://doi.org/10.2337/diabetes.51.12.3391>.
73. Aouadi M, Vangala P, Yawo JC, Tencerova M, Nicoloso SM, Cohen JL, et al. Lipid storage by adipose tissue macrophages regulates systemic glucose tolerance. *Am J Physiol Endocrinol Metab*. 2014;307:E374–383. <https://doi.org/10.1152/ajpendo.00187.2014>.
74. Prieur X, Mok CY, Velagapudi VR, Núñez V, Fuentes L, Montaner D, et al. Differential lipid partitioning between adipocytes and tissue macrophages modulates macrophage lipotoxicity and M2/M1 polarization in obese mice. *Diabetes*. 2011;60:797–809. <https://doi.org/10.2337/db10-0705>.
75. Vandanmagsar B, Youm YH, Ravussin A, Galgani JE, Stadler K, Mynatt RL, et al. The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nat Med*. 2011;17:179–88. <https://doi.org/10.1038/nm.2279>.
76. Lee JY, Sohn KH, Rhee SH, Hwang D. Saturated fatty acids, but not unsaturated fatty acids, induce the expression of cyclooxygenase-2 mediated through Toll-like receptor 4. *J Biol Chem*. 2001;276:16683–9. <https://doi.org/10.1074/jbc.M011695200>.
77. Lee JY, Ye J, Gao Z, Youn HS, Lee WH, Zhao L, et al. Reciprocal modulation of Toll-like receptor-4 signaling pathways involving MyD88 and phosphatidylinositol 3-kinase/AKT by saturated and polyunsaturated fatty acids. *J Biol Chem*. 2003;278:37041–51. <https://doi.org/10.1074/jbc.M305213200>.
78. Davis JE, Gabler NK, Walker-Daniels J, Spurlock ME. Tlr-4 deficiency selectively protects against obesity induced by diets high in saturated fat. *Obes (Silver Spring)*. 2008;16:1248–55. <https://doi.org/10.1038/oby.2008.210>.
79. Lancaster GL, Langley KG, Berglund NA, Kammoun HL, Reibe S, Estevez E, et al. Evidence that TLR4 is not a receptor for saturated fatty acids but mediates lipid-induced inflammation by reprogramming macrophage metabolism. *Cell Metab*. 2018;27:1096–110. <https://doi.org/10.1016/j.cmet.2018.03.014>. e1095.
80. Han MS, Jung DY, Morel C, Lakhani SA, Kim JK, Flavell RA, et al. JNK expression by macrophages promotes obesity-induced insulin resistance and inflammation. *Science*. 2013;339:218–22. <https://doi.org/10.1126/science.1227568>.
81. Solinas G, Vilcu C, Neels JG, Bandyopadhyay GK, Luo JL, Naugler W, et al. JNK1 in hematopoietically derived cells contributes to diet-induced inflammation and insulin resistance without affecting obesity. *Cell Metab*. 2007;6:386–97. <https://doi.org/10.1016/j.cmet.2007.09.011>.
82. Hill DA, Lim HW, Kim YH, Ho WY, Foong YH, Nelson VL, et al. Distinct macrophage populations direct inflammatory versus physiological changes in adipose tissue. *Proc Natl Acad Sci USA*. 2018;115:E5096–E5105. <https://doi.org/10.1073/pnas.1802611115>.
83. Kosteli A, Sugaru E, Haemmerle G, Martin JF, Lei J, Zechner R, et al. Weight loss and lipolysis promote a dynamic immune response in murine adipose tissue. *J Clin Invest*. 2010;120:3466–79. <https://doi.org/10.1172/JCI42845>.
84. Kraakman MJ, Kammoun HL, Allen TL, Deswaerte V, Henstridge DC, Estevez E, et al. Blocking IL-6 trans-signaling prevents high-fat diet-induced adipose tissue macrophage recruitment but does not improve insulin resistance. *Cell Metab*. 2015;21:403–16. <https://doi.org/10.1016/j.cmet.2015.02.006>.
85. Lee BC, Kim MS, Pae M, Yamamoto Y, Eberlé D, Shimada T, et al. Adipose natural killer cells regulate adipose tissue macrophages to promote insulin resistance in obesity. *Cell Metab*. 2016;23:685–98. <https://doi.org/10.1016/j.cmet.2016.03.002>.
86. O'sullivan TE, Rapp M, Fan X, Weizman OE, Bhardwaj P, Adams NM, et al. Adipose-resident group 1 innate lymphoid cells promote obesity-associated insulin resistance. *Immunity*. 2016;45:428–41. <https://doi.org/10.1016/j.immuni.2016.06.016>.
87. Stolarczyk E, Vong CT, Perucha E, Jackson I, Cawthorne MA, Wargent ET, et al. Improved insulin sensitivity despite increased visceral adiposity in mice deficient for the immune cell transcription factor T-bet. *Cell Metab*. 2013;17:520–33. <https://doi.org/10.1016/j.cmet.2013.02.019>.
88. Kallies A, Good-Jacobson KL. Transcription factor T-bet orchestrates lineage development and function in the immune system. *Trends Immunol*. 2017;38:287–97. <https://doi.org/10.1016/j.it.2017.02.003>.
89. Li C, Wang G, Sivasami P, Ramirez RN, Zhang Y, Benoist C, et al. Interferon- α -producing plasmacytoid dendritic cells drive the loss of adipose tissue regulatory T cells during obesity. *Cell Metab*. 2021;33:1610–23. <https://doi.org/10.1016/j.cmet.2021.06.007>. e1615.
90. Fabbiano S, Suárez-Zamorano N, Rigo D, Veyrat-Durebex C, Stevanovic Dokic A, Colin DJ, et al. Caloric restriction leads to browning of white adipose tissue through type 2 immune signaling. *Cell Metab*. 2016;24:434–46. <https://doi.org/10.1016/j.cmet.2016.07.023>.
91. Weisberg SP, Hunter D, Huber R, Lemieux J, Slaymaker S, Vaddi K, et al. CCR2 modulates inflammatory and metabolic effects of high-fat feeding. *J Clin Invest*. 2006;116:115–24. <https://doi.org/10.1172/JCI24335>.
92. Cannon B, Nedergaard J. Nonshivering thermogenesis and its adequate measurement in metabolic studies. *J Exp Biol*. 2011;214:242–53. <https://doi.org/10.1242/jeb.050989>.
93. Seale P, Conroe HM, Estall J, Kajimura S, Frontini A, Ishibashi J, et al. Prdm16 determines the thermogenic program of subcutaneous white adipose tissue in mice. *J Clin Invest*. 2011;121:96–105. <https://doi.org/10.1172/JCI44271>.
94. Wu J, Boström P, Sparks LM, Ye L, Choi JH, Giang AH, et al. Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human. *Cell*. 2012;150:366–76. <https://doi.org/10.1016/j.cell.2012.05.016>.
95. Seale P, Bjork B, Yang W, Kajimura S, Chin S, Kuang S, et al. PRDM16 controls a brown fat/skeletal muscle switch. *Nature*. 2008;454:961–7. <https://doi.org/10.1038/nature07182>.
96. Sanchez-Gurmaches J, Hung CM, Sparks CA, Tang Y, Li H, Guertin DA. PTEN loss in the Myf5 lineage redistributes body fat and reveals subsets of white

- adipocytes that arise from Myf5 precursors. *Cell Metab.* 2012;16:348–62. <https://doi.org/10.1016/j.cmet.2012.08.003>.
97. Shao M, Wang QA, Song A, Vishwanath L, Busbuso NC, Scherer PE, et al. Cellular origins of beige fat cells revisited. *Diabetes.* 2019;68:1874–85. <https://doi.org/10.2337/db19-0308>.
 98. Himms-Hagen J, Melnyk A, Zingaretti MC, Ceresi E, Barbatelli G, Cinti S. Multilocular fat cells in WAT of CL-316243-treated rats derive directly from white adipocytes. *Am J Physiol Cell Physiol.* 2000;279:C670–681. <https://doi.org/10.1152/ajpcell.2000.279.3.C670>.
 99. Ukropec J, Anunciado RP, Ravussin Y, Hulver MW, Kozak LP. UCP1-independent thermogenesis in white adipose tissue of cold-acclimated Ucp1^{-/-} mice. *J Biol Chem.* 2006;281:31894–908. <https://doi.org/10.1074/jbc.M606114200>.
 100. Kazak L, Chouchani ET, Jedrychowski MP, Erickson BK, Shinoda K, Cohen P, et al. A creatine-driven substrate cycle enhances energy expenditure and thermogenesis in beige fat. *Cell.* 2015;163:643–55. <https://doi.org/10.1016/j.cell.2015.09.035>.
 101. Ikeda K, Kang Q, Yoneshiro T, Camporez JP, Maki H, Homma M, et al. UCP1-independent signaling involving SERCA2b-mediated calcium cycling regulates beige fat thermogenesis and systemic glucose homeostasis. *Nat Med.* 2017;23:1454–65. <https://doi.org/10.1038/nm.4429>.
 102. Guan HP, Li Y, Jensen MV, Newgard CB, Steppan CM, Lazar MA. A futile metabolic cycle activated in adipocytes by antidiabetic agents. *Nat Med.* 2002;8:1122–8. <https://doi.org/10.1038/nm780>.
 103. Kazak L, Chouchani ET, Lu GZ, Jedrychowski MP, Bare CJ, Mina AI, et al. Genetic depletion of adipocyte creatine metabolism inhibits diet-induced thermogenesis and drives obesity. *Cell Metab.* 2017;26:660–71. <https://doi.org/10.1016/j.cmet.2017.08.009>. e663.
 104. Qiu Y, Nguyen KD, Odegaard JI, Cui X, Tian X, Locksley RM, et al. Eosinophils and type 2 cytokine signaling in macrophages orchestrate development of functional beige fat. *Cell.* 2014;157:1292–308. <https://doi.org/10.1016/j.cell.2014.03.066>.
 105. Rao RR, Long JZ, White JP, Svensson KJ, Lou J, Lokurkar I, et al. Meteorin-like is a hormone that regulates immune-adipose interactions to increase beige fat thermogenesis. *Cell.* 2014;157:1279–91. <https://doi.org/10.1016/j.cell.2014.03.065>.
 106. Lee MW, Odegaard JI, Mukundan L, Qiu Y, Molofsky AB, Nussbaum JC, et al. Activated type 2 innate lymphoid cells regulate beige fat biogenesis. *Cell.* 2015;160:74–87. <https://doi.org/10.1016/j.cell.2014.12.011>.
 107. Brestoff JR, Artis D. Immune regulation of metabolic homeostasis in health and disease. *Cell.* 2015;161:146–60. <https://doi.org/10.1016/j.cell.2015.02.022>.
 108. Hu B, Jin C, Zeng X, Resch JM, Jedrychowski MP, Yang Z, et al. gammadelta T cells and adipocyte IL-17RC control fat innervation and thermogenesis. *Nature.* 2020;578:610–4. <https://doi.org/10.1038/s41586-020-2028-z>.
 109. Lynch L, Hogan AE, Duquette D, Lester C, Banks A, LeClair K, et al. iNKT Cells Induce FGF21 for Thermogenesis and Are Required for Maximal Weight Loss in GLP1 Therapy. *Cell Metab.* 2016;24:510–9. <https://doi.org/10.1016/j.cmet.2016.08.003>.
 110. Hui X, Gu P, Zhang J, Nie T, Pan Y, Wu D, et al. Adiponectin enhances cold-induced browning of subcutaneous adipose tissue via promoting M2 macrophage proliferation. *Cell Metab.* 2015;22:279–90. <https://doi.org/10.1016/j.cmet.2015.06.004>.
 111. Medrikova D, Sijmonsma TP, Sowodniok K, Richards DM, Delacher M, Sticht C, et al. Brown adipose tissue harbors a distinct sub-population of regulatory T cells. *PLoS One.* 2015;10:e0118534. <https://doi.org/10.1371/journal.pone.0118534>.
 112. Kälin S, Becker M, Ott VB, Serr I, Hosp F, Mollah M, et al. A Stat6/Pten axis links regulatory T cells with adipose tissue function. *Cell Metab.* 2017;26:475–92. <https://doi.org/10.1016/j.cmet.2017.08.008>. e477.
 113. Goto T, Nakukool S, Yoshitake R, Hanafusa Y, Tokiwa S, Li Y, et al. Proinflammatory cytokine interleukin-1beta suppresses cold-induced thermogenesis in adipocytes. *Cytokine.* 2016;77:107–14. <https://doi.org/10.1016/j.cyto.2015.11.001>.
 114. Valladares A, Roncero C, Benito M, Porras A. TNF-alpha inhibits UCP-1 expression in brown adipocytes via ERKs. Opposite effect of p38MAPK. *FEBS Lett.* 2001;493:6–11. [https://doi.org/10.1016/S0014-5793\(01\)02264-5](https://doi.org/10.1016/S0014-5793(01)02264-5).
 115. Kumari M, Wang X, Lantier L, Lyubetskaya A, Eguchi J, Kang S, et al. IRF3 promotes adipose inflammation and insulin resistance and represses browning. *J Clin Invest.* 2016;126:2839–54. <https://doi.org/10.1172/JCI86080>.
 116. Chiang SH, Bazuine M, Lumeng CN, Geletka LM, Mowers J, White NM, et al. The protein kinase IKKepsilon regulates energy balance in obese mice. *Cell.* 2009;138:961–75. <https://doi.org/10.1016/j.cell.2009.06.046>.
 117. Li G, Xie C, Lu S, Nichols RG, Tian Y, Li L, et al. Intermittent fasting promotes white adipose browning and decreases obesity by shaping the gut microbiota. *Cell Metab.* 2017;26:801. <https://doi.org/10.1016/j.cmet.2017.10.007>.
 118. Di Francesco A, Di Germanio C, Bernier M, de Cabo R. A time to fast. *Science.* 2018;362:770–5. <https://doi.org/10.1126/science.aau2095>.
 119. Camell CD, Günther P, Lee A, Goldberg EL, Spadaro O, Youm YH, et al. Aging induces an Nlrp3 inflammasome-dependent expansion of adipose B cells that impairs metabolic homeostasis. *Cell Metab.* 2019;30:1024–39. <https://doi.org/10.1016/j.cmet.2019.10.006>. e1026.
 120. Camell CD, Sander J, Spadaro O, Lee A, Nguyen KY, Wing A, et al. Inflammasome-driven catecholamine catabolism in macrophages blunts lipolysis during ageing. *Nature.* 2017;550:119–23. <https://doi.org/10.1038/nature24022>.
 121. Lumeng CN, Liu J, Geletka L, Delaney C, Delproposto J, Desai A, et al. Aging is associated with an increase in T cells and inflammatory macrophages in visceral adipose tissue. *J Immunol.* 2011;187:6208–16. <https://doi.org/10.4049/jimmunol.1102188>.
 122. Goldberg EL, Shchukina I, Youm YH, Ryu S, Tsusaka T, Young KC, et al. IL-33 causes thermogenic failure in aging by expanding dysfunctional adipose ILC2. *Cell Metab.* 2021. <https://doi.org/10.1016/j.cmet.2021.08.004>.
 123. Geer EB, Shen W. Gender differences in insulin resistance, body composition, and energy balance. *Gend Med.* 2009;6:60–75. <https://doi.org/10.1016/j.genm.2009.02.002>.
 124. Macotela Y, Boucher J, Tran TT, Kahn CR. Sex and depot differences in adipocyte insulin sensitivity and glucose metabolism. *Diabetes.* 2009;58:803–12. <https://doi.org/10.2337/db08-1054>.
 125. Olszak T, An D, Zeissig S, Vera MP, Richter J, Franke A, et al. Microbial exposure during early life has persistent effects on natural killer T cell function. *Science.* 2012;336:489–93. <https://doi.org/10.1126/science.1219328>.
 126. Alquier T, Poitout V. Considerations and guidelines for mouse metabolic phenotyping in diabetes research. *Diabetologia.* 2018;61:526–38. <https://doi.org/10.1007/s00125-017-4495-9>.
 127. Caspar-Bauguil S, Cousin B, Galinier A, Segafredo C, Nibbelink M, André M, et al. Adipose tissues as an ancestral immune organ: site-specific change in obesity. *FEBS Lett.* 2005;579:3487–92. <https://doi.org/10.1016/j.febslet.2005.05.031>.
 128. Kotas ME, Lee HY, Gillum MP, Annicelli C, Guigni BA, Shulman GI, et al. Impact of CD1d deficiency on metabolism. *PLoS One.* 2011;6:e25478. <https://doi.org/10.1371/journal.pone.0025478>.
 129. Satoh M, Andoh Y, Clingan CS, Ogura H, Fujii S, Eshima K, et al. Type II NKT cells stimulate diet-induced obesity by mediating adipose tissue inflammation, steatohepatitis and insulin resistance. *PLoS One.* 2012;7:e350568. <https://doi.org/10.1371/journal.pone.0030568>.
 130. Wu L, Parekh VV, Gabriel CL, Bracy DP, Marks-Shulman PA, Tamboli RA, et al. Activation of invariant natural killer T cells by lipid excess promotes tissue inflammation, insulin resistance, and hepatic steatosis in obese mice. *Proc Natl Acad Sci USA.* 2012;109:E1143–1152. <https://doi.org/10.1073/pnas.1200498109>.
 131. Ji Y, Sun S, Xia S, Yang L, Li X, Qi L. Short term high fat diet challenge promotes alternative macrophage polarization in adipose tissue via natural killer T cells and interleukin-4. *J Biol Chem.* 2012;287:24378–86. <https://doi.org/10.1074/jbc.M112.371807>.
 132. Ohmura K, Ishimori N, Ohmura Y, Tokuhara S, Nozawa A, Horii S, et al. Natural killer T cells are involved in adipose tissues inflammation and glucose intolerance in diet-induced obese mice. *Arterioscler Thromb Vasc Biol.* 2010;30:193–9. <https://doi.org/10.1161/ATVBAHA.109.198614>.
 133. Mantell BS, Stefanovic-Racic M, Yang X, Dedousis N, Sipula IJ, O'Doherty RM. Mice lacking NKT cells but with a complete complement of CD8⁺ T-cells are not protected against the metabolic abnormalities of diet-induced obesity. *PLoS One.* 2011;6:e19831. <https://doi.org/10.1371/journal.pone.0019831>.
 134. Lynch L. Adipose invariant natural killer T cells. *Immunology.* 2014;142:337–46. <https://doi.org/10.1111/imm.12269>.
 135. Huh JY, Park J, Kim JI, Park YJ, Lee YK, Kim JB. Deletion of CD1d in adipocytes aggravates adipose tissue inflammation and insulin resistance in obesity. *Diabetes.* 2017;66:835–47. <https://doi.org/10.2337/db16-1122>.
 136. Burzyn D, Benoist C, Mathis D. Regulatory T cells in nonlymphoid tissues. *Nat Immunol.* 2013;14:1007–13. <https://doi.org/10.1038/ni.2683>.
 137. Bapat SP, Myoung Suh J, Fang S, Liu S, Zhang Y, Cheng A, et al. Depletion of fat-resident Treg cells prevents age-associated insulin resistance. *Nature.* 2015;528:137–41. <https://doi.org/10.1038/nature16151>.
 138. Beppu LY, Mooli RGR, Qu X, Marrero GJ, Finley CA, Fooks AN, et al. Tregs facilitate obesity and insulin resistance via a Blimp-1/IL-10 axis. *JCI Insight.* 2021;6. <https://doi.org/10.1172/jci.insight.140644>.
 139. Rajbhandari P, Thomas BJ, Feng AC, Hong C, Wang J, Vergnes L, et al. IL-10 signaling remodels adipose chromatin architecture to limit thermogenesis and energy expenditure. *Cell.* 2018;172:218–33. <https://doi.org/10.1016/j.cell.2017.11.019>. e217.
 140. Cretney E, Xin A, Shi W, Minnich M, Masson F, Miasari M, et al. The transcription factors Blimp-1 and IRF4 jointly control the differentiation and function of effector regulatory T cells. *Nat Immunol.* 2011;12:304–11. <https://doi.org/10.1038/ni.2006>.
 141. Wu D, Han JM, Yu X, Lam AJ, Hoeppli RE, Pesenacker AM, et al. Characterization of regulatory T cells in obese omental adipose tissue in humans. *Eur J Immunol.* 2019;49:336–47. <https://doi.org/10.1002/eji.201847570>.
 142. Fang W, Deng Z, Benadjaoud F, Yang D, Yang C, Shi GP. Regulatory T cells promote adipocyte beiging in subcutaneous adipose tissue. *FASEB J.* 2020;34:9755–70. <https://doi.org/10.1096/fj.201902518R>.

143. O'Rourke RW, White AE, Metcalf MD, Winters BR, Diggs BS, Zhu X, et al. Systemic inflammation and insulin sensitivity in obese IFN-gamma knockout mice. *Metabolism*. 2012;61:1152–61. <https://doi.org/10.1016/j.metabol.2012.01.018>.
144. Tramunt B, Smati S, Grandgeorge N, Lenfant F, Arnal JF, Montagner A, et al. Sex differences in metabolic regulation and diabetes susceptibility. *Diabetologia*. 2020;63:453–61. <https://doi.org/10.1007/s00125-019-05040-3>.
145. Giles DA, Moreno-Fernandez ME, Stankiewicz TE, Graspeuntner S, Cappelletti M, Wu D, et al. Thermoneutral housing exacerbates nonalcoholic fatty liver disease in mice and allows for sex-independent disease modeling. *Nat Med*. 2017;23:829–38. <https://doi.org/10.1038/nm.4346>.
146. Ganeshan K, Nikkanen J, Man K, Leong YA, Sogawa Y, Maschek JA, et al. Energetic trade-offs and hypometabolic states promote disease tolerance. *Cell*. 2019;177:399–413. <https://doi.org/10.1016/j.cell.2019.01.050>. e312.
147. Man K, Bowman C, Braverman KN, Escalante V, Tian Y, Bisanz JE, et al. A thermogenic fat-epithelium cell axis regulates intestinal disease tolerance. *Proc Natl Acad Sci USA*. 2020;117:32029–37. <https://doi.org/10.1073/pnas.2012003117>.
148. Kokolus KM, Capitano ML, Lee CT, Eng JW, Waight JD, Hylander BL, et al. Baseline tumor growth and immune control in laboratory mice are significantly influenced by subthermoneutral housing temperature. *Proc Natl Acad Sci USA*. 2013;110:20176–81. <https://doi.org/10.1073/pnas.1304291110>.
149. Ganeshan K, Chawla A. Warming the mouse to model human diseases. *Nat Rev Endocrinol*. 2017;13:458–65. <https://doi.org/10.1038/nrendo.2017.48>.
150. Kleinert M, Clemmensen C, Hofmann SM, Moore MC, Renner S, Woods SC, et al. Animal models of obesity and diabetes mellitus. *Nat Rev Endocrinol*. 2018;14:140–62. <https://doi.org/10.1038/nrendo.2017.161>.
151. Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR, et al. Genetic studies of body mass index yield new insights for obesity biology. *Nature*. 2015;518:197–206. <https://doi.org/10.1038/nature14177>.
152. Cohen P, Kajimura S. The cellular and functional complexity of thermogenic fat. *Nat Rev Mol Cell Biol*. 2021;22:393–409. <https://doi.org/10.1038/s41580-021-00350-0>.

AUTHOR CONTRIBUTIONS

All three authors have contributed to conceptualizing and writing and are responsible for the content of this article.

COMPETING INTERESTS

The authors declare no competing interests.

ADDITIONAL INFORMATION

Correspondence and requests for materials should be addressed to Kevin Man, Axel Kallies or Ajithkumar Vasanthakumar.

Reprints and permission information is available at <http://www.nature.com/reprints>



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2021