



Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Fidelito, G;Watt, MJ;Taylor, RA

Title:

Personalized Medicine for Prostate Cancer: Is Targeting Metabolism a Reality?

Date:

2022-01-21

Citation:

Fidelito, G., Watt, M. J. & Taylor, R. A. (2022). Personalized Medicine for Prostate Cancer: Is Targeting Metabolism a Reality?. *Frontiers in Oncology*, 11, <https://doi.org/10.3389/fonc.2021.778761>.

Persistent Link:

<https://hdl.handle.net/11343/302579>

License:

[CC BY](#)



# Personalized Medicine for Prostate Cancer: Is Targeting Metabolism a Reality?

## OPEN ACCESS

### Edited by:

Jennifer H Gunter,  
Queensland University of Technology,  
Australia

### Reviewed by:

Cyril Corbet,  
Fonds National de la Recherche  
Scientifique (FNRS), Belgium  
Jianbo Li,  
Case Western Reserve University,  
United States  
Charles Liam Bidgood,  
Queensland University of Technology,  
Australia

### \*Correspondence:

Renea A. Taylor  
renea.taylor@monash.edu  
Matthew J. Watt  
matt.watt@unimelb.edu.au

### †ORCID:

Gio Fidelito  
orcid.org/0000-0001-7278-2644  
Matthew J. Watt  
orcid.org/0000-0003-4064-4188  
Renea A. Taylor  
orcid.org/0000-0003-2609-2380

‡These authors have contributed  
equally to this work

### Specialty section:

This article was submitted to  
Genitourinary Oncology,  
a section of the journal  
Frontiers in Oncology

**Received:** 17 September 2021

**Accepted:** 21 December 2021

**Published:** 21 January 2022

### Citation:

Fidelito G, Watt MJ and Taylor RA  
(2022) Personalized Medicine for  
Prostate Cancer: Is Targeting  
Metabolism a Reality?  
Front. Oncol. 11:778761.  
doi: 10.3389/fonc.2021.778761

Gio Fidelito<sup>1†</sup>, Matthew J. Watt<sup>1\*†‡</sup> and Renea A. Taylor<sup>2,3,4\*†‡</sup>

<sup>1</sup> Department of Anatomy & Physiology, The University of Melbourne, Melbourne, VIC, Australia, <sup>2</sup> Department of Physiology, Biomedicine Discovery Institute, Cancer Program, Monash University, Melbourne, VIC, Australia, <sup>3</sup> Prostate Cancer Research Program, Cancer Research Division, Peter MacCallum Cancer Centre, Melbourne, VIC, Australia, <sup>4</sup> Sir Peter MacCallum Department of Oncology, University of Melbourne, Melbourne, VIC, Australia

Prostate cancer invokes major shifts in gene transcription and metabolic signaling to mediate alterations in nutrient acquisition and metabolic substrate selection when compared to normal tissues. Exploiting such metabolic reprogramming is proposed to enable the development of targeted therapies for prostate cancer, yet there are several challenges to overcome before this becomes a reality. Herein, we outline the role of several nutrients known to contribute to prostate tumorigenesis, including fatty acids, glucose, lactate and glutamine, and discuss the major factors contributing to variability in prostate cancer metabolism, including cellular heterogeneity, genetic drivers and mutations, as well as complexity in the tumor microenvironment. The review draws from original studies employing immortalized prostate cancer cells, as well as more complex experimental models, including animals and humans, that more accurately reflect the complexity of the *in vivo* tumor microenvironment. In synthesizing this information, we consider the feasibility and potential limitations of implementing metabolic therapies for prostate cancer management.

**Keywords:** prostate neoplasia, lipid metabolism, obesity, metabolism, patient-derived xenograft, metabolic targeting, metabolic heterogeneity

## INTRODUCTION

Urological cancers accounted for 13.1% of 19.3 million new cancer incidence worldwide in 2020 (1). Prostate cancer is the most commonly diagnosed urologic cancer, followed by bladder, kidney, testis, and penile cancers (1) and frequently occurs in men over 65 years of age (2). More than 80% of men are diagnosed with localized disease, and the majority of these patients will have indolent tumors that are slow to progress, with low risk of experiencing prostate cancer-specific death (3). For these men, active surveillance, curative intent surgery or radiotherapy, are mostly effective with 10-year disease-specific survival rate of >90% (4). However, approximately one third of patients will experience disease progression and develop metastases, most commonly to bone, but also to other soft tissues such as liver and lung (5, 6). For these men, androgen deprivation therapy (ADT) is standard of care and while initially effective at reducing tumor burden, residual cancer cells adapt to low systemic androgen levels and therapy resistant metastatic castrate-resistant prostate cancer (mCRPC) develops, where

tumorigenesis is driven by adaptive androgen receptor (AR) changes and intra-tumoral steroid biosynthesis (7). There are limited therapeutic options in managing this advanced stage disease, necessitating the development of novel targeted therapies and/or neo-adjuvant therapies that either prevent progression or treat mCRPC (Figure 1).

The hallmarks of cancer, proposed by Hanahan and Weinberg (8), comprise a series of biological capabilities acquired during the multistep development of human tumors, of which 'deregulated cellular energetics' is one. Cancer invokes an increase in energy production to sustain proliferation, and metabolic 'rewiring' is often invoked to maintain this requirement. Alterations in metabolic reprogramming include adaptation in nutrient acquisition, preferential utilization of substrates, and transcriptional changes that alter intracellular metabolic signaling pathways. Exploiting such metabolic reprogramming is proposed to enable the development of targeted therapies in cancers (9), leading to an explosion of interest in the field of cancer metabolism.

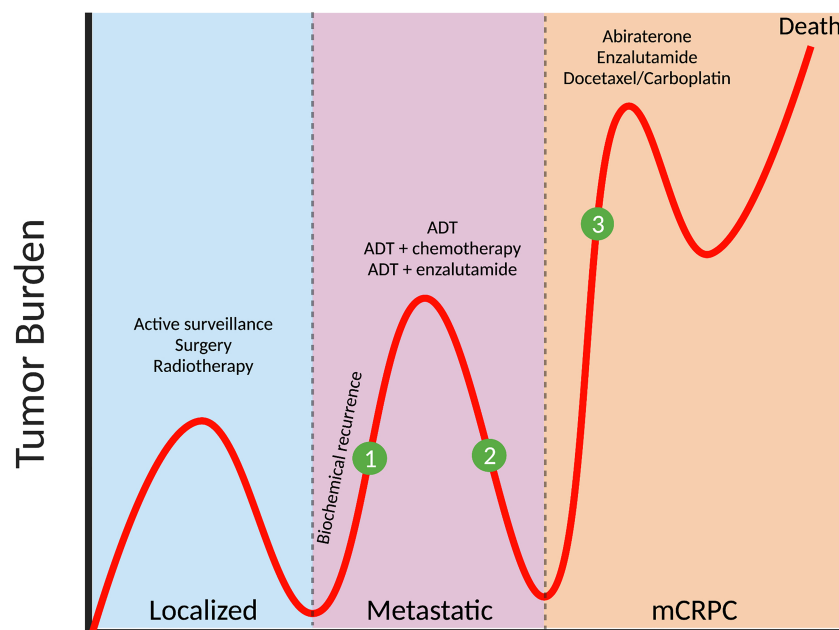
Metabolic inhibitors have been used for cancer therapies for many years, including the anti-metabolite class of chemotherapy (10, 11), and other agents have been developed for the treatment of advanced breast cancer, colorectal cancer, and hematological malignancies (12). This firmly establishes the principle that metabolic vulnerabilities can be effectively targeted for cancer treatment. However, to date, there are no metabolic inhibitors

approved for use in prostate cancer, which we posit is due to a knowledge gap in understanding the molecular and cellular reprogramming and associated changes in substrate utilization in human tumors, and the marked heterogeneity of this disease.

Herein, we will discuss how metabolism is reprogrammed in prostate cancer, in both localized and mCRPC, which likely have different metabolic needs. We will focus on literature employing studies in immortalized prostate cancer cells and expand to more complex environments, including animal models and human studies. We will then outline the factors contributing to variability in prostate cancer metabolism, including genetic drivers and alterations in the tumor microenvironment (TME), and lastly discuss the feasibility of metabolic targeting in patients and potential limitations in prostate cancer management.

## PROSTATE CANCER METABOLISM

The prostate gland secretes large amounts of citrate (~1000-fold than blood plasma) as the major constituent of prostatic fluid (13). The accumulation of zinc within the prostate gland by ZIP1 (SLC39A1) competitively inhibits mitochondrial aconitase (ACO2) activity, which hinders citrate oxidation and Tricarboxylic Acid (TCA) cycle flux (14–16). Hence, unlike other well-differentiated tissues, which rely on oxidative



**FIGURE 1** | Prostate cancer progression and potential stages for intervention with metabolic therapies. The majority of patients (>80%) are diagnosed with localized prostate cancer, with treatment including active surveillance (for low-risk tumors), or surgery/radiotherapy (for intermediate- and high-risk tumors). In one third of patients, biochemical recurrence (defined as a rise in prostate specific antigen, PSA, and indicative of active tumor growth) occurs and metastases develop at distant organs, and androgen-deprivation therapy (ADT) is administered. While initially effective, tumors eventually progress to metastatic castrate-resistant prostate cancer (mCRPC) and treatments include abiraterone, enzalutamide and chemotherapy such as docetaxel or carboplatin. Clinical intervention with relevant metabolic inhibitors, that are designed to slow tumor growth, could be applied (1) at the time of biochemical recurrence, thereby delaying the need for ADT, (2) in combination with ADT to target metabolic vulnerabilities induced by androgen withdrawal or (3) to treat mCRPC in combination with, or after existing therapies.



Enhanced aerobic glycolysis in response to androgen withdrawal is also observed in *in vivo* models. A metabolomic screen of an orthotopic xenograft model of TRAMP-C1 prostate cancer demonstrated increased glycolysis in tumors following androgen deprivation (31) while, *in vivo* and *ex vivo* metabolic imaging using hyperpolarized 1- $^{13}\text{C}$ pyruvate in TRAMP tumors also points towards elevated glycolysis and higher lactate dehydrogenase (LDH) activity in the castrate setting (30).

Evidence from human studies similarly show different glucose utilization across the spectrum of prostate cancer, which is best illustrated by  $^{18}\text{F}$ -fluorodeoxyglucose ( $^{18}\text{F}$ -FDG) cancer diagnostic imaging in patients.  $^{18}\text{F}$ -FDG is taken up by tissues and 'trapped', and its accumulation is reflective of the tissues glycolytic activity (32). Notably, the diagnostic utility of  $^{18}\text{F}$ -FDG imaging is limited to localized high-risk tumors and metastatic disease, indicating increased glucose uptake in rapidly growing malignant tissues and not indolent localized disease [as reviewed in (33)]. In addition, proteins that regulate glucose metabolism were increased in both localized and metastatic lesions of prostate cancer, including HIF-1 $\alpha$ , GLUT1, HK2, PFKFB3, PFKFB4, PKM2, PDK1 (29, 34–37). Functional analysis of glucose metabolism showed increased *de novo* lipogenesis in localized prostate cancer tissues compared to patient-matched benign tissues (38); however, this was not accompanied by increased glucose oxidation, indicating that much of the additional citrate produced in the TCA cycle is exported into the cytosol for lipogenesis (38).

Lactate is produced *via* the reduction of pyruvate and is classically viewed as the by-product of excess glycolysis; however, it is becoming increasingly recognized as an important mediator of tumorigenesis in some cancers. Lactate is used to fuel the TCA cycle in some malignancies (e.g., non-small cell lung cancer) (39) and inhibiting lactate influx into cells through the monocarboxylate transporter 1 (MCT1) reduces the metastatic potential of melanoma (40). Serum lactate dehydrogenase (LDH) is often increased in patients with high-grade prostate cancer and is associated with increased risk of mortality and disease progression in patients with metastatic prostate cancer (41, 42). Consistent with these observations, clinical studies utilizing hyperpolarized  $^{13}\text{C}$ -pyruvate imaging reported a positive correlation between prostate cancer Gleason grade and the conversion of pyruvate to lactate (43). Interestingly, monocarboxylate transporter 4 (MCT4), the protein responsible for lactate efflux from cells, is increased in localized and metastatic tumors (29) and RNAi-mediated silencing of MCT1/4 in prostate cancer cells decreased cell growth (44), suggesting lactate production and its intracellular utilization are important for tumorigenesis. A more comprehensive investigation of lactate metabolism in prostate cancer is clearly warranted. Finally, the pentose phosphate pathway (PPP) is a glucose catabolic pathway that appears to be important in prostate tumor growth in AR/SREBP/6PGD-dependent manner (45). However, whether PPP plays a significant role in prostate cancer by generating nucleotide precursor or sustaining the NADPH pool for lipogenesis and redox homeostasis is yet to be elucidated.

## Glutamine

Glutamine is a nonessential amino acid and the most abundant amino acid in the circulation ( $\sim 500\ \mu\text{M}$ ). Glutamine functions as a carbon donor for lipogenesis *via* reductive carboxylation, a nitrogen donor for non-essential amino acid production and nucleotide biosynthesis (46), and as a fuel source (47–51). Glutamine anaplerosis starts with glutamine conversion into glutamate by glutaminase (GLS) then further conversion into  $\alpha$ -ketoglutarate (AKG) to feed the TCA cycle by the actions of glutamate dehydrogenase (GLUD) and several transaminases, including glutamate–oxaloacetate transaminase (GOT), glutamate–pyruvate transaminase (GPT), and phosphoserine transaminase (PSAT) (46). While fourteen proteins are known to transport extracellular glutamine into cells, SLC1A5/ASCT2 is thought to be the major transporter, and its expression is upregulated in various cancers (52, 53). Glutamine can also donate its alpha nitrogen to serine, glycine, alanine, or aspartate following deamidation to glutamate (54). Serine feeds into one-carbon metabolism, which centrally integrates many pathways that are dysregulated within prostate cancer, strengthening the argument for targeting glutamine metabolism (55, 56). Additionally, enhanced aspartate metabolism has been implicated with epithelial to mesenchymal transition while increased levels of alanine has been identified within prostate cancer biopsies (57, 58).

Several lines of evidence demonstrate an important role for glutamine in prostate cancer growth and progression. ASCT2 is expressed in prostate cancer cells (e.g., LNCaP, VCaP, PC3, and DU145) (53, 59, 60) and approaches that reduce ASCT2 expression/function suppress glutamine uptake and hamper cell proliferation (53, 59). In a similar manner, GLS expression is higher in prostate cancer cells (e.g., LNCaP, 22Rv1, DU145, and PC-3) as compared with non-malignant prostate epithelial cells (e.g., RWPE-1) (60–62), and selective inhibition of GLS reduced proliferation and survival (60–63).

Key findings in cultured cells have been recapitulated in mouse models. ASCT2 mRNA expression is decreased upon castration and increased in CRPC (59) and knockdown of ASCT2 suppresses growth and metastatic burden in PC3 xenografts in mice (59), although rates of glutamine uptake and downstream metabolism were not assessed in this study. GLS expression is increased post-castration in LNCaP and LAPC4 xenografts (61), and pharmacological inhibition of GLS1 reduces the tumor burden in PC3, but not LNCaP xenografts (61), highlighting the dependency on glutamine metabolism in AR-negative, hormone-insensitive prostate cancer (61). Consistent with this notion, analysis of TRAMP tumors utilizing  $[\text{U}-^{13}\text{C}]$  glutamine metabolic tracing reported upregulation of glutaminolysis to replenish TCA cycle intermediates and upregulation of GLS1 activity in castrate-resistant compared to androgen-dependent tumors (30).

The importance of glutamine metabolism in human prostate cancer is unknown. ASCT2 and GLS1 mRNA expression is high in human prostate cancer (59, 63, 64) and ASCT2 expression is significantly associated with shorter time to biochemical recurrence in recurrent prostate cancer (64). Temporal ASCT2 expression is also observed in human tumors, with decreased expression upon ADT treatment (1–6 months and 7–12 months)

and increased expression in recurrent tumors (59). In addition, expression of the GLS1 enzyme undergoes a shift in isoform from kidney-type glutaminase (KGA) to the more active isoform, glutaminase C (GAC). This shift occurs progressively from localized to mCRPC and neuroendocrine prostate cancer (NEPC) (61). While these observations signal an important role for glutamine metabolism in advanced stages of prostate cancer (*i.e.*, mCRPC and NEPC), studies evaluating glutamine uptake, glutaminolysis and ATP production in human prostate cancer are clearly needed.

## Fatty Acids

Fatty acids are essential for the generation of structural cell membranes, energy production, and cellular signaling. Fatty acids are derived from adipose tissue lipolysis or from triglycerides stored in chylomicrons and very-low density lipoproteins, where they are transported from the circulation into cells. Several cell types, most notably hepatocytes and adipocytes, are capable of synthesizing fatty acids using other substrates, such as glucose and acetate, through a process called *de novo lipogenesis*. Fatty acids are the dominant metabolic substrate in most tissues where they undergo mitochondrial  $\beta$ -oxidation to generate acetyl-CoA, which feeds into the TCA cycle and oxidative metabolism.

Emerging evidence demonstrates an important role for fatty acid metabolism in prostate cancer. Fatty acid uptake is increased in immortalized prostate cancer cells (38, 65), which is often accompanied with increased energy production from fatty acid oxidation (65, 66). Treatment of prostate cancer cells with etomoxir, an inhibitor of fatty acid oxidation, reduces cell viability and proliferation, reinforcing the importance of this metabolic substrate for cancer progression (65, 67). Aside from the direct energy-generating mitochondrial fatty acid oxidation, peroxisomal fatty acid oxidation also supports prostate cancer growth (68, 69).

As mentioned above, prostate cancer is exceedingly lipogenic, highlighted by accelerated *de novo* synthesis of fatty acids driven by enhanced activity of sterol regulatory element-binding protein (SREBP) (70, 71), which induces the transcription of many genes involved in lipid metabolism, including ACLY, ACACA, FASN, SCD1 and LDLR (72). Studies employing pharmacological and genetic manipulation of key regulatory enzymes of lipid metabolism in immortalized cell lines and xenografts have demonstrated the importance of several lipid metabolism pathways in prostate cancer progression including increased *de novo* lipogenesis (*i.e.*, *via* ACLY, ACC and FASN inhibition) (73–76), triacylglycerol storage (DGAT1) (77), cholesterol metabolism (SOAT1, HMGCS1, HMGCR, and SCARB1) (78–80), lipolysis (MAGL) (81), and fatty acid elongation (ELOVL5 and ELOVL7) (82, 83). Similarly, 2,4-dienoyl-CoA reductase (DECR1) and enoyl-CoA delta isomerase 2 (ECI2), auxiliary enzymes responsible for the degradation of unsaturated fatty acids, are also essential for prostate cancer growth and therapy resistance (84–86). Finally, studies using tandem mass spectrometry lipidomics have reported marked alterations in the prostate lipidome with cancer (38, 82, 87, 88), indicating the likelihood that other nodes of lipid metabolism are regulated in prostate cancer development and metastasis.

While studies in cells and mice provide a reasonably compelling narrative that distinguishes lipid metabolism as a hallmark of prostate cancer, studies in primary human tissue are limited. Our team recently performed functional metabolic analysis in freshly procured human prostate tissue. Fatty acid uptake, fatty acid storage into complex lipids and cellular membranes, and *de novo* lipogenesis were upregulated in malignant compared to benign prostate tissues (38). Further studies identified fatty acid translocase (FAT/CD36) as a key fatty acid transport protein in prostate cancer while inhibition of FAT/CD36 with a monoclonal antibody attenuated tumor growth in a prostate patient-derived xenograft (PDX) and PDX-derived organoids. While this study identified a role for altered lipid metabolism in localized disease, further studies are required to ascertain whether these, and other changes in lipid metabolism, occur in metastatic disease. Additionally, whether there are further alterations in fatty acid utilization in the setting of mCRPC, where AR activity is amplified, is yet to be determined. In this context, a recent study employing transcriptomics and proteomics in prostate cancer cell lines and patient samples identified several lipid-mediated transporters and increased rates of fatty acid, cholesterol, and low-density lipoprotein uptake with androgen stimulation (89). Hence, any potential therapeutic benefit is likely to require cotargeting of lipid supply.

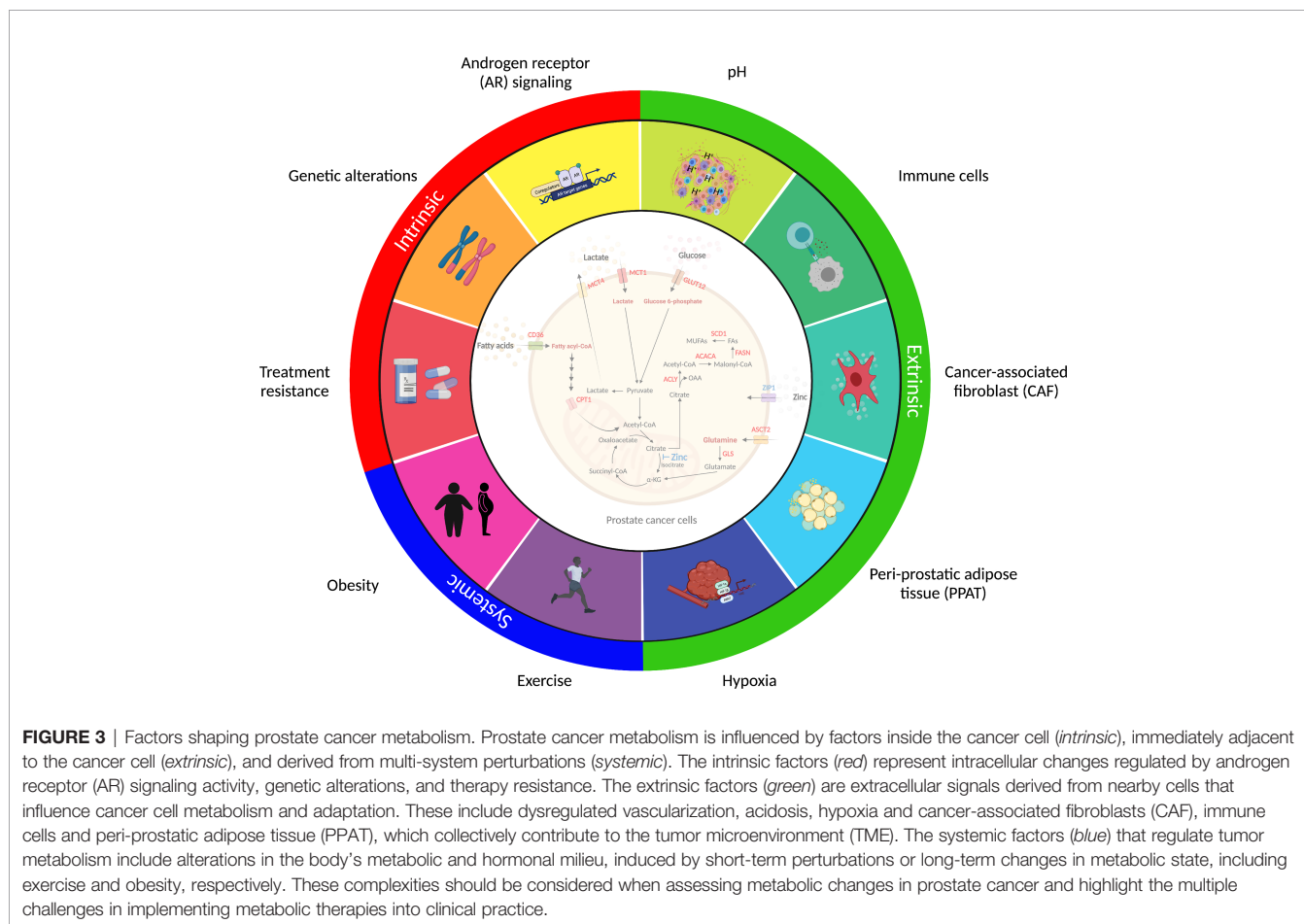
Efforts to elucidate the metabolic landscape of prostate cancer have highlighted the importance of glucose, glutamine, and fatty acid in prostate cancer growth and progression, and it is evident that there is a ‘*metabolic switch*’ from normal prostate epithelium to prostate cancer (90). However, the differences in metabolic regulation between localized and mCRPC tumors are less well defined. This highlights the need for comprehensive studies evaluating multiple substrates in a more complex system that reflect clinical tumors. The current advancement in patient-derived organoids (PDO) generation protocols (91, 92) and the creation of several PDX collections (93–96) will enable complex studies in identifying targetable metabolic vulnerabilities in different disease stages. However, a limitation of all *in vitro* studies is that metabolite concentrations in the TME are unknown. A widely held view is that commonly used cell culture medium (*e.g.*, RPMI, MEM, DMEM) contain significantly higher concentrations of glucose and amino acids than what is physiologically available, and often do not contain free fatty acids. Acknowledgement of this limitation and a better understanding of the constituents of the TME in different disease stages is required to move the field forward (see **Table 1**).

## FACTORS INFLUENCING PROSTATE CANCER METABOLISM

Prostate cancer displays marked heterogeneity from a molecular, morphological and clinical perspective and consideration of the factors that influence metabolic selection is essential to better understand the metabolic requirements of human prostate tumors in their native environment (**Figure 3**).

**TABLE 1 |** Methodological considerations.**Limitations of experimental models used to assess metabolism in prostate cancer**

- Most studies assessing metabolic regulation have been conducted in immortalized prostate cancer cells, including PC3, DU145 and LNCaP, which facilitate simple physiological and/or genetic manipulation and high throughput analysis, but bare limited resemblance to the complexity or heterogeneity of human tumors.
- Exposure of cells *in vitro* to supraphysiological nutrient levels in the culture medium unlikely recapitulate the condition in tumors, although it is noted that the concentration of metabolic substrates in the tumor microenvironment (TME) are currently unknown.
- *In vivo* studies using genetically engineered mouse models of prostate cancer overcome some of these issues, however the mutations do not replicate the genomic and phenotypic heterogeneity observed clinically.
- The use of human tissues or clinical studies are often impracticable due to limited access to patients under carefully controlled conditions and the technical difficulty in assessing tissue-specific metabolism *in vivo*.
- To overcome this limitation, prostate cancer patient-derived xenografts (PDXs) capture the heterogenous nature of tumor of origin. However, despite its perceived superiority over other approaches, PDXs lack stroma and immunological contribution.
- Combined approaches that integrate these complementary models are required to understand the metabolic landscape of prostate cancer and identify promising therapeutic strategies.



## Genetic Drivers of Metabolism

Specific oncogenic mutations can promote metabolic phenotypes in some cancers [as reviewed in (97)]. This is unequivocally the case in melanoma, where *BRAF*<sup>V600E</sup> mutations that account for ~80% of melanomas drive a metabolic program with a preference towards Warburg metabolism (98). Inhibition of oncogenic *BRAF* using drugs such as vemurafenib, dabrafenib or encorafenib cause

profound reductions in glucose uptake and improve patient outcomes (99). While the use of oncogene-driven mouse models has been helpful in linking specific genomic alterations with aberrant metabolic phenotypes in some tumor types (100–103), the profound number of molecular aberrations and heterogeneity observed in human tumors make it challenging to identify single DNA or gene alterations that dictate metabolic regulation.

Acquisition of genomic alterations underpins prostate tumorigenesis. Comprehensive genomic characterization of prostate cancer has identified recurrent alterations in genes involved in androgen signalling, DNA repair, and PI3K signalling, such as *TP53*, *SPOP*, *PTEN*, *AR*, *FOXA1*, *MYC*, *ATM* and *APC*. However, the incidence of significantly mutated genes follows a long-tail distribution, where the frequent alterations are only detected in ~5-10% of cases, and many other genes are mutated in <3% of cases (104). This underpins a complex genetic landscape in prostate cancer and heterogeneous nature of the disease. There is limited evidence showing induction of metabolic remodelling by individual oncogenes, such as *MYC* amplification, which promotes fatty acid synthesis and accelerates prostate cancer progression (105, 106). However, the absence of a dominant and frequent genetic mutation in prostate cancer indicates that a 'common' oncogenic-driven metabolic phenotype is unlikely to exist, although this remains to be fully explored.

## Neuroendocrine Prostate Cancer

The prominent pathology in prostate cancer is adenocarcinoma; however, in rare cases (<1%), NEPC tumors occur and present with an AR-null phenotype. While these are uncommon at diagnosis, there is increasing prevalence of therapy-induced NEPC that develops as an aggressive form of mCRPC. Treatment of NEPC presents an unmet clinical challenge in managing advanced prostate cancer. Emergence of an AR-null, NEPC phenotype is characterized by the expression of neuroendocrine markers such as synaptophysin, CD56, and chromogranin, with the absence of AR and AR-regulated gene expression (107). The genomic loss of tumor suppressors, dysregulation of specific transcription factors, and epigenetic modifications have been linked to the gain of neuroendocrine-like properties [as reviewed in (108)].

The metabolic regulation in NEPC disease states requires independent investigation. A previous study identified the requirement of increased serine biosynthesis following the loss of *PKC $\lambda$ /I* to fuel the methionine salvage pathway, which in turn augmented NEPC differentiation through DNA methylation (56). This highlights the role of metabolism in epithelial cell differentiation, beyond energy production. NEPC is characterized by increased glucose uptake and glucokinase expression compared to adenocarcinoma, despite the suppression of *GLUT12* (109). Transcriptomic analysis of NEPC PDX and patient specimens identified elevated glycolysis and lactate production as the metabolic feature of NEPC (110); however, these and other metabolic processes in NEPC are yet to be quantified using appropriate tracer methodologies.

## Prostate Metabolism and the Tumor Microenvironment

Prostate tumor cells reside in close proximity to neighboring cells within the tumor microenvironment (TME). The major components of the TME include cancer-associated fibroblasts (CAFs), endothelial cells, mesenchymal cells, as well as immune

cells such as mast cells, T cells, macrophages and monocytes. Each of these cell types secrete metabolites, hormones, extracellular vesicles and cytokines that could impact local metabolism. Several characteristic changes in the TME can impact metabolism, including dysregulated vascularization that involves disorganized and leaky blood vessels with low pericytes coverage, which in turn creates a hypoxic and acidic environment. Hypoxia has been implicated in the metabolic reprogramming of cancer cells, and upregulation of HIF-1 $\alpha$  plays an important role in the regulation of glycolysis (111). Additionally, CAFs themselves undergo a significant shift from oxidative phosphorylation to aerobic glycolysis, altering substrate availability for nearby cancer cells (112). There are also remarkable alterations in immune responses and the inflammatory environment in the TME, creating an immunosuppressive milieu. Interestingly, this relationship is likely to be bidirectional, with evidence that the low pH induced by excess lactate production in cancer cells reduces T cell infiltration. In addition, the microbiome is an important modulator of various host processes such as metabolism and immunity, and microbiome dysbiosis is associated with tumor development, disease progression, and treatment response and resistance in prostate cancer (113). The complexity of intra-tumoral paracrine signaling is exacerbated by remarkable heterogeneity in the cellular composition between individual human tumors, as demonstrated by single-cell transcriptomic analysis (114). These findings highlight the need to develop methods to sample and define the components of the prostate TME, with a view to understanding the factors controlling tumor metabolism and, perhaps, determining targetable metabolic vulnerabilities.

## Prostate Cancer Metabolism and Obesity

Obesity is a global epidemic affecting 281 million men (115) and more than 40% of men aged between 45-74 are obese (116), an age when the majority of prostate cancer diagnoses occur (117). While there is limited evidence that obesity is an initiator of prostate cancer (118, 119), epidemiologic evidence indicates that obese patients develop aggressive tumors with poor clinical outcomes (120, 121), although there is some conjecture with respect to mCRPC (122). Studies in rodents mostly confirm progression towards an aggressive phenotype in obesity [as reviewed in (123)] and many plausible mechanisms have been proposed to explain the link between obesity and aggressive prostate cancer (123), including increased free fatty acid supply, hyperinsulinemia, hypertriglyceridemia, altered endocrine signaling and low-grade inflammation (123). Notably, definitive evidence supporting these putative obesity-related drivers of prostate cancer progression is lacking. While not directly related to obesity, higher dietary saturated fat intake is associated with prostate cancer lethality (105) and raises the possibility that dietary interventions that reduce saturated fat intake and/or interventions to desaturate fatty acids might be efficacious in managing prostate cancer.

Periprostatic adipose tissue (PPAT) covers the prostate anteriorly and patients with more PPAT have worse cancer

prognosis (124), leading to the view that PPAT secreted factors stimulate tumorigenesis, particularly in obesity (123). Studies employing co-culture of prostate cancer cell lines and adipocytes (125), or the addition of PPAT secreted factors to prostate cancer cells (126, 127) support this possibility; however, co-grafting of patient-matched PPAT and localized prostate cancer PDX did not enhance prostate cancer tumorigenesis in mice (126). Nevertheless, changes in fatty acid delivery or adipose-secreted proteins (*i.e.*, adipokines) from PPAT are factors that may impact prostate cancer metabolism.

## Exercise and Prostate Cancer

Observational studies indicate that exercise and physical activity are associated with decreased risk of prostate cancer incidence, and lower overall prostate cancer mortality. Notably, vigorous activity is associated with a reduced risk of advanced, high Gleason grade group, or fatal prostate cancer in men over 65 years of age (128). While the mechanisms underlying potential anti-tumorigenic effects of exercise remain elusive, several have been proposed and include reduced circulating insulin, insulin-like growth factor 1 and proinflammatory cytokines, reduced tumor vascularization, AR adaptations, reduced cholesterol, production of unknown ‘exercise circulating factors’ contained in exosomes and reprogramming of metabolic and immunological dysregulation (129, 130). Overall, local, systemic and external influences play a significant role in metabolic regulation and prostate tumorigenesis, although there remains much to be learnt in this space.

## SYSTEMIC THERAPIES FOR PROSTATE CANCER

Hormone therapy is standard of care for patients with advanced prostate cancer, involving the use of gonadotropin-releasing hormone (GnRH) agonists or antagonists to suppress testicular testosterone synthesis. The use of androgen-targeted agents, such as enzalutamide (AR antagonist) and abiraterone (inhibitor of cytochrome P450 (CYP) C17 to block androgen synthesis), are used clinically to treat mCRPC (131). Meanwhile, Rucaparib (132) and Olaparib (133) (PARP inhibitor) have been recently approved for men with mCRPC harboring deleterious mutation of homologous recombination repair genes. While these discoveries improve current management of prostate cancer, the need for new therapeutics or adjuvant therapies continues as mCRPC remains lethal, and NEPC tumors are refractory to hormone therapy.

Metabolic changes, including insulin resistance, dyslipidemia, diabetes, and cardiovascular morbidity have been associated with ADT (134). Recent studies examining metabolomic profiles of men receiving ADT reported a reduction in acyl-carnitines and ketone bodies, indicating ADT-induced systemic changes in fatty acid metabolism (135, 136). Meanwhile, low-carbohydrate diets reversed this alteration in fatty acid metabolism while slightly increasing androgen suppression (137). This emphasizes the importance of diet in maximizing ADT therapeutic activity

while minimizing its effects on altering metabolism. In addition, patients are often prescribed with exercise, anti-hypertensive, anti-hyperlipidemic and anti-hyperglycemic medications to attenuate the effects of ADT, and it is possible that these interventions induce metabolic changes that improve cancer outcomes, although the evidence for this is limited (discussed below).

Aside from the impact of ADT on systemic metabolism, it has been postulated that ADT induces metabolic vulnerabilities in the tumor itself that can be therapeutically targeted using combination approaches. The use of metabolic inhibitor(s) as an adjuvant therapy have improved the efficacy of existing therapies and prevented the development of resistance in several tumors (138). In prostate cancer, metabolic adaptations occur in prostate cancer cells following ADT, as well as androgen-targeted therapies, including enzalutamide or abiraterone, suggesting the possibility of co-treatment strategies (139, 140) (**Figure 1**). This was exemplified in prostate cancer PDXs where a synergistic effect was demonstrated following treatment of ADT (through castration) plus metformin (141). Thus, the possibility of metabolic targeting in combination with ADT should be further explored.

## PUTATIVE METABOLIC TARGETING IN PROSTATE CANCER

Effective targeting of cancer metabolism relies on suppressing or modulating metabolic pathways identified as cancer ‘dependent’ and the use of metabolic agents is thereby limited by the defined therapeutic window of efficacy and toxicity in cancerous and non-cancerous cells. While there are no metabolic inhibitors approved for clinical use in patients with prostate cancer, several agents targeting *de novo* lipogenesis, fatty acid oxidation and glutamine oxidation are in pre-clinical or early phase clinical trials.

### De Novo Lipogenesis Inhibitors

The lipogenic phenotype of prostate cancer raises the possibility of targeting *de novo* lipogenesis. In this context, fatty acid synthase (FASN) is a rate-limiting enzyme in this process, and several FASN inhibitors, including TVB-3166 and TVB-2640, suppressed tumor growth by 15% in 22Rv1 xenografts (142), and notably, induced up to 97% tumor growth inhibition in combination with paclitaxel (142). Another FASN inhibitor, IPI-9119, showed anti-tumorigenic activity in human mCRPC organoids and 22Rv1 and LNCaP-95 xenograft models (143). Phase I studies of TVB-2640, the first FASN inhibitor to enter clinical trials for prostate cancer, indicated a favorable tolerability profile as either monotherapy or in combination with taxane in four heavily pre-treated prostate cancer patients (144). Clinical studies are warranted to evaluate the clinical utility of FASN inhibitors in mCRPC.

Moreover, several drugs that target other enzymes in the *de novo* lipogenesis pathway are in clinical trials for other diseases, such as the ACC inhibitors Firsocostat (Gilead) and PF-

05175157 (Pfizer) for non-alcoholic fatty liver disease (145), and derivatives of these compounds could conceivably be adopted for treatment of prostate cancer. Indeed, PF-05175157 showed promising results in reducing proliferation and inducing apoptosis in localized prostate cancer patient-derived explants (88).

### Fatty Acid Oxidation Inhibitors

Etomoxir is an irreversible inhibitor of carnitine palmitoyl transferase 1, which is the protein that transports fatty acids into the mitochondria for eventual oxidation. Treating mice with etomoxir reduced tumor growth in VCaP xenografts, without changing body weight or inducing systemic toxicity (67); however, etomoxir caused hepatotoxicity in patients with heart failure leading to the premature termination of a phase II clinical trial (146). While etomoxir is unlikely to progress to clinical trials for prostate cancer, two angina medications, ranolazine and perhexiline, may prove to be efficacious. Ranolazine is an FDA-approved partial inhibitor of fatty acid oxidation (147), while perhexiline is an TGA-approved competitive inhibitor of CPT1 (148). While neither drug reduces tumor growth alone, combining either compound with enzalutamide significantly decreased tumor growth *in vitro* and *in vivo* (149). Moreover, perhexiline alone showed no anti-tumorigenic activity in patient-derived explants, while cotreatment of perhexiline with the HSP90 inhibitor, AUY922, significantly reduced proliferation and increased apoptosis (150). These observations indicate that inhibitors of fatty acid oxidation may sensitize prostate cancer to other therapies, albeit through unknown mechanisms, and could be rapidly translated to the clinic.

### Glutaminolysis Inhibitors

CB-839, an oral glutaminase inhibitor, showed encouraging safety and tolerability results in a phase 1 study conducted in patients with advanced and/or treatment-refractory solid tumors, including breast cancer, lung cancer, renal cell carcinoma and mesothelioma (151). Preclinical studies in DU145 cells and xenografts indicated a synergistic effect of CB-839 in combination with talazoparib (PARP inhibitor) (152), leading to an upcoming phase II open label study of CB-839 and talazoparib in patients with mCRPC (NCT04824937).

### HMG-CoA Reductase Inhibitors

Statins are a class of drugs that inhibit the activity of HMG-CoA reductase and are widely used to treat patients with hypercholesterolemia. While observational studies demonstrate that statin use is associated with reduced cancer-specific mortality in patients with mCRPC receiving ADT (153), the results from one randomized trial indicates that short-term statin use does not impact tumor proliferation or serum prostate-specific antigen (PSA) compared to placebo (154). Similarly, statins alone did not reduce tumor burden in LNCaP xenograft and PDX trials; however, combination therapy with a re-purposed SREBP2 inhibitor, dipyrindamole, significantly reduced tumor growth (155). Future studies exploring the

safety and efficacy of this, and other combinations, in clinical studies are yet to be seen.

### Metformin

Metformin is the current first-line treatment of type 2 diabetes. While the exact mechanisms of action of metformin are still incompletely resolved, the anticancer potential of metformin is indicated through the capacity to activate AMPK and inhibit the cell cycle and epithelial-mesenchymal transition [*as reviewed in* (156)]. However, epidemiology studies showed no effects in reducing prostate cancer incidence and minimal improvement in overall survival (157). Multiple clinical trials are currently underway to assess the therapeutic utility of metformin as a monotherapy, or in combination with androgen targeted agents (enzalutamide and abiraterone) in managing CRPC.

## PROSPECTS AND CHALLENGES FOR IMPLEMENTING METABOLIC THERAPIES IN PROSTATE CANCER

Prostate cancer is slow growing by nature, providing sufficient time to implement therapies to delay progression or manage aggressive disease. For patients with intermediate risk disease, the median time to biochemical recurrence is ~4.25 years (158), necessitating the need for initiation of ADT, and in some patients, radiotherapy. Current clinical practice is to combine ADT with androgen-targeted therapy or chemotherapy, as this approach has been shown to increase overall survival (159). While effective in the short term, CRPC inevitably develops in ~5-8 years (160), which is then associated with a median survival ranging from 13-30 months (161-163). Overall, the time from diagnosis to end-stage disease for most patients is ~10-15 years, providing ample time for therapeutic intervention (**Figure 1**). This makes prostate cancer distinct to other more rapidly progressing cancers.

Of course, the overarching challenge in developing and utilizing 'metabolic therapies' for prostate cancer is to determine the appropriate strategy for the appropriate patient at the appropriate time, which as outlined above will vary between localized, metastatic and CRPC (see *Prostate Cancer Metabolism* section). We are, however, some way off implementing precise, actionable therapies as the focus of current research in cancer metabolism is predominantly pre-clinical and there is an urgent need for clinically based metabolic research. One emerging methodology, not yet applied to prostate cancer, is the use of intraoperative <sup>13</sup>C metabolic tracer infusions in human cancer patients, which overcomes limitations of *ex vivo* studies and by integrating systemic, TME and spatial parameters that shape metabolic phenotypes (164).

The clinical trajectory described above is generalized for patients with intermediate-risk prostate cancer, although in reality, each patient has individual prognostic features that dictate disease progression. Risk-stratification for prostate cancer is critical to guide appropriate treatment decision-

making. Towards this, it is worth considering whether there are subsets of patients who might benefit from metabolic therapies, either based on the reliance of an essential metabolic substrate, or specific tumor subtypes with common genomic aberrations or pathology. However, this has not been demonstrated, likely because of the remarkable heterogeneity of prostate cancer, diversity in metabolic substrate fluxes described in human tumors, and lack of appropriate biomarkers. In this context, mass spectrometry metabolic imaging is being refined to detect 'metabolic signatures' of prostate cancer, with evidence indicating that such imaging may aid in understanding biological processes and to help cancer diagnosis, prognosis and monitor response to therapies (165, 166).

A major challenge for the field is to define when metabolic therapies could be clinically applied. One option is during early-stage disease, following curative intent surgery or radiation when PSA levels are beginning to slowly rise, indicative of residual disease that is progressing. It is envisaged that metabolic therapies designed to reduce nutrient supply and/or ATP production could slow growth and delay the need for ADT. Alternatively, there is interest in the potential for metabolic therapies to be used to treat CRPC, because significant energy is required for the growth of highly aggressive therapy resistant tumors (Figure 1). More generally, it has been suggested that a better understanding of the association between metabolism and prostate cancer may lead to cancer prevention, although such strategies are opaque.

Overall, there is very little evidence from preclinical models or clinical studies that targeting a single metabolic pathway will be sufficient to slow prostate tumor progression. Firstly, this requires modulation of a single substrate, enzyme or metabolic pathway to limit tumor growth or increase tumor susceptibility to an adjunct therapy. In this context, metabolic inhibition, commonly leads to compensatory upregulation of other fuel utilization pathways to maintain pro-tumorigenic energy demands. For example, our work showed that this was the case with fatty acid transport inhibition, whereby blocking FAT/CD36 induced an increase in *de novo* lipogenesis in localized disease (38). Similarly, others showed that inhibition of FASN led to the upregulation of genes involved in steroid biosynthesis and increased intracellular cholesterol (143, 167). Thus, we posit that targeting dual processes will most likely be required for effective metabolic intervention in prostate cancer. Further to this, most tissues in the body readily utilize each of the substrates commonly used in prostate cancer, with evidence of dependencies in some tissues (e.g. glucose for red blood cells and brain). Hence, approaches that direct metabolic therapies to the tumor will be essential to minimize the likelihood of off-target effects. Such approaches are feasible as evidenced by the implementation of radioligand-therapy targeted to prostate-specific antigens in the clinic.

## CONCLUSIONS

Prostate cancer invokes major shifts in gene transcription and metabolic signaling to mediate alterations in nutrient

acquisition and metabolic substrate selection when compared to normal tissues. Exploiting such metabolic reprogramming is proposed to enable the development of targeted therapies for prostate cancer, yet there are several challenges to be overcome before this becomes a reality. Firstly, several metabolic substrates have been identified in prostate cancer, including (but not limited to) fatty acids, glucose, lactate and glutamine, all of which are 'required' substrates in prostate cancer. Thus, identifying the most appropriate substrate to be targeted, and in which type of prostate cancer, remains unclear. Somewhat related, there is a gap in our knowledge of metabolism in human tumors. The majority of studies that have defined metabolic regulation of prostate cancer have been limited to cell culture or genetically modified mouse models, which does not accurately reflect the complexity of the *in vivo* tumor microenvironment and the impact that this induces on prostate metabolism (Table 1). Thirdly, prostate cancer is notoriously heterogeneous and there is currently insufficient evidence to indicate that subgroups of patients or tumor subtypes, based on genomic aberrations or pathology, share common metabolic vulnerabilities. Hence, there is an urgent need for these gaps to be addressed before metabolic therapies can be designed and incorporated into clinical practice.

## AUTHOR CONTRIBUTIONS

All authors (GF, MW, and RT) conceived the idea for the review, searched the literature, drafted the manuscript and provided critical revision of the manuscript for intellectual content. GF generated figures. MW and RT obtained funding and provided supervision. All authors contributed to the article and approved the submitted version.

## FUNDING

This work was supported by the Prostate Cancer Foundation of Australia (ID: PCFA-NCG 3313, awarded to MW, RT), the Diabetes Australia Research Trust (awarded to MW) and the Cancer Council of Victoria (APP1160217, awarded to MW, RT). MW was supported by the National Health and Medical Research Council NHMRC of Australia (ID: APP1077703), and RT by the Victorian Cancer Agency (MCRF15023). GF was supported by the Melbourne Research Scholarship (University of Melbourne).

## ACKNOWLEDGMENTS

We thank Dr. David Pook and Dr. Weranja Ranasinghe for helpful discussions. Figures were created with BioRender.com.

## REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin* (2021) 71:209–49. doi: 10.3322/caac.21660
- Rawla P. Epidemiology of Prostate Cancer. *World J Oncol* (2019) 10(2):63–89. doi: 10.14740/wjon1191
- Litwin MS, Saigal CS, editors. Prostate Cancer. In: *Urologic Diseases in America*, vol. 78. Washington, DC: US Department of Health and Human Services, Public Health Service, National Institutes of Health, National Institute of Diabetes and Digestive and Kidney Diseases p. 73–96.
- Hamdy FC, Donovan JL, Lane JA, Mason M, Metcalfe C, Holding P, et al. 10-Year Outcomes After Monitoring, Surgery, or Radiotherapy for Localized Prostate Cancer. *N Engl J Med* (2016) 375(15):1415–24. doi: 10.1056/NEJMoa1606220
- Barbosa FG, Queiroz MA, Nunes RF, Viana PCC, Marin JFG, Cerri GG, et al. Revisiting Prostate Cancer Recurrence With PSMA PET: Atlas of Typical and Atypical Patterns of Spread. *Radiographics* (2019) 39(1):186–212. doi: 10.1148/rg.2019180079
- Gandaglia G, Abdollah F, Schiffmann J, Trudeau V, Shariat SF, Kim SP, et al. Distribution of Metastatic Sites in Patients With Prostate Cancer: A Population-Based Analysis. *Prostate* (2014) 74(2):210–6. doi: 10.1002/pros.22742
- Jernberg E, Bergh A, Wikstrom P. Clinical Relevance of Androgen Receptor Alterations in Prostate Cancer. *Endocr Connect* (2017) 6(8):R146–R61. doi: 10.1530/EC-17-0118
- Hanahan D, Weinberg RA. Hallmarks of Cancer: The Next Generation. *Cell* (2011) 144(5):646–74. doi: 10.1016/j.cell.2011.02.013
- Faubert B, Solmonson A, DeBerardinis RJ. Metabolic Reprogramming and Cancer Progression. *Science* (2020) 368(6487):eaaw5473. doi: 10.1126/science.aaw5473
- Kozminski P, Halik PK, Chesori R, Gniazdowska E. Overview of Dual-Acting Drug Methotrexate in Different Neurological Diseases, Autoimmune Pathologies and Cancers. *Int J Mol Sci* (2020) 21(10):3483. doi: 10.3390/ijms21103483
- Longley DB, Harkin DP, Johnston PG. 5-Fluorouracil: Mechanisms of Action and Clinical Strategies. *Nat Rev Cancer* (2003) 3(5):330–8. doi: 10.1038/nrc1074
- Luengo A, Gui DY, Vander Heiden MG. Targeting Metabolism for Cancer Therapy. *Cell Chem Biol* (2017) 24(9):1161–80. doi: 10.1016/j.chembiol.2017.08.028
- Costello LC, Franklin RB. A Comprehensive Review of the Role of Zinc in Normal Prostate Function and Metabolism; and its Implications in Prostate Cancer. *Arch Biochem Biophys* (2016) 611:100–12. doi: 10.1016/j.abb.2016.04.014
- Liu Y, Franklin RB, Costello LC. Prolactin and Testosterone Regulation of Mitochondrial Zinc in Prostate Epithelial Cells. *Prostate* (1997) 30(1):26–32. doi: 10.1002/(sici)1097-0045(19970101)30:1<26::Aid-pros4>3.0.Co;2-j
- Franklin RB, Ma J, Zou J, Guan Z, Kukoyi BI, Feng P, et al. Human ZIP1 is a Major Zinc Uptake Transporter for the Accumulation of Zinc in Prostate Cells. *J Inorganic Biochem* (2003) 96(2-3):435–42. doi: 10.1016/s0162-0134(03)00249-6
- Costello LC, Liu Y, Franklin RB, Kennedy MC. Zinc Inhibition of Mitochondrial Aconitase and its Importance in Citrate Metabolism of Prostate Epithelial Cells. *J Biol Chem* (1997) 272(46):28875–81. doi: 10.1074/jbc.272.46.28875
- Costello LC, Franklin RB. Prostate Epithelial Cells Utilize Glucose and Aspartate as the Carbon Sources for Net Citrate Production. *Prostate* (1989) 15(4):335–42. doi: 10.1002/pros.2990150406
- Franklin RB, Feng P, Milon B, Desouki MM, Singh KK, Kajdacsy-Balla A, et al. Hzip1 Zinc Uptake Transporter Down Regulation and Zinc Depletion in Prostate Cancer. *Mol Cancer* (2005) 4:32. doi: 10.1186/1476-4598-4-32
- Gregorio EP, Alexandrino AP, Schuquel ITA, da Costa WF, Rodrigues MAF. Seminal Citrate is Superior to PSA for Detecting Clinically Significant Prostate Cancer. *Int Braz J Urol* (2019) 45(6):1113–21. doi: 10.1590/S1677-5538.IBJU.2018.0730
- Singh KK, Desouki MM, Franklin RB, Costello LC. Mitochondrial Aconitase and Citrate Metabolism in Malignant and Nonmalignant Human Prostate Tissues. *Mol Cancer* (2006) 5:14. doi: 10.1186/1476-4598-5-14
- Kline EE, Treat EG, Averna TA, Davis MS, Smith AY, Sillerud LO. Citrate Concentrations in Human Seminal Fluid and Expressed Prostatic Fluid Determined via 1H Nuclear Magnetic Resonance Spectroscopy Outperform Prostate Specific Antigen in Prostate Cancer Detection. *J Urol* (2006) 176(5):2274–9. doi: 10.1016/j.juro.2006.07.054
- Latonen L, Afyounian E, Jylha A, Nattinen J, Aapola U, Annala M, et al. Integrative Proteomics in Prostate Cancer Uncovers Robustness Against Genomic and Transcriptomic Aberrations During Disease Progression. *Nat Commun* (2018) 9(1):1176. doi: 10.1038/s41467-018-03573-6
- Lunt SY, Vander Heiden MG. Aerobic Glycolysis: Meeting the Metabolic Requirements of Cell Proliferation. *Annu Rev Cell Dev Biol* (2011) 27:441–64. doi: 10.1146/annurev-cellbio-092910-154237
- Vaupel P, Multhoff G. Revisiting the Warburg Effect: Historical Dogma Versus Current Understanding. *J Physiol* (2021) 599(6):1745–57. doi: 10.1113/JP278810
- Massie CE, Lynch A, Ramos-Montoya A, Boren J, Stark R, Fazli L, et al. The Androgen Receptor Fuels Prostate Cancer by Regulating Central Metabolism and Biosynthesis. *EMBO J* (2011) 30(13):2719–33. doi: 10.1038/emboj.2011.158
- Moon JS, Jin WJ, Kwak JH, Kim HJ, Yun MJ, Kim JW, et al. Androgen Stimulates Glycolysis for De Novo Lipid Synthesis by Increasing the Activities of Hexokinase 2 and 6-Phosphofructo-2-Kinase/Fructose-2,6-Bisphosphatase 2 in Prostate Cancer Cells. *Biochem J* (2011) 433(1):225–33. doi: 10.1042/BJ20101104
- White MA, Tsouko E, Lin C, Rajapakse K, Spencer JM, Wilkenfeld SR, et al. GLUT12 Promotes Prostate Cancer Cell Growth and is Regulated by Androgens and CaMKK2 Signaling. *Endocr Relat Cancer* (2018) 25(4):453–69. doi: 10.1530/ERC-17-0051
- Vaz CV, Alves MG, Marques R, Moreira PI, Oliveira PF, Maia CJ, et al. Androgen-Responsive and Nonresponsive Prostate Cancer Cells Present a Distinct Glycolytic Metabolism Profile. *Int J Biochem Cell Biol* (2012) 44(11):2077–84. doi: 10.1016/j.biocel.2012.08.013
- Pertega-Gomes N, Felisbino S, Massie CE, Vizcaino JR, Coelho R, Sandi C, et al. A Glycolytic Phenotype is Associated With Prostate Cancer Progression and Aggressiveness: A Role for Monocarboxylate Transporters as Metabolic Targets for Therapy. *J Pathol* (2015) 236(4):517–30. doi: 10.1002/path.4547
- Sun J, Bok RA, DeLos Santos J, Upadhyay D, DeLos Santos R, Agarwal S, et al. Resistance to Androgen Deprivation Leads to Altered Metabolism in Human and Murine Prostate Cancer Cell and Tumor Models. *Metabolites* (2021) 11(3):139. doi: 10.3390/metabol11030139
- Martin PL, Yin JJ, Seng V, Casey O, Corey E, Morrissey C, et al. Androgen Deprivation Leads to Increased Carbohydrate Metabolism and Hexokinase 2-Mediated Survival in Pten/Tp53-Deficient Prostate Cancer. *Oncogene* (2017) 36(4):525–33. doi: 10.1038/ncr.2016.223
- Smith TA. FDG Uptake, Tumour Characteristics and Response to Therapy: A Review. *Nucl Med Commun* (1998) 19(2):97–105. doi: 10.1097/00006231-199802000-00002
- Jadvar H. Molecular Imaging of Prostate Cancer With 18F-Fluorodeoxyglucose PET. *Nat Rev Urol* (2009) 6(6):317–23. doi: 10.1038/nrurol.2009.81
- Jans J, van Dijk JH, van Schelven S, van der Groep P, Willems SH, Jonges TN, et al. Expression and Localization of Hypoxia Proteins in Prostate Cancer: Prognostic Implications After Radical Prostatectomy. *Urology* (2010) 75(4):786–92. doi: 10.1016/j.urology.2009.08.024
- Wang J, Li J, Li X, Peng S, Li J, Yan W, et al. Increased Expression of Glycolytic Enzymes in Prostate Cancer Tissues and Association With Gleason Scores. *Int J Clin Exp Pathol* (2017) 10(11):11080–9.
- Zhong H, Semenza GL, Simons JW, De Marzo AM. Up-Regulation of Hypoxia-Inducible Factor 1alpha is an Early Event in Prostate Carcinogenesis. *Cancer Detect Prev* (2004) 28(2):88–93. doi: 10.1016/j.cdp.2003.12.009
- Wang L, Xiong H, Wu F, Zhang Y, Wang J, Zhao L, et al. Hexokinase 2-Mediated Warburg Effect is Required for PTEN- and P53-Deficiency-Driven Prostate Cancer Growth. *Cell Rep* (2014) 8(5):1461–74. doi: 10.1016/j.celrep.2014.07.053
- Watt MJ, Clark AK, Selth LA, Haynes VR, Lister N, Rebello R, et al. Suppressing Fatty Acid Uptake has Therapeutic Effects in Preclinical Models

- of Prostate Cancer. *Sci Transl Med* (2019) 11(478):eaa5758. doi: 10.1126/scitranslmed.aau5758
39. Faubert B, Li KY, Cai L, Hensley CT, Kim J, Zacharias LG, et al. Lactate Metabolism in Human Lung Tumors. *Cell* (2017) 171(2):358–71 e9. doi: 10.1016/j.cell.2017.09.019
  40. Tasdogan A, Faubert B, Ramesh V, Ubellacker JM, Shen B, Solmonson A, et al. Metabolic Heterogeneity Confers Differences in Melanoma Metastatic Potential. *Nature* (2020) 577(7788):115–20. doi: 10.1038/s41586-019-1847-2
  41. Mori K, Kimura S, Parizi MK, Enikeev DV, Glybochko PV, Seebacher V, et al. Prognostic Value of Lactate Dehydrogenase in Metastatic Prostate Cancer: A Systematic Review and Meta-Analysis. *Clin Genitourin Cancer* (2019) 17(6):409–18. doi: 10.1016/j.clgc.2019.07.009
  42. Li F, Xiang H, Pang Z, Chen Z, Dai J, Chen S, et al. Association Between Lactate Dehydrogenase Levels and Oncologic Outcomes in Metastatic Prostate Cancer: A Meta-Analysis. *Cancer Med* (2020) 9(19):7341–51. doi: 10.1002/cam4.3108
  43. Granlund KL, Tee SS, Vargas HA, Lyashchenko SK, Reznik E, Fine S, et al. Hyperpolarized MRI of Human Prostate Cancer Reveals Increased Lactate With Tumor Grade Driven by Monocarboxylate Transporter 1. *Cell Metab* (2020) 31(1):105–14 e3. doi: 10.1016/j.cmet.2019.08.024
  44. Pereira-Nunes A, Simoes-Sousa S, Pinheiro C, Miranda-Goncalves V, Granja S, Baltazar F. Targeting Lactate Production and Efflux in Prostate Cancer. *Biochim Biophys Acta Mol Basis Dis* (2020) 1866(11):165894. doi: 10.1016/j.bbadis.2020.165894
  45. Gillis JL, Hinneh JA, Ryan NK, Irani S, Moldovan M, Quek LE, et al. A Feedback Loop Between the Androgen Receptor and 6-Phosphogluconate Dehydrogenase (6PGD) Drives Prostate Cancer Growth. *Elife* (2021) 10:e62592. doi: 10.7554/eLife.62592
  46. Yang L, Venneti S, Nagrath D. Glutaminolysis: A Hallmark of Cancer Metabolism. *Annu Rev BioMed Eng* (2017) 19:163–94. doi: 10.1146/annurev-bioeng-071516-044546
  47. Fan J, Kamphorst JJ, Mathew R, Chung MK, White E, Shlomi T, et al. Glutamine-Driven Oxidative Phosphorylation is a Major ATP Source in Transformed Mammalian Cells in Both Normoxia and Hypoxia. *Mol Syst Biol* (2013) 9:712. doi: 10.1038/msb.2013.65
  48. Zhang W, Li H, Ogando DG, Li S, Feng M, Price FW Jr, et al. Glutaminolysis is Essential for Energy Production and Ion Transport in Human Corneal Endothelium. *EBioMedicine* (2017) 16:292–301. doi: 10.1016/j.ebiom.2017.01.004
  49. Agostini M, Romeo F, Inoue S, Niklison-Chirou MV, Elia AJ, Dinsdale D, et al. Metabolic Reprogramming During Neuronal Differentiation. *Cell Death Differ* (2016) 23(9):1502–14. doi: 10.1038/cdd.2016.36
  50. DeBerardinis RJ, Mancuso A, Daikhin E, Nissim I, Yudkoff M, Wehrli S, et al. Beyond Aerobic Glycolysis: Transformed Cells can Engage in Glutamine Metabolism That Exceeds the Requirement for Protein and Nucleotide Synthesis. *Proc Natl Acad Sci USA* (2007) 104(49):19345–50. doi: 10.1073/pnas.0709747104
  51. Oburoglu L, Tardito S, Fritz V, de Barros SC, Merida P, Craveiro M, et al. Glucose and Glutamine Metabolism Regulate Human Hematopoietic Stem Cell Lineage Specification. *Cell Stem Cell* (2014) 15(2):169–84. doi: 10.1016/j.stem.2014.06.002
  52. Scopelliti AJ, Font J, Vandenberg RJ, Boudker O, Ryan RM. Structural Characterisation Reveals Insights Into Substrate Recognition by the Glutamine Transporter ASCT2/SLC1A5. *Nat Commun* (2018) 9(1):38. doi: 10.1038/s41467-017-02444-w
  53. Bhutia YD, Ganapathy V. Glutamine Transporters in Mammalian Cells and Their Functions in Physiology and Cancer. *Biochim Biophys Acta* (2016) 1863(10):2531–9. doi: 10.1016/j.bbamcr.2015.12.017
  54. Jiang J, Srivastava S, Zhang J. Starve Cancer Cells of Glutamine: Break the Spell or Make a Hungry Monster? *Cancers (Basel)* (2019) 11(6):804. doi: 10.3390/cancers11060804
  55. Corbin JM, Ruiz-Echevarria MJ. One-Carbon Metabolism in Prostate Cancer: The Role of Androgen Signaling. *Int J Mol Sci* (2016) 17(8):1208. doi: 10.3390/ijms17081208
  56. Reina-Campos M, Linares JF, Duran A, Cordes T, L'Hermitte A, Badur MG, et al. Increased Serine and One-Carbon Pathway Metabolism by PKC*lambda*/iota Deficiency Promotes Neuroendocrine Prostate Cancer. *Cancer Cell* (2019) 35(3):385–400 e9. doi: 10.1016/j.ccell.2019.01.018
  57. Chen Y, Wang K, Liu T, Chen J, Lv W, Yang W, et al. Decreased Glucose Bioavailability and Elevated Aspartate Metabolism in Prostate Cancer Cells Undergoing Epithelial-Mesenchymal Transition. *J Cell Physiol* (2020) 235(7–8):5602–12. doi: 10.1002/jcp.29490
  58. Tessem MB, Swanson MG, Keshari KR, Albers MJ, Joun D, Tabatabai ZL, et al. Evaluation of Lactate and Alanine as Metabolic Biomarkers of Prostate Cancer Using 1H HR-MAS Spectroscopy of Biopsy Tissues. *Magn Reson Med* (2008) 60(3):510–6. doi: 10.1002/mrm.21694
  59. Wang Q, Hardie RA, Hoy AJ, van Geldermalsen M, Gao D, Fazli L, et al. Targeting ASCT2-Mediated Glutamine Uptake Blocks Prostate Cancer Growth and Tumour Development. *J Pathol* (2015) 236(3):278–89. doi: 10.1002/path.4518
  60. Cardoso HJ, Figueira MI, Vaz CV, Carvalho TMA, Bras LA, Madureira PA, et al. Glutaminolysis is a Metabolic Route Essential for Survival and Growth of Prostate Cancer Cells and a Target of 5alpha-Dihydrotestosterone Regulation. *Cell Oncol (Dordr)* (2021) 44(2):385–403. doi: 10.1007/s13402-020-00575-9
  61. Xu L, Yin Y, Li Y, Chen X, Chang Y, Zhang H, et al. A Glutaminase Isoform Switch Drives Therapeutic Resistance and Disease Progression of Prostate Cancer. *Proc Natl Acad Sci USA* (2021) 118(13):e2012748118. doi: 10.1073/pnas.2012748118
  62. Zhang J, Mao S, Guo Y, Wu Y, Yao X, Huang Y. Inhibition of GLS Suppresses Proliferation and Promotes Apoptosis in Prostate Cancer. *Biosci Rep* (2019) 39(6):BSR20181826. doi: 10.1042/BSR20181826
  63. Pan T, Gao L, Wu G, Shen G, Xie S, Wen H, et al. Elevated Expression of Glutaminase Confers Glucose Utilization via Glutaminolysis in Prostate Cancer. *Biochem Biophys Res Commun* (2015) 456(1):452–8. doi: 10.1016/j.bbrc.2014.11.105
  64. Li R, Younes M, Frolov A, Wheeler TM, Scardino P, Ohori M, et al. Expression of Neutral Amino Acid Transporter ASCT2 in Human Prostate. *Anticancer Res* (2003) 23(4):3413–8.
  65. Balaban S, Nassar ZD, Zhang AY, Hosseini-Beheshti E, Centenera MM, Schreuder M, et al. Extracellular Fatty Acids Are the Major Contributor to Lipid Synthesis in Prostate Cancer. *Mol Cancer Res* (2019) 17(4):949–62. doi: 10.1158/1541-7786.MCR-18-0347
  66. Schlaepfer IR, Glode LM, Hitz CA, Pac CT, Boyle KE, Maroni P, et al. Inhibition of Lipid Oxidation Increases Glucose Metabolism and Enhances 2-Deoxy-2-[(18F)]Fluoro-D-Glucose Uptake in Prostate Cancer Mouse Xenografts. *Mol Imaging Biol* (2015) 17(4):529–38. doi: 10.1007/s11307-014-0814-4
  67. Schlaepfer IR, Rider L, Rodrigues LU, Gijon MA, Pac CT, Romero L, et al. Lipid Catabolism via CPT1 as a Therapeutic Target for Prostate Cancer. *Mol Cancer Ther* (2014) 13(10):2361–71. doi: 10.1158/1535-7163.MCT-14-0183
  68. Yu G, Cheng CJ, Lin SC, Lee YC, Frigo DE, Yu-Lee LY, et al. Organelle-Derived Acetyl-CoA Promotes Prostate Cancer Cell Survival, Migration, and Metastasis via Activation of Calmodulin Kinase II. *Cancer Res* (2018) 78(10):2490–502. doi: 10.1158/0008-5472.CAN-17-2392
  69. Kumar-Sinha C, Shah RB, Laxman B, Tomlins SA, Harwood J, Schmitz W, et al. Elevated  $\alpha$ -Methylacyl-CoA Racemase Enzymatic Activity in Prostate Cancer. *Am J Pathol* (2004) 164(3):787–93. doi: 10.1016/s0002-9440(10)63167-7
  70. Ettinger SL, Sobel R, Whitmore TG, Akbari M, Bradley DR, Gleave ME, et al. Dysregulation of Sterol Response Element-Binding Proteins and Downstream Effectors in Prostate Cancer During Progression to Androgen Independence. *Cancer Res* (2004) 64(6):2212–21. doi: 10.1158/0008-5472.can-2148-2
  71. Huang WC, Li X, Liu J, Lin J, Chung LW. Activation of Androgen Receptor, Lipogenesis, and Oxidative Stress Converged by SREBP-1 is Responsible for Regulating Growth and Progression of Prostate Cancer Cells. *Mol Cancer Res* (2012) 10(1):133–42. doi: 10.1158/1541-7786.MCR-11-0206
  72. Guo D, Bell EH, Mischel P, Chakravarti A. Targeting SREBP-1-Driven Lipid Metabolism to Treat Cancer. *Curr Pharm Des* (2014) 20(15):2619–26. doi: 10.2174/13816128113199990486
  73. Zhang H, Liu S, Cai Z, Dong W, Ye J, Cai Z, et al. Down-Regulation of ACACA Suppresses the Malignant Progression of Prostate Cancer Through Inhibiting Mitochondrial Potential. *J Cancer* (2021) 12(1):232–43. doi: 10.7150/jca.49560
  74. Brusselmanns K, De Schrijver E, Verhoeven G, Swinnen JV. RNA Interference-Mediated Silencing of the Acetyl-CoA-Carboxylase-Alpha

- Gene Induces Growth Inhibition and Apoptosis of Prostate Cancer Cells. *Cancer Res* (2005) 65(15):6719–25. doi: 10.1158/0008-5472.CAN-05-0571
75. De Schrijver E, Brusselmans K, Heyns W, Verhoeven G, Swinnen JV. RNA Interference-Mediated Silencing of the Fatty Acid Synthase Gene Attenuates Growth and Induces Morphological Changes and Apoptosis of LNCaP Prostate Cancer Cells. *Cancer Res* (2003) 63(13):3799–804.
  76. Shah S, Carrière WJ, Li J, Campbell SL, Kopinski PK, Lim HW, et al. Targeting ACLY Sensitizes Castration-Resistant Prostate Cancer Cells to AR Antagonism by Impinging on an ACLY-AMPK-AR Feedback Mechanism. *Oncotarget* (2016) 7(28):43713–30. doi: 10.18632/oncotarget.9666
  77. Nardi F, Franco OE, Fitchev P, Morales A, Vickman RE, Hayward SW, et al. DGAT1 Inhibitor Suppresses Prostate Tumor Growth and Migration by Regulating Intracellular Lipids and Non-Centrosomal MTOC Protein Gm130. *Sci Rep* (2019) 9(1):3035. doi: 10.1038/s41598-019-39537-z
  78. Liu Y, Wang Y, Hao S, Qin Y, Wu Y. Knockdown of Sterol O-Acyltransferase 1 (SOAT1) Suppresses SCD1-Mediated Lipogenesis and Cancer Progression in Prostate Cancer. *Prostaglandins Other Lipid Mediat* (2021) 153:106537. doi: 10.1016/j.prostaglandins.2021.106537
  79. Ashida S, Kawada C, Inoue K. Stromal Regulation of Prostate Cancer Cell Growth by Mevalonate Pathway Enzymes HMGCS1 and HMGR. *Oncol Lett* (2017) 14(6):6533–42. doi: 10.3892/ol.2017.7025
  80. Gordon JA, Noble JW, Midha A, Derakhshan F, Wang G, Adomat HH, et al. Upregulation of Scavenger Receptor B1 Is Required for Steroidogenic and Nonsteroidogenic Cholesterol Metabolism in Prostate Cancer. *Cancer Res* (2019) 79(13):3320–31. doi: 10.1158/0008-5472.CAN-18-2529
  81. Nomura DK, Lombardi DP, Chang JW, Niessen S, Ward AM, Long JZ, et al. Monoacylglycerol Lipase Exerts Dual Control Over Endocannabinoid and Fatty Acid Pathways to Support Prostate Cancer. *Chem Biol* (2011) 18(7):846–56. doi: 10.1016/j.chembiol.2011.05.009
  82. Centenera MM, Scott JS, Machiels J, Nassar ZD, Miller DC, Zinonos I, et al. ELOVL5 Is a Critical and Targetable Fatty Acid Elongase in Prostate Cancer. *Cancer Res* (2021) 81(7):1704–18. doi: 10.1158/0008-5472.CAN-20-2511
  83. Tamura K, Makino A, Hullin-Matsuda F, Kobayashi T, Furihata M, Chung S, et al. Novel Lipogenic Enzyme ELOVL7 Is Involved in Prostate Cancer Growth Through Saturated Long-Chain Fatty Acid Metabolism. *Cancer Res* (2009) 69(20):8133–40. doi: 10.1158/0008-5472.CAN-09-0775
  84. Itkonen HM, Brown M, Urbanucci A, Tredwell G, Ho Lau C, Barfeld S, et al. Lipid Degradation Promotes Prostate Cancer Cell Survival. *Oncotarget* (2017) 8(24):38264–75. doi: 10.18632/oncotarget.16123
  85. Blomme A, Ford CA, Mui E, Patel R, Ntala C, Jamieson LE, et al. 2,4-Dienoyl-CoA Reductase Regulates Lipid Homeostasis in Treatment-Resistant Prostate Cancer. *Nat Commun* (2020) 11(1):2508. doi: 10.1038/s41467-020-16126-7
  86. Nassar ZD, Mah CY, Dehairs J, Burvenich IJ, Irani S, Centenera MM, et al. Human DECR1 is an Androgen-Repressed Survival Factor That Regulates PUFA Oxidation to Protect Prostate Tumor Cells From Ferroptosis. *Elife* (2020) 9:e54166. doi: 10.7554/eLife.54166
  87. Li J, Ren S, Piao HL, Wang F, Yin P, Xu C, et al. Integration of Lipidomics and Transcriptomics Unravels Aberrant Lipid Metabolism and Defines Cholesteryl Oleate as Potential Biomarker of Prostate Cancer. *Sci Rep* (2016) 6:20984. doi: 10.1038/srep20984
  88. Butler LM, Mah CY, Machiels J, Vincent AD, Irani S, Mutuku SM, et al. Lipidomic Profiling of Clinical Prostate Cancer Reveals Targetable Alterations in Membrane Lipid Composition. *Cancer Res* (2021) 81(19):4981–93. doi: 10.1158/0008-5472.CAN-20-3863
  89. Tousignant KD, Rockstroh A, Taherian Fard A, Lehman ML, Wang C, McPherson SJ, et al. Lipid Uptake Is an Androgen-Enhanced Lipid Supply Pathway Associated With Prostate Cancer Disease Progression and Bone Metastasis. *Mol Cancer Res* (2019) 17(5):1166–79. doi: 10.1158/1541-7786.MCR-18-1147
  90. Lin C, Salzillo TC, Bader DA, Wilkenfeld SR, Awad D, Pulliam TL, et al. Prostate Cancer Energetics and Biosynthesis. *Adv Exp Med Biol* (2019) 1210:185–237. doi: 10.1007/978-3-030-32656-2\_10
  91. Servant R, Garioni M, Vlajnic T, Blind M, Püeschel H, Müller DC, et al. Prostate Cancer Patient-Derived Organoids: Detailed Outcome From a Prospective Cohort of 81 Clinical Specimens. *J Pathol* (2021) 254:543–55. doi: 10.1002/path.5698
  92. Puca L, Bareja R, Prandi D, Shaw R, Benelli M, Karthaus WR, et al. Patient Derived Organoids to Model Rare Prostate Cancer Phenotypes. *Nat Commun* (2018) 9(1):2404. doi: 10.1038/s41467-018-04495-z
  93. Karkampouna S, La Manna F, Benjak A, Kiener M, De Menna M, Zoni E, et al. Patient-Derived Xenografts and Organoids Model Therapy Response in Prostate Cancer. *Nat Commun* (2021) 12(1):1117. doi: 10.1038/s41467-021-21300-6
  94. Palanisamy N, Yang J, Shepherd PDA, Li-Ning-Tapia EM, Labanca E, Manyam GC, et al. The MD Anderson Prostate Cancer Patient-Derived Xenograft Series (MDA PCa PDX) Captures the Molecular Landscape of Prostate Cancer and Facilitates Marker-Driven Therapy Development. *Clin Cancer Res* (2020) 26(18):4933–46. doi: 10.1158/1078-0432.CCR-20-0479
  95. Lin D, Wyatt AW, Xue H, Wang Y, Dong X, Haegert A, et al. High Fidelity Patient-Derived Xenografts for Accelerating Prostate Cancer Discovery and Drug Development. *Cancer Res* (2014) 74(4):1272–83. doi: 10.1158/0008-5472.CAN-13-2921-T
  96. Risbridger GP, Clark AK, Porter LH, Toivanen R, Bakshi A, Lister NL, et al. The MURAL Collection of Prostate Cancer Patient-Derived Xenografts Enables Discovery Through Preclinical Models of Uro-Oncology. *Nat Commun* (2021) 12(1):5049. doi: 10.1038/s41467-021-25175-5
  97. Kim J, DeBerardinis RJ. Mechanisms and Implications of Metabolic Heterogeneity in Cancer. *Cell Metab* (2019) 30(3):434–46. doi: 10.1016/j.cmet.2019.08.013
  98. Shi X, Tasdogan A, Huang F, Hu Z, Morrison SJ, DeBerardinis RJ. The Abundance of Metabolites Related to Protein Methylation Correlates With the Metastatic Capacity of Human Melanoma Xenografts. *Sci Adv* (2017) 3(11):eaao5268. doi: 10.1126/sciadv.aao5268
  99. Parmenter TJ, Kleinschmidt M, Kinross KM, Bond ST, Li J, Kaadige MR, et al. Response of BRAF-Mutant Melanoma to BRAF Inhibition is Mediated by a Network of Transcriptional Regulators of Glycolysis. *Cancer Discovery* (2014) 4(4):423–33. doi: 10.1158/2159-8290.CD-13-0440
  100. Davidson SM, Papagiannakopoulos T, Olenchok BA, Heyman JE, Keibler MA, Luengo A, et al. Environment Impacts the Metabolic Dependencies of Ras-Driven Non-Small Cell Lung Cancer. *Cell Metab* (2016) 23(3):517–28. doi: 10.1016/j.cmet.2016.01.007
  101. Yuneva MO, Fan TW, Allen TD, Higashi RM, Ferraris DV, Tsukamoto T, et al. The Metabolic Profile of Tumors Depends on Both the Responsible Genetic Lesion and Tissue Type. *Cell Metab* (2012) 15(2):157–70. doi: 10.1016/j.cmet.2011.12.015
  102. Ying H, Kimmelman AC, Lyssiotis CA, Hua S, Chu GC, Fletcher-Sananikone E, et al. Oncogenic Kras Maintains Pancreatic Tumors Through Regulation of Anabolic Glucose Metabolism. *Cell* (2012) 149(3):656–70. doi: 10.1016/j.cell.2012.01.058
  103. Mayers JR, Torrence ME, Danai LV, Papagiannakopoulos T, Davidson SM, Bauer MR, et al. Tissue of Origin Dictates Branched-Chain Amino Acid Metabolism in Mutant Kras-Driven Cancers. *Science* (2016) 353(6304):1161–5. doi: 10.1126/science.aaf5171
  104. Armenia J, Wankowicz SAM, Liu D, Gao J, Kundra R, Reznik E, et al. The Long Tail of Oncogenic Drivers in Prostate Cancer. *Nat Genet* (2018) 50(5):645–51. doi: 10.1038/s41588-018-0078-z
  105. Labbe DP, Zadra G, Yang M, Reyes JM, Lin CY, Cacciatore S, et al. High-Fat Diet Fuels Prostate Cancer Progression by Rewiring the Metabolome and Amplifying the MYC Program. *Nat Commun* (2019) 10(1):4358. doi: 10.1038/s41467-019-12298-z
  106. Singh KB, Hahm ER, Kim SH, Wendell SG, Singh SV. A Novel Metabolic Function of Myc in Regulation of Fatty Acid Synthesis in Prostate Cancer. *Oncogene* (2021) 40(3):592–602. doi: 10.1038/s41388-020-01553-z
  107. Epstein JI, Amin MB, Beltran H, Lotan TL, Mosquera JM, Reuter VE, et al. Proposed Morphologic Classification of Prostate Cancer With Neuroendocrine Differentiation. *Am J Surg Pathol* (2014) 38(6):756–67. doi: 10.1097/PAS.0000000000000208
  108. Beltran H, Hruszkewycz A, Scher HI, Hildesheim J, Isaacs J, Yu EY, et al. The Role of Lineage Plasticity in Prostate Cancer Therapy Resistance. *Clin Cancer Res* (2019) 25(23):6916–24. doi: 10.1158/1078-0432.CCR-19-1423
  109. Bakht MK, Lovnicki JM, Tubman J, Stringer KF, Chiaramonte J, Reynolds MR, et al. Differential Expression of Glucose Transporters and Hexokinases in Prostate Cancer With a Neuroendocrine Gene Signature: A Mechanistic Perspective for (18)F-FDG Imaging of PSMA-Suppressed Tumors. *J Nucl Med* (2020) 61(6):904–10. doi: 10.2967/jnumed.119.231068
  110. Choi SYC, Ettinger SL, Lin D, Xue H, Ci X, Nabavi N, et al. Targeting MCT4 to Reduce Lactic Acid Secretion and Glycolysis for Treatment of

- Neuroendocrine Prostate Cancer. *Cancer Med* (2018) 7:3385–92. doi: 10.1002/cam4.1587
111. Semenza GL. Targeting HIF-1 for Cancer Therapy. *Nat Rev Cancer* (2003) 3(10):721–32. doi: 10.1038/nrc1187
  112. Li Z, Sun C, Qin Z. Metabolic Reprogramming of Cancer-Associated Fibroblasts and its Effect on Cancer Cell Reprogramming. *Theranostics* (2021) 11(17):8322–36. doi: 10.7150/thno.62378
  113. Jaye K, Li CG, Bhuyan DJ. The Complex Interplay of Gut Microbiota With the Five Most Common Cancer Types: From Carcinogenesis to Therapeutics to Prognoses. *Crit Rev Oncol Hematol* (2021) 165:103429. doi: 10.1016/j.critrevonc.2021.103429
  114. Chen S, Zhu G, Yang Y, Wang F, Xiao YT, Zhang N, et al. Single-Cell Analysis Reveals Transcriptomic Remodellings in Distinct Cell Types That Contribute to Human Prostate Cancer Progression. *Nat Cell Biol* (2021) 23(1):87–98. doi: 10.1038/s41556-020-00613-6
  115. Abarca-Gómez L, Abdeen ZA, Hamid ZA, Abu-Rmeileh NM, Acosta-Cazares B, Acuin C, et al. Worldwide Trends in Body-Mass Index, Underweight, Overweight, and Obesity from 1975 to 2016: A Pooled Analysis of 2416 Population-Based Measurement Studies in 128·9 Million Children, Adolescents, and Adults. *The Lancet* (2017) 390(10113):2627–42. doi: 10.1016/s0140-6736(17)32129-3
  116. Australian Bureau of Statistics. *National Health Survey: First Results 2017-2018 (2018) [Updated 28 July 2021]*. Available at: <https://www.abs.gov.au/statistics/health/health-conditions-and-risks/national-health-survey-first-results/latest-release>.
  117. Australian Institute of Health and Welfare. *Cancer Data in Australia [Internet]. Canberra : AIHW [Updated 2021 June 08]. CAN 122*. Available at: <https://www.aihw.gov.au/reports/cancer/cancer-data-in-australia>.
  118. Wilson KM, Giovannucci EL, Mucci LA. Lifestyle and Dietary Factors in the Prevention of Lethal Prostate Cancer. *Asian J Androl* (2012) 14(3):365–74. doi: 10.1038/aja.2011.142
  119. Khandekar MJ, Cohen P, Spiegelman BM. Molecular Mechanisms of Cancer Development in Obesity. *Nat Rev Cancer* (2011) 11(12):886–95. doi: 10.1038/nrc3174
  120. Discacciati A, Orsini N, Wolk A. Body Mass Index and Incidence of Localized and Advanced Prostate Cancer—a Dose-Response Meta-Analysis of Prospective Studies. *Ann Oncol* (2012) 23(7):1665–71. doi: 10.1093/annonc/mdr603
  121. Langlais CS, Cowan JE, Neuhaus J, Kenfield SA, Van Blarigan EL, Broering JM, et al. Obesity at Diagnosis and Prostate Cancer Prognosis and Recurrence Risk Following Primary Treatment by Radical Prostatectomy. *Cancer Epidemiol Biomarkers Prev* (2019) 28(11):1917–25. doi: 10.1158/1055-9965.EPI-19-0488
  122. Martini A, Shah QN, Waingankar N, Sfakianos JP, Tsao CK, Necchi A, et al. The Obesity Paradox in Metastatic Castration-Resistant Prostate Cancer. *Prostate Cancer Prostatic Dis* (2021). doi: 10.1038/s41391-021-00418-0
  123. Taylor RA, Lo J, Asci N, Watt MJ. Linking Obesogenic Dysregulation to Prostate Cancer Progression. *Endocr Connect* (2015) 4(4):R68–80. doi: 10.1530/EC-15-0080
  124. Bhindi B, Trottier G, Elharram M, Fernandes KA, Lockwood G, Toi A, et al. Measurement of Peri-Prostatic Fat Thickness Using Transrectal Ultrasonography (TRUS): A New Risk Factor for Prostate Cancer. *BJU Int* (2012) 110(7):980–6. doi: 10.1111/j.1464-410X.2012.10957.x
  125. Laurent V, Toulet A, Attane C, Milhas D, Dauvillier S, Zaidi F, et al. Periprostatic Adipose Tissue Favors Prostate Cancer Cell Invasion in an Obesity-Dependent Manner: Role of Oxidative Stress. *Mol Cancer Res* (2019) 17(3):821–35. doi: 10.1158/1541-7786.MCR-18-0748
  126. Lo JC, Clark AK, Asci N, Frydenberg M, Risbridger GP, Taylor RA, et al. Obesity Does Not Promote Tumorigenesis of Localized Patient-Derived Prostate Cancer Xenografts. *Oncotarget* (2016) 7(30):47650–62. doi: 10.18632/oncotarget.10258
  127. Laurent V, Guerard A, Mazerolles C, Le Gonidec S, Toulet A, Nieto L, et al. Periprostatic Adipocytes Act as a Driving Force for Prostate Cancer Progression in Obesity. *Nat Commun* (2016) 7:10230. doi: 10.1038/ncomms10230
  128. Kenfield SA, Stampfer MJ, Giovannucci E, Chan JM. Physical Activity and Survival After Prostate Cancer Diagnosis in the Health Professionals Follow-Up Study. *J Clin Oncol* (2011) 29(6):726–32. doi: 10.1200/JCO.2010.31.5226
  129. Koelwyn GJ, Zhuang X, Tammela T, Schietinger A, Jones LW. Exercise and Immunometabolic Regulation in Cancer. *Nat Metab* (2020) 2(9):849–57. doi: 10.1038/s42255-020-00277-4
  130. Galvao DA, Taaffe DR, Spry N, Gardiner RA, Taylor R, Risbridger GP, et al. Enhancing Active Surveillance of Prostate Cancer: The Potential of Exercise Medicine. *Nat Rev Urol* (2016) 13(5):258–65. doi: 10.1038/nrurol.2016.46
  131. Davis ID, Martin AJ, Stockler MR, Begbie S, Chi KN, Chowdhury S, et al. Enzalutamide With Standard First-Line Therapy in Metastatic Prostate Cancer. *N Engl J Med* (2019) 381(2):121–31. doi: 10.1056/NEJMoa1903835
  132. Abida W, Patnaik A, Campbell D, Shapiro J, Bryce AH, McDermott R, et al. Rucaparib in Men With Metastatic Castration-Resistant Prostate Cancer Harboring a BRCA1 or BRCA2 Gene Alteration. *J Clin Oncol* (2020) 38(32):3763–72. doi: 10.1200/JCO.20.01035
  133. de Bono J, Mateo J, Fizazi K, Saad F, Shore N, Sandhu S, et al. Olaparib for Metastatic Castration-Resistant Prostate Cancer. *N Engl J Med* (2020) 382(22):2091–102. doi: 10.1056/NEJMoa1911440
  134. Faris JE, Smith MR. Metabolic Sequelae Associated With Androgen Deprivation Therapy for Prostate Cancer. *Curr Opin Endocrinol Diabetes Obes* (2010) 17(3):240–6. doi: 10.1097/MED.0b013e3283391fd1
  135. Chi JT, Lin PH, Tolstikov V, Oyekunle T, Chen EY, Bussberg V, et al. Metabolomic Effects of Androgen Deprivation Therapy Treatment for Prostate Cancer. *Cancer Med* (2020) 9(11):3691–702. doi: 10.1002/cam4.3016
  136. Saylor PJ, Karoly ED, Smith MR. Prospective Study of Changes in the Metabolomic Profiles of Men During Their First Three Months of Androgen Deprivation Therapy for Prostate Cancer. *Clin Cancer Res* (2012) 18(13):3677–85. doi: 10.1158/1078-0432.CCR-11-3209
  137. Chi JT, Lin PH, Tolstikov V, Oyekunle T, Alvarado GCG, Ramirez-Torres A, et al. The Influence of Low-Carbohydrate Diets on the Metabolic Response to Androgen-Deprivation Therapy in Prostate Cancer. *Prostate* (2021) 81(10):618–28. doi: 10.1002/pros.24136
  138. Fendt SM, Frezza C, Erez A. Targeting Metabolic Plasticity and Flexibility Dynamics for Cancer Therapy. *Cancer Discovery* (2020) 10(12):1797–807. doi: 10.1158/2159-8290.CD-20-0844
  139. Zhou J, Wang Y, Wu D, Wang S, Chen Z, Xiang S, et al. Orphan Nuclear Receptors as Regulators of Intratumoral Androgen Biosynthesis in Castration-Resistant Prostate Cancer. *Oncogene* (2021) 40(15):2625–34. doi: 10.1038/s41388-021-01737-1
  140. Auchus RJ, Sharifi N. Sex Hormones and Prostate Cancer. *Annu Rev Med* (2020) 71:33–45. doi: 10.1146/annurev-med-051418-060357
  141. Yang B, Damodaran S, Khemees TA, Filon MJ, Schultz A, Gawdzik J, et al. Synthetic Lethal Metabolic Targeting of Androgen Deprived Prostate Cancer Cells With Metformin. *Mol Cancer Ther* (2020) 19(11):2278–87. doi: 10.1158/1535-7163.MCT-19-1141
  142. Heuer TS, Ventura R, Mordec K, Lai J, Fridlib M, Buckley D, et al. FASN Inhibition and Taxane Treatment Combine to Enhance Anti-Tumor Efficacy in Diverse Xenograft Tumor Models Through Disruption of Tubulin Palmitoylation and Microtubule Organization and FASN Inhibition-Mediated Effects on Oncogenic Signaling and Gene Expression. *EBioMedicine* (2017) 16:51–62. doi: 10.1016/j.ebiom.2016.12.012
  143. Zadra G, Ribeiro CF, Chetta P, Ho Y, Cacciatore S, Gao X, et al. Inhibition of De Novo Lipogenesis Targets Androgen Receptor Signaling in Castration-Resistant Prostate Cancer. *Proc Natl Acad Sci USA* (2019) 116(2):631–40. doi: 10.1073/pnas.1808834116
  144. Falchook G, Infante J, Arkenau HT, Patel MR, Dean E, Borazanci E, et al. First-In-Human Study of the Safety, Pharmacokinetics, and Pharmacodynamics of First-in-Class Fatty Acid Synthase Inhibitor TVB-2640 Alone and With a Taxane in Advanced Tumors. *EClinicalMedicine* (2021) 34:100797. doi: 10.1016/j.eclinm.2021.100797
  145. Looma R, Kayali Z, Noureddin M, Ruane P, Lawitz EJ, Bennett M, et al. GS-0976 Reduces Hepatic Steatosis and Fibrosis Markers in Patients With Nonalcoholic Fatty Liver Disease. *Gastroenterology* (2018) 155(5):1463–73 e6. doi: 10.1053/j.gastro.2018.07.027
  146. Holubarsch CJ, Rohrbach M, Karrasch M, Boehm E, Polonski L, Ponikowski P, et al. A Double-Blind Randomized Multicenter Clinical Trial to Evaluate the Efficacy and Safety of Two Doses of Etomoxir in Comparison With Placebo in Patients With Moderate Congestive Heart Failure: The ERGO

- (Etomoxir for the Recovery of Glucose Oxidation) Study. *Clin Sci (Lond)* (2007) 113(4):205–12. doi: 10.1042/CS20060307
147. Bhandari B, Subramanian L. Ranolazine, a Partial Fatty Acid Oxidation Inhibitor, its Potential Benefit in Angina and Other Cardiovascular Disorders. *Recent Pat Cardiovasc Drug Discovery* (2007) 2(1):35–9. doi: 10.2174/15748900779606095
  148. Ashrafi H, Horowitz JD, Frenneaux MP. Perhexiline. *Cardiovasc Drug Rev* (2007) 25(1):76–97. doi: 10.1111/j.1527-3466.2007.00006.x
  149. Flaig TW, Salzmann-Sullivan M, Su LJ, Zhang Z, Joshi M, Gijon MA, et al. Lipid Catabolism Inhibition Sensitizes Prostate Cancer Cells to Antiandrogen Blockade. *Oncotarget* (2017) 8(34):56051–65. doi: 10.18632/oncotarget.17359
  150. Nassar ZD, Mah CY, Centenera MM, Irani S, Sadowski MC, Scott JS, et al. Fatty Acid Oxidation Is an Adaptive Survival Pathway Induced in Prostate Tumors by HSP90 Inhibition. *Mol Cancer Res* (2020) 18(10):1500–11. doi: 10.1158/1541-7786.MCR-20-0570
  151. Harding JJ, Telli ML, Munster PN, Le MH, Molineaux C, Bennett MK, et al. Safety and Tolerability of Increasing Doses of CB-839, a First-in-Class, Orally Administered Small Molecule Inhibitor of Glutaminase, in Solid Tumors. *J Clin Oncol* (2015) 33(15\_suppl):2512–. doi: 10.1200/jco.2015.33.15\_suppl.2512
  152. Emberley ED, Bennett M, Chen J, Gross M, Huang T, Makkouk A, et al. Abstract 3509: The Glutaminase Inhibitor CB-839 Synergizes With CDK4/6 and PARP Inhibitors in Pre-Clinical Tumor Models. *Cancer Res* (2018) 78(13 Supplement):3509–. doi: 10.1158/1538-7445.Am2018-3509
  153. Yang H, Pang L, Hu X, Wang W, Xu B, Zhang X, et al. The Effect of Statins on Advanced Prostate Cancer Patients With Androgen Deprivation Therapy or Abiraterone/Enzalutamide: A Systematic Review and Meta-Analysis. *J Clin Pharm Ther* (2020) 45(3):488–95. doi: 10.1111/jcpt.13092
  154. Murtola TJ, Syvala H, Tolonen T, Helminen M, Riikonen J, Koskimaki J, et al. Atorvastatin Versus Placebo for Prostate Cancer Before Radical Prostatectomy—A Randomized, Double-Blind, Placebo-Controlled Clinical Trial. *Eur Urol* (2018) 74(6):697–701. doi: 10.1016/j.eururo.2018.06.037
  155. Longo J, Mullen PJ, Yu R, van Leeuwen JE, Masoomian M, Woon DTS, et al. An Actionable Sterol-Regulated Feedback Loop Modulates Statin Sensitivity in Prostate Cancer. *Mol Metab* (2019) 25:119–30. doi: 10.1016/j.molmet.2019.04.003
  156. Zingales V, Distefano A, Raffaele M, Zanghi A, Barbagallo I, Vanella L. Metformin: A Bridge Between Diabetes and Prostate Cancer. *Front Oncol* (2017) 7:243. doi: 10.3389/fonc.2017.00243
  157. Ahn HK, Lee YH, Koo KC. Current Status and Application of Metformin for Prostate Cancer: A Comprehensive Review. *Int J Mol Sci* (2020) 21(22):8540. doi: 10.3390/ijms21228540
  158. Martinez PF, Belisle DF, Cristallo C, Tobia I, Damia O, Villamil W, et al. [Comparison of the Biochemical Recurrence Risk Between Radical Prostatectomy and Brachytherapy in the Treatment of Low- and Intermediate-Risk Prostate Cancer]. *Arch Esp Urol* (2015) 68(8):655–60.
  159. Sweeney CJ, Chen YH, Carducci M, Liu G, Jarrard DF, Eisenberger M, et al. Chemohormonal Therapy in Metastatic Hormone-Sensitive Prostate Cancer. *N Engl J Med* (2015) 373(8):737–46. doi: 10.1056/NEJMoa1503747
  160. Tamada S, Iguchi T, Kato M, Asakawa J, Kita K, Yasuda S, et al. Time to Progression to Castration-Resistant Prostate Cancer After Commencing Combined Androgen Blockade for Advanced Hormone-Sensitive Prostate Cancer. *Oncotarget* (2018) 9(97):36966–74. doi: 10.18632/oncotarget.26426
  161. Aly M, Leval A, Schain F, Liwing J, Lawson J, Vago E, et al. Survival in Patients Diagnosed With Castration-Resistant Prostate Cancer: A Population-Based Observational Study in Sweden. *Scand J Urol* (2020) 54(2):115–21. doi: 10.1080/21681805.2020.1739139
  162. Halabi S, Lin CY, Kelly WK, Fizazi KS, Moul JW, Kaplan EB, et al. Updated Prognostic Model for Predicting Overall Survival in First-Line Chemotherapy for Patients With Metastatic Castration-Resistant Prostate Cancer. *J Clin Oncol* (2014) 32(7):671–7. doi: 10.1200/JCO.2013.52.3696
  163. Moreira DM, Howard LE, Sourbeer KN, Amarasekara HS, Chow LC, Cockrell DC, et al. Predicting Time From Metastasis to Overall Survival in Castration-Resistant Prostate Cancer: Results From SEARCH. *Clin Genitourin Cancer* (2017) 15(1):60–6 e2. doi: 10.1016/j.clgc.2016.08.018
  164. Vander Linden C, Corbet C. Reconciling Environment-Mediated Metabolic Heterogeneity With the Oncogene-Driven Cancer Paradigm in Precision Oncology. *Semin Cell Dev Biol* (2020) 98:202–10. doi: 10.1016/j.semcdb.2019.05.016
  165. Morse N, Jamaspishvili T, Simon D, Patel PG, Ren KYM, Wang J, et al. Reliable Identification of Prostate Cancer Using Mass Spectrometry Metabolomic Imaging in Needle Core Biopsies. *Lab Invest* (2019) 99(10):1561–71. doi: 10.1038/s41374-019-0265-2
  166. Randall EC, Zadra G, Chetta P, Lopez BGC, Syamala S, Basu SS, et al. Molecular Characterization of Prostate Cancer With Associated Gleason Score Using Mass Spectrometry Imaging. *Mol Cancer Res* (2019) 17(5):1155–65. doi: 10.1158/1541-7786.MCR-18-1057
  167. Ventura R, Mordec K, Waszczuk J, Wang Z, Lai J, Fridlib M, et al. Inhibition of *De Novo* Palmitate Synthesis by Fatty Acid Synthase Induces Apoptosis in Tumor Cells by Remodeling Cell Membranes, Inhibiting Signaling Pathways, and Reprogramming Gene Expression. *EBioMedicine* (2015) 2(8):808–24. doi: 10.1016/j.ebiom.2015.06.020

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Fidelito, Watt and Taylor. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.