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Utility of clinical comprehensive genomic characterization for diagnostic categorization in patients presenting with hypocellular bone marrow failure syndromes

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ABSTRACT

Bone marrow failure (BMF) related to hypoplasia of hematopoietic elements in the bone marrow is a heterogeneous clinical entity with a broad differential diagnosis including both inherited and acquired causes. Accurate diagnostic categorization is critical to optimal patient care and detection of genomic variants in these patients may provide this important diagnostic and prognostic information. We performed real-time, accredited (ISO15189) comprehensive genomic characterization including targeted sequencing and whole exome sequencing in 115 patients with BMF syndromes (median age 24 years, range: 3 months - 81 years). In patients with clinical diagnoses of inherited BMF syndromes, acquired BMF syndromes or clinically unclassifiable BMF we detected variants in 52% (12 of 23), 53% (25 of 47) and 56% (25 of 45) respectively. Genomic characterization resulted in a change of diagnosis in 30 of 115 (26%) including the identification of germline causes for 3 of 47 and 16 of 45 cases with pre-test diagnoses of acquired and clinically unclassifiable BMF respectively. The observed clinical impact of accurate diagnostic categorization included choice to perform allogeneic stem cell transplantation, disease-specific targeted treatments, identification of at-risk family members and influence of sibling allogeneic stem cell donor choice. Multiple novel pathogenic variants and copy number changes were identified in our cohort including in *TERT*, *FANCA*, *RPS7* and *SAMD9*. Whole exome sequence analysis facilitated the identification of variants in two genes not typically associated with a primary clinical manifestation of BMF but also demonstrated reduced sensitivity for detecting low level acquired variants. In conclusion, genomic characterization can improve diagnostic categorization of patients presenting with hypoplastic BMF syndromes and should be routinely performed in this group of patients.

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Introduction

Bone marrow failure (BMF) related to hypoplasia of hematopoietic elements in the bone marrow is a heterogeneous clinical entity with a broad differential diagnosis. BMF is most commonly an acquired disorder related to an exogenous mar-

row insult such as drugs (e.g., cytotoxic chemotherapy), toxins or immune destruction of hemopoietic progenitors (acquired aplastic anemia [aAA]). Less commonly, BMF may be the result of an inherited/genetic cause such as Fanconi anemia (FA), Shwachman-Diamond syndrome (SDS), Diamond-Blackfan anemia (DBA) and other rare genetic disorders. Moreover, myelodysplastic syndrome (MDS) can present in the minority of cases (approximately 10%) with marrow hypoplasia (hMDS) rather than the typical hypercellular form with the notable exception that most children and adolescents with MDS will be categorized as refractory cytopenia of childhood (RCC) with the majority of these cases being hypocellular.¹

An accurate diagnosis of the precise etiology and subtype of BMF is critical to providing appropriate clinical care. However, the clinical and morphological features of the bone marrow biopsy of the various BMF entities (both acquired and genetic) are non-specific with significant overlap limiting the ability to differentiate the underlying type of BMF. Whilst investigations such as telomere length assessment and chromosomal fragility testing can be helpful in suggesting an underlying diagnosis (of dyskeratosis congenita [DKC] and FA respectively), these techniques have significant technical limitations.^{2,3} In addition, they do not provide the clinical utility of knowing a specific causative genetic variant that can provide extra information regarding risk stratification, prediction of inheritance patterns and screening of asymptomatic/pre-symptomatic family members.

Genomic evaluation using next generation sequencing (NGS) is a powerful technique to identify variants in a large number of genes in order to resolve the diagnostic and therapeutic challenges faced by physicians treating BMF. Indeed, literature to date suggests a significant contribution to diagnostic rate and reclassification using NGS in patients with inherited/genetic BMF syndromes (IBMFS).⁴ Moreover, in the acquired setting, detection of variants in patients with aAA may inform of the risk of transformation to hematological malignancy and response to immunosuppression^{5,6} as well as aiding the diagnostic categorization between aAA and hMDS.⁴ In some diagnostic laboratories, detection of germline variants is now being performed by whole exome sequencing (WES) as a single approach applicable to a broad range of phenotypes, whereas targeted panel testing is more typically used in the acquired/hematological malignancy setting due to a requirement for greater sensitivity to detect subclonal variants.

Here we describe the results of the Melbourne Genomic Health Alliance Bone Marrow Failure Flagship - a multi-institutional study of 115 patients which aimed to prospectively assess the diagnostic impact of comprehensive genomic evaluation including both WES and targeted NGS panel testing in patients presenting with BMF characterized by bone marrow hypoplasia.

Methods

Recruitment and ethics

Eligible patients were recruited from four participating institutions in Victoria, Australia. Inclusion criteria were (i) age ≥ 3 months, (ii) a clinicopathological diagnosis (based on current guidelines)^{1,3} of either aAA, IBMFS, hMDS or a BMF syndrome characterized by marrow hypoplasia/aplasia but not able to be

definitively categorized. All patients received pre-test counselling and assessment. Standardized pre-test counselling was provided by the treating hematologist or participants were referred to the study hematologist for the purpose of the study. Participants received written information in the form of the Patient Information Consent Form (PICF). Genetic counsellors and clinical geneticists were available at each of the study sites for referral if required. Research was conducted after Institutional Review Board ethics approval (HREC/13/MH/326 and HREC/17/PMCC/163) and all research was conducted in accordance with the Declaration of Helsinki.

Genomic characterization

All patients underwent baseline testing with (i) WES and (ii) targeted panel sequencing with a hybridization-based NGS BMF panel (a subset of the Peter MacCallum Cancer Center PanHaem panel).⁹ Sequence variants alone were analyzed from WES and both sequence variants as well as genome-wide copy number changes were assessed from the targeted BMF panel. Genes analyzed by either WES or targeted panel sequencing are shown in Figure 1 and listed in the *Online Supplementary Table S1*. Details of bioinformatic processing of sequence data are provided in the *Online Supplementary Appendix*. Briefly, both aligned WES and targeted panel data underwent variant calling using a typical germline variant caller (GATK HaplotypeCaller¹⁰) and GATK recommended best practices.^{11,12} In addition the targeted panel aligned sequence data was processed through a dedicated bioinformatics pipeline which included variant calling with GATK4/Mutect2 (<https://software.broadinstitute.org/gatk/>) in order to improve detection of low level acquired variants. Copy number variation (CNV) was estimated by comparing read counts from targeted panel data to a reference pool comprising normal samples with similar technical artefacts and visualized using CNspecter.¹³

Depending on the clinical context and genomic findings, patients underwent further characterization with RNA-studies (including whole transcriptome RNA sequencing), Sanger sequencing and/or droplet digital PCR (ddPCR) (see the *Online Supplementary Appendix*). Sequencing was performed on peripheral blood, bone marrow, buccal epithelial cells or cultured skin fibroblasts depending on the clinical context.

All variants were described according to Human Genome Variation Society (HGVS) nomenclature and germline variants classified according to the American College of Medical Genetics guidelines for the interpretation of sequence variants.¹⁴ Accredited (International Organization for Standardization [ISO] 15189) diagnostic reports were issued to referrers for all patients in real-time. In addition, genomic results were reviewed in centralized multi-disciplinary case conferences attended by the treating clinicians, molecular hematopathologists, medical scientists, clinical geneticists and genetic counsellors.

Results

Cohort description

115 patients were recruited from May 2017 to August 2018. The median age of the overall cohort was 24 years (range: 3 months - 81 years) with a male to female (M:F) ratio of 1:1.4. Upon enrolment, the referring clinical team were asked to assign a BMF subtype based on the available clinicopathological features to capture the “pre-test diagnosis” for each patient, noting that these had been assigned on varied degrees of evidence. Any genomic evaluation completed before enrolment was considered part of “standard-of-care” investigations and were performed

Table 1. Causative genomic variants in patients with the clinical diagnosis of an inherited bone marrow failure syndrome.

Patient ID	Age/ Sex	Pre-test diagnosis	Post-test diagnosis	Germline variants	Acquired variants	Comment
14 [#]	4M	DBA	DBA	<i>RPS19</i> c.251_252del; p.(Arg84Lysfs*69) [het]	–	Variant previously described in DBA ²⁵
25 [#]	14F	DBA	DBA	<i>RPL35A</i> whole gene copy loss [het]	–	<i>RPL35A</i> deletion previously described in DBA ²⁶
31 [#]	16M	DBA	DBA	<i>RPS19</i> exon 2-3 copy loss [het]	–	<i>RPS19</i> copy number losses previously described in DBA ²⁷
76 [#]	33M	DKC	DKC	<i>TINF2</i> c.844C>T; p.(Arg282Cys) [het]	–	Variant previously described in DKC ^{28,29}
44 [#]	35M	FA	FA	<i>RAD51C</i> c.773G>A; p.(Arg258His) [hom]	–	Variant previously described in FA-like syndrome ³⁰⁻³²
85 [#]	33F	FA	FA	<i>FANCI</i> c.3184C>T; p.(Gln1062*) [het] <i>FANCI</i> c.3041G>A; p.(Cys1014Tyr) [het]	–	Variant previously described in FA ^{33,34}
2	9F	SDS	SDS	<i>SBDS</i> c.183_184delinsCT; p.(Lys62*) [het] <i>SBDS</i> c.258+2T>C; p.? [het]	–	Established pathogenic variants in SDS
102	32F	SDS	SDS	<i>SBDS</i> c.183_184delinsCT; p.(Lys62*) [het] <i>SBDS</i> c.258+2T>C; p.? [het]	–	Established pathogenic variants in SDS
107 [#]	18F	SCN	SCN	<i>ELANE</i> c.684C>A; p.(Tyr228*) [het]	<i>CSF3R</i> c.2308C>T; p.(Gln770*)	Variant previously described in SCN ^{35,36} Acquired mutation detected at VAF of 2.9%
18	18F	SCN	SCN	<i>HAX1</i> c.91del; p.(Glu31Lysfs*54) [hom]	–	Variant previously described in SCN ^{37,38}
32 [#]	5F	SCN	SCN	<i>SRP54</i> c.349_351del; p.(Thr117del) [het]	–	Variant previously described in SCN ^{39,40}
104 [#]	20F	SCN	SCN	<i>SRP54</i> c.349_351del; p.(Thr117del) [het]	–	Variant previously described in SCN ^{39,40}

#Case description provided in the *Online Supplementary Appendix*. DBA: Diamond-Blackfan anemia; DKC: Dyskeratosis congenita; FA: Fanconi anemia; SCN: severe congenital neutropenia; SDS: Shwachman-Diamond syndrome; VAF: variant allele frequency; het: heterozygous; hom: homozygous. Reference transcripts – *CSF3R*, NM_156039.3; *ELANE*, NM_001972.2; *FANCI*, NM_001113378.1; *HAX1*, NM_006118.3; *RAD51C*, NM_058216.2; *RPS19*, NM_001022.3; *RPL35A*, NM_000996.2; *SBDS*, NM_016038.2; *SRP54*, NM_003136.3; *TINF2*, NM_001099274.1.

according to individual treating teams/institutional guidelines. For analysis purposes, patients were divided into three different pre-test diagnosis cohorts (i) patients with a pre-diagnosis of an IBMFS (n=23; median age 10 [range: 2-36 years]; M:F ratio 1:1.88), (ii) patients with a pre-test diagnosis of aAA or hMDS (n=47; median age 32 [range: 7-80 years]; M:F ratio 1:1.24) and (iii) patients with a pre-test diagnosis of clinically unclassifiable BMF (UBMF) (n=45; median age 25 [range: 1-81 years]; M:F ratio 1:1.37) (Figure 2). If the referring clinical team could not definitively assign a pre-test diagnosis of IBMFS (including subtype) or aAA/hMDS, the patient was assigned to the UBMF category. After comprehensive genomic characterization was completed, multidisciplinary re-review of all clinical and pathological features incorporating newly acquired genomic data was performed and a “post-test diagnosis” was assigned. Clinical management of patients after assignment of post-test diagnosis was captured in regard to (i) proceeding to allogeneic stem cell transplant, (ii) screening of relatives for identified causative variants and (iii) disease-specific interventions.

Genomic characterization identifies causative variants in the majority of patients with a pre-test diagnosis of an IBMFS

In patients with a pre-test diagnosis of an IBMFS, genomic characterization confirmed an IBMFS by detecting causative germline variants in 52.2% (12 of 23). The genomic variants detected in this cohort are listed in Table 1. In all cases where a causative germline variant was identified, the post-test diagnosis was consistent with the pre-test diagnosis. One acquired *CSF3R* variant was detected in this group in a patient with severe congenital neutropenia (SCN) and a germline *ELANE* variant who had life-long treatment with G-CSF. A causative

germline variant was not identified in patients with a pre-test diagnosis of SCN in 71.4% (10 of 14) of cases despite genomic characterization.

Despite no change in the pre-test diagnosis in this group, the observed clinical impacts of rendering a precise genomic diagnosis to this group of patients with established clinical diagnoses included one patient (#76) where the identification of a specific variant in *TINF2* permitted screening of his pre-symptomatic children for DKC. In addition, this group included an adult patient (#44) with the observation of a homozygous variant in *RAD51C* (Arg258His) resulting in a Fanconi-like syndrome (associated with positive chromosomal fragility studies) with subsequent implications for breast and ovarian cancer risk screening in siblings. Further clinicopathological details for cases are presented in the *Online Supplementary Appendix*.

Both acquired and inherited variants were detected in patients with a pre-test diagnosis of aAA/hMDS

In patients with a pre-test diagnosis of aAA/hMDS, acquired variants were detected in 53.2% of patients (25 of 47) (see Table 2). Forty-one of 47 patients had a pre-test diagnosis of aAA (22 of 41 [53.7%] with variants) and 6 of 47 had a pre-test diagnosis of hMDS (3 of 6 [50%] with variants). Variant allele frequency (VAF) of acquired variants in this group ranged from 0.35%-56.1%. Acquired variants were detected in *PIGA* (n=15), *ASXL1* (n=6), *BCOR* (n=4), *RUNX1* (n=4), *CBL* (n=3), *CSMD1* (n=3), *DNMT3A* (n=2), *TET2* (n=2), *U2AF1* (n=2), *SRSF2* (n=1), *SF3B1* (n=1), *TERT* (n=1), *IKZF1* (n=1), *IDH2* (n=1) and *BCORL1* (n=1). Three patients in this group had multiple subclonal *PIGA* variants detectable, a phenomenon previously described in patients with paroxysmal nocturnal hemoglobinuria

Table 2. Variants detected in patients with a clinicopathological diagnosis of acquired aplastic anemia or hypoplastic myelodysplastic syndrome.

Patient ID	Age/ Sex	Pre-test diagnosis	Post-test diagnosis	Germline variants	Acquired variants	Acquired VAF
22	19F	aAA	aAA	–	<i>ASXL1</i> c.3785del; p.(Ser1262Thrfs*18)	2.8%
30	17M	aAA	aAA	–	<i>ASXL1</i> c.1864_1865del; p.(Leu622Alafs*12) <i>CSMD1</i> c.1124G>A; p.(Cys375Tyr)	1.6% 1.8%
37	28M	aAA	aAA	–	<i>BCORL1</i> c.1429dup; p.(Thr477Asnfs*58) <i>CSMD1</i> c.4377del; p.(Gly1460Valfs*22)	3.3% 1.7%
39 [†]	27F	aAA	IBMFS (FA)	<i>FANCA</i> c.2980A>G; p.(Ser994Gly) [hom]	–	100%
46	31F	aAA	aAA	–	<i>PIGA</i> c.986dup; p.(Ser330Lysfs*9) <i>PIGA</i> c.849-2A>G; p.?	1.6% 3.9%
47 [†]	64M	aAA	IBMFS (APS)	<i>SAMD9L</i> c.2956C>T; p.(Arg986Cys) [het]	<i>CBL</i> c.1259G>C; p.(Arg420Pro)	42.9% 7.5%
87 [†]	49M	aAA	IBMFS (Telomeropathy)	<i>TERT</i> c.1223T>C; p.(Leu408Pro) [het]	<i>BCOR</i> c.3549_3561dup; p.(Val1188Metfs*27) <i>TERT</i> c.-124C>T	52.4% 16% –
41	50F	aAA	aAA	–	<i>PIGA</i> c.749T>C; p.(Leu250Pro) <i>PIGA</i> c.684del; p.(Ile228Metfs*19) <i>PIGA</i> c.1347dup; p.(Ile450Tyrfs*3) <i>PIGA</i> c.105_106del; p.(Ile35Metfs*5)	18.1% 6.9% 4.3% 1.0%
42	47F	aAA	aAA	–	<i>RUNX1</i> c.317G>T; p.(Trp106Leu) <i>ASXL1</i> c.2329G>T; p.(Glu777*) <i>ASXL1</i> c.2053G>T; p.(Gly685*)	41.7% 6.0% 4.2%
43	24M	aAA	aAA	–	<i>PIGA</i> c.491C>T; p.(Ser164Leu)	2.4%
49	22M	aAA	aAA	–	<i>PIGA</i> c.715+2T>G; p.?	13.0%
55	71F	aAA	aAA	–	<i>PIGA</i> c.944G>A; p.(Cys315Tyr)	1.9%
57	36F	aAA	aAA	–	<i>DNMT3A</i> c.2645G>A; p.(Arg882His)	8.6%
58	49F	aAA	aAA	–	<i>BCOR</i> c.4537C>T; p.(Arg1513*)	1.8%
65	73M	aAA	MDS	–	<i>PIGA</i> c.715+1G>T; p.? <i>TET2</i> c.488del; p.(Phe163Serfs*20) <i>IDH2</i> c.419G>A; p.(Arg140Gln) <i>SF3B1</i> c.1997A>G; p.(Lys666Arg) <i>U2AF1</i> c.470A>C; p.(Gln157Pro) <i>ASXL1</i> c.1934dup; p.(Gly646Trpfs*12)	56.1% 10.6% 8.0% 8.3% 24.3% 22.5%
66	36F	aAA	aAA	–	<i>BCOR</i> c.1971C>A; p.(Tyr657*)	2.0%
71	38M	aAA	aAA	–	<i>PIGA</i> c.715+1G>A; p.?	2.6%
78	37F	aAA	aAA	–	<i>PIGA</i> c.811_812del; p.(Leu271Glyfs*11)	1.9%
89	37F	aAA	MDS	–	<i>RUNX1</i> c.790_791del; p.(Gln264Glufs*335) <i>RUNX1</i> c.602G>A; p.(Arg201Gln) <i>CBL</i> c.1259G>A; p.(Arg420Gln) <i>CBL</i> c.616C>T; p.(Arg206*) <i>ASXL1</i> c.2555C>G; p.(Ser852*)	8.2% 1.4% 16.6% 9.9% 40.2%
101	38F	aAA	aAA	–	<i>TET2</i> c.4910_4911insTC; p.(Ser1638Hisfs*58)	0.52%
108	24F	aAA	MDS	–	<i>U2AF1</i> c.101C>T; p.(Ser34Phe) <i>PIGA</i> c.548G>T; p.(Cys183Phe) <i>PIGA</i> c.525_526del; p.(Cys176*) <i>PIGA</i> c.227_229delinsTT; p.(Asn761Ilefs*19)	8.3% 1.8% 0.35% 0.47%
114	40F	aAA	aAA	–	<i>BCOR</i> c.4504G>A; p.(Asp1502Asn)	0.89%
45	58M	hMDS	hMDS	–	<i>DNMT3A</i> c.2525A>G; p.(Gln842Arg)	19.8%
9	19M	hMDS	hMDS	–	<i>IKZF1</i> c.476A>G; p.(Asn159Ser)	7.1%
111	75M	hMDS	hMDS	–	<i>RUNX1</i> c.602G>A; p.(Arg201Gln) <i>SRSF2</i> c.284_304del; p.(Pro95_Ser101del) <i>CSMD1</i> c.2257C>T; p.(Arg753Cys)	32.5% 50.3% 1.5%

[†]Case description provided in the *Online Supplementary Appendix*. aAA: acquired aplastic anemia; FA: Fanconi anemia; APS: ataxia-pancytopenia syndrome; MDS: myelodysplastic syndrome; hMDS, hypocellular MDS; VAF: variant allele frequency; het: heterozygous; hom: homozygous. Reference transcripts – *ASXL1*, NM_015338.5; *BCOR*, NM_017745.5; *BCORL1*, NM_021946.4; *CBL*, NM_005188.3; *CSMD1*, NM_033225.5; *DNMT3A*, NM_022552.4; *FANCA*, NM_000135.2; *IDH2*, NM_002168.2; *IKZF1*, NM_006060.4; *PIGA*, NM_002641.3; *RUNX1*, NM_001754.4; *SAMD9L*, NM_152703.2; *SF3B1*, NM_012433.2; *SRSF2*, NM_003016.4; *TERT*, NM_198253.2; *TET2*, NM_001127208.2; *U2AF1*, NM_006758.2.

Severe Congenital Neutropenia				
<i>CSF3R</i>	<i>G6PC3</i>	<i>HAX1</i>	<i>SRP54*</i>	<i>VPS45</i>
<i>ELANE</i>	<i>GFI1</i>	<i>JAGN1</i>	<i>TCIRG1</i>	<i>WAS</i>
Shwachman-Diamond Syndrome				
<i>SBDS</i>		<i>DNAJC21*</i>		
Diamond-Blackfan Anemia				
<i>GATA1</i>	<i>RPL15</i>	<i>RPS7</i>	<i>RPS19</i>	<i>RPS26</i>
<i>RPL5</i>	<i>RPL26</i>	<i>RPS10</i>	<i>RPS24</i>	<i>RPS29</i>
<i>RPL11</i>	<i>RPL35A</i>			
Fanconi Anemia				
<i>BRCA2</i>	<i>FANCA</i>	<i>FANCD2</i>	<i>FANCG</i>	<i>PALB2</i>
<i>BRIP1</i>	<i>FANCB</i>	<i>FANCE</i>	<i>FANCI</i>	<i>RAD51C</i>
<i>ERCC4</i>	<i>FANCC</i>	<i>FANCF</i>	<i>FANCL</i>	<i>SLX4</i>
Telomeropathies				
<i>ACD</i>	<i>DKC1</i>	<i>PARN</i>	<i>NHP2</i>	<i>TINF2</i>
<i>CTC1</i>	<i>NOP10</i>	<i>RTEL1</i>	<i>TERT</i>	<i>WRAP53</i>
Predisposition to Hematological Malignancy				
<i>ANKRD26*</i>	<i>DDX41</i>	<i>ETV6*</i>	<i>GATA2</i>	<i>RUNX1</i>
Other IBMFS				
<i>ATR</i>	<i>ERCC6L2</i>	<i>RBM8A</i>	<i>SH2D1A</i>	<i>SRP72</i>
<i>AK2</i>	<i>FANCM</i>	<i>SAMD9*</i>	<i>SMARCD2*</i>	<i>THPO*</i>
	<i>MPL</i>	<i>SAMD9L*</i>	<i>SMARCAL1</i>	

aAA/hMDS			
<i>ASXL1</i>	<i>DNMT3A</i>	<i>JAK3</i>	<i>SF3B1</i>
<i>BCOR</i>	<i>ERBB2</i>	<i>KIT</i>	<i>SRSF2</i>
<i>BCORL1</i>	<i>EZH2</i>	<i>KRAS</i>	<i>TET2</i>
<i>CALR</i>	<i>GATA2</i>	<i>MPL</i>	<i>TP53</i>
<i>CBL</i>	<i>IDH1</i>	<i>NRAS</i>	<i>U2AF1</i>
<i>CSF3R</i>	<i>IDH2</i>	<i>PIGA</i>	<i>WT1</i>
<i>CSMD1</i>	<i>IKZF1</i>	<i>RUNX1</i>	<i>ZRSR2</i>
	<i>JAK2</i>	<i>SETBP1</i>	

Figure 1. Genes involved in inherited and acquired bone marrow failure syndromes. Genes causative of an inherited bone marrow failure syndrome (IBMFS), and genes associated with a risk of transformation to hematological malignancy or response to immunosuppression in patients with acquired aplastic anemia (aAA) and hypoplastic myelodysplastic syndrome (hMDS) were assessed in this study. Four genes, *CSF3R*, *GATA2*, *MPL* and *RUNX1*, were assessed in both germline and acquired settings. Genes targeted by whole exome sequencing (WES) only (i.e., not included on targeted panel) are denoted with an asterisk. Genes *KMT2D* and *PSTPIP1* are not represented in this figure; variants detected in these two genes were considered 'off-target' findings.

(PNH).¹⁵ No patients in this group had a clinical diagnosis of overt PNH. Over the limited follow-up available, 3 of 25 patients with acquired variants progressed with an aggressive hematological malignancy (acute myeloid leukemia or high-grade MDS) during the study period compared to 0 of 22 patients with no acquired variant detected ($P=0.2368$, Fisher's exact test).

Six of 47 (12.8%) patients in this group had a change of diagnostic categorization as a result of genomic characterization. Three of these six patients had a pre-test diagnosis of aAA and were re-categorized as having a myelodysplastic syndrome (transformed from underlying aAA) after genomic characterization. In addition to the genomic testing results themselves, the re-categorization of these patients was also the result of a re-assessment of the patient's disease that was prompted by the availability of genomic testing (including repeat of investigations such as morphological assessment of bone marrow biopsy). The other three patients had a change in pre-test diagnosis from aAA/hMDS to an IBMFS. These cases include a 64-year-old male (#47) who presented with pancytopenia and was found to have bone marrow hypoplasia and morphological dysplasia as well as an acquired pathogenic *CBL* variant. A pathogenic *SAMD9L* variant was identified during germline testing. Detailed clinical review revealed a family history of early onset ataxia in a number of family members and subsequently the patient was diagnosed with ataxia-pancytopenia syndrome – one presentation of the *SAMD9L* group of inherited disor-

ders¹⁶ (see the *Online Supplementary Appendix*). In addition, a 49-year-old male (#87) presented with pancytopenia and severe bone marrow hypoplasia and was diagnosed and treated for aAA. Genomic testing identified a germline variant in *TERT* (Leu408Pro). Identification of the *TERT* variant prompted telomere length testing and the patient was found to have telomere lengths <1st percentile. In addition, an acquired *TERT* promoter variant was detected in this patient adding further evidence to the pathogenicity of the germline variant. The patient had no other clinical features of DKC (see the *Online Supplementary Appendix*). Finally, a 27-year-old female (#39) was diagnosed with aAA after presenting with pancytopenia. She was commenced on eltrombopag but had no improvement in her blood counts. Genomic characterization revealed a homozygous *FANCA* variant (Ser994Gly) that resulted in exon skipping (demonstrated by RNA sequencing) with further testing demonstrating equivocal chromosomal fragility but failure of monoubiquitination in *FANCD2* consistent with a diagnosis of FA (see the *Online Supplementary Appendix*).

Genomic characterization clarifies diagnostic categorization in patients with clinically unclassifiable BMF

In patients with a pre-test diagnosis of UBMF, genomic characterization identified a causative germline or acquired variant in 25 of 45 (55.6%). In 24 of 25 patients this resulted in a change in the diagnostic categorization

Table 3. Variants detected in patients with a clinical diagnosis of undifferentiated bone marrow failure syndromes.

Patient ID	Age/ Sex	Pre-test diagnosis	Post-test diagnosis	Germline variants	Acquired variants	Comment
13 ^a	1F	UBMF	IBMFS (DBA)	<i>RPS19</i> c.184C>T; p.(Arg62Trp) [het]	–	Variant previously described in DBA ⁴¹
67 ^a	2F	UBMF	IBMFS (DBA)	<i>RPS19</i> c.184C>T; p.(Arg62Trp) [het]	–	Variant previously described in DBA ⁴¹
106 ^a	4F	UBMF	IBMFS (DBA)	<i>RPS7</i> c.75+1G>T; p.? [het]	–	Novel variant predicted to cause abnormal splicing
60 ^a	1M	UBMF	IBMFS (DKC)	<i>TERT</i> c.3148A>G; p.(Lys1050Glu) [het] <i>TERT</i> c.1670T>C; p.(Leu557Pro) [het]	–	Variant previously described in telomeropathy ² Novel variant. Impairment of TERT function demonstrated <i>in vitro</i> .
96 ^a	34M	UBMF	IBMFS (FA)	<i>FANCA</i> c.2852G>A; p.(Arg951Gln) [het] <i>FANCA</i> c.3971C>T; p.(Pro1324Leu) [het]	–	Variants previously described in FA ⁴³⁻⁴⁷
118 ^a	21F	UBMF	IBMFS (GATA2 syndrome)	<i>GATA2</i> c.351C>G; p.(Thr117=) [het]	<i>SETBP1</i> c.2602G>A; p.(Asp868Asn)	Variant previously described in GATA2 syndrome ⁴⁸
53 ^a	11M	UBMF	IBMFS (FA-like syndrome)	<i>FANCM</i> c.1972C>T; p.(Arg658*) [het] <i>FANCM</i> c.5101C>T; p.(Gln1701*) [het]	–	
20 ^a	5F	UBMF	IBMFS (SAMD9 syndrome)	<i>SAMD9</i> c.4057A>G; p.(Lys1353Glu) [het]	–	Confirmed <i>de novo</i>
62 ^a	2M	UBMF	IBMFS (SAMD9 syndrome)	<i>SAMD9</i> c.2318T>C; p.(Ile773Thr) [het]	–	Confirmed <i>de novo</i>
23 ^a	7M	UBMF	IBMFS (SAMD9 syndrome)	<i>SAMD9</i> c.3821A>G; p.(Tyr1274Cys) [het]	–	Confirmed <i>de novo</i>
51	2F	UBMF	IBMFS (SDS)	<i>SBDS</i> c.183_184delinsCT; p.(Lys62*) [het] <i>SBDS</i> c.258+2T>C; p.? [het]	–	Established pathogenic variants
69 ^a	1M	UBMF	IBMFS (SDS-like syndrome)	<i>DNAJC21</i> c.972_983+1414del [het]	–	Novel variant resulting in exon 7 skipping on RNA sequencing.
81 ^a	13M	UBMF	IBMFS (DDX41 syndrome)	<i>DDX41</i> c.435-2_435-1delinsCA; p.? [het]	–	Variant previously described in DDX41 syndrome ⁴¹
82 ^a	69F	UBMF	IBMFS (DDX41 syndrome)	<i>DDX41</i> c.517G>A; p.(Gly173Arg) [het]	<i>DDX41</i> c.1574G>A; p.(Arg525His) <i>DNMT3A</i> c.1903C>T; p.(Arg635Trp) <i>TET2</i> c.2366_2402del; p.(Asn789Metfs*12)	Variant previously described in DDX41 syndrome ¹⁸
103	2F	UBMF	IBMFS (Kabuki Syndrome)	<i>KMT2D</i> c.9218del; p.(Val3073Glufs*46) [het]	–	Novel variant. Cytopenias observed as a result of autoimmune destruction.
48	40F	UBMF	IBMFS (Autoimmune cytopenias associated with PSTPIP1 syndrome)	<i>PSTPIP1</i> c.748G>A; p.(Glu250Lys) [het]	–	Variant previously described in Hyperzincaemia/hypercalproteinaemia ⁴⁹
4 ^a	5M	UBMF	RCC		<i>RUNX1</i> c.496C>T; p.(Arg166*)	
79	53F	UBMF	Therapy-related myeloid neoplasm		<i>TP53</i> c.833C>T; p.(Pro278Leu) <i>TP53</i> c.818G>A; p.(Arg273His) <i>TP53</i> c.592del; p.(Glu198Lysfs*49) <i>TET2</i> c.5437C>T; p.(Gln1813*)	
80	81M	UBMF	hMDS		<i>DNMT3A</i> c.2634dup; p.(Asn879Glnfs*42) <i>TET2</i> c.4145A>G; p.(His1382Arg)	
94	73F	UBMF	hMDS		<i>TET2</i> c.3415del; p.(Ile1139Leufs*13) <i>TET2</i> c.4072T>G; p.(Cys1358Gly)	
52 ^a	41F	UBMF	MDS with isolated del(5q)		5q deletion (5q14.3-33.2)	
98	53F	UBMF	UBMF		<i>DNMT3A</i> c.2191T>A; p.(Phe731Ile)	
112	65M	UBMF	MDS		<i>BCORL1</i> c.2699del; p.(Asp900Alafs*25) <i>DNMT3A</i> c.2141C>G; p.(Ser714Cys)	
63	69M	UBMF	aAA		<i>BCOR</i> c.4848T>G; p.(Tyr1616*)	
88	68F	UBMF	Therapy-related myeloid neoplasm		<i>TET2</i> c.4075C>T; p.(Arg1359Cys)	

^aCase description provided in the *Online Supplementary Appendix*. aAA: acquired aplastic anemia; DBA: Diamond-Blackfan anemia; DKC: Dyskeratosis congenita; FA: Fanconi anemia; hMDS: hypoplastic myelodysplastic syndrome; MDS: myelodysplastic syndrome; SDS: Shwachman-Diamond syndrome; UBMF: undifferentiated bone marrow failure; het: heterozygous; RCC: Refractory cytopenia of childhood. Reference transcripts – *BCOR*, NM_017745.5; *BCORL1*, NM_021946.4; *DDX41*, NM_016222.2; *DNAJC21*, NM_194283.3; *DNMT3A*, NM_022552.4; *FANCA*, NM_000135.2; *FANCM*, NM_020937.2; *GATA2*, NM_032638.4; *KMT2D*, NM_003482.3; *PSTPIP1*, NM_003978.4; *RPS19*, NM_001022.3; *RPS7*, NM_001011.3; *RUNX1*, NM_001754.4; *SAMD9*, NM_017654.3; *SBDS*, NM_016038.2; *SETBP1*, NM_015559.2; *TERT*, NM_198253.2; *TET2*, NM_001127208.2; *TP53*, NM_000546.5.

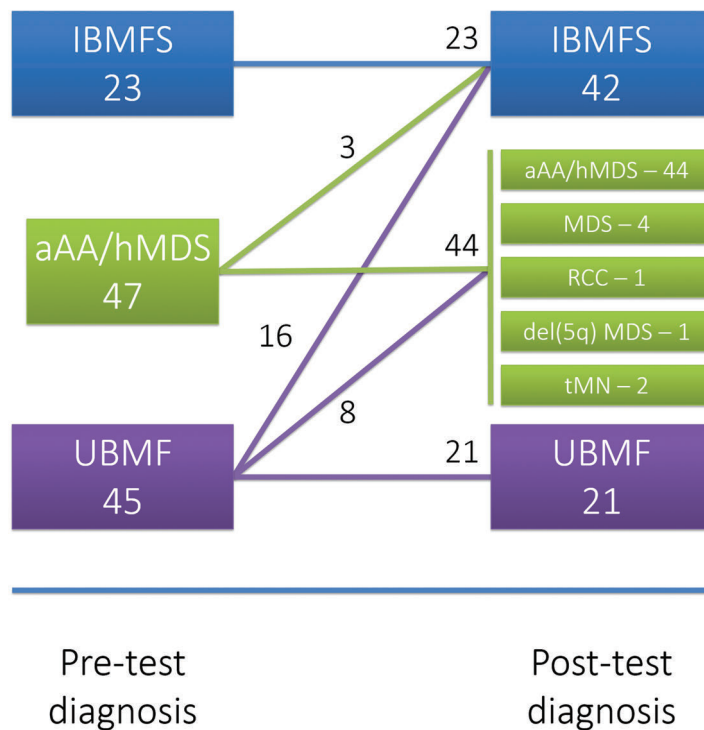


Figure 2. Pre- and post-test diagnosis classification of patients. Numbers of patients classified into each diagnostic group, (i) inherited bone marrow failure syndrome (IBMFS), (ii) acquired aplastic anemia (aAA) or hypoplastic MDS (hMDS) or (iii) clinically unclassifiable bone marrow failure (UBMF) as evaluated prior to genomic testing (pre-test diagnosis) and again following genomic testing (post-test diagnosis) with transitions indicated by the connecting lines (in addition three patients changed diagnosis within the aAA/hMDS category from aAA to MDS). RCC: refractory cytopenia of childhood; tMN: therapy related myeloid neoplasm.

from a pre-test diagnosis of UBMF to a specific post-test diagnosis. Therefore genomic characterization resulted in diagnostic categorization in 24 of 45 (53.3%) of patients in this diverse group of patients. Variants detected in this cohort are listed in Table 3. Causative germline variants were detected in 16 patients and acquired variants were detected in nine patients. The diverse inherited and acquired diagnoses included FA, SDS, DKC, DBA, aAA, hMDS, therapy-related myeloid neoplasm (tMN) and MDS with isolated del(5q).

Three patients in this group received a post-test diagnosis of hMDS or MDS after genomic characterization. This was the result of re-interpretation of the clinical and pathological features in light of supportive molecular evidence for these diagnoses. Notably, one patient retained a post-test diagnosis of UBMF despite having a *DNMT3A* mutation detected due to inadequate pathological and clinical features to support a diagnosis of aAA, hMDS, MDS or tMN.

Five patients in this group presented with hypoplastic BMF and a monosomy 7 detectable by either conventional karyotyping, fluorescence *in situ* hybridization or copy number analysis by targeted sequencing. Three of these (#20, #23, #62) had *SAMD9* variants that were all confirmed to be *de novo* through testing of parental samples. Multiple somatic reversion events were noted in these patients including acquired sequence variants as well as progressive resolution of monosomy 7 over time (see the *Online Supplementary Appendix*). One patient (#118) with monosomy 7 had a synonymous *GATA2* variant (Thr117=) which has previously been associated with *GATA2* haploinsufficiency syndrome.¹⁷

Two patients in this group had pathogenic *DDX41* variants identified. One of these patients was a 69-year-old female (#82) who presented with severe neutropenia and was found to have a severely hypocellular bone marrow. Genomic characterization revealed a *DDX41* missense variant (Gly173Arg) which has been described previously in

five patients presenting with a similar clinical phenotype.¹⁸ In addition, targeted panel sequencing in this patient identified a hotspot somatic *DDX41* Arg525His variant present at approximately 3% VAF.

Finally, this group included a 5-year-old male (#4) with a pre-test diagnosis of UBMF with onset just after birth. Targeted panel sequencing identified a *RUNX1* nonsense variant (Arg166*) at low VAF (4.8%). This variant was quantitatively monitored over time using ddPCR and was noted to be acquired, steadily increasing and reaching 11.8% over a 12-month period. After multidisciplinary review it was decided to proceed to an allogeneic bone marrow transplant in this patient.

Contribution to diagnostic categorization from targeted panel sequencing versus WES

All germline variants in genes common to the targeted panel and whole exome were detected. In eight patients, variants were detected in the whole exome in genes not covered in the targeted panel. These included variants in *SAMD9*, *SAMD9L*, *DNAJC21*, *SRP54*, *KMT2D* and *PSTPIP1*. These genes can be divided into two groups: (i) genes where the primary phenotype of pathogenic variants is an IBMFS (*SAMD9*, *SAMD9L*, *DNAJC21* and *SRP54*) and (ii) genes where BMF is either an infrequent or not the primary clinical manifestation (*KMT2D* and *PSTPIP1*). Mutations in genes in this latter category were detected through an extended analysis of the WES data given the absence of other causative mutations found in these two patients and the remaining strong suspicion of a genetic etiology.

In total, 65 acquired variants were detected across the entire cohort. All of these variants were detected by targeted panel sequencing with a bioinformatic pipeline optimized for acquired variant detection. Only 16 of 65 (24.6%) of these variants were detected by WES using a standard germline bioinformatics pipeline. Of the 49

acquired variants not called by the WES pipeline, 24 were visible after manual inspection of aligned reads in whole exome data based on variant detection in the targeted panel data. The mean depth of targeted panel sequencing was 795x (range: 576-1,025) with 98.9% of bases sequenced to $\geq 100x$ coverage, compared to a mean depth of 198x (range: 117-260) and 77.5% $\geq 100x$ coverage for WES. The VAF of the variants that were not detected by WES (and associated pipeline) were all less than 15%.

Discussion

We have described a cohort of 115 patients presenting with a diverse range of hypocellular BMF that have undergone prospective comprehensive genomic evaluation for germline and acquired causes. In our cohort we were able to identify pathogenic germline or acquired variants in 54% (62 of 115) of the entire cohort including in 52% (12 of 23) of patients with a clinical diagnosis of IBMFS, 53% (25 of 47) of patients with a clinical diagnosis of aAA/hMDS and 56% (25 of 45) of patients with clinically unclassifiable BMF. Overall, genomic characterization resulted in a change in diagnostic categorization in 30 of 115 (26.1%) patients. The observed clinical impacts in our patient cohort after comprehensive genomic characterization included the choice to perform allogeneic stem cell transplantation, disease-specific treatments (such as corticosteroid for DBA and lenalidomide for MDS with del(5q)), identification of at-risk family members, enhanced screening of patients for secondary malignancy and influence of sibling allogeneic stem cell donor choice. We have also described multiple novel genomic findings including novel pathogenic variants in *TERT* (Leu408Pro and Leu557Pro), *RPS7* (c.75+1G>T) and *SAMD9* (Lys1353Glu, Tyr1274Cys and Ile773Thr) and have confirmed recent novel findings of other groups including (i) the association of biallelic *FANCM* variants with chemotherapy toxicity in the absence of an overt FA phenotype,¹⁹ (ii) the association of *DDX41* Gly173Arg with a hypocellular MDS phenotype¹⁸ and (iii) the presence of acquired reversion variants in *SAMD9/SAMD9L*.²⁰

The diverse spectrum of inherited and acquired diseases identified in our cohort highlights the diagnostic utility of comprehensive genomic characterization for patients with a clinical presentation of BMF. Despite the range of underlying diagnoses that may manifest as hypoplastic BMF there may be minimal or no specific features on conventional (non-genomic) diagnostic testing to indicate the specific underlying diagnosis. In addition, clinical features such as abnormal digits or oral leukoplakia cannot be relied upon to suggest an underlying genetic cause as they may be absent or unrecognized due to lack of physician familiarity with these rare diseases. Moreover, as more literature and larger cohorts emerge with genetic annotation, the clinical phenotypes associated with specific genes are expanding. For example, the expansion of the phenotype of pathogenic variants in *DDX41* to include a hypocellular MDS-like phenotype in the case of Gly173Arg¹⁸ rather than the initial description of presentation in later life with acute myeloid leukemia after minimal antecedent overt hematological abnormality.²¹ Indeed, in our cohort we observed a pathogenic *DDX41* variant in a 13-year-old male (#81) with isolated severe stable

thrombocytopenia and no evidence of hematological malignancy.

A significant proportion of patients (47.8%) with a clinical diagnosis of IBMFS did not have causative variants detected. The vast majority (10 of 11) of these were in patients with SCN. This may be explained by the significant baseline rate of “variant-negative” cases in this condition described in the literature to date²² as well as the potential for other causes of neutropenia. Given these patients had no pathogenic variants detectable despite comprehensive genomic analysis (including WES), one possible future strategy for identifying causative variants may be to evaluate the non-coding regions of existing pathogenic genes (e.g., *ELANE*, *HAX1*) for deep intronic variants resulting in splicing abnormalities that are missed by methods focused on coding regions.

In the acquired BMF setting, we noted six patients who were re-categorized as either hMDS, MDS, tMN or refractory cytopenia of childhood (RCC) after genomic characterization. Whilst there are no absolutely specific molecular abnormalities that distinguish these disorders from aAA complicated by clonal hematopoiesis or age-related clonal hematopoiesis, the presence of the genomic abnormalities detected were sufficient for the treating team to re-evaluate the clinical and morphological features of these six cases and assign post-diagnoses of hMDS/MDS/RCC/tMN. This reflects the way in which this type of genomic data is increasingly integrated into diagnostic decision making in a real-world context, especially in areas where morphological interpretation is challenging and inter-observer agreement is poor.^{23,24} Another practical aspect regarding diagnostic workup is the limited availability of specific non-genomic tests (e.g., telomere length analysis) in the accredited setting which may have contributed to a variable rate of performance of these tests in the cohort.

In our cohort 39% (45 of 115) of patients were categorized as clinically UBMF before genomic testing. The relatively large number of patients in this category may be the result of numerous factors including a referral bias for patients to this study with atypical clinicopathological features in order to access novel genomic technologies. In addition, patients could also be included at an early time-point in their disease course and it is possible that repeated history and physical examination over time in patients (including assessing response to therapy) would have enabled more definitive categorization before genomic testing. Moreover, there is an increasing recognition of the possibility of occult IBMFS in patients diagnosed as acquired BMF (e.g., aAA), indeed in our cohort three patients harbored occult IBMFS. This recognition, in turn, may lead to a desire by treating clinical teams to more comprehensively exclude IBMFS rather than relying on a clinical diagnosis of a purely acquired BMFS - hence their inclusion in a UBMF category. Reliable clinical and laboratory features to identify those with a low likelihood of IBMFS (e.g., the presence of a PNH clone, severity/time course/persistence of neutropenia in SCN) would be of significant value in resource-limited settings in order to avoid expensive and time consuming genetic testing in this potentially expanding subgroup. The approach to genomic characterization in our cohort included both targeted BMF panel sequencing and WES for all patients. Our choice of targeted panel sequencing in addition to WES was based on the requirement for enhanced sensitivity

due to the clinical context of BMF mandating assessment for both germline and acquired genomic lesions. In addition to the detection of acquired variants in aAA/hMDS (which may be present at VAF less than 5%), clinically relevant reversion variants in inherited conditions such as *SAMD9/SAMD9L* can also exist which require sensitive testing methodologies. As expected (given the greater depth of sequencing able to be cost-effectively achieved), targeted panel sequencing was clearly superior at detecting low-level acquired variants when compared to WES. Using a bioinformatics pipeline optimized for acquired variant detection (GATK4/Mutect2) we were able to detect acquired variants down to a VAF of <0.5%. Despite their very low level, detection of these variants can be highly instructive for diagnostic categorization by demonstration of clonality (to distinguish from reactive conditions) as well as providing a baseline genomic profile for future monitoring of clonal evolution. Conversely, one potential benefit of WES for investigation of germline variants is in the context of phenotypic uncertainty in order to identify pathogenic variants across a broad gene list. In our cohort, two patients had pathogenic variants detected in genes which are not typically associated with BMF (*KMT2D* and *PSTPIP1*). The other genes originally not present in the targeted panel due to their recent description (*SAMD9*, *SAMD9L*, *DNAJC21*) have been added to a future iteration of the targeted panel. However the need for panel re-design and subsequent costly laboratory validation highlights a recognized drawback of targeted panel sequencing. Nevertheless, given the relative specificity of the clinical presentation and the requirement for sensitive detection of acquired variants, targeted panel sequencing is a methodological choice highly suited for this clinical context, compared to which, WES offers minimal additional clinical utility. Copy number analysis was possible using either of these sequencing strategies and facilitated the identification of pathogenic copy number losses (*RPL35A*, *RPS19*) as well as clinically relevant acquired monosomies. RNA sequencing was valuable for determining the effect of putative splice variants on mRNA transcripts (*FANCA*, *DNAJC21*).

In conclusion, we have described the impact of genomic characterization on diagnostic categorization in a cohort of patients with hypoplastic BMF. In addition to multiple

novel genomic findings, this personalized genomic approach is of significant utility to rapidly providing accurate diagnostic categorization to inform clinical decision-making in this diverse and challenging group of diseases. The incorporation of a multidisciplinary review further strengthens the impact and clinical value of this approach.

Disclosures

No conflicts of interest to disclose.

Contributions

PB designed the research study, analyzed data, curated and interpreted genetic variation for clinical reports, recruited and managed patients, participated in multidisciplinary meetings and wrote the paper. LF analyzed data, curated and interpreted genetic variation for clinical reports, recruited and managed patients, participated in multidisciplinary meetings and wrote the paper. GR analyzed data, curated and interpreted genetic variation for clinical reports, performed experiments, participated in multidisciplinary meetings and wrote the paper. ET, JL and MM analyzed data, curated and interpreted genetic variation for clinical reports, participated in multidisciplinary meetings and wrote the paper. DH, AG, FM, GL, JS, PBar, JR and JW recruited and managed patients, participated in multidisciplinary meetings and wrote the paper. EW, EL and MM participated in multidisciplinary meetings and wrote the paper. CG designed the research study and wrote the paper. DR designed the research study, recruited and managed patients, participated in multidisciplinary meetings and wrote the paper.

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