

Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Gaw, S;Harford, A;Pettigrove, V;Sevicke-Jones, G;Manning, T;Ataria, J;Cresswell, T;Dafforn, KA;Leusch, FDL;Moggridge, B;Cameron, M;Chapman, J;Coates, G;Colville, A;Death, C;Hageman, K;Hassell, K;Hoak, M;Gadd, J;Jolley, DF;Karami, A;Kotzakoulakis, K;Lim, R;McRae, N;Metzeling, L;Mooney, T;Myers, J;Pearson, A;Saaristo, M;Sharley, D;Stuthe, J;Sutherland, O;Thomas, O;Tremblay, L;Wood, W;Boxall, ABA;Rudd, MA;Brooks, BW

Title:

Towards Sustainable Environmental Quality: Priority Research Questions for the Australasian Region of Oceania

Date:

2019-11-01

Citation:

Gaw, S., Harford, A., Pettigrove, V., Sevicke-Jones, G., Manning, T., Ataria, J., Cresswell, T., Dafforn, K. A., Leusch, F. D. L., Moggridge, B., Cameron, M., Chapman, J., Coates, G., Colville, A., Death, C., Hageman, K., Hassell, K., Hoak, M., Gadd, J., ... Brooks, B. W. (2019). Towards Sustainable Environmental Quality: Priority Research Questions for the Australasian Region of Oceania. *Integrated Environmental Assessment and Management*, 15 (6), pp.917-935. <https://doi.org/10.1002/ieam.4180>.

Persistent Link:

<https://hdl.handle.net/11343/249954>

License:

CC BY

Workshop Synthesis

Towards Sustainable Environmental Quality: Priority Research Questions for the Australasian Region of Oceania

Sally Gaw,*[†] Andrew Harford,[‡] Vincent Pettigrove,[§] Graham Sevicke-Jones,^{||} Therese Manning,[#]
James Ataria,^{††} Tom Cresswell,^{‡‡} Katherine A Dafforn,^{§§} Frederic DL Leusch,^{||||} Bradley Moggridge,^{###}
Marcus Cameron,^{†††} John Chapman,^{‡‡‡} Gary Coates,^{§§§} Anne Colville,^{|||||} Claire Death,^{####}
Kimberly Hageman,^{††††} Kathryn Hassell,[§] Molly Hoak,^{‡‡‡‡} Jennifer Gadd,^{§§§§} Dianne F Jolley,^{|||||||}
Ali Karami,^{#####} Konstantinos Kotzakoulakis,^{‡‡‡‡‡} Richard Lim,^{|||||||} Nicole McRae,[†] Leon Metzeling,^{†††††}
Thomas Mooney,[‡] Jackie Myers,[§] Andrew Pearson,^{§§§§§} Minna Saaristo,^{|||||||} Dave Sharley,^{#####}
Julia Stuthe,^{††††††} Oliver Sutherland,^{‡‡‡‡‡‡} Oliver Thomas,^{§§§§§§} Louis Tremblay,^{††} ^{|||||||}
Waitangi Wood,^{#####} Alistair BA Boxall,^{†††††††} Murray A Rudd,^{‡‡‡‡‡‡} and Bryan W Brooks^{§§§§§§§§}

[†]School of Physical and Chemical Sciences, University of Canterbury, Christchurch, New Zealand

[‡]Department of the Environment and Energy, Australian Government, Darwin, Australia

[§]Aquatic Environmental Stress Research Centre, RMIT University, Bundoora, Victoria, Australia

^{||}Environment Southland Regional Council, Invercargill, New Zealand

[#]Environmental Risk Sciences, Sydney, Australia

^{††}Cawthron Institute, Nelson, New Zealand

^{‡‡}Australia's Nuclear Science and Technology Organisation, Lucas Heights, Australia

^{§§}Department of Environmental Sciences, Macquarie University, North Ryde, Australia

^{||||}Australian Rivers Institute and School of Environment and Science, Griffith University, Brisbane, Australia

^{###}Institute for Applied Ecology, University of Canberra, Canberra, Australia

^{†††}Auckland Council, Auckland, New Zealand

^{‡‡‡}Office of Environment and Heritage, New South Wales, Australia

^{§§§}Te Rūnanga o Ngāi Tahu, Christchurch, New Zealand

^{|||||}School of Life Sciences, University of Technology Sydney, Sydney, Australia

^{####}Faculty of Veterinary Science, University of Melbourne, Victoria, Australia

^{††††}Department of Chemistry and Biochemistry, Utah State University, Logan, Utah, USA

^{‡‡‡‡}School of Biosciences, The University of Melbourne, Parkville, Victoria, Australia

^{§§§§}National Institute of Atmospheric and Water Research, Auckland, New Zealand

^{|||||||}Faculty of Science, University of Technology Sydney, Sydney, Australia

^{#####}Environmental Futures Research Institute, Griffith University, Brisbane, Australia

^{†††††}Environment Protection Authority, Victoria, Australia

^{‡‡‡‡‡}Macquarie University, Sydney, Australia

^{§§§§§}Ministry for Primary Industries, Wellington, New Zealand

^{|||||||}School of Biological Sciences, Monash University, Melbourne, Australia

^{#####}Bio2Lab, Melbourne Innovation Centre, Greensborough, Australia

^{††††††}CSIRO Publishing, Clayton, Victoria, Australia

^{‡‡‡‡‡‡}Independent researcher, Nelson, New Zealand

^{§§§§§§}School of Applied Chemistry and Environmental Science, RMIT University, Melbourne, Victoria, Australia

^{|||||||}School of Biological Sciences, University of Auckland, Auckland, New Zealand

^{#####}Tau Iho I Te Po Trust, Kaeo, New Zealand

^{†††††††}Environment Department, University of York, York, United Kingdom

^{‡‡‡‡‡‡‡}World Maritime University, Malmö, Sweden

^{§§§§§§§}Baylor University, Waco, Texas, USA



ABSTRACT

Environmental challenges persist across the world, including the Australasian region of Oceania, where biodiversity hotspots and unique ecosystems such as the Great Barrier Reef are common. These systems are routinely affected by multiple stressors from anthropogenic activities, and increasingly influenced by global megatrends (e.g., the food–energy–water nexus, demographic transitions to cities) and climate change. Here we report priority research questions from the Global Horizon Scanning Project, which aimed to identify, prioritize, and advance environmental quality research needs from an Australasian perspective,

This article contains online-only Supplemental Data.

* Address correspondence to sally.gaw@canterbury.ac.nz

Published 5 July 2019 on wileyonlinelibrary.com/journal/ieam.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

within a global context. We employed a transparent and inclusive process of soliciting key questions from Australasian members of the Society of Environmental Toxicology and Chemistry. Following submission of 78 questions, 20 priority research questions were identified during an expert workshop in Nelson, New Zealand. These research questions covered a range of issues of global relevance, including research needed to more closely integrate ecotoxicology and ecology for the protection of ecosystems, increase flexibility for prioritizing chemical substances currently in commerce, understand the impacts of complex mixtures and multiple stressors, and define environmental quality and ecosystem integrity of temporary waters. Some questions have specific relevance to Australasia, particularly the uncertainties associated with using toxicity data from exotic species to protect unique indigenous species. Several related priority questions deal with the theme of how widely international ecotoxicological data and databases can be applied to regional ecosystems. Other timely questions, which focus on improving predictive chemistry and toxicology tools and techniques, will be important to answer several of the priority questions identified here. Another important question raised was how to protect local cultural and social values and maintain indigenous engagement during problem formulation and identification of ecosystem protection goals. Addressing these questions will be challenging, but doing so promises to advance environmental sustainability in Oceania and globally. *Integr Environ Assess Manag* 2019;15:917–935. © 2019 The Authors. *Integrated Environmental Assessment and Management* published by Wiley Periodicals, Inc. on behalf of Society of Environmental Toxicology & Chemistry (SETAC)

Keywords: Multiple stressors and mixtures Risk assessment Contaminants of emerging concern Indigenous knowledge Cultural values

INTRODUCTION

Achieving sustainable environmental quality and ecosystem integrity is a critical goal shared by diverse stakeholders around the world. Unimpaired and diverse ecosystems conserve biodiversity and provide essential

ecosystem services, while being more resilient when natural and anthropogenic disasters occur (Alexander et al. 2016). The United Nations Sustainable Development Goals aim to protect the planet and realize prosperity for all people, including future generations (UN 2015). Within

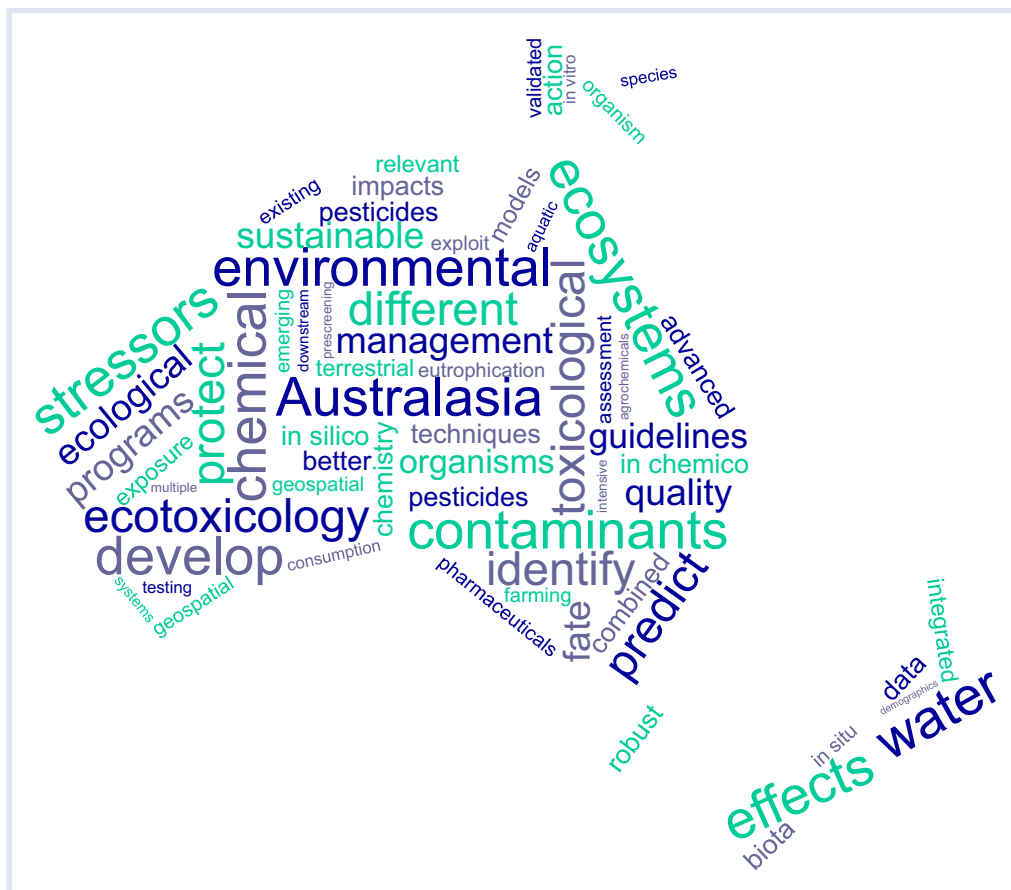


Figure 1. Word cloud of priority research questions from the Australasian Region of Oceania.

this framework are interconnected goals that inherently rely on achieving more sustainable environmental quality and ecosystem integrity. But achieving these goals depends on effective environmental management efforts informed by the best available scientific knowledge and technological advancements. Integration of robust environmental risk assessment with ecosystem protection goals is therefore critical in light of global megatrends (e.g., the food–energy–water nexus, demographic transitions to cities) and climate change that present unique challenges for policy makers and environmental and health professionals. These sustainable management challenges are complex, particularly given environmental, political, and economic contexts that exist among and within global regions.

Intersections of biodiversity, environmental variability, and anthropogenic stressors are pronounced in the Australasian region of Oceania. Countries in the region have iconic landscapes with unique flora and fauna. The ability to participate in outdoor activities, including hiking, camping, fishing, and swimming, is treasured in Australia and New Zealand and considered to be part of their national identities (Garner 2013; McCrone 2017). Biodiversity hotspots are prevalent, as are freshwater and marine ecosystems (e.g., the Great Barrier Reef), which are susceptible to stress from anthropogenic activities, including climate change (Adams et al. 2016). Interconnections among stressors from landscape development and urbanization across freshwater to marine gradients are widespread in this region (e.g., Mayer-Pinto et al. 2015; Weeks et al. 2016), where the vast majority of human populations reside within 50 km from the coast.

Climate change is significantly affecting the island nations of Oceania (Caritas 2018) and magnifying the importance of understanding how multiple physical and chemical stressors impact biodiversity and ecosystem services (Weeks et al. 2016). However, information on the influences of natural and anthropogenic stressors, particularly chemical contaminants, is scarce for native species. There are also relatively few ecotoxicology data sets of relevance to the tropical conditions of Papua New Guinea and much of the northern parts of Australia. Degradation of water quality is of particular concern for Mori and Aboriginal communities. There is growing appreciation of the spiritual and cultural values and developmental aspirations of indigenous communities; momentum is building to incorporate these in environmental policies and decision making (Bark et al. 2015; Harmsworth et al. 2016; Ataria et al. 2018). Unfortunately, identifying global priority environmental quality research needs to attain these ecosystem protection goals and effectively implement policy instruments has remained elusive on a regional scale. Horizon-scanning approaches for identifying key research questions may be part of developing sustainable solutions.

Horizon scanning using a key questions approach has emerged from the conservation sciences, public health, and other disciplines as an effective means to identify important research needs through engagement of diverse

stakeholders (Sutherland and Woodroof 2009; Boxall et al. 2012; Rudd et al. 2018). The Global Horizon Scanning Project (GHSP) was initiated with the Society of Environmental Toxicology and Chemistry (SETAC) to identify priority research questions that advance understanding of how environmental stressors impact environmental quality (Brooks et al. 2013). This initiative is collecting and prioritizing the most important current and emerging research questions related to environmental quality as recognized by scientists and engineers from multiple disciplines working in government, academia, and business around the globe. For example, priority research questions were recently reported from Latin America (Furley et al. 2018), Europe (Van den Brink et al. 2018), and North America (Fairbrother et al. 2019). Here we specifically present results from the GHSP project focused on the Australasian region of Oceania. The scope of these questions was intended to be of relevance to Australasia, within a global context. We anticipate these priority research questions will be indispensable in informing and structuring research agendas by the government and business communities in the future.

METHODS

In the present study, we followed previously reported methods (Boxall et al. 2012; Furley et al. 2018; Van den Brink et al. 2018) to identify priority research questions. Prior to holding a workshop in Nelson, New Zealand, in 2015, members of SETAC and other scientists from Oceania were asked to submit research questions which, in their view, were priority environmental quality research needs to address. Consistent with methods employed in other studies (Sutherland et al. 2011) and SETAC geographic regions (Furley et al. 2018; Van den Brink et al. 2018), participants were provided criteria for an ideal question, which should address important gaps in knowledge; be answerable through a realistic research design; have a factual answer that does not depend on value judgments; cover a spatial and temporal scale that could realistically be addressed by a research team; not be answerable by “it all depends,” “yes,” or “no”; and if related to impact and interventions, the research question should contain a subject, an intervention, and a measurable outcome. In total, 78 questions were received and are presented in Supplemental Data.

Before the workshop, questions were partitioned among 6 themes, including contaminants of emerging concern; environmental chemistry: analysis, fate, and exposure; multiple stressors and mixtures; risk assessment, regulations, and guidelines; spotlight on Australasia; and tools for improving risk assessment. These 6 themes were used to structure an expert workshop held in Nelson, New Zealand as part of the SETAC Australasia meeting in 2015 at which the questions were discussed. During the workshop, 20 priority research questions were identified by participants from academic, business, the indigenous community, and government sectors. We specifically examine each of these priority research questions in the sections that follow (Table 1).

Table 1. Top 20 priority research questions from the Australasian portion of the Global Horizon Scanning Project by theme

Themes and priority research questions
Contaminants of emerging concern
What are the most appropriate toxicological approaches to develop regulatory guidelines specifically for contaminants of emerging concern that address multimodes of action and sublethal effects?
How can we identify and prioritize contaminants (traditional and emerging stressors) for sustainable management of ecosystems within different biogeographic regions?
How can we identify and examine the environmental fate and toxicity of ingredients other than the stated “active” components in commercial formulations, individually and in chemical mixtures?
Environmental chemistry: Analysis, fate, and exposure
How can we develop robust chemical assays and models to replace, refine, and reduce biological testing?
How do we better understand the linkages between the structural and physicochemical properties of substances to predictively model fate and bioavailability in different environments?
How do we develop better broad-screening analytical and information-processing techniques that do not require preselection of target contaminants?
How do we use chemistry to better design sustainable waste management?
How can we ensure sustainable supplies of clean water, energy development, and food security while simultaneously minimizing ecological impacts and protecting environmental quality?
Multiple stressors and mixtures
What are the combined impacts of various agrochemicals (e.g., veterinary medicines, pesticides) and eutrophication from intensive terrestrial farming operations on the health of aquatic and terrestrial organisms?
What are the effects of changing demographics, economic development, consumption patterns, and climate (e.g., ocean acidity, water temperature) on chemical emissions, environmental fate, and ecotoxicology of contaminants and multiple stressors?
What are the combined effects of very low levels of multiple contaminants (e.g., pesticides, natural resource extraction contaminants, salinity, pharmaceuticals and personal care products, endocrine-disrupting chemicals) with different modes of action on aquatic and terrestrial organisms and ecosystems?
Risk assessment, regulations and guidelines
What water quality guidelines are needed to protect temporary waters and associated ecosystems from the influences of development?
What are the effects of short magnitude, frequency, and duration (e.g., intermittent, episodic) exposures to contaminants and other stressors, and how can these scenarios be effectively incorporated into water quality guidelines?
How can we measure ecosystem resilience to and recovery following exposure to stressors?
Spotlight on Australasia
Are there differences in toxicological thresholds among native and nonnative organisms, and how can species sensitivity information from nonresident species be used to predict adverse outcomes and protect our unique biota and ecosystems?
How do we incorporate and protect cultural and social values (relating to humans, biota, and ecosystems) to empower citizen, societal, and indigenous engagement in the research, management, and legislation of priority environmental contaminants?
Tools for improving risk assessment
How do we exploit, collate, and integrate existing environmental toxicology, chemistry, and geospatial data to help develop robust risk assessment?
How can prescreening techniques (e.g., in silico, in vitro) be developed, advanced, and validated to identify and predict whole organism effects?
How can ecotoxicology information be integrated more closely during interpretation of ecological data?
How do we advance ecotoxicology testing to be more relevant to ecological systems?

CONTAMINANTS OF EMERGING CONCERN

What are the most appropriate toxicological approaches to develop regulatory guidelines specifically for contaminants of emerging concern (CECs) that address multimodes of action and sublethal effects?

Measures of effect are selected during problem formulation in ecological risk assessment to support assessment endpoints that are aligned with ecosystem protection goals (Suter 2006). Historically, these measures of effect include a limited number of model organisms and endpoints (survival, growth, reproduction) linked to adverse outcomes of importance to the population level and environmental management. Single-species ecotoxicity information for a specific chemical is then routinely utilized to develop species sensitivity distributions from which water quality criteria, standards, or guidelines are derived around the world (Posthuma et al. 2001). Recent revisions of Australia and New Zealand Environment and Conservation Council (ANZECC, now referred to as the Australian and New Zealand Governments [ANZG] 2018) guidelines, and the use of multiple lines of evidence in weight-of-evidence assessments, represent global steps forward consistent with global trends.

Although recent years have seen an increase in the use of chronic toxicity testing with Australasian species, a historical overreliance on a limited number of model organisms and endpoints has potentially undermined management activities related to sustainable environmental quality and ecosystem integrity. Much of the available ecotoxicology information has been primarily comprised of acute lethality responses of several species (e.g., *Daphnia* sp.) from the Northern Hemisphere. Sublethal responses to chemical stressors were primarily available for cladoceran reproduction, microalgal growth rate, and juvenile fish growth. Similar model organisms and endpoints also have been employed for whole effluent (aka “direct toxicity assessments”) and ambient toxicity testing (USEPA 1991). However, assays based on these model organisms and endpoints were often not developed to account for mutagenicity, teratogenicity, and other adverse outcomes that result from diverse molecular initiation events (MIEs; Ankley et al. 2010). Other ecologically important endpoints, including developmental and behavioral responses, are increasingly receiving attention in terms of potential importance (e.g., Saaristo et al. 2018).

Early research with endocrine-disrupting and -modulating chemicals (EDCs) recognized some of the limitations of these traditional tools to assess environmental quality and to derive guideline values protective of aquatic systems. For example, a 6-order-of-magnitude difference exists between adverse effects on cladoceran (Clubbs and Brooks 2007) versus fish reproduction (Kidd et al. 2007) elicited by the human estrogen agonist 17 α -ethinylestradiol because invertebrates do not possess a functional estrogen receptor (Ankley et al. 2016). After almost a decade of health and ecological research on EDCs, Ankley et al. (2007) identified that such lessons learned from these chemicals were

important to understanding risks of pharmaceuticals in the environment. Subsequently, efforts such as the development of adverse outcome pathways (AOPs; Ankley et al. 2010), informed by comparative pharmacology and toxicology research (LaLone et al. 2016; Brooks 2018), have been advancing the use of pathway-based predictive approaches in ecological risk assessment.

In parallel, buoyed by release of Toxicity Testing in the 21st Century (NRC 2007), the Tox21 and ToxCast programs were launched (Dix et al. 2006). These have screened thousands of chemicals with hundreds of in vitro assays, largely adapted from drug discovery and safety testing programs, to identify likely MIEs associated with many untested chemicals. These and related next-generation risk assessment efforts are breaking new ground (Cote et al. 2016). For example, identification of diverse MIEs associated with chemical properties is supporting development of next-generation computational toxicology models to identify problematic (and useful) substances and to sustainably design less hazardous chemicals. More recent applications include employing these in vitro systems for prioritizing environmental assessments (Li et al. 2017) and performing cross-species extrapolation (LaLone et al. 2018), or tracking movements of multiple individuals simultaneously using ToxTrac (Rodriguez et al. 2018). Such efforts promise to continue to further advance environmental risk assessment practices (Milleneuve et al. 2019).

Integrating comparative toxicology information and mechanistic tools such as high-throughput assays with regulatory guideline development and environmental monitoring and assessment represents important research needs. In the case of pharmaceuticals, for example, short-term standardized ecotoxicity test model species and endpoints are often not adequate to define chronic toxicity (Brooks 2018). Herein, therapeutic hazard values (Brooks 2014) and minimal selective concentrations and associated predicted no-effect concentrations for the development of antibiotic resistance by microorganisms in particular (Bengtsson-Palme and Larsson 2016) represent recent approaches to identify water concentrations supporting more robust ecological and human health water quality assessments, respectively, and to further support environmental diagnostic applications. However, integrative, comparative, and predictive toxicology research must be advanced to understand ecologically important effects caused by new and poorly studied chemicals.

How can we identify and prioritize contaminants (traditional and emerging stressors) for sustainable management of ecosystems within different biogeographic regions?

Like other regions of the globe, Australasian ecosystems are subject to a variety of chemical and other stressors, which challenges stressor identification research and practice. However, due to the smaller economies of the Australasian region, it is especially not feasible to have ever-expanding monitoring lists for contaminants, and care is needed to avoid needless selection of priority contaminants based on data from different biogeographic regions of Australasia. Risk-based frameworks for identification and

prioritization of contaminants that incorporate local ecosystem-specific vulnerability to contaminants and Australasia-specific use of chemicals are urgently needed. Factors contributing to Australasia-specific use of chemicals include regulatory decisions, patents, demographics, land use, and climate, along with human and animal disease and pest profiles (Daughton 2014; Kookana et al. 2014; Gaw and Brooks 2016). These factors will change over time, and prioritization schemes and ultimately regulatory and monitoring regimes will need to be sufficiently agile and adaptive to examine substances currently in commercial use. Solid waste and wastewater management practices in Australasia will also determine priority substances in the region. In addition to anthropogenic chemical contaminants, transformation products and endogenous biomolecules, including toxins from harmful algal blooms (HABs), need to be assessed. Other important stressors that also need to be taken into consideration include changing land use, urbanization, climate change, and biological stressors such as predation, overexploitation, and invasive species. Globally, the need for contaminant prioritization has been identified for pharmaceuticals and personal care products (Boxall et al. 2012), microplastics (Eerkes-Medrano et al. 2015), and pesticides and their transformation products (Sinclair et al. 2006). Ultimately, risk-based identification and prioritization frameworks for contaminants, which are currently used by Australasian chemical management authorities, need to be diligently updated to reflect contemporary uses and potential exposure. They also need to be further developed to be broader than single classes of contaminants and to incorporate nonchemical stressors.

How can we identify and examine the environmental fate and toxicity of ingredients other than the stated “active” components in commercial formulations, individually and in chemical mixtures?

Ecotoxicity testing is generally focused on known active components as pure substances rather than as components of commercial formulations and chemical mixtures. Many products contain ingredients other than the stated active components to enhance the stability or performance of the product. Examples include adjuvants added to pesticides, coloring agents and preservatives added to soaps, fragrances added to cleaning products, and a wide range of excipients added to pharmaceutical products. These “other” or “inert” ingredients have the potential to alter the environmental fate and toxicity of the active components in commercial formulations as well as in other contaminants and may also present their own inherent hazards and risks (Cox and Sorgan 2006). For example, glyphosate formulations containing surfactants were more toxic than glyphosate on its own (Vincent and Davidson 2015). Such ingredients may not be listed, especially for proprietary formulations, making it difficult to identify and prioritize components of formulations for study. Identification of potentially problematic ingredients in products other than the active ingredient will lead to improved risk assessment and ultimately to safer products.

ENVIRONMENTAL CHEMISTRY: ANALYSIS, FATE, AND EXPOSURE

How can we develop robust chemical assays and models to replace, refine, and reduce biological testing?

Globally there is a focus on reducing biological testing to reduce the numbers of animals used in testing and to minimize the costs and time involved (e.g., Hutchinson et al. 2016). Additionally, the ever-increasing volume and classes of chemicals in widespread use makes comprehensive biological testing unfeasible. Consequently, *in silico* toxicology efforts that commonly employ quantitative structure–activity relationships (QSARs) have become critical for early tier assessments of industrial chemicals (Myatt et al. 2018). The AOP approach has been proposed as a tool to help assess the safety of chemicals that, when coupled with robust computational toxicology, will reduce reliance on biological testing (Burden et al. 2015). Importantly, more research efforts should be targeted at predictively identifying chemical properties that result in MIEs with adverse outcomes at the organism and population levels. Also as noted above, one such attempt is the United States Environmental Protection Agency’s (USEPA) ToxCast program, which employs computational and high-throughput screening (HTS) tools for prioritizing environmental contaminants (Dix et al. 2006; Cote et al. 2016). In fact, molecular docking (McRobb et al. 2014) and quantum mechanics approaches are advancing the science beyond traditional $\log K_{ow}$ based QSAR approaches (Kostal 2018).

How do we better understand the linkages between the structural and physicochemical properties of substances to predictively model fate and bioavailability in different environments?

Structural and physicochemical properties of compounds are used in risk assessments to identify priority persistent and bioaccumulative compounds (Howard and Muir 2010). Many of the algorithms used in risk assessments were developed for hydrophobic organic compounds under temperate conditions. There is increasing evidence that these “rules of thumb” developed for neutral hydrophobic compounds may not be sufficiently predictive of the fate and bioavailability of hydrophilic compounds and do not predict the behavior of ionizable compounds. For example, the octanol–water partition coefficient $\log K_{ow}$ is used as an indicator of enhanced accumulation, with molecules that have $\log K_{ow}$ values greater than 3 predicted to accumulate. However, some uncharged molecules with low $\log K_{ow}$ values have also been shown to accumulate in organisms (e.g., Emnet et al. 2015). Similarly, K_{ow} -based approaches have limitations for ionizable chemicals such as pharmaceuticals and per- and polyfluoroalkyl substances (PFAS), which partition by nonhydrophobic mechanisms (e.g., ion exchange, protein binding; Armitage et al. 2017). There is a need to undertake a metaanalysis of the available data on the linkages between the structural and physicochemical properties of substances and their environmental fate and bioavailability. Basic and applied research will be necessary to improve

predictive models for properties that fall outside of the mechanistic domain of historic hydrophobic contaminants.

How do we develop better broad-screening analytical and information-processing techniques that do not require preselection of target contaminants?

“You only find what you are looking for” is a truism of environmental monitoring (Waller and Allen 2008). Widely available analytical techniques require preselection of target analytes and commonly include extensive sample preparation. This approach means that environmental monitoring programs selectively include known contaminants for which robust analytical methods exist and may not provide data on the priority contaminants for a particular time or location (Daughton 2014; Gaw and Brooks 2016). Analysis costs associated with screening just 1 water sample, for example, can be prohibitive when using multiple traditionally available analytical methods for diverse classes of contaminants. In addition, it can be difficult to establish whether there are no data for a particular contaminant because it is not present in the environment or because there are no suitable analytical methods and standards. Although new approaches using high-resolution mass spectrometry are being developed to enable nontarget analysis of organic compounds (Samanipour et al. 2016; Hollender et al. 2017), these techniques are not yet routine and provide information only on organic classes of contaminants. In contrast, ecosystems are exposed to complex mixtures that contain nutrients and metals, in addition to synthetic and naturally produced organic compounds. Advancing development and availability of robust nontarget screening techniques would significantly enhance environmental protection and would specifically support a number of the other top 20 research questions identified here.

How do we use chemistry to better design sustainable waste management?

Global pollution is now recognized as being responsible for the loss of more human lives each year than all wars or cancers (Landrigan et al. 2018). Human population growth and urbanization results in product use and chemical consumption being concentrated in cities faster than environmental management systems and interventions are being developed (Brooks 2018). For example, solid waste generation, which is currently estimated at 10 billion tons per year in urban areas, will continue to grow and become increasingly concentrated, particularly in developing and middle-income countries (Wilson et al. 2015). In Australia, although per capita waste generation has decreased, the mass of solid waste produced continues to increase, with a 7 % increase over a recent 11-y period (over the period of 2006–2007 to 2016–2017; National Waste Report 2018). New Zealand is one of the highest generators of household waste in the Organisation for Economic Co-operation and Development (OECD 2019). Similarly, wastewater production is concentrated in cities, yet 80 % of the global sewage production is released untreated to the environment (WWAP 2017). Key sustainable development

goals aim to increase sustainable cities and communities as well as responsible consumption (UN 2015), which will require development and implementation of innovative waste management programs. Advancing green engineering to reduce waste generation, increasing beneficial reuse and recovery from diverse waste streams, and stimulating sustainable molecular design of chemical ingredients and products that maintain function but are less hazardous and degrade faster (Coish et al. 2016) represent important opportunities to meet sustainability goals while stimulating innovation and reducing chemical risks to public health and the environment. In fact, designing a future without waste and associated environmental pollution was recently identified as a grand challenge for environmental engineering (NASEM 2018). To realize this challenge, environmental toxicology, chemistry, and engineering will need to advance transdisciplinary research cooperation with ecology, public health, and other disciplines.

How can we ensure sustainable supplies of clean water, energy development, and food security while simultaneously minimizing ecological impacts and protecting environmental quality?

This question represents perhaps the grandest challenge of the 21st century. Increasing populations and levels of development across the globe are driving the need for sustainable supplies of clean water, energy development, and food security (UN 2015). In fact, the US National Academy of Science also identified the production of sustainable supplies of food, energy, and water as a grand challenge for environmental engineering in the 21st century (NASEM 2018). However, there is a need to ensure that any new technological advances to address a particular issue do not result in risk trade-offs that have adverse impacts to environmental quality and ecosystem integrity. For example, sources of clean energy are being heavily promoted to mitigate climate change and poor air quality. In 2018 six solar panels were installed every minute in Australia, with 1 of every 5 households hosting rooftop solar generation (CER 2019). Over the next 10 y the use of solar technologies is expected to accelerate, and improved solar energy capture and storage materials are being developed. There is the potential for these materials to become sources for CECs and to enter waste streams as they are decommissioned and replaced. Therefore, as we move toward a circular economy, we must be mindful of the implications of new technologies for environmental quality. Better integration of robust predictive and comparative toxicology within life cycle assessment represents an important research opportunity.

MULTIPLE STRESSORS AND MIXTURES

What are the combined impacts of various agrochemicals (e.g., veterinary medicines, pesticides) and eutrophication from intensive terrestrial farming operations on the health of aquatic and terrestrial organisms?

Primary industry is a key economic driver in the Australasian region of Oceania. Intensive and industrial agricultural

practices have resulted in increased levels of pollutants being discharged to the environment increasing the potential to impact associated ecosystems and adjacent landscapes. Agrochemicals and veterinary medicines often co-occur in nutrient-enriched ecosystems, yet ecotoxicology studies of these contaminants across nutrient gradients are rare (Brooks et al. 2008). Traditionally, ecological risk assessment of agrochemicals has been conducted on a chemical-by-chemical basis, but the cumulative effects of these chemicals with veterinary medicines, with other stressors (Gustavsson et al. 2017), or within eutrophic systems (Baxter et al. 2016) has not been robustly addressed. Common ecotoxicity assays with plants and algae often employ media with nutrient-enriched concentrations and stoichiometric conditions that deviate from environmentally relevant conditions (Brooks et al. 2015). Further, nutrient-enriched conditions can promote development of HABs and associated production of algal toxins, which are now recognized to confound stressor identification approaches for anthropogenic contaminants (Brooks et al. 2016).

More complex laboratory and (semi)controlled field studies are needed to assess the potential additive, antagonistic, or synergistic effects of these complex stressor mixtures. As one example, Taylor et al. (2018) recently demonstrated the usefulness of employing coupled field studies with experimental stream mesocosm experiments to identify ecological thresholds associated with P enrichment. Unfortunately, similar studies have rarely examined influences of agrochemicals or veterinary medicines, a number of which are actually pesticidal, on stream ecosystems across nutrient gradients. Aided by answering other priority research questions identified in the current paper, developing fundamental understanding of the specific Mode of Action (MOAs) of these chemicals will help determine their combined effects. However, the data generated need to be supported within ecological risk assessment models that are able to accurately predict cumulative effects, including ecosystems services (Syberg et al. 2017). Herein, future research at the intersections of ecological stoichiometry and toxicology (i.e., how nutrition can affect the toxicity of contaminants, how contaminants can influence nutrient dynamics, or how nutrients can influence toxins production) promises to support an understanding of interactive effects of anthropogenic contaminants and algal toxins in nutrient-enriched systems (Conine and Frost 2016). Similarly, advances in ecological genomics are poised to support environmental assessment of complex stressors in the field (Yang et al. 2018; Zhang et al. 2018).

What are the effects of changing demographics, economic development, consumption patterns, and climate (e.g., ocean acidity, water temperature) on chemical emissions, environmental fate, and ecotoxicology of contaminants and multiple stressors?

Anthropogenic stressors, including increased population, economic activity, and changing consumption patterns, are contributing to rapid environmental change (Steffen et al.

2015). The identified global megatrends of increased urbanization, diverging population trends, changing disease burdens, and accelerating technological growth will determine the types and quantities of chemicals released regionally (e.g., Kookana et al. 2014). Our current paradigms for environmental fate and toxicity of contaminants will be challenged by the anticipated increase in environmental pollution (EEA 2015) and the consequences of climate change. Global climate change is anticipated to alter both the environmental variables (e.g., temperature, precipitation, salinity, pH) that determine the environmental fate and toxicity of chemicals as well as the resilience of organisms to cope with exposure to chemical stressors (Hooper et al. 2013). Risk assessment tools and environmental surveillance systems will need to be sufficiently adaptive to identify and prioritize emerging threats, particularly those that arise due to a combination of chemical and physical stressors, some of which will be driven by global climate changes (Landis et al. 2013). Given the inherent difficulties in replicating “real world” conditions for experiments, our predictive modeling tools will need to be refined to ensure that a precautionary approach can be taken to managing risk in a rapidly changing world.

What are the combined effects of very low levels of multiple contaminants (e.g., pesticides, natural resource extraction contaminants, salinity, pharmaceuticals and personal care products, endocrine-disrupting chemicals) with different modes of action on aquatic and terrestrial organisms and ecosystems?

Understanding environmental consequences of chemical mixtures remains one of the most challenging issues in achieving sustainable environmental quality (Van den Brink et al. 2018; Fairbrother et al. 2019). With increasing urbanization, multiple land uses are interfacing in peri-urban watersheds, which inherently increases the likelihood of diverse contaminants from urban, agricultural, and industrial activities that co-occur in complex mixture scenarios. Guidelines derived for individual stressors may not be sufficiently protective when ecosystems are exposed to multiple stressors. For example, changes in benthic community distributions have been reported at concentrations below individual metal guideline values (Tremblay et al. 2017). Salinization is particularly relevant to regions in Australasia, yet influences of salinity gradients on contaminants with diverse modes of action are poorly understood among species (Canedo-Arguelles et al. 2018). Various toxicity identification evaluation (TIE) protocols, response-directed fractionation procedures, and effects-directed analyses have been developed to identify causative chemical stressors within surface waters and sediments. However, it is particularly important to define strengths and limitations of historical bioassays employed for such activities, particularly when low levels of biologically active contaminants with diverse MIEs are considered. In recent years, bioassay tools with increasing mechanistic specificity have become important for diagnostic applications (Escher et al. 2014) beyond the

traditional morphometric aquatic toxicity responses introduced above that are employed in TIEs (USEPA 1991). Unprecedented opportunities are emerging with use of high-throughput *in vitro*, transgenic fish lines, and *in situ* toxicogenomic platforms when coupled with targeted and nontargeted chemical analyses (Bradley et al. 2017) in the field (Blackwell et al. 2017; Bradley et al. 2017; Perkins et al. 2017). However, metabolic transformation of contaminants and other basic scientific limitations remain when extrapolating *in vitro* to *in vivo* effects and even comparing responses among the 2 most common fish models (Corrales et al. 2016; Steele et al. 2018). Advancing AOP efforts for mixtures and predictive modeling of these complex low-level constituents will be important. The funnel hypothesis (Warne and Hawker 1995) postulates that, as the numbers of chemicals present at equipotent concentrations increases, the likelihood of additive combined effects increases. Efforts are needed to identify whether, when, and what specific MOAs drive divergence from such theoretical constructs of low-level mixture toxicity. It is thus not surprising that understanding the environmental implications of chemical mixtures was also identified as a priority research question in GHSP efforts from Europe (van den Brink et al. 2018), Latin America (Furley et al. 2018), and North America (Fairbrother et al. 2019). Clearly, this area deserves future attention.

RISK ASSESSMENT, REGULATIONS, AND GUIDELINES

What water quality guidelines are needed to protect temporary waters and associated ecosystems from the influences of development?

Temporary waters (i.e., intermittent, ephemeral, and seasonal) are common in temperate, arid, and semiarid landscapes of Australia and many other regions around the world. Sheldon and Fellows (2010) reported that up to 95 % of Australia's river channels are temporary, while a large proportion of the standing inland waters are also classified as temporary. Consequently, when these waters are present they are an extremely important source of water for the ecosystems of inland Australia and other regions. To date, much of the research has focused on the effects of extraction and sustainable use of temporary waters (Acuña et al. 2014; Datry et al. 2014), provision of their ecosystem services (Boulton 2014), and the importance of wetting and drying cycles for ecosystem health (Leigh 2013). However, there is a recognized need to better address changes in water quality arising from urbanization, agriculture, and mining (e.g., Queensland, Ramsay et al. 2012; South Australia, Botwe et al. 2015).

Due to the nature of these temporary waters, they are likely to experience pulse-exposure scenarios, but there are limited data sets that are useful for determining water quality guideline values for episodic exposures to contaminants. Moreover, many temporary waters have been converted to perennial or near-perennial waters by effluent discharges (Brooks et al. 2006), which represent important

systems for environmental management with changing climatic conditions (Luthy et al. 2015). Although there are controls in Australia and some other countries on water quality in discharges and/or receiving waters for perennial or near-perennial waters, no specific guidance exists in any set of guidelines or regulations on the combined impact of conversion from temporary to nontemporary status together with alteration of water quality. Understanding and managing environmental quality impairments in these temporary waters represents a timely research need for parts of the Australasian region of Oceania and other global systems experiencing urbanization and climate change.

What are the effects of short magnitude, frequency, and duration (e.g., intermittent, episodic) exposures to contaminants and other stressors, and how can these scenarios be effectively incorporated into water quality guidelines?

Water quality criteria, standards, and guidelines are developed to protect various uses of surface waters. Through these efforts, threshold concentrations of contaminants (e.g., metals, pesticides, ammonia) and other stressors (e.g., depressed dissolved O, increased temperature) are identified and then applied, particularly in developed countries. Such regulatory “bright lines,” representing specific concentrations of individual contaminants, have historically been intended to be protective of, and ideally predictive of, ecological integrity. Presently, these numeric values are most commonly derived from probabilistic analyses of results from single-species toxicity assays, which are intended to identify concentration–response thresholds, instead of individual species or community effects from episodic exposures that inherently vary in magnitude, frequency, and duration (Posthuma et al. 2001). For example, King et al. (2016) recently reported ecological structure and function responses to environmentally realistic episodic pulses of a common herbicide using outdoor stream mesocosms. Clearly, an advanced understanding of responses to episodic and intermittent chemical exposures is needed. Such information, while requiring innovative mechanistic coupling of toxicokinetics and toxicodynamics, and ecological genomics in the field, promises to reduce uncertainties associated with laboratory-to-field extrapolation during derivation of water quality guidelines.

How can we measure ecosystem resilience to and recovery following exposure to stressors?

Stochastic events influence ecosystem services and biodiversity, which are among the most common protection goals identified during problem formulation of ecological risk assessments. Such stochasticity inherently affects interpretation of stressor–response observations in the field and implementation of environmental management decisions. Although the diversity–stability hypothesis and functional redundancies have long been considered, both theoretically and empirically, and debated (McCann 2000) in ecology and ecotoxicology, identifying functional traits within

assemblages and other ecosystem characteristics that impart resilience to natural and anthropogenic stressors remains decidedly challenging. In fact, 2017 has been described as the year of the disaster, with numerous billion-dollar events reported throughout the world (NOAA 2018). Herein, ecosystem services, when not compromised, represent key management objectives for disaster risk reduction and climate change adaptation (Monty et al. 2016; Renaud et al. 2016), and are appropriately included in the United Nation's Sendai Framework for Disaster Risk Reduction for 2015 to 2030 (UNDRR 2015). For example, rapid global declines of terrestrial and aquatic species present a profound manifestation of cumulative threats to biodiversity. In Australasia, degradation of the Great Barrier Reef has prompted extensive efforts to define cumulative stressors and advance resilience-based management (Anthony et al. 2013). In New Zealand, large earthquakes in the Canterbury region resulted in loss of habitat and measurable stress on aquatic organisms (Potter et al. 2015; Chandurvelan et al. 2016). Similarly, the Rena oil spill, New Zealand's largest maritime environmental disaster, impacted hundreds of kilometers of coastline in 2011 (Schiel et al. 2016). In such cases, influences of rare species on ecosystems functions require additional study (Leitao et al. 2016). With the prospects of climate change further compounding multiple stressor effects on aquatic and terrestrial ecosystems, it appears clear that developing an advanced understanding of ecosystem resiliency prior to and following disasters and in the face of cumulative stressors has never been more important.

SPOTLIGHT ON AUSTRALASIA

Are there differences in toxicological thresholds among native and nonnative organisms, and how can species sensitivity information from nonresident species be used to predict adverse outcomes and protect our unique biota and ecosystems?

The iconic aquatic and terrestrial species unique to Oceania in general and Australasia in particular hold deep cultural significance to indigenous communities and are important to the recreational, commercial, and conservation sectors. However, most of the toxicity estimates are derived from studies that use North American and European species; very little toxicity data exist using Oceania species, with some notable exceptions. Consequently, the Australian and New Zealand Water Quality Management Strategy (ANZECC 2000), and the new revised guidelines took the pragmatic approach of deriving Default Water Quality Guideline Values using any available data that passed predefined quality control criteria. However, this approach makes the considerable assumption that native Oceania species are of a similar sensitivity to that of nonnative species. This assumption has not been comprehensively tested because there have been no broad-scale systematic comparisons on toxicity data from native Oceania species and

nonnative species. It is important to note that a similar question was recently identified from Latin America (Furley et al. 2018). Advancing comparative and predictive toxicology research promises to help us understand differences among species sensitivities to contaminants with diverse mechanisms of action (Brooks 2018).

There have been many toxicity tests developed for native species in Australasia. The earliest of these native-species suites were developed to satisfy the research needs for controversial issues. For example, in the early 1990s, the National Pulp Mills Research Programme identified a number of temperate Australian species to assess the toxicity of pulp mill effluents and test "greener" technology options (Crossland and Abel 1992; Stauber et al. 1994). In New Zealand, a standard suite of 3 marine and 4 freshwater tests on native species was developed by the National Institute of Water and Atmospheric Research (Hall and Golding 1998), and sensitivities of these species were compared with those of nonnative species for 4 reference toxicants. A suite of standardized tropical freshwater toxicity tests was developed by the Environmental Research Institute of the Supervising Scientist for the regulation of the Ranger Uranium Mine, which is adjacent to the World Heritage-listed Kakadu National Park (Riethmuller et al. 2003). Both of these industries were faced with significant public opposition, but the development of native-species toxicity tests helped decision makers reassure the public that environmental issues were being addressed appropriately.

In more recent years, a member of the business community invested in the development of a suite of toxicity tests using native tropical marine species to improve the environmental management of their industrial effluents by using biological effects data (van Dam et al. 2018). The motivation for this was to address a gap that existed for tropical species because most toxicity tests were developed by first-world nations in temperate environments (van Dam et al. 2008). Such research investments have subsequently benefited other industries that have capitalized on the availability of the tropical tests (e.g., Gissi et al. 2018), which has enabled valuable tropical-versus-temperate comparisons (Peters et al. 2019). Ad hoc toxicity testing using culturally significant fishes (e.g., Inanga, *Galaxias maculatus*; McRae et al. 2018) and invertebrates (e.g., freshwater mussels, clams, and crayfish; Clearwater et al. 2014) has been developed in New Zealand and Australia (e.g., Markich and Camilleri 1997). The sensitivities of native and nonnative species to certain contaminants have been compared in some cases. For example, Hagen and Douglas (2014) asked this question but could find sufficient data for only 3 chemicals, that is, 4-chlorophenol, phenol, and ammonia. They concluded that there were no differences in species sensitivity that warranted the application of safety factors. However, until a sufficient Oceania data set for a broader set of chemicals is available, this question will remain unaddressed. Here again, advancing comparative ecotoxicology research in this area is a priority.

How do we incorporate and protect cultural and social values (relating to humans, biota, and ecosystems) to empower citizen, societal, and indigenous engagement in the research, management, and legislation of priority environmental contaminants?

Indigenous peoples are key to many environmental management projects and decisions globally, where their status ranges from disadvantaged minorities to the dominant cultural group within their respective communities and country. Indigenous peoples carry with them distinctive and localized cultural and environmental knowledge, based on thousands of years' experience (Stevenson 1996). However, mechanisms to incorporate their indigenous knowledge, cultural values, and traditional management systems into decision-making processes remain poorly formulated in most global legislatures, business decisions, and academic programs. This is the case despite numerous international and regional, legally and nonlegally binding instruments (Convention on Biological Diversity 1992; UN 1992, 2007) and statutory national obligations (legislative and policy level; Palmer 2008) requiring appropriate and meaningful indigenous peoples' involvement. Further, ignorance of inherent challenges around the application of indigenous knowledge, existing power relations, and contextual nuances of Indigenous knowledge have also hampered access to, and an articulation of indigenous knowledge in, environmental management and decision-making processes (Briggs 2005).

Oceania, like other global regions, has a diverse range of indigenous peoples, each with their own unique history, experiences, and challenges with respect to articulating their voice around environmental contaminants. Unfortunately, indigenous knowledge and values (IK&V) are not well represented in assessment and management approaches in environmental issues. Applying an indigenous knowledge lens considers the whole of environmental change in determining the impact of contaminants (Kookana et al. 2013). In addition to considering the impact of contaminants to indigenous people's environments, biodiversity, and culture (Ataria et al. 2016), the impact of practices that disrupt ecological patterns and services are also critical to consider, particularly for those communities that are reliant on natural resources for their physical and cultural existence.

The collaboration of traditional knowledge and research is needed between communities and indigenous peoples. Advancing forward it will be imperative to manage environmental quality as both strive to advance their knowledge systems to protect environmental quality and natural resources. Engagement protocols differ across all indigenous peoples globally. However, the environmental science and engineering communities can assist in cocreating protocols in close consultation with the relevant indigenous peoples that are specific to regions, are equitable, empower mutual benefit, and are enduring. Indigenous people assert an inherent expectation to be involved in caring for, protecting, and rejuvenating their traditional land, freshwater, marine, and atmospheric

environments. To some it is a cultural obligation as custodians, whereas to others it is a means of maintaining their identity by reinstating and retaining their cultural practice and heritage and by empowering their developmental aspirations for future generations. Here we call for concerted global research efforts to integrate IK&V during problem formulation and, more specifically, identification of ecosystem protection goals within environmental risk assessment and management efforts.

TOOLS FOR IMPROVING RISK ASSESSMENT

How do we exploit, collate, and integrate existing environmental toxicology, chemistry, and geospatial data to help develop robust risk assessment?

Natural ecosystems are increasingly degraded as a result of exposure to multiple stressors that vary over space and time. We now know that the global reach of anthropogenic stressors is beyond what was previously predicted, with persistent pollutants such as PCBs, polybrominated diphenyl ethers (PBDEs), and microplastics found in the remote Arctic and deep sea trenches (Schlining et al. 2013; Van Cauwenberghe et al. 2013; Obbard et al. 2014; Jamieson et al. 2017). To address these challenges, we have increasing access to physical, biological, and chemical measurements from new remote sensing tools and their integration into geographical information systems (Dafforn et al. 2016). Moreover, advances in molecular analysis have allowed us to capture more holistic information about the health of entire ecosystems, from microbial to macrobiotic scales, and to go beyond impacts on structure to understand consequences for ecosystem function and services (Chariton et al. 2016). The advent of real-time technologies such as the MinION for DNA/RNA sequencing and the microfluidic lab-on-a-chip provides us with more opportunities for improved spatiotemporal analyses (Campana and Wlodkowic 2018). The availability of these data and new geospatial and ecogenomic bioassessment tools has the potential to increase our capacity for ranking and understanding stressor impacts and crucially to allow us to differentiate stressors impacts when present in combination.

At the same time, we are experiencing technological advances and associated information booms, with many decades of ecotoxicological testing and biomonitoring information collected and added to databases following regulatory requirements. Numerous databases around the world hold information about different chemical stressors as well as potential biological responses. For example, the Pesticide Properties DataBase has approximately 2300 pesticide active substances and >700 metabolites stored alongside response metrics related to human and environmental health (Lewis et al. 2016). Other large collections of biological data such as GENBANK (Benson et al. 2010), TRY (Kattge et al. 2011), D3 (Hintze et al. 2013), COMADRE, and COMPADRE (Salguero-Gómez et al. 2015) offer information related to genetics, functional plant ecology, grassland

ecology, and plant and animal demography alongside metadata from, for example, ecoregions that can be used to ask globally relevant questions (Salguero-Gómez et al. 2015) and be integrated within risk assessment frameworks.

Machine learning techniques could be used to harness the power of such extensive data sets into risk assessment. For example, molecular tools such as transcriptomics have been integrated with machine learning techniques to identify and classify priority EDCs (Ornostay et al. 2013). Similarly, artificial neural networks have been used to select biomarkers on the basis of key response variables (Bradley 2012). Decision tree models based on environmental metadata have been used to predict benthic macroinvertebrate distributions (D'Heygere et al. 2003). Environmental metadata using a Random Forests machine learning algorithm have likewise been used to reveal nonlinear relationships and critical thresholds for cyanobacterial blooms (Nelson et al. 2018), which is significant because HABs now represent the greatest water quality threat in some ecosystems (Brooks et al. 2017).

Overall, our predictive power has exponentially increased, allowing us to move beyond the current norm of single-stressor assessments, done at small spatial scales and with few receptors, to enhanced risk assessment (Van den Brink et al. 2016). However, there are still hurdles to overcome before we can harness and exploit this Big Data to its fullest. We need to 1) improve our techniques for data validation to remove errors in, for example, specimen identifications for DNA barcoding; 2) improve the availability of data not just through openness but also by targeting underrepresented taxonomic and geographic groupings; 3) improve standardization so that data are comparable over space and time; and 4) invest in real-time technologies that provide direct measures of impact rather than providing proxies (Dafforn et al. 2016).

How can prescreening techniques (e.g., in silico, in vitro) be developed, advanced, and validated to identify and predict whole organism effects?

The rate of discovery and synthesis of new chemicals has grown exponentially in the last decades, exceeding our ability to empirically determine the toxicity of new compounds using conventional (whole animal) toxicity testing methods. This means that more and more chemicals are put into global circulation without a thorough understanding of their potential toxic impacts. Too often, the chemicals substituted for problematic substances display unacceptable toxicity profiles (Rosal et al. 2010; Björnsdotter et al. 2017). Unfortunately, conventional toxicity testing provides too narrow a funnel (in terms of time, cost, and ultimately, throughput) to assess the risk of the vast number of new compounds designed daily by chemical, pharmaceutical, and agricultural industries. Clearly, a higher throughput approach is required.

This is where *in silico* modeling and *in vitro* pretesting methods offer a way forward. Using these HTS techniques, which can screen thousands of chemicals every day, toxicity

testing can be prioritized and focused on those molecules most likely to pose a threat to humans and/or ecosystems (Collins et al. 2008). This is the paradigm shift foreshadowed in the Tox21 vision for toxicity testing in the 21st century (NRC 2007), and which relies on the AOP concept (Ankley et al. 2010) to translate a key initiating event at the molecular or cellular level (either modeled *in silico* or measured *in vitro*) to the adverse outcome of consequence (e.g., survival, reproduction, development, behavior) that is our focus of concern (Ankley et al. 2016).

Although tremendous progress has been achieved in adapting and validating *in vitro* tools to environmental monitoring and risk assessment (e.g., in Australasia, Coleman et al. 2008; Mispagel et al. 2009; Chinathamby et al. 2013; Bain et al. 2014; Escher et al. 2014; Leusch et al. 2014; Scott et al. 2014; Roberts et al. 2015; Boehler et al. 2017; Neale, Achard et al. 2017; Neale, Altenburger et al. 2017; Chen et al. 2018; Leusch et al. 2018), some fundamental questions still need to be systematically addressed before these techniques can become reliable predictors of whole animal level effects:

- 1) Refine quantitative *in vitro* to *in vivo* extrapolation (QI-VIVE): Although there is a clear correlation between *in vitro* response and *in vivo* effects for some endpoints such as acute toxicity (Kaiser 1998; Tanneberger et al. 2013; Natsch et al. 2018) and receptor-mediated endocrine effects (Sonneveld et al. 2006, 2011; Henneberg et al. 2014), toxicokinetic factors (absorption, distribution, metabolism, and excretion) still pose a difficult challenge for QI-VIVE (Blaauboer 2015; Meek and Lipscomb 2015), although groundbreaking studies suggest that this may soon be within reach (Rotroff et al. 2010; Wetmore 2015).
- 2) Fully map relevant AOPs: There is still much work to be done to map key events (KEs) to connect the dots between the molecular or cellular initiating event and the ultimate apical consequence to produce comprehensive AOPs, for both humans and ecosystems (Ankley et al. 2016). In combination with QI-VIVE, this mapping would ultimately allow us to produce quantitative AOPs.
- 3) How much is too much? *In vitro* assays are often exquisitely sensitive and able to detect activity even in clean samples. In whole organisms, a small amount of dysfunction at the molecular and cellular level can often be compensated for by defense and repair mechanisms to avoid any higher level consequence. Until we can quantitatively extrapolate from *in vitro* to *in vivo* (steps 1 and 2 above) and quantify the repair ability for each type of dysfunction, it will be difficult to accurately link an *in vitro* response to an *in vivo* adverse effect. In the meantime, several different approaches have been proposed to produce effects-based trigger (EBT) values, including reading across from current chemical guidelines (Escher et al. 2015, 2018) or *de novo* derivation (Brand et al. 2013; Jarošová et al. 2014) and how to use them in a practical context (Leusch and Snyder 2015; Ron et al. 2017).

Clearly, there are still some unanswered questions in how we use *in silico* models and *in vitro* bioassays. But these new

tools also offer a unique and necessary solution to overhaul the single-chemical risk assessment approach that relies on the traditional aquatic models and endpoints discussed above and to properly screen the sheer number of chemicals that make our modern lifestyles possible without negatively impacting human health and the environment. Further, advancing these diagnostic tools, particularly when coupled with nontarget analytical methods, promises to support efforts to answer other priority research questions identified here.

How can ecotoxicology information be integrated more closely during interpretation of ecological data?

Two closely related questions focus on the necessity of more closely integrating research among ecology and ecotoxicology, which in many parts of the world remain separate fields of study. Whereas basic ecology studies in terrestrial and aquatic systems are fundamentally important for conservation, including understanding ecosystem services and biodiversity, translational ecological efforts remain critical for environmental assessment and management (Saaristo et al. 2018). Interpretation of field data sets can be challenging due to ecosystems commonly being exposed to multiple stressors, which may be known or unknown. Subsequently, identifying underlying causative relationships among complex stressors requires multidisciplinary perspectives. For example, failure to consider chemical stressors beyond nutrient enrichment during basic ecological and biogeochemical studies in systems influenced by agriculture and urbanization can confound interpretation of findings. For decades, researchers have called for close integration among ecology and ecotoxicology research pursuits (Cairns 1988; Zala and Penn 2004; Melvin and Wilson 2013; Arnold et al. 2014).

More recent contributions in community and stream ecology (Rohr et al. 2006; Rosi-Marshall and Royer 2012; Bernhardt et al. 2017), behavioral ecology (Saaristo et al. 2018), and ecophysiology (Cooke et al. 2013) consistently echo these earlier sentiments. Beyond applied studies aimed at stressor identification, anthropogenic chemicals, particularly specifically acting contaminants (e.g., pesticides, pharmaceuticals), can serve as experimental scalpels to dissect basic structural and functional relationships. For example, mesocosm studies by Fairchild et al. (1994) with pesticides partitioned direct from indirect community interactions. Environmental studies with pharmaceuticals have yielded unique comparative ecophysiology information (Owen et al. 2007). Addressing several of the questions identified in earlier sections aimed at advancing integrated research in ecological threshold analyses, environmental genomics, quantitative AOPs, and integrative, comparative, and predictive toxicology, when coupled within mainstream experimental and theoretical ecology, promises reciprocal and transformational basic and applied benefit, particularly as global ecosystems continue to be influenced by complex stressors.

How do we advance ecotoxicology testing to be more relevant to ecological systems?

Prospective ecotoxicology assays are employed by businesses and government agencies to assess the safety of substances prior to their introduction to the market or to assess contaminants of potential concern before they are released to the environment. Industrial operations have also been required to synthesize predicted effluents for safety assessments when changing their waste treatment or introducing new ones. Historical products in commerce may also be prioritized for more detailed safety assessment. Whereas retrospective ecotoxicological studies often include *in vitro* and *in vivo* models to examine field-collected water, sediment, or soil in laboratory settings, *in situ* studies with caged organisms, and surveillance of biological conditions in the field, micro- and mesocosm studies are employed for both prospective and retrospective efforts in an attempt to bridge laboratory-to-field information. For decades, researchers have noted challenges from lower to higher scales of biological complexity due to increasing endpoint variability (and societal relevance) and environmental stochasticity as one moves from the laboratory model to ecosystem-level perturbations (Dickson et al. 1992; La Point and Waller 2000).

Predictive coupling of laboratory with field perturbations remains a grand challenge in environmental science. However, it remains important to ensure the quality of data produced from standardized model systems, while advancing innovative and exploratory ecotoxicological research that may not be intended or amenable to directly be integrated within environmental assessments (Moermond et al. 2017). Such challenges were considered during a recent SETAC Pellston Workshop on “Improving Usability of Ecotoxicology in Regulatory Decision Making, August 2015” which has documented the need for ecotoxicological data sets that are reliable and relevant (Rudén et al. 2017). Beyond the traditional biological indices approaches, recent progress in ecological threshold analysis (Baker and King 2010), ecological genomics (Zhang et al. 2018), and species traits (Van den Brink et al. 2013) are improving field studies. Future research in mechanistic and comparative ecotoxicology, if integrated with ecology, is poised to support more robust experimental designs and extrapolations across levels of biological organization, although uptake of recent advances within prospective and retrospective regulatory activities remains differential around the world. Therefore, employing reasonable and defensible weight-of-evidence approaches will remain important (Suter 2016).

CONCLUSIONS

The Australasian region of Oceania faces increasingly diverse environmental challenges associated with multiple stressor influences on environmental quality. The current analysis represents an initial attempt within Oceania to develop a research agenda aimed at advancing toward more sustainable environmental quality and ecosystem integrity. Through a transparent, bottom-up, multidisciplinary, and

multistakeholder process, we identified 20 priority questions to support future environmental research. As noted recently (Van den Brink et al. 2018), step changes are needed for basic and applied studies of environmental stressors, and their management, if we are to achieve the United Nation's Sustainable Development Goals (UN 2015). We agree, as evidenced by the interconnections among priority research questions reported herein.

Several questions identified the need to improve predictive environmental exposure and toxicology tools for risk assessment and to reduce and replace animal testing. Similarly, the development of robust nontarget analytical screening techniques to determine priority contaminants in ecosystems exposed to complex mixtures was identified as an urgent need. Strategically advancing these areas will assist in addressing other questions related to multiple stressors (e.g., chemicals, salinity, acidification), susceptibility of regional flora and fauna, management of unique ecosystems (e.g., ephemeral water bodies), and stress from global megatrends (e.g., urbanization, the food–energy–water nexus) and climate change. The importance of understanding the comparative sensitivities of regionally unique species was also reported from Latin America (Furley et al. 2018). Incorporating and protecting cultural and social values to empower citizens, especially indigenous peoples' engagement during research, management, and policy development, was further identified as a key research opportunity. In this regard, ongoing efforts within Australasia are incorporating cultural knowledge during identification of ecosystems protection goals (i.e., the Whanganui River and other systems in New Zealand have been granted the same legal rights as a person), which represents an interesting model that could benefit elsewhere.

We expect the top 20 questions identified here will be complementary to and assist advancement of national prioritization efforts such as the Australian Science and Research Priorities and Practical Challenges (Australian Government 2015) and the New Zealand National Science Challenges (MBIE 2016). For example, 5 of the 11 Australian Science Research Priorities (e.g., Environmental Change, Energy, Soil, Water, Food) include Practical Challenges to address sustainable environmental quality and ecosystem integrity. Similarly in New Zealand, Science Challenges relevant to sustainable environmental quality include Biological Heritage, The Deep South, Sustainable Seas, and Our Land and Water. Expertise and capacity within the Australasia chapter of SETAC and other scientific disciplines in Oceania are well positioned to support these efforts (a brief history of SETAC Australasia can be found in the Supplemental Data). Answering the 20 priority research questions will not be trivial, but will support basic and applied research innovation and advancement of robust practices to achieve more sustainable environmental quality within the region and other parts of the world.

Acknowledgment—Financial support was provided by Baylor University, Tony and Donna Robert, the University of York, and the US Environmental Protection Agency. In-kind support was provided by the Society of Environmental

Toxicology and Chemistry (SETAC) and SETAC Australasia. We deeply acknowledge all experts who anonymously submitted questions, which made this exercise possible. We would like to thank the anonymous reviewers and Gary Ankley (US Environmental Protection Agency) for their thoughtful comments and efforts towards improving our manuscript.

Disclaimer—The authors declare no conflicts of interest. The peer-review process for this article was managed by the Editorial Board without the involvement of S Gaw.

SUPPLEMENTAL DATA

Background information and the full set of questions submitted:

- 1) A Brief History of the SETAC Australasia Chapter
- 2) Author-submitted Questions

REFERENCES

- Acuña V, Datry T, Marshall J, Barceló D, Dahm CN, Ginebreda A, McGregor G, Sabater S, Tockner K, Palmer MA. 2014. Why should we care about temporary waterways? *Science* 343(6175):1080–1081.
- Adams VM, Spindler RE, Kingsford RT. 2016. Thinking globally, acting locally—Conservation lessons from Oceania. *Pac Conserv Biol* 22(2):85–89.
- Alexander S, Aronson J, Whaley O, Lamb D. 2016. The relationship between ecological restoration and the ecosystem services concept. *Ecol Soc* 21(1).
- Ankley G, Escher B, Hartung T, Shah I. 2016. Pathway-based approaches for environmental monitoring and risk assessment. *Chem Res Toxicol* 29(11):1789–1790.
- Ankley GT, Brooks BW, Huggett DB, Sumpter JP. 2007. Repeating history: Pharmaceuticals in the environment. *Environ Sci Technol* 41(24):8211–8217.
- Ankley GT, Bennett RS, Erickson RJ, Hoff DJ, Hornung MW, Johnson RD, Mount DR, Nichols JW, Russom CL, Schmieder PK et al. 2010. Adverse outcome pathways: A conceptual framework to support ecotoxicology research and risk assessment. *Environ Toxicol Chem* 29(3):730–741.
- Anthony KRN, Dambacher JM, Walshe T, Beeden R. 2013. A framework for understanding cumulative impacts, supporting environmental decisions and informing resilience-based management of the Great Barrier Reef World Heritage Area. Townsville (AU): Australian Institute of Marine Science. 110 p.
- [ANZECC, ARMCANZ] Australian and New Zealand Environment and Conservation Council and Agriculture and Resource Management Council of Australia and New Zealand. 2000. Australian and New Zealand guidelines for fresh and marine water quality. Canberra (ACT): Australiatable. 103 p.
- [ANZG] Australian and New Zealand Governments. 2018. Guidelines for fresh and marine water quality. Canberra (AU). [accessed 2019 Jun 5]. <https://www.waterquality.gov.au/anz-guidelines/about>
- Armitage JM, Erickson RJ, Luckenbach T, Ng CA, Prosse RS, Arnot JA, Schirmer K, Nichols JW. 2017. Assessing the bioaccumulation potential of ionizable organic compounds: Current knowledge and research priorities. *Environ Toxicol Chem* 36:882–897.
- Arnold KE, Brown AR, Ankley GT, Sumpter JP. 2014. Medicating the environment: Assessing risks of pharmaceuticals to wildlife and ecosystems. *Philos Trans R Soc B* 369.
- Ataria J, Baker J, Goven J, Langer ER, Leckie A, Ross M, Horswell J. 2016. From Tapu to Noa - Māori cultural views on biowastes management: A focus on biosolids. Centre for Integrated Biowaste Research Report No. 16-01. March 2016. Wellington (NZ): Centre for Integrated Biowaste Research. 8 p.
- Australian Government. 2015. Science and research priorities. Canberra (ACT). 4 p. [accessed 2019 Jan 6]. www.industry.gov.au/sites/g/files/net3906/ff/2018-10/science_and_research_priorities_2015.pdf

- Bain PA, Williams M, Kumar A. 2014. Assessment of multiple hormonal activities in wastewater at different stages of treatment. *Environ Toxicol Chem* 33(10):2297–2307.
- Baker ME, King RS. 2010. A new method for detecting and interpreting biodiversity and ecological community thresholds. *Methods Ecol Evol* 1(1):25–37.
- Bark RH, Barber M, Jackson S, Maclean K, Pollino C, Moggridge B. 2015. Operationalising the ecosystem services approach in water planning: A case study of indigenous cultural values from the Murray–Darling Basin, Australia. *Int J Biodivers Sci* 11(3):239–249.
- Baxter L, Brain RA, Lissimore L, Solomon KR, Hanson ML, Prosser RS. 2016. Influence of light, nutrients, and temperature on the toxicity of atrazine to the algal species *Raphidocelis subcapitata*: Implications for the risk assessment of herbicides. *Ecotoxicol Environ Saf* 132:250–259.
- Bengtsson-Palme J, Larsson DJ. 2016. Concentrations of antibiotics predicted to select for resistant bacteria: Proposed limits for environmental regulation. *Environ Int* 86:140–149.
- Benson DA, Karsch-Mizrachi I, Lipman DJ, Ostell J, Rapp BA, Wheeler DL. 2000. GenBank. *Nucleic Acids Res* 28(1):15–18.
- Bernhardt ES, Blaszczyk JR, Ficken CD, Fork ML, Kaiser KE, Seybold C. 2017. Control points in ecosystems: Moving beyond the hot spot hot moment concept. *Ecosystems* 20(4):665–682.
- Björnsdotter MK, Jonker W, Legradi J, Kool J, Ballesteros-Gómez A. 2017. Bisphenol A alternatives in thermal paper from the Netherlands, Spain, Sweden and Norway. Screening and potential toxicity. *Sci Total Environ* 601–602:210–221.
- Blaauboer BJ. 2015. The long and winding road of progress in the use of in vitro data for risk assessment purposes: From “carnation test” to integrated testing strategies. *Toxicology* 332:4–7.
- Blackwell BR, Ankley GT, Corsi SR, DeCicco LA, Houck KA, Judson RS, Li S, Martin MT, Murphy E, Schroeder AL et al. 2017. An “EAR” on environmental surveillance and monitoring: A case study on the use of exposure-activity ratios (EARs) to prioritize sites, chemicals, and bioactivities of concern in Great Lakes waters. *Environ Sci Technol* 51:8713–8724.
- Boehler S, Strecker R, Heinrich P, Prochazka E, Northcott GL, Ataria JM, Leusch FDL, Braunbeck T, Tremblay LA. 2017. Assessment of urban stream sediment pollutants entering estuaries using chemical analysis and multiple bioassays to characterise biological activities. *Sci Total Environ* 593:498–507.
- Botwe PK, Barmuta LA, Magierowski R, McEvoy P, Goonan P, Carver S. 2015. Temporal patterns and environmental correlates of macro-invertebrate communities in temporary streams. *PLoS ONE* 10(11):e0142370.
- Boxall ABA, Rudd M, Brooks BW, Caldwell D, Choi K, Hickmann S, Innes E, Ostapkyk K, Staveley J, Verslycke T et al. 2012. Pharmaceuticals and personal care products in the environment: What are the big questions? *Environ Health Perspect* 120(9):1221–1229.
- Bradley BP. 2012. Finding biomarkers is getting easier. *Ecotoxicology* 21(3):631–636.
- Bradley PM, Journey CA, Romanok KM, Barber LB, Buxton HT, Foreman WT, Furlong ET, Glassmeyer ST, Hladik ML, Iwanowicz LR et al. 2017. Expanded target-chemical analysis reveals extensive mixed-organic-contaminant exposure in U.S. streams. *Environ Sci Technol* 51:4792–4802.
- Brand W, de Jongh CM, van der Linden SC, Mennes W, Puijker LM, van Leeuwen CJ, van Wezel AP, Schriks M, Heringa MB. 2013. Trigger values for investigation of hormonal activity in drinking water and its sources using CALUX bioassays. *Environ Int* 55(0):109–118.
- Briggs J. 2005. The use of indigenous knowledge in development: Problems and challenges. *Prog Dev Stud* 5(2):99–114.
- Brooks BW. 2014. Fish on Prozac (and Zoloft): Ten years later. *Aquat Toxicol* 151:61–67.
- Brooks BW. 2018. Urbanization, environment and pharmaceuticals: Advancing comparative physiology, pharmacology and toxicology. *Conserv Physiol* 6(1):cox079.
- Brooks BW, Ankley GT, Boxall ABA, Rudd M. 2013. Towards sustainable environmental quality: A call to prioritize global research needs. *Integr Environ Assess Manag* 9:179–180.
- Brooks BW, Ankley GT, Hobson JF, Lazorchak JM, Meyerhoff RD, Solomon KR. 2008. Assessing the aquatic hazards of veterinary medicines. In: Crane M, Barrett K, Boxall A, editors. *Effects of veterinary medicines in the environment*. Boca Raton (FL): CRC. p 97–128.
- Brooks BW, Fulton BA, Hanson ML. 2015. Aquatic toxicology studies with macrophytes and algae should balance experimental pragmatism with environmental realism. *Sci Total Environ* 536:406–407.
- Brooks BW, Lazorchak JM, Howard MDA, Johnson MV, Morton SL, Perkins DAK, Reavie ED, Scott GI, Smith SA, Steevens JA. 2016. Are harmful algal blooms becoming the greatest inland water quality threat to public health and aquatic ecosystems? *Environ Toxicol Chem* 35:6–13.
- Brooks BW, Lazorchak JM, Howard MDA, Johnson MV, Morton SL, Perkins DAK, Reavie ED, Scott GI, Smith SA, Steevens JA. 2017. In some places, in some cases and at some times, harmful algal blooms are the greatest threat to inland water quality. *Environ Toxicol Chem* 36:1125–1127.
- Brooks BW, Riley TM, Taylor RD. 2006. Water quality of effluent-dominated ecosystems: ecotoxicological, hydrological, and management considerations. *Hydrobiologia* 556(1):365–379.
- Boulton AJ. 2014. Conservation of ephemeral streams and their ecosystem services: What are we missing? *Aquat Conserv* 24(6):733–738.
- Burden N, Sewell F, Andersen ME, Boobis A, Chipman JK, Cronin MT, Hutchinson TH, Kimber I, Whelan M. 2015. Adverse outcome pathways can drive non-animal approaches for safety assessment. *J Appl Toxicol* 35(9):971–975.
- Cairns J. 1988. Putting the eco into ecotoxicology. *Regul Toxicol Pharmacol* 8:226–238.
- Campana O, Wlodkovic D. 2018. Ecotoxicology goes on a chip: Embracing miniaturized bioanalysis in aquatic risk assessment. *Environ Sci Technol* 52(3):932–946.
- Cañedo-Argüelles M, Ben Kefferd B, Schäfer R. 2018. Salt in freshwaters: Causes, effects and prospects – Introduction to the theme issue. *Philos Trans R Soc B* 374:20180002.
- Caritas. 2018. State of the environment for Oceania 2018. Wellington (NZ). [accessed 2019 Jan 9]. <https://caritas.org.nz/state-environment>
- [CER]. Clean Energy Research. 2019. Clean Energy Council. 82 p. [accessed 2019 Jun 17]. <https://assets.cleanenergycouncil.org.au/documents/resources/reports/clean-energy-australia/clean-energy-australia-report-2019.pdf>
- Chandurvelan R, Marsden ID, Glove CN, Gaw S. 2016. Biomarker responses of mussels exposed to earthquake disturbances. *Estuarine Coastal Shelf Sci* 182:98–111.
- Chariton AA, Sun M, Gibson J, Webb JA, Leung KMY, Hickey CW, Hose GC. 2016. Emergent technologies and analytical approaches for understanding the effects of multiple stressors in aquatic environments. *Mar Freshwater Res* 67(4):414–428.
- Chen JL, Ravindran S, Swift S, Singhal N. 2018. Changes in estrogenicity and micropollutant concentrations across unit processes in a biological wastewater treatment system. *Water Sci Technol* 77(6):1673–1682.
- Chinathamby K, Allinson M, Shiraishi F, Lopata AL, Nugegoda D, Pettigrove V, Allinson G. 2013. Screening for potential effects of endocrine-disrupting chemicals in peri-urban creeks and rivers in Melbourne, Australia using mosquitofish and recombinant receptor-reporter gene assays. *Environ Sci Pollut Res* 20(3):1831–1841.
- Clearwater SJ, Hickey CW, Thompson KJ. 2014. The effect of chronic exposure to phosphorus-inactivation agents on freshwater biota. *Hydrobiologia* 728:51–65.
- Clubbs RL, Brooks BW. 2007. *Daphnia magna* responses to a vertebrate estrogen receptor agonist and an antagonist: A multigenerational study. *Ecotoxicol Environ Saf* 67(3):385–398.
- Coish P, Brooks BW, Gallagher EP, Kavanagh TJ, Voutchkova-Kostal A, Zimmerman JB, Anastas PT. 2016. Current status and future challenges in the molecular design for reduced hazard. *ACS Sustainable Chem Eng* 4:5900–5906.
- Coleman HM, Khan SJ, Watkins G, Stuetz RM. 2008. Fate and analysis of endocrine disrupting chemicals in some sewage treatment plants in Australia. *Water Sci Technol* 58(11):2187–2194.
- Collins F, Gray GN, Bucher JR. 2008. Transforming environmental health protection. *Science* 319:906–907.

- Conine AL, Frost PC. 2016. Variable toxicity of silver nanoparticles to *Daphnia magna*: Effects of algal particles and animal nutrition. *Ecotoxicology* 26:118–126.
- Cooke SJ, Sack L, Franklin CE, Farrell AP, Beardall J, Wikelski M, Chown SL. 2013. What is conservation physiology? Perspectives on an increasingly integrated and essential science. *Conserv Physiol* 1:1–23.
- Corrales J, Kristofco LA, Steele WB, Saari GN, Kostal J, Williams ES, Mills M, Gallagher EP, Kavanagh TJ, Simcox N, Shen LQ. 2016. Toward the design of less hazardous chemicals: Exploring comparative oxidative stress in two common animal models. *Chem Res Toxicol* 30(4):893–904.
- Cote I, Andersen ME, Ankley GT, Barone S, Birnbaum LS, Boelkeheide K, Bois FY, Burgoon LD, Chiu WA, Crawford-Brown D et al. 2016. The next generation of risk assessment multi-year study – Highlights of findings, applications to risk assessment, and future directions. *Environ Health Perspect* 124:1671–1682.
- Cox C, Surgan M. 2006. Unidentified inert ingredients in pesticides: Implications for human and environmental health. *Environ Health Perspect* 114(12):1803–1806.
- Crossland C, Abel K. 1992. New pulp mills in Australia: A perspective. *Mar Pollut Bull* 25(5–8):181–185.
- D'Heygere T, Goethals PLM, De Pauw N. 2003. Use of genetic algorithms to select input variables in decision tree models for the prediction of benthic macroinvertebrates. *Ecol Modell* 160(3):291–300.
- Dafforn KA, Johnston EL, Ferguson A, Humphrey CL, Monk W, Nichols SJ, Simpson SL, Tulbure MG, Baird DJ. 2016. Big data opportunities and challenges for assessing multiple stressors across scales in aquatic ecosystems. *Mar Freshwater Res* 67(4):393–413.
- Datry T, Larned ST, Fritz KM, Bogan MT, Wood PJ, Meyer EI, Santos AN. 2014. Broad-scale patterns of invertebrate richness and community composition in temporary rivers: Effects of flow intermittence. *Ecography* 37(1):94–104.
- Daughton CG. 2014. The Matthew effect and widely prescribed pharmaceuticals lacking environmental monitoring: Case study of an exposure-assessment vulnerability. *Sci Total Environ* 466:315–325.
- Dickson KL, Waller WT, Kennedy JH, Ammann LP. 1992. Assessing the relationship between ambient toxicity and instream biological response. *Environ Toxicol Chem* 11:1307–1322.
- Dix DJ, Houck KA, Martin MT, Richard AM, Setzer RW, Kavlock RJ. 2006. The ToxCast program for prioritizing toxicity testing of environmental chemicals. *Toxicol Sci* 95(1):5–12.
- [EEA] European Environment Agency. 2015. The European environment state and outlook. 2015 Assessment of global megatrends. Copenhagen (DK). 140 p. [accessed 2019 Jan 9]. <https://www.eea.europa.eu/soer-2015/global/action-download-pdf>
- Eerkes-Medrano D, Thompson RC, Aldridge DC. 2015. Microplastics in freshwater systems: A review of the emerging threats, identification of knowledge gaps and prioritization of research needs. *Water Res* 75:63–82.
- Emnet P, Gaw S, Northcott G, Storey B, Graham L. 2015. Personal care products and steroid hormones in the Antarctic coastal environment associated with two Antarctic research stations, McMurdo Station and Scott Base. *Environ Res* 136:331–342.
- Escher BI, Ait-Aïssa S, Behnisch PA, Brack W, Brion F, Brouwer A, Buchinger S, Crawford SE, Du Pasquier D, Hamers T et al. 2018. Effect-based trigger values for in vitro and in vivo bioassays performed on surface water extracts supporting the environmental quality standards (EQS) of the European Water Framework Directive. *Sci Total Environ* 628–629:748–765.
- Escher BI, Allinson M, Altenburger R, Bain A, Balaguer P, Busch W, Crago J, Denslow ND, Dopp E, Hilscherova K et al. 2014. Benchmarking organic micropollutants in wastewater, recycled water and drinking water with in vitro bioassays. *Environ Sci Technol* 48(3):1940–1956.
- Escher BI, Neale PA, Leusch FDL. 2015. Effect-based trigger values for in vitro bioassays: Reading across from existing water quality guideline values. *Water Res* 81:137–148.
- Fairbrother A, Muir D, Solomon KR, Ankley GT, Rudd MA, Boxall AB, Apell JN, Armbrust KL, Blalock BJ, Bowman SR, Campbell LM. 2019. Toward sustainable environmental quality: Priority research questions for North America. *Environ Toxicol Chem* 38(8):1606–1624.
- Fairchild JF, La Point TW, Schwartz TR. 1994. Effects of herbicide and insecticide mixture in aquatic mesocosms. *Arch Environ Contam Toxicol* 27:527–533.
- Furley TH, Brodeur J, Silva de Assis HC, Carriquiriborde P, Chagas KR, Corrales J, Denadai M, Fuchs J, Mascarenhas R, Miglioranza KS et al. 2018. Toward sustainable environmental quality: Identifying priority research questions for Latin America. *Integr Environ Assess Manag* 14(3):344–357.
- Garner WVN. 2013. Born in a tent: How camping makes us Australian. Sydney (AU): NewSouth Publishing. 228 p.
- Gaw S, Brooks BW. 2016. Changing tides: Adaptive monitoring, assessment, and management of pharmaceutical hazards in the environment through time. *Environ Toxicol Chem* 35(4):1037–1042.
- Gissi F, Stauber J, Binet M, Trenfield M, van Dam J, Jolley D. 2018. Assessing the chronic toxicity of nickel to a tropical marine gastropod and two crustaceans. *Ecotoxicol Environ Saf* 159:284–292.
- Gustavsson BM, Magner J, Almroth BC, Eriksson MK, Sturve J, Backhaus T. 2017. Chemical monitoring of Swedish coastal waters indicates common exceedances of environmental thresholds, both for individual substances as well as their mixtures. *Mar Pollut Bull* 122:409–419.
- Hagen TG, Douglas RW. 2014. Comparative chemical sensitivity between marine Australian and Northern Hemisphere ecosystems: Is an uncertainty factor warranted for water-quality-guideline setting? *Environ Toxicol Chem* 33(5):1187–1192.
- Hall J, Golding L. 1998. Standard methods for whole effluent toxicity testing: Development and application. Wellington (NZ): NIWA. MFE 80205. 47 p.
- Harmsworth G, Awatere S, Robb M. 2016. Indigenous Māori values and perspectives to inform freshwater management in Aotearoa-New Zealand. *Ecol Soc* 21(4).
- Henneberg A, Bender K, Blaha L, Giebner S, Kuch B, Köhler H-R, Maier D, Oehlmann J, Richter D, Scheurer M et al. 2014. Are in vitro methods for the detection of endocrine potentials in the aquatic environment predictive for in vivo effects? Outcomes of the Projects SchussenAktiv and SchussenAktivplus in the Lake Constance W, Germany. *PLoS ONE* 9(6):e98307.
- Hintze C, Heydel F, Hoppe C, Cunze S, König A, Tackenberg O. 2013. D3: The dispersal and diaspore database—baseline data and statistics on seed dispersal. *Perspect Plant Ecol* 15(3):180–192.
- Hollender J, Schymanski EL, Singer HP, Ferguson PL. 2017. Nontarget screening with high resolution mass spectrometry in the environment: Ready to go? *Environ Sci Technol* 51:11505–11512.
- Hooper MJ, Ankley GT, Cristol DA, Maryoung LA, Noyes PD, Pinkerton KE. 2013. Interactions between chemical and climate stressors: A role for mechanistic toxicology in assessing climate change risks. *Environ Toxicol Chem* 32(1):32–48.
- Howard PH, Muir DCG. 2010. Identifying new persistent and bioaccumulative organics among chemicals in commerce. *Environ Sci Technol* 44:2277–2285.
- Hutchinson TH, Wheeler JR, Gourmelon A, Burden N. 2016. Promoting the 3Rs to enhance the OECD fish toxicity testing framework. *Regul Toxicol Pharmacol* 76:231–233.
- Jamieson AJ, Malkocs T, Piernney SB, Fujii T, Zhang Z. 2017. Bioaccumulation of persistent organic pollutants in the deepest ocean fauna. *Nature Ecol Evol* 1:0051.
- Jarošová B, Bláha L, Giesy JP, Hilscherová K. 2014. What level of estrogenic activity determined by in vitro assays in municipal waste waters can be considered as safe? *Environ Int* 64:98–109.
- Kaiser KLE. 1998. Correlations of *Vibrio fischeri* bacteria test data with bioassay data for other organisms. *Environ Health Perspect* 106:583–591.
- Kattge J, Diaz S, Lavorel S, Prentice IC, Leadley P, Bönlisch G, Garnier E, Westoby M, Reich PB, Wright IJ, Cornelissen JHC. 2011. TRY—A global database of plant traits. *Global Change Biol* 17(9):2905–2935.
- Kidd KA, Blanchfield PJ, Mills KH, Palace VP, Evans RE, Lazorchak JM, Flick RW. 2007. Collapse of a fish population after exposure to a synthetic estrogen. *Proc Natl Acad Sci USA* 104(21):8897–8901.
- King RS, Brain RA, Back JA, Becker C, Wright MV, Toteu Djomte V, van dam Scott WC, Virgil SR, Brooks BW, Hosmer AJ et al. 2016. Effects of pulsed atrazine exposures on autotrophic community structure, biomass, and

- production in field-based stream mesocosms. *Environ Toxicol Chem* 35(3):660–675.
- Kookana RS, Moggridge B, Mihini R, Gray B, Northcott G, Boxall A. 2013. Pharmaceuticals and personal care products in the environment: Cultural and spiritual perspectives. *Integr Environ Assess Manag* 9(1):164–166.
- Kookana RS, Williams M, Boxall AB, Larsson DJ, Gaw S, Choi K, Yamamoto H, Thatikonda S, Zhu YG, Carriquiriborde P. 2014. Potential ecological footprints of active pharmaceutical ingredients: An examination of risk factors in low-, middle- and high-income countries. *Philos Trans R Soc B* 369(1656):20130586.
- Kostal J, Voutchkova-Kostal A. 2015. CADRE-SS, an in silico tool for predicting skin sensitization potential based on modeling of molecular interactions. *Chem Res Toxicol* 29:58–64.
- LaLone CA, Villeneuve DL, Doering JA, Blackwell BR, Transue TR, Simmons CW, Swintek J, Degitz SJ, Williams AJ, Ankley GT. 2018. Evidence for cross species extrapolation of mammalian-based high-throughput screening assay results. *Environ Sci Technol* 52(23):13960–13971. doi: 10.1021/acs.est.8b04587
- LaLone CA, Villeneuve DL, Lyons D, Helgen HW, Robinson SL, Swintek JA, Saari TW, Ankley GT. 2016. Editor's highlight: Sequence alignment to predict across species susceptibility (SeqAPASS): A web-based tool for addressing the challenges of cross-species extrapolation of chemical toxicity. *Toxicol Sci* 153:228–245.
- Landis W, Durda J, Brooks M, Chapman P, Menzie C, Stahl R, Stauber J. 2013. Ecological risk assessment in the context of global climate change. *Environ Toxicol Chem* 32:79–92.
- La Point TW, Waller WT. 2000. Field assessments in conjunction with WET testing. *Environ Toxicol Chem* 19:14–24.
- Landrigan PJ, Fuller R, Acosta NJ, Adeyi O, Arnold R, Baldé AB, Bertollini R, Bose-O'Reilly S, Boufford JL, Brayne PN, Chiles T. 2018. The Lancet Commission on pollution and health. *The Lancet* 391(10119):462–512.
- Leigh C. 2013. Dry-season changes in macroinvertebrate assemblages of highly seasonal rivers: responses to low flow, no flow and antecedent hydrology. *Hydrobiologia* 703(1):95–112.
- Leitao RP, Zuanon J, Villegier S, Williams SE, Baraloto C, Fortunel C, Mendonca FP, Mouillot D. 2016. Rare species contribute disproportionately to the function structure of species assemblages. *Proc R Soc B* 283:20160084.
- Leusch FDL, Khan SJ, Gagnon MM, Quayle P, Trinh T, Coleman H, Rawson C, Chapman HF, Blair P, Nice H et al. 2014. Assessment of wastewater and recycled water quality: A comparison of lines of evidence from in vitro, in vivo and chemical analyses. *Water Res* 50(0):420–431.
- Leusch FDL, Neale PA, Ansal C, Aneck-Hahn NH, Balaguer P, Bruchet A, Escher BI, Esperanza M, Grimaldi M, Leroy G et al. 2018. Analysis of endocrine activity in drinking water, surface water and treated wastewater from six countries. *Water Res* 139:10–18.
- Leusch FDL, Snyder SA. 2015. Bioanalytical tools: Half a century of application for potable reuse. *Environ Sci Water Res Technol* 1(5):606–621.
- Lewis KA, Tzilivakis J, Warner DJ, Green A. 2016. An international database for pesticide risk assessments and management. *Hum Ecol Risk Assess* 22(4):1050–1064.
- Li S, Villeneuve DL, Berninger JP, Blackwell BR, Cavallin JE, Hughes MN, Jensen KM, Jorgenson Z, Kahl MD, Schroeder AL et al. 2017. An integrated approach for identifying priority contaminants in the Great Lakes Basin – Investigations in the Lower Green Bay/Fox River and Milwaukee Estuaries areas of concern. *Sci Total Environ* 579:825–837.
- Luthy RG, Sedlak DL, Plumlee MH, Austin D, Resh VH. 2015. Wastewater-effluent-dominated streams as ecosystem-management tools in a drier climate. *Front Ecol Environ* 13:477–485.
- Markich S, Camilleri C. 1997. Investigation of metal toxicity to tropical biota: Recommendations for revision of the Australian water quality guidelines. Darwin (AU): Australian Department of Environment and Energy. 94 p. Supervising Scientist Report SSR127.
- Mayer-Pinto M, Johnston EL, Hutchings PA, Marzinielli EM, Ah Yong ST, Birch G, Booth DJ, Creese RG, Doblin MA, Figueira W et al. 2015. Sydney Harbour: A review of anthropogenic impacts on the biodiversity and ecosystem function of one of the world's largest natural harbours. *Mar Freshwater Res* 66(12):1088–1105.
- [MBIE] Ministry of Business Innovation and Employment. 2016. National science challenges. Wellington (NZ). [accessed 2019 Jan 6]. <https://www.mbie.govt.nz/science-and-technology/science-and-innovation/funding-information-and-opportunities/investment-funds/national-science-challenges/>
- McCann KS. 2000. The diversity–stability debate. *Nature* 405:228–233.
- McCrone J. 2017 Dec 2. Feeling Kiwi: A brief history of the development of the NZ identity. Stuff [Wellington NZ]. [accessed 2019 Jan 6]. <https://www.stuff.co.nz/the-press/news/99074312/feeling-kiwi-a-brief-history-of-the-development-of-the-nz-identity>
- McRae NK, Glover CN, Burket SR, Brooks BW, Gaw S. 2018. Acute exposure to an environmentally relevant concentration of diclofenac elicits oxidative stress in the culturally important galaxiid fish *Galaxias maculatus*. *Environ Toxicol Chem* 37(1):224–235.
- McRobb FM, Kufareva I, Abagyan R. 2014. In silico identification and pharmacological evaluation of novel endocrine disrupting chemicals that act via the ligand-binding domain of the estrogen receptor α . *Toxicol Sci* 141(1):188–197.
- Meek ME, Lipscomb JC. 2015. Gaining acceptance for the use of in vitro toxicity assays and QIVIVE in regulatory risk assessment. *Toxicology* 332: 112–123.
- Melvin SD, Wilson SP. 2013. The utility of behavioral studies for aquatic toxicology testing: A meta-analysis. *Chemosphere* 93(10):2217–2223.
- Mispagel C, Allinson G, Allinson M, Shiraiishi F, Nishikawa M, Moore MR. 2009. Observations on the estrogenic activity and concentration of 17 β -estradiol in the discharges of 12 wastewater treatment plants in Southern Australia. *Arch Environ Contam Toxicol* 56(4):631–637.
- Moermond C, Beasley A, Breton R, Junghans M, Laskowski R, Solomon K, Zahner H. 2017. Assessing the reliability of ecotoxicological studies: An overview of current needs and approaches. *Integr Environ Assess Manag* 13:640–651.
- Monty F, Murti R, Furuta N. 2016. Helping nature help us: Transforming disaster risk reduction through ecosystem management. Gland (CH): IUCN. 82 p.
- Myatt GU, Ahlberg E, Akahori Y, Allen D, Amberg A, Anger LT, Aptula A, Auerbach S, Beilke L, Bellion P, Benigni R et al. 2018. In silico toxicology protocols. *Regul Pharmacol Toxicol* 96:1–17.
- [NASEM] National Academies of Sciences, Engineering, and Medicine. 2018. Environmental engineering for the 21st century: Addressing grand challenges. Washington (DC): Natl Academies. 124 p.
- Natsch A, Laue H, Haupt T, von Niederhäusern V, Sanders G. 2018. Accurate prediction of acute fish toxicity of fragrance chemicals with the RTgill-W1 cell assay. *Environ Toxicol Chem* 37(3):931–941.
- Neale PA, Achard MES, Escher BI, Leusch FDL. 2017. Exploring the oxidative stress response mechanism triggered by environmental water samples. *Environ Sci Processes Impacts* 19(9):1126–1133.
- Neale PA, Altenburger R, Ait-Aïssa S, Brion F, Busch W, de Aragão Umbuzeiro G, Denison MS, Du Pasquier D, Hilscherová K, Hollert H et al. 2017. Development of a bioanalytical test battery for water quality monitoring: Fingerprinting identified micropollutants and their contribution to effects in surface water. *Water Res* 123:734–750.
- Nelson NG, Muñoz-Carpena R, Philips EJ, Kaplan D, Sucsy P, Hendrickson J. 2018. Revealing biotic and abiotic controls of harmful algal blooms in a shallow subtropical lake through statistical machine learning. *Environ Sci Technol* 52(6):3527–3535.
- New Zealand. 2018. New Zealand legislation containing Treaty of Waitangi (1840) clauses – correct and update this reference.
- [NOAA] National Oceanic and Atmospheric Administration. 2018. Billion-dollar weather and climate disasters. Asheville (NC): National Centers for Environmental Information (NCEI) US. [accessed 2019 Jan 6]. <https://www.ncdc.noaa.gov/billions/>
- [NRC] National Research Council. 2007. Toxicity testing in the 21st century: A vision and a strategy. Washington (DC). 216 p.
- Obbard RW, Sadris, Wong YQ, Khitun AA, Baker I, Thompson RC. Global warming releases microplastic legacy frozen in Arctic Sea ice. *Earth's Future* 2(6):315–320.
- [OECD] Organisation for Economic Co-operation and Development. 2019. Municipal waste (indicator). Paris (FR). [accessed on 2019 Jun 17]. doi: 10.1787/89d5679a-en

- Ornostay A, Cowie AM, Hindle M, Baker CJO, Martyniuk CJ. 2013. Classifying chemical mode of action using gene networks and machine learning: A case study with the herbicide Linuron. *Comp Biochem Physiol Part D: Genomics Proteomics* 8(4):263–274.
- Owen SF, Giltrow E, Huggett DB, Hutchinson TH, Saye J, Winter MJ, Sumpter JP. 2007. Comparative physiology, pharmacology and toxicology of beta-blockers: Mammals versus fish. *Aquat Toxicol* 82:145–162.
- Palmer MSR. 2008. The Treaty of Waitangi in New Zealand's law and constitution. Wellington (NZ): Victoria University Press.
- Peters A, Merrington G, Leverett D, Wilson I, Schlekot C, Garman E. 2019. Comparison of the chronic toxicity of nickel to temperate and tropical freshwater species. *Environ Toxicol Chem* 38(6):1211–1220.
- Perkins EJ, Habib T, Escalon BL, Cavallin JE, Thomas L, Weberg M, Hughes MN, Jensen KM, Kahl MD, Villeneuve DL et al. 2017. Prioritization of contaminants of emerging concern in wastewater treatment plant discharges using chemical:gene interactions in caged fish. *Environ Sci Technol* 51:8701–8712.
- Pickin J, Randell P, Trinh J, Grant B. 2019. National Waste Report. Canberra (ACT): Australian Department of the Environment and Energy. 126 p. [accessed 2019 Jun 4]. <http://www.environment.gov.au/protection/waste-resource-recovery/national-waste-reports/national-waste-report-2018>
- Posthuma K, Suter GW, Traas TP. 2001. Species sensitivity distributions in ecotoxicology. Boca Raton (FL): CRC. 616 p.
- Potter SH, Becker JS, Johnston DM, Rossiter KP. 2015. An overview of the impacts of the 2010–2011 Canterbury earthquakes. *Int J Disast Risk Re* 14:6–14.
- Renaud FG, Sudmeier-Rieux K, Estrella M, Nehren U, editors. 2016. Ecosystem-based disaster risk reduction and adaptation in practice. New York (NY): Springer. 598 p.
- Riethmuller N, Camilleri C, Franklin N, Hogan AC, King A, Koch A, Markich SJ, Turlay C, van Dam R. 2003. Ecotoxicological testing protocols for Australian tropical freshwater ecosystems. Darwin (AU): Australian Department of Environment and Energy. 140 p. Supervising Scientist Report 173.
- Roberts J, Bain PA, Kumar A, Hepplewhite C, Ellis DJ, Christy AG, Beavis SG. 2015. Tracking multiple modes of endocrine activity in Australia's largest inland sewage treatment plant and effluent-receiving environment using a panel of in vitro bioassays. *Environ Toxicol Chem* 34(10):227–2281.
- Rodriguez A, Zhang H, Klaminder J, Brodin T, Andersson PL, Andersson M. 2018. ToxTrac: A fast and robust software for tracking organisms. *Methods Ecol Evol* 9(3):460–464.
- Rohr JR, Kerby JL, Sih A. 2006. Community ecology as a framework for predicting contaminant effects. *Trends Ecol Evol* 21:606–613.
- Rosal R, Rodea-Palomares I, Boltes K, Fernández-Piñas F, Leganés F, Petre A. 2010. Ecotoxicological assessment of surfactants in the aquatic environment: Combined toxicity of docusate sodium with chlorinated pollutants. *Chemosphere* 81(2):288–293.
- Rosi-Marshall EJ, Royer TV. 2012. Pharmaceutical compounds and ecosystem function: An emerging research challenge for aquatic ecologists. *Ecosystems* 15:867–880.
- Rotroff DM, Wetmore BA, Dix DJ, Ferguson SS, Clewell HJ, Houck A, LeCluyse EL, Andersen ME, Judson RS, Smith CM et al. 2010. Incorporating human dosimetry and exposure into high-throughput in vitro toxicity screening. *Toxicol Sci* 117(2):348–358.
- Rudd MA, Moore AF, Rochberg D, Bianchi-Fossati L, Brown MA, D'Onofrio D, Furman CA, Garcia J, Jordan B, Kline J, Risse LM. 2018. Climate research priorities for policy-makers, practitioners, and scientists in Georgia, USA. *Environ Manage* 62(2):190–209.
- Rudén C, Adams J, Agerstrand M, Brock TCM, Buonsante V, Poulsen V, Schlekot CE, Wheeler JR, Henry TR. 2017. Assessing the relevance of ecotoxicological studies for regulatory decision-making. *Integr Environ Assess Manag* 13:652–663.
- Saaristo M, Brodin T, Balshine S, Bertram MG, Brooks BW, Ehlman SM, McCallum ES, Sih A, Sundin J, Wong BBM, Arnold KE. 2018. Direct and indirect effects of chemical contaminants on the behaviour, ecology and evolution of wildlife. *Proc R Soc B* 285:20181297.
- Salguero-Gómez R, Jones OR, Archer CR, Buckley YM, Che-Castaldo J, Caswell H, Hodgson D et al. 2015. The Compadre Plant matrix database: An open online repository for plant demography. *J Ecol* 103(1):202–218.
- Samanipour S, Langford K, Reid MJ, Thomas KV. 2016. A two stage algorithm for target and suspect analysis of produced water via gas chromatography coupled with high resolution time of flight mass spectrometry. *J Chromatogr A* 1463:153–161.
- Schiel DR, Ross PM, Battershill CN. 2016. Environmental effects of the MV *Rena* shipwreck: Cross-disciplinary investigations of oil and debris impacts on a coastal ecosystem. *New Zeal J Mar Freshwat Res* 50:1–9.
- Schlining K, von Thun S, Kuhn L, Schlining B, Lundsten L, Stout NJ, Chaney L, Connor J. 2013. Debris in the deep: Using a 22-year video annotation database to survey marine litter in Monterey Canyon, central California, USA. *Deep Sea Res Part I* 79(0):96–105.
- Scott PD, Bartkow M, Blockwell SJ, Coleman HM, Khan SJ, Lim R, McDonald JA, Nice H, Nuggeoda D, Pettigrove V et al. 2014. An assessment of endocrine activity in Australian rivers using chemical and in vitro analyses. *Environ Sci Pollut Res* 21(22):12951–12967.
- Sheldon F, Fellows CS. 2010. Water quality in two Australian dryland rivers: Spatial and temporal variability and the role of flow. *Mar Freshwater Res* 61(8):864–874.
- Sinclair CJ, Boxall ABA, Parsons SA, Thomas MR. 2006. Prioritization of pesticide environmental transformation products in drinking water supplies. *Environ Sci Technol* 40:7283–7289.
- Sonneveld E, Pieterse B, Schoonen WG, van der Burg B. 2011. Validation of in vitro screening models for progestagenic activities: Inter-assay comparison and correlation with in vivo activity in rabbits. *Toxicol In Vitro* 25(2):545–554.
- Sonneveld E, Ritco JA, Jansen HJ, Pieterse B, Brouwer A, Schoonen WG, van der Burg B. 2006. Comparison of in vitro and in vivo screening models for androgenic and estrogenic activities. *Toxicol Sci* 89(1):173–187.
- Stauber JL, Gunthorpe L, Munday BL, Deavin J, Ahsanullah M. 1994. Validation of new marine bioassays using synthetic treated and untreated BEKM effluents. Canberra (AU): CSIRO. p 1–58. National Pulp Mills Research Program Technical Report Series No 8.
- Steele WB, Kristofc LA, Corrales J, Saar GN, Haddad P, Gallagher EP, Kavanagh TJ, Kostal J, Zimmerman JB, Voutchkova-Kostal A et al. 2018. Comparative behavioral toxicology with two common larval fish models: Exploring relationships among modes of action and locomotor responses. *Sci Total Environ* 640:1587–1600.
- Steffen W, Richardson K, Rockström J, Cornell SE, Fetzer I, Bennett M, Biggs R, Carpenter SR, De Vries W, De Wit CA et al. 2015. Planetary boundaries: Guiding human development on a changing planet. *Science* 347(6223):1259855.
- Stevenson MG. 1996. Indigenous knowledge in environmental assessment. *Artic* 49(3):278–291.
- Suter GW. 2006. Ecological risk assessment and ecological epidemiology for contaminated sites. *Hum Ecol Risk Assess* 12(1):31–38.
- Suter II GW. 2016. Ecological risk assessment. Boca Raton (FL): CRC Press. 680 p.
- Sutherland WJ, Woodroof H. 2009. The need for environmental horizon scanning. *Trends Ecol Evol* 24(10):523–527.
- Sutherland WJ, Bardsley S, Bennun L, Clout M, Côté IM, Depledge MH, Dicks LV, Dobson AP, Fellman L, Fleishman E, Gibbons DW. 2011. Horizon scan of global conservation issues for 2011. *Trends Ecol Evol* 26(1):10–16.
- Syberg K, Backhaus T, Banta G, Bruce P, Gustavsson M, Munns WR, Ramo R, Selck H, Gunnarsson JS. 2017. Toward a conceptual approach for assessing risks from chemical mixtures and other stressors to coastal ecosystem services. *Integr Environ Assess Manag* 13:376–386.
- Tanneberger K, Knöbel M, Busser FJM, Sinnige TL, Hermens JLM, Schirmer K. 2013. Predicting fish acute toxicity using a fish gill cell line-based toxicity assay. *Environ Sci Technol* 47(2):1110–1119.
- Taylor JM, Back JA, Brooks BW, King RS. 2018. Spatial, temporal, and experimental: Three study-design cornerstones for establishing defensible numeric criteria for freshwater ecosystems. *J Appl Ecol* 55:2114–2123.
- Tremblay LA, Clark D, Sinner J, Ellis JI. 2017. Integration of community structure data reveals observable effects below sediment guideline thresholds in a large estuary. *Environ Sci: Processes Impacts* 19(9):1134–1141.

- [UN] United Nations. 1992. Rio Declaration.
- [UN] United Nations. 2007. Declaration on the Rights of Indigenous People.
- [UN] United Nations. 2015. Transforming our world: The 2030 Agenda for Sustainable Development. http://www.un.org/ga/search/view_doc.asp?symbol=A/RES/70/1&Lang=E70/1
- [UNDRR] UN Office for Disaster Risk Reduction. 2015. Sendai Framework for Disaster Risk Reduction 2015-2030. [accessed 2019 May 1]. <https://www.unisdr.org/we/coordinate/sendai-framework>
- [UNEP] United Nations Environment Program. 2015. Global waste management outlook. New York (NY).
- [USEPA] United States Environmental Protection Agency. 1991. Methods for aquatic toxicity identification evaluation: Phase 1 toxicity characterization procedures. 2nd ed. Washington (DC): Office of Research and Development. EPA-600-6-91-003. 87 p.
- Van Cauwenberghe L, Vanreusel A, Mees J, Janssen CR. 2013. Microplastic pollution in deep-sea sediments. *Environ Pollut* 182:495–499.
- van Dam JW, Trenfield MA, Stretten C, Harford AJ, Parry D, van Dam RA. 2018. Assessing chronic toxicity of aluminium, gallium and molybdenum in tropical marine waters using a novel bioassay for larvae of the hermit crab *Coenobita variabilis*. *Ecotoxicol Environ Saf* 165:349–356.
- van Dam RA, Harford AJ, Houston MA, Hogan AC, Negri AP. 2008. Tropical marine toxicity testing in Australia: A review and recommendations. *Australas J Ecotoxicol* 14(2/3):55.
- Van den Brink PJ, Baird DJ, Baveco HJM, Fochs A. 2013. The use of traits-based approaches and eco(toxico)logical models to advance the ecological risk assessment framework for chemicals. *Integr Environ Assess Manag* 9:47–57.
- Van den Brink PJ, Boxall AB, Maltby L, Brooks BW, Rudd MA, Backhaus T, Spurgeon D, Verougstraete V, Ajao C, Ankley GT et al. 2018. Toward sustainable environmental quality: Priority research questions for Europe. *Environ Toxicol Chem* 37(9):2281–2295.
- Van den Brink PJ, Choung CB, Landis W, Mayer-Pinto M, Pettigrove V, Scanes P, Smith R, Stauber J. 2016. New approaches to the ecological risk assessment of multiple stressors. *Mar Freshwater Res* 67(4):429–439.
- van der Oost R, Sileno G, Suárez-Muñoz M, Nguyen MT, Besselink H, Brouwer A. 2017. SIMONI (Smart Integrated Monitoring) as a novel bio-analytical strategy for water quality assessment: Part I—model design and effect-based trigger values. *Environ Toxicol Chem* 36(9):2385–2399.
- Villeneuve DL, Coady K, Escher BI, Mihaich E, Murphy CA, Schlekot T, Garcia-Reyero N. 2019. High-throughput screening and environmental risk assessment: State of the science and emerging applications. *Environ Toxicol Chem* 38:12–26.
- Vincent K, Davidson C. 2015. The toxicity of glyphosate alone and glyphosate–surfactant mixtures to western toad (*Anaxyrus boreas*) tadpoles. *Environ Toxicol Chem* 34(12):2791–2795.
- Waller WT, Allen HJ. 2008. Acute and chronic toxicity. In: Jorgensen SE, Fath BD, editors. Encyclopedia of ecology. Oxford (UK): Elsevier. p 32–43.
- Warne MSJ, Hawker DW. 1995. The number of components in a mixture determines whether synergistic and antagonistic or additive toxicity predominate: The funnel hypothesis. *Ecotoxicol Environ Saf* 31(1):23–28.
- Weeks ES, Death RG, Foote K, Anderson-Lederer R, Joy MK, Boyce P. 2016. Conservation science statement. The demise of New Zealand's freshwater flora and fauna: A forgotten treasure. *Pac Conserv Biol* 22(2):110–115.
- Wetmore BA. 2015. Quantitative in vitro-to-in vivo extrapolation in a high-throughput environment. *Toxicology* 332:94–101.
- Wilson DC, Rodic L, Modak P, Soos R, Carpintero A, Velis K, Iyer M, Simonett O. 2015. Global waste management outlook. Osaka (JP): UNEP. 331 p.
- [WWAP]. United Nations World Water Assessment Programme. 2017. The United Nations World Water Development Report 2017: Wastewater, The Untapped Resource. Paris (FR): UNESCO. 180 p.
- Yang J, Jeppe KJ, Pettigrove VJ, Zhang X. 2018. eDNA metabarcoding supporting community assessment of environmental stressor in a field-based sediment microcosm study. *Environ Sci Technol* 52:14469–14479.
- Zala SM, Penn DJ. 2004. Abnormal behaviours induced by chemical pollution: A review of the evidence and new challenges. *Anim Behav* 68(4):649–664.
- Zhang X, Xia P, Wang P, Yang J, Baird DJ. 2018. Omics advances in ecotoxicology. *Environ Sci Technol* 52:3842–3851.