

Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Zoneff, E;Wang, Y;Jackson, C;Smith, O;Duchi, S;Onofrillo, C;Farrugia, B;Moulton, SE;Williams, R;Parish, C;Nisbet, DR;Caballero-Aguilar, LM

Title:

Controlled oxygen delivery to power tissue regeneration

Date:

2024-12-01

Citation:

Zoneff, E., Wang, Y., Jackson, C., Smith, O., Duchi, S., Onofrillo, C., Farrugia, B., Moulton, S. E., Williams, R., Parish, C., Nisbet, D. R. & Caballero-Aguilar, L. M. (2024). Controlled oxygen delivery to power tissue regeneration. *Nature Communications*, 15 (1), <https://doi.org/10.1038/s41467-024-48719-x>.

Persistent Link:

<https://hdl.handle.net/11343/353697>

License:

[CC BY](#)










Controlled oxygen delivery to power tissue regeneration

Received: 7 November 2023

Accepted: 13 May 2024

Published online: 22 May 2024

 Check for updates

Elizabeth Zoneff^{1,2}, Yi Wang ^{1,2}, Colin Jackson ^{3,4}, Oliver Smith ^{3,4},
Serena Duchi^{5,6}, Carmine Onofrillo^{5,6}, Brooke Farrugia², Simon E. Moulton ^{6,7,8},
Richard Williams ⁹, Clare Parish ¹⁰, David R. Nisbet ^{1,2,11}  &
Lilith M. Caballero-Aguilar^{1,2,6} 

Oxygen plays a crucial role in human embryogenesis, homeostasis, and tissue regeneration. Emerging engineered regenerative solutions call for novel oxygen delivery systems. To become a reality, these systems must consider physiological processes, oxygen release mechanisms and the target application. In this review, we explore the biological relevance of oxygen at both a cellular and tissue level, and the importance of its controlled delivery via engineered biomaterials and devices. Recent advances and upcoming trends in the field are also discussed with a focus on tissue-engineered constructs that could meet metabolic demands to facilitate regeneration.

The base unit of the body are cells, and cells require energy. As tissues and organs become more complex, the utilities that ensure their metabolic demands are met increase in capacity and sophistication. A challenge hindering the translation of tissue engineering and regenerative medicine into clinical practice is ensuring an engineered solution for the mass transport of nutrients and molecular oxygen to cells.

For engineered tissues, the size of the bioengineered tissue that can be fabricated ex-situ or in situ is limited by poor oxygen transport from physiological environments into newly implanted constructs. In the absence of a vascular system, a reliance on the diffusion of molecules throughout bioengineered tissues or bioscaffolds might not be rapid enough to satisfy the vital oxygen demand¹. Challenges arise when trying to control oxygen tension, as it decreases towards the center of the graft or bioscaffold. Over time, the effects of hypoxia and accumulation of waste products, can culminate in central cell death, resulting in a necrotic core^{2,3}.

Mimicking the complex geometries of the vascular network and achieving complete integration with host tissue is a vital future step

towards truly engineered tissue. This is because it not only enables the distribution of oxygen, but also facilitates enhanced nutrient exchange and removal of metabolic waste. Efforts to replicate the host tissue vasculature involve stimulating blood vessel infiltration into the tissue through growth factor up-regulation and/or supplementation and/or fabricating perfusable template channels that can direct endothelial progenitors to form organized vascular networks^{4,5}. However, not only do these require time to mature, but incomplete angiogenesis is also associated with ischemic conditions, and the fragile networks observed from overvascularisation can be similarly detrimental⁶.

Oxygen is transported from gases within the alveola to tissues by hemoglobin. Inspired by this, an alternative approach is the development of oxygen-generating biomaterials and devices that have the potential to provide a depot of oxygen species to replenish oxygen on demand. For instance, tissue scaffolds that have been functionalised with peroxides and perfluorocarbons show evidence of controlled and prolonged oxygen delivery (>12 days in vitro)^{7,8}. Although there remain several challenges that need to be addressed before oxygen-

¹The Graeme Clark Institute, The University of Melbourne, Parkville, Melbourne, VIC, Australia. ²Department of Biomedical Engineering, Faculty of Engineering and Information Technology, The University of Melbourne, Parkville, Melbourne, VIC, Australia. ³Research School of Chemistry, Australian National University, Canberra, ACT, Australia. ⁴ARC Centre of Excellence in Synthetic Biology, Australian National University, Canberra, ACT, Australia. ⁵Department of Surgery, Faculty of Medicine, Dentistry and Health Science, The University of Melbourne, Melbourne, VIC, Australia. ⁶Aikenhead Centre for Medical Discovery, St. Vincent's Hospital, Melbourne, VIC, Australia. ⁷Department of Engineering Technologies, Swinburne University of Technology, Melbourne, VIC, Australia. ⁸Iverson Health Innovation Research Institute, Swinburne University of Technology, Melbourne, VIC, Australia. ⁹IMPACT, School of Medicine, Deakin University, Geelong, VIC, Australia. ¹⁰The Florey Institute, The University of Melbourne, Melbourne, VIC, Australia. ¹¹Melbourne Medical School, Faculty of Medicine, Dentistry and Health Science, The University of Melbourne, Melbourne, VIC, Australia.  e-mail: david.nisbet@unimelb.edu.au; l.caballeroaguilar@unimelb.edu.au

generating biomaterials can become clinically widespread, recent developments are exceptionally promising Fig. 1.

In this review, we explore the fundamental biological role of oxygen and subsequently focus on past and current efforts to deliver oxygen to engineered tissues for regeneration to alleviate the limitations of passive diffusion. Both biological and non-biological approaches to oxygen delivery are considered, and the future scope of both biomolecule engineering and materials engineering are discussed in this context.

The pivotal role of oxygen in mammalian events

Oxygen regulation undergoes dynamic adjustments throughout embryogenesis and maturation events. Through the years, the study of physiological processes has provided stepping stones in understanding how oxygen is regulated from the cellular level to organogenesis and tissue repair. The understanding of this sophisticated relationship between tissue development and oxygen has led to new evidence-based clinical practices; for example, during embryonic development, there has been a shift from the previously endorsed low oxygen tension (5% O₂, or 38 mmHg)^{9,10} due to recent contrasting evidence^{11,12}. This example underscores the need for ongoing exploration and understanding of oxygen's role in key events such as cellular respiration, hypoxia, development, tissue damage, and repair. We provide a brief introduction to these events in the following subsections.

Function of oxygen in development

Oxygen is tightly regulated through human development and adult homeostasis. First identified in the 1970s, the link between mammalian embryogenesis and oxygen levels encompasses two primary phases. Initial low oxygen levels (8–38 mmHg) support embryogenesis, promoting cell proliferation and maintain embryonic cells in an undifferentiated state (Fig. 2). This phase is guided by hypoxia-inducible factors (HIF) and transcription genes like vascular endothelial growth factor (VEGF). Oxygen levels then rise almost 2-fold after placental formation, optimizing maternal-fetal oxygen and nutrient transfer and securing adequate oxygen delivery to the growing organism¹³.

During early post-natal development, aerobic respiration is the primary oxygen source, with the respiratory system facilitating oxygen transport to organs and tissues at various degrees. Oxygen levels fluctuate between 7.5 and 100 mmHg in mature and healthy mammalian tissues¹⁴. Hypoxia and hyperoxia significantly influence development, and dysregulation of the oxygen gradients can hinder organogenesis¹⁵, with imbalances leading to potential developmental challenges¹⁶, respiratory issues¹⁷, low birth weight¹⁸, and impairments in cerebrovascular function and neurogenesis¹⁹. This highlights the delicate balance between oxygen-driving gradients and tissue homeostasis.

Oxygen upon tissue damage and its role in repair

The immediate response of mature tissue to injury involves immune cell activation and increased blood flow to the injury site. Oxygen is a cornerstone of cellular activities associated with regeneration: (i) in the proliferation phase, it is required for glycoprotein biosynthesis²⁰, and (ii) during the maturation or remodeling phase, it is required as a co-factor for hydroxyproline (key to forming fibrillar collagens)²¹. The enzymes that are essential for collagen stability, prolyl and lysyl hydroxylases, utilize oxygen as a co-substrate, after hydroxylation, the hydroxylysine residues undergo oxygen dependent- glycosylation, which facilitates cross-linking between collagen molecules. An imbalance of oxygen during the hydroxylation process can lead to unstable collagen formation (for more information about this process, refer to: ref. 22). In the repair of tissue that relies heavily on collagen formation and where the oxygen tension is lower than in other tissues, such as in cartilaginous tissue, the role of available oxygen may be critical^{23,24}.

Failure to repair can open the path for degenerative diseases and chronic pain.

After injury resolution, the healing-promoting inflammatory microenvironments typically return to homeostasis. However, persistent stimuli can escalate acute inflammation towards a chronic state, potentially leading to degenerative diseases²⁵. Inflammation can paradoxically induce hypoxia (despite the increased demand for oxygen following an injury), a cycle frequently seen in ischemic injuries, where inflammation is both accompanied and triggered by hypoxia^{26,27}. Injury-driven hypoxia is common in ischemic injuries, blood vessel damage and thrombotic scenarios.

In diseases characterized by prolonged inflammation, severe oxygen deprivation can occur in affected tissues. For example, in inflammatory bowel disease, the intestinal epithelial cells undergo an inflammatory-induced hypoxia²⁸. In such hypoxic scenarios, HIFs, which are mediators acting through post-translational hydroxylation of proline residues via prolyl hydroxylase (PHD) enzymes, stabilize by translocating to the cell nucleus where they bind hypoxia response elements (HREs)²⁶ (Fig. 2). Indeed, recent studies have demonstrated that HIF stabilization through targeted PHD inhibition is an emerging strategy to treat diseases like inflammatory bowel disease.

The challenge of providing oxygen to cells for tissue repair

In native tissues, oxygen distribution is guided by structured capillary networks, creating gradients that cater to the varying blood and oxygen demands of different organs. As a result, some are avascular while others are highly vascularized^{29–31}. In bioengineered tissues, mimetics of native oxygen gradients remain under investigation. Two key challenges are (i) the lack of interconnection between an engineered vasculature and the host tissue and (ii) the absence of an adequate oxygen supply post-implantation. The limited oxygen diffusion length from native capillaries (typically ~100–200 μm)³², often constrains oxygenation of cells nestled deep within newly implanted scaffolds. This problem aggravates within large scaffolds, thereby limiting the reach of tissue engineering constructs in repairing large defects³³.

However, straightforward augmentation of oxygen supply in the constructs will not yield a sustainable general solution because high concentrations of oxygen (hyperoxia) can inadvertently lead to ROS overproduction³⁴ (Fig. 3). Cellular oxygen demand also varies among cell types. For example, the oxygen demand of hepatocytes (rate of consumption 0.3 nmol/s/10⁶) has been reported to be up to 5-fold higher than that of fibroblasts (rate of consumption 0.05 nmol/s/10⁶) due to their high metabolic activity³⁵. In addition, this oxygen-metabolic dependent demand fluctuates according to the cell's cycle phase (e.g., stromal vs terminally differentiated)³⁶ and the cell number within the scaffold³⁷. The interplay of hypoxia and hyperoxia introduces another layer of complexity to tissue engineering. Short bursts of hyperoxia have been shown to protect against hypoxic damage, a phenomenon termed 'ischemic preconditioning'³⁸. This compensatory mechanism, however, has a narrow therapeutic window as prolonged exposure remains detrimental. Thus, oxygen delivery in the context of tissue engineering must optimize oxygen levels to meet cellular demands while navigating the delicate balance between hypoxia and hyperoxia.

Biological molecules as oxygen-controlling strategies

The term "biomolecule" refers to any molecule that is naturally derived from an organism, including, but not limited to, proteins, peptides, nucleic acids and their derivatives, fatty acids, and lipids. In recent years, tissue engineering has focused on developing and identifying materials that can stabilize and provide controlled release of various biomolecules in support of grafted cells or to elicit a desired response from host tissue. The most proven biomolecules in this context are hemoproteins, which can reversibly bind oxygen, and other proteins

involved in intracellular oxygen regulation, such as those that scavenge ROS.

Hemoglobin and hemoglobin-engineered molecules

Hemoglobin is the most widely recognized oxygen-binding biomolecule because of its role in respiration. It is expressed in red blood cells (RBCs), where it binds oxygen in the lungs and circulates throughout the body via the bloodstream, releasing oxygen in tissue capillaries³⁹.

Hemoglobin functions as a heterotetramer (a protein complex with four non-identical subunits) that consists of two $\alpha\beta$ dimers ($\alpha1\beta1$, $\alpha2\beta2$). Each subunit coordinates one heme cofactor at an identical coordination site, so every functional hemoglobin complex can reversibly bind up to four oxygen molecules simultaneously (Fig. 4).

The reversibility of oxygen binding makes hemoglobin an attractive target for biomedical oxygen delivery. Hemoglobin-based oxygen carriers (HBOCs) have been investigated as blood substitutes in order to reduce dependence on allogeneic RBC transfusions^{40,41}. This was initiated by Amberson et al. in 1934, who isolated hemoglobin from hemolysed beef RBCs to create a Ringer-Locke solution that, when provided as a blood substitute into cats, allowed survival for up to 36 h⁴². Though promising, many initial HBOCs suffered from toxicity and side effects. Clinical studies of cell-free HBOCs have caused renal toxicity in patients⁴³, due to the formation of hemoglobin $\alpha\beta$ heterodimers that are cleared through renal catabolism⁴⁴.

The first HBOC approved for clinical trial by the FDA was HemAssist, which utilized a fumarate crosslinker to crosslink the two α -subunits of hemoglobin purified from human blood banks. It was

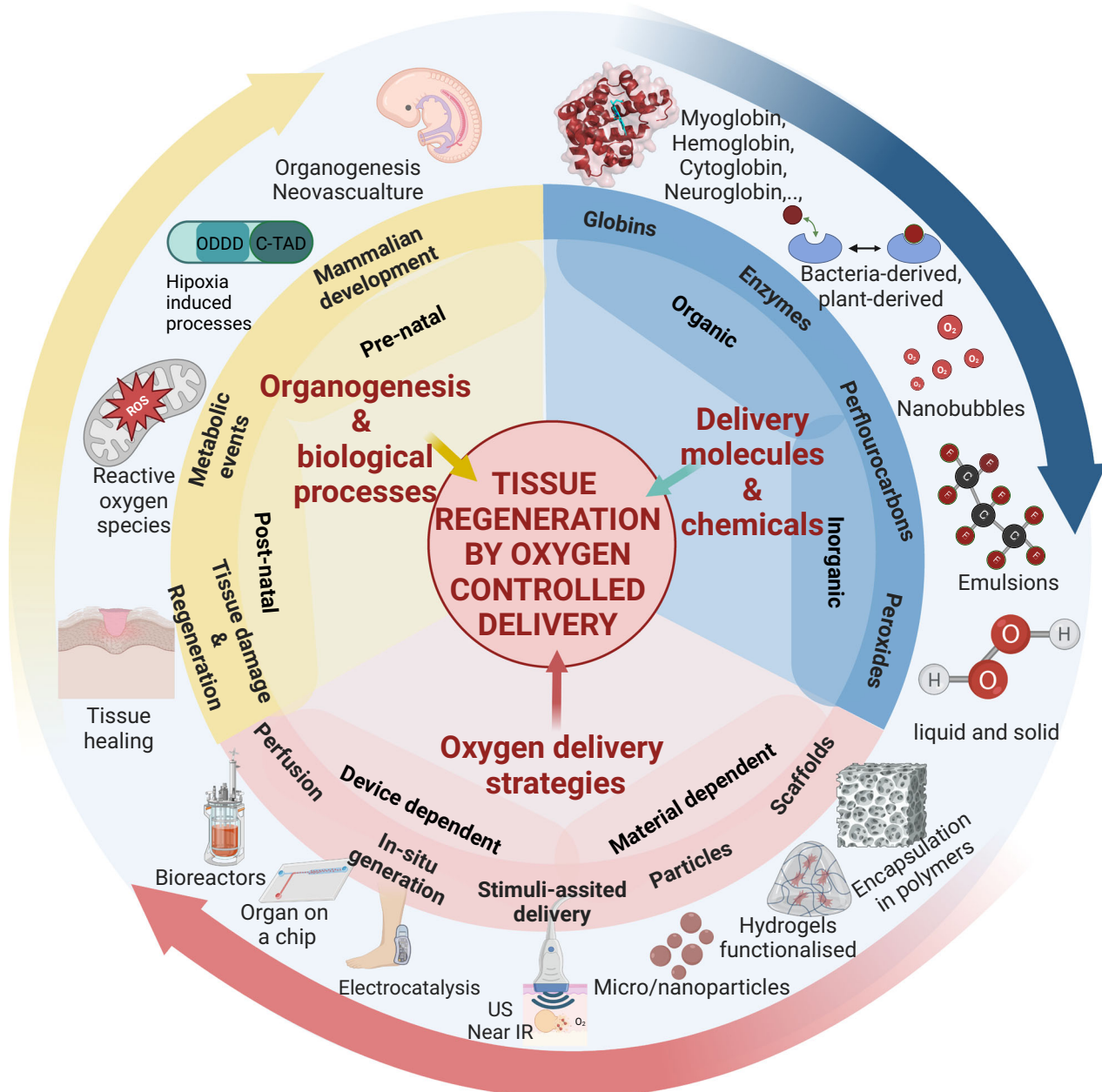


Fig. 1 | The key role of oxygen in driving tissue regeneration is an interplay between biological considerations, delivery from biological molecules and chemicals, and the delivery strategy. Organogenesis and biological processes can be classified according to the developmental stage (pre-natal and post-natal), with events such as tissue regeneration occurring at the post-natal stage. Delivery

strategies can be device- or material-dependent, with some strategies overlapping these two. Molecules and chemicals that produce oxygen can be naturally derived (organic) or inorganic (such as peroxides). Figure created with BioRender.com, released under a Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International license.

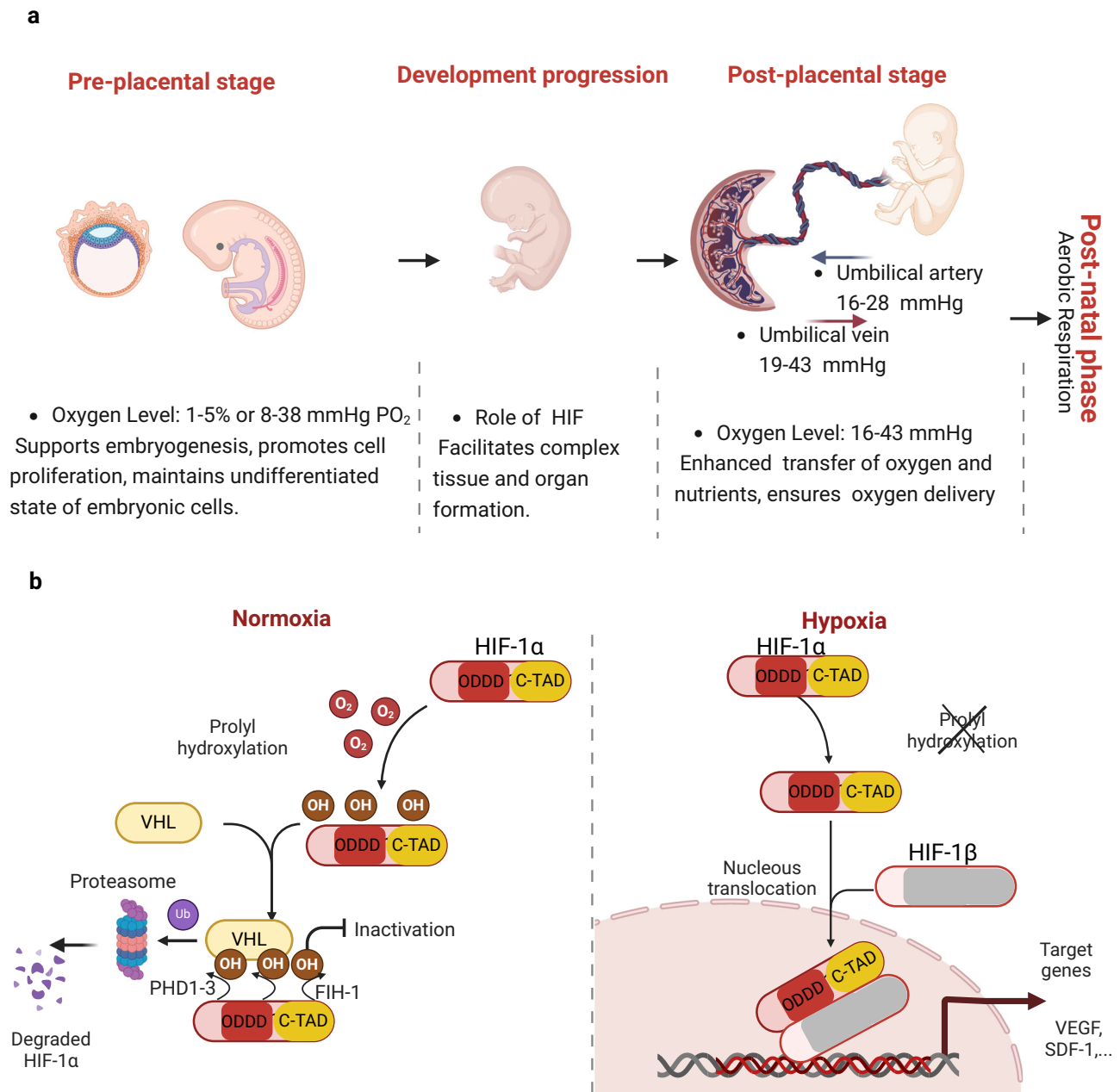


Fig. 2 | Oxygen in development and HIF-1 α regulation. **a** The role of oxygen in development starts from embryonic development up to post-natal phase. Early hypoxic conditions trigger HIF1- α accumulation, thus targeting genes associated with vascularization (such as VEGF) to encourage organ formation and placental-fetus interactions. After the placenta is formed, oxygen levels rise up to 38 mmHg. **b** During normoxia, HIF-1 α is regulated by prolyl hydroxylation. The HIF1- α subunits contain three sites for hydroxylation (two on prolyl residues within the oxygen-dependent degradation domain (ODDD) and one on an asparaginyl residue within the C-terminal transactivation domain (C-TAD)). Upon oxygen-dependent Fe(II), oxygen-, and 2-oxoglutarate, PHDs facilitate prolyl hydroxylation. This is then

recognized by the von Hippel-Lindau protein (pVHL), which then marks it for degradation through a process called ubiquitination, leading to its breakdown by the proteasome. Separately, the asparaginyl hydroxylation process is driven by an enzyme called factor-inhibiting HIF (FIH) acting on a specific site in the C-TAD. In hypoxia, when the hydroxylation is suppressed or PHDs are inhibited, HIF-1 α is translocated into the nucleus, where it heterodimerises with HIF1-b subunit allowing binding to hypoxia-response elements (HREs). Adapted from¹²⁴. Figure created with BioRender.com, released under a Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International license.

ultimately discontinued after a Phase III trial due to an increased risk of pancreatitis and myocardial infarction⁴⁵. Other potential issues include nitric oxide (NO)-scavenging and met-hemoglobin formation, causing hypertension and vasoconstriction. Recent advances in HBOCs include its use to promote wound healing, particularly for diabetic ulcers. For instance, 'Granulox' is a topical spray consisting of purified porcine hemoglobin that has clinically demonstrated improved healing in poorly oxygenated wounds⁴⁶. Globin-modified biomaterials have also been tested for their neuroprotective/regenerative potential in

preclinical models (namely traumatic brain injury and stroke). These have included regional reperfusion using HemoAct (a core-shell structured hemoglobin-albumin cluster)⁴⁷ and delivery of HBOCs, including modified nanoparticles thereof containing the catalase enzyme to provide additive benefits⁴⁸ and polynitroxylated polyethylene glycol (PEG)-ylated hemoglobin. However, in the latter, as with previous studies of HBOCs, deployment in larger species (*pig*) resulted in hypertension and cardiac arrest, questioning the future utility of this approach or need for further development⁴⁹.

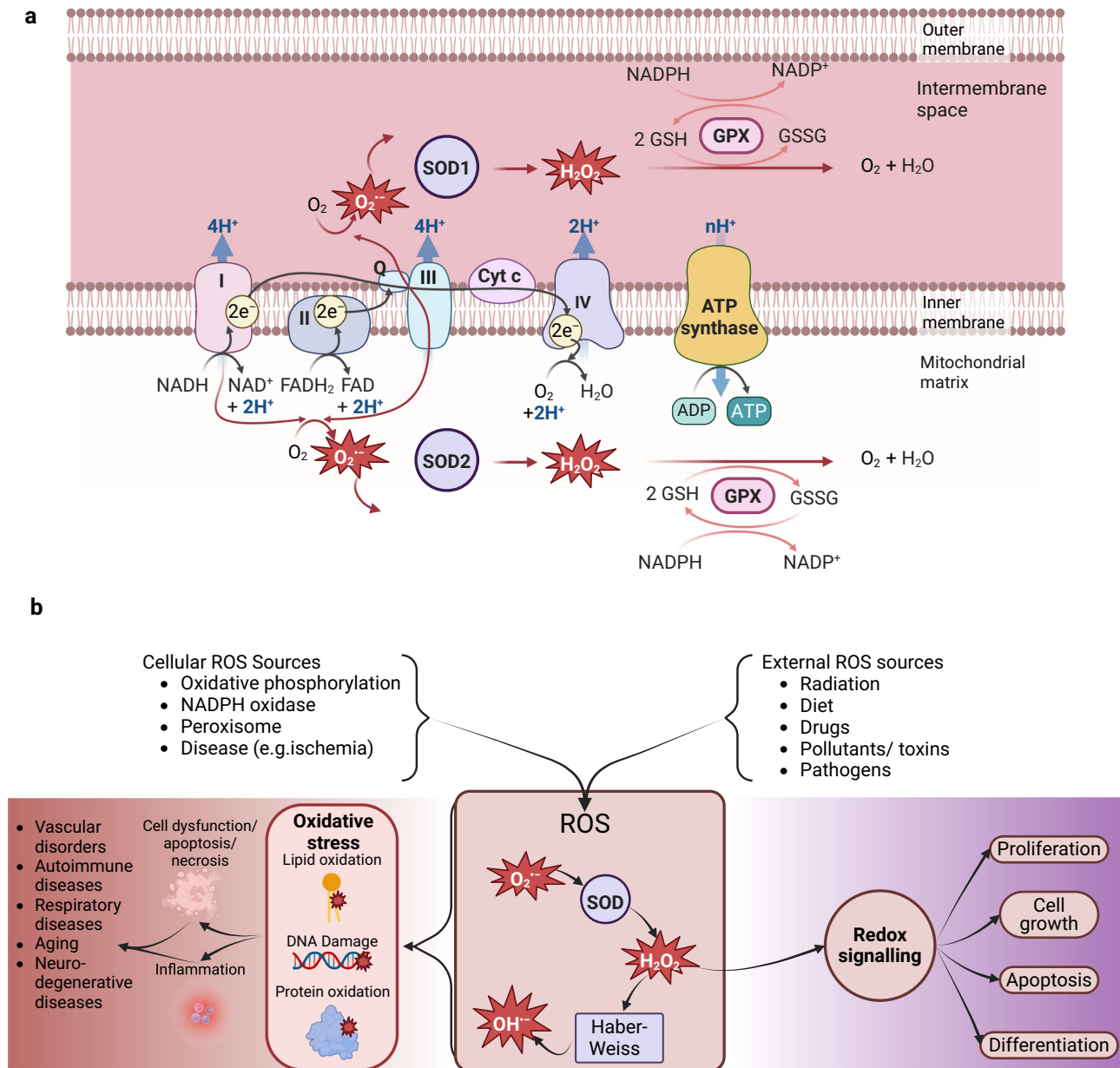


Fig. 3 | Oxidative phosphorylation, ROS generation, pathology, and redox signaling. **a** During oxidative phosphorylation, electron carriers NAD⁺ and FAD feed electrons to the ETC at either complex I or II. The electrons are shuttled through four membrane-bound protein complexes, referred to as complexes I-IV, which undergo a series of redox reactions. Through this process electrons from complexes I and II also enter the 'Q-cycle', where they are passed to another electron carrier, ubiquinone (Q), to facilitate transfer to cytochrome c (via complex III as an intermediary). Diatomic oxygen binds to Complex IV, where it accepts the electrons carried by cytochrome C and is reduced to water. Energy released through these reactions facilitates an electrochemical proton gradient, or mitochondrial membrane potential, within the intermembrane space. To relieve this membrane potential, protons can reenter into the mitochondrial matrix through

another protein complex, ATP synthase, which harnesses the mechanical energy generated by the movement of protons to overcome energy constrictions limiting the conversion of adenosine diphosphate (ADP) into ATP. "Leaky" electrons result in the generation of ROS. Adapted from ref. 125. **b** Oxygen is involved in both redox signaling and pathology in the form of ROS. A number of sources, both endogenous and exogenous, are capable of generating cellular ROS. H₂O₂ is involved in the signal transduction of pathways that control cell growth, proliferation, apoptosis, and differentiation. However, most ROS species, particularly HO⁻, facilitate oxidative stress, cell damage, inflammation and, subsequently, disease. Figure created with BioRender.com, released under a Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International license.

To alleviate these limitations, current work on HBOCs focuses on (i) reducing the toxicity of free hemoglobin, (ii) limiting oxidation to met-hemoglobin without comprising oxygen binding and (iii) exploring HBOC modifications such as polymerization, crosslinking and genetic engineering to increase hemoglobin half-life while in intravascular circulation⁵⁰. For example, Jansman et al., have developed a nanoparticle solution that encapsulates hemoglobin in poly(lactide-co-glycolide) (PLGA) decorated with cerium oxide, which acts as an

enzyme mimic to scavenge ROS. To improve the circulating half-life of the nanoparticle and reduce sequestering into the liver or spleen, the surface was further functionalised with components derived from RBC membranes⁵¹. In vitro studies of the nanoparticles demonstrated biocompatibility, ROS scavenging capability, preserved protein function and limited endothelial cell uptake. However, further investigation using appropriate in vivo models is required to elucidate whether such materials will avoid the clinical challenges of earlier HBOCs.

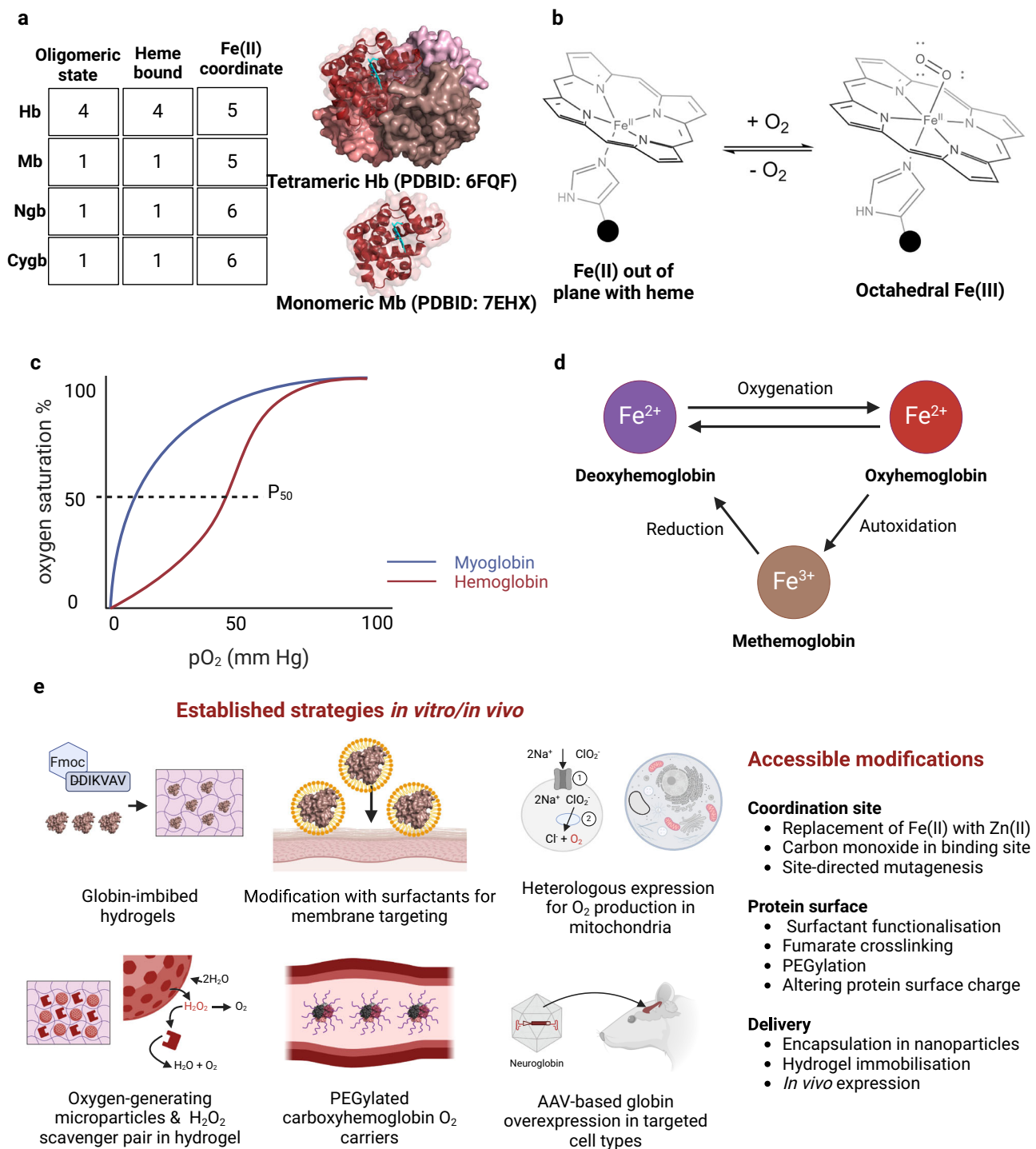


Fig. 4 | General biochemical and redox properties of globins and delivery strategies. **a** The oligomeric state and heme coordination properties of all four human globins. The X-ray crystal structure of tetrameric hemoglobin (Hb) displays the b1 subunit as a ribbon structure, while the entire structure of monomeric myoglobin (Mb) is shown as a ribbon, positioned relative to Hb b1. The coordinated heme is highlighted in cyan in both structures. **b** Structural properties of five-coordinate heme-bound Fe(II) (Hb and Mb) binding to oxygen. In the deoxy conformation, Fe(II) sits out of plane from the tetradentate heme. Oxygen binding

facilitates shift to stable, tetrahedral geometry in oxyhemoglobin. **c** oxygen binding properties of Hb and Mb. Hb exhibits cooperative binding as the oligomers transition from T-state to R-state after one oxygen binds. Mb exhibits hyperbolic binding because it functions as a monomer. **d** Redox properties of oxygen coordination to heme. **e** Strategies adopted to modify or delivery globins for use as an artificial blood supply. Figure created with BioRender.com, released under a Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International license.

A secondary function of hemoglobin is to facilitate the transport of waste gas from tissue to the lungs. This property of hemoglobin remains somewhat overlooked in the context of HBOCs, and could present an opportunity for further investigation, especially when considering the need for metabolic waste sequestration, removal, and

the effect on the acid-base balance in engineered tissues⁵². One area where carbon dioxide (CO₂, waste/by-product of metabolism) content has been investigated is in applications such as organ preservation via machine perfusion⁵³ and severe hemorrhagic shock models⁵⁴. Here, glutaraldehyde polymerized HBOCs were utilized as blood substitutes.

In the latter example, Chang's group^{55–57}, crosslinked the poly-hemoglobin (polyHb) with antioxidant enzymes SOD and CAT, and carbonic anhydrase (CA). CA catalyzes the hydration of CO₂, to boost efficiency of CO₂ transport away from tissues. The polyHb-SOD-CAT-CA complex demonstrated lower intracellular pCO₂ (68.6 ± 3 mmHg) compared with lactated Ringer's solution (98 ± 4.5 mmHg) and polyHb (90.1 ± 4.0 mmHg)⁵⁴. Such work illustrates that despite the potential of HBOCs to concurrently transport oxygen and waste CO₂, hemoglobin on its own may not be sufficient to maintain acid-base balance (hemoglobin transport accounts for only 23% of sequestered CO₂ in the body, with the 70% of the remaining catalyzed by CA⁵⁷). However, caution should be taken when chemically modifying and crosslinking enzymes, as such modifications can result in a decrease in functional activity.

Myoglobin and myoglobin-engineered molecules

Myoglobin is another important hemoprotein that is endogenous to most vertebrates and reversibly binds oxygen. Myoglobin shares the eight alpha-helical structural motif of hemoglobin subunits but is distinct in that it is a monomer functioning as an oxygen reservoir. It has higher oxygen affinity than hemoglobin, with P₅₀ (pO₂ at half oxygen saturation) values of approximately 2–3 mmHg⁵⁸ and 27 mmHg⁵⁹, respectively. Expressed in cardiac and skeletal muscle, myoglobin binds oxygen transferred from arterial hemoglobin because of this affinity difference. Oxygen release from myoglobin to the mitochondria of myocytes will occur when demand is high, like during aerobic exercise⁶⁰.

Myoglobin can be exploited for oxygen storage to mediate hypoxic conditions in bioengineered tissues, usually through complexation or immobilization. Some examples of myoglobin being applied in this context include studies of its covalent immobilization and inclusion within sol/gel films^{61,62}. Engineered myoglobin was also encapsulated within peptide-based hydrogels to deliver oxygen to transplanted neural progenitor cells⁶³. After 28 days post-transplantation in mice, there was significant increase in graft volume and innervation for groups horse and sperm whale myoglobin. However, the high oxygen-affinity (P₅₀ = 0.007 mmHg) sperm whale myoglobin mutant demonstrated the greatest impact, suggesting that there is a delicate balance between preventing hypoxia and over-supplying oxygen (e.g., for arterial PO₂ >120 mmHg)⁶⁴. Armstrong et al., had engineered a myoglobin-polymer-surfactant complex designed to bind to cell membranes and directly deliver oxygen when cellular levels were critically low. When applied to human MSCs in an in vitro model of cartilage, it was found that the technology not only alleviated hypoxia, but also improved the overall distribution and composition of the tissue⁶⁵.

An important consideration to make when engineering structural and functional changes to myoglobin is the relationship between stability and activity. It is well known that myoglobin will auto-oxidize to inactive metmyoglobin, which can subsequently react with hydrogen peroxide to form ferryl heme. This highly reactive species can initiate lipid peroxidation⁶⁶. Future work would benefit from exploring methods to reduce myoglobin auto-oxidation and conducting longer in vivo studies to determine protein retention and stability.

Neuroglobin

Within the last two decades, a newly discovered globin has been isolated from mouse and human neuronal tissue⁶⁷. With a similar oxygen affinity to myoglobin (P₅₀ = 1–10 mmHg), in vivo experiments indicate its role in protecting the brain and nervous system from hypoxia and oxidative stress⁶⁸.

Overexpression of neuroglobin increases neuron survival and viability after oxidative stress⁶⁹. Studies where neuroglobin-expressing adeno-associated viral vectors were administered into the cortex of a rat stroke model showed reduced infarct size (49–52% reduction

compared with control groups) and improved functional outcomes⁷⁰. Furthermore, when neuroglobin was overexpressed in mice hearts, ischemic injury was significantly reduced (reduction of ~25–30%). These findings suggest that neuroglobin possesses an enhanced ability to protect cells against oxidative stress in comparison to myoglobin⁷¹. Further investigation into neuroglobin and other under-studied hemoproteins, such as cytoglobin, could therefore be beneficial to the field of tissue engineering.

Enzyme systems modulating oxygen delivery

Physiological reactions involving oxygen or electron transfer tend to produce ROS as by-products (see Box 1).

Clearance of ROS, when they form in the ETC, is managed by enzymes such as SOD and CAT. CAT is capable of both sequestering H₂O₂ and generating oxygen, both of which are advantageous in the context of some oxygenated biomaterials. For example, hydrophobic oxygen generating microparticles (HOGMPs), composed of calcium peroxide-laden poly(caprolactone) (PCL) microparticles, can generate toxic levels of H₂O₂ as a by-product. By co-entrapment of the HOGMP with CAT in a hydrogel, H₂O₂ levels are reduced under hypoxic conditions, improving the viability and metabolic activity of cells cultured within the hydrogel for up to seven days⁷². Clearly, coadministration of oxygen- or redox-functional enzymes can alleviate challenges caused by oxygen delivery systems in vivo, in addition to facilitating delivery themselves.

Exogenous enzymes from bacteria, archaea and plants have also been explored as a means to deliver oxygen to hypoxic tissues. For example, a prokaryotic chlorite O₂-lyase enzyme, which catalyzes the conversion of chlorite (ClO₂⁻) into oxygen, can be heterologously expressed at high levels in human cells. Coexpression of a chlorite transporter to facilitate chlorite import was shown to initiate molecular oxygen production inside cells in response to extracellular chlorite. For example, when 5 million chlorite O₂-lyase expressing HeLa cells were subjected to a pulse of 1 mM sodium chlorite, 0.359 ± 0.01 μmol of oxygen was generated over a period of 30s⁷³. This coexpression technology, SNORCL, is an example of how the functional diversity of oxygen-related enzymes in all kingdoms of life can be taken advantage of for oxygen delivery if appropriate candidates are identified.

Other biomolecules

Serum albumins are transport proteins found in plasma that can bind diverse hydrophobic ligands such as water, hormones, fatty acids, cations, bilirubin and thyroxine. Notably, albumin has high affinity for heme, and can act as a hemoprotein for oxygen transportation⁷⁴. A limitation of heme-bound albumin is that the protein lacks the distal histidine of the globins, leading to rapid oxidation of the heme complex. To alleviate this, site-directed mutagenesis of the binding pocket to introduce a proximal histidine, as in the globin superfamily, can generate a heme-specific cleft and facilitate reversible oxygen binding, albeit with lower oxygen binding affinities (P₅₀ = 18–134 mmHg) compared to hemoglobin and myoglobin⁷⁵. Given the functional and structural diversity of oxygen-binding hemoproteins, there is clear potential to use evolved, engineered or de novo proteins to generate biomaterials capable of appropriate oxygen delivery depending on the application.

A recent approach to introducing oxygen into tissue-engineered constructs involves leveraging the properties of *Synechococcus elongatus*, a photoautotrophic cyanobacterium that can transform carbon dioxide into oxygen through photosynthesis, across a broad excitation spectrum (440–680 nm). *S. elongatus* has facilitated oxygenation in cardiac tissues during episodes of ischemic stroke⁷⁶. The application of *S. elongatus* has been extended to the domain of bio-printing, providing oxygenation for cell constructs under hypoxic conditions⁷⁷.

BOX 1

Oxygen and ROS in cell signaling and pathology

The primary cellular organelle that utilizes oxygen is the mitochondrion during oxidative phosphorylation, the ETC and ATP synthesis (see Fig. 3). Consequently, it is considered a major producer of ROS within the cell. It is hypothesized that singular electrons that leak from Complex I and III are available to then reduce molecular oxygen to superoxide ($O_2^{\cdot-}$). Notably, nicotinamide adenine dinucleotide phosphate (NADPH) oxidases are also a significant source of cellular $O_2^{\cdot-}$ ¹²⁶. There are a number of antioxidant systems that sequester ROS, such as by superoxide dismutase (SOD), which catalyzes the dismutation of $O_2^{\cdot-}$ into a more stable product - hydrogen peroxide (H_2O_2). H_2O_2 is then reduced to water by catalase (CAT) or glutathione peroxidase (GPX). H_2O_2 is an important signal transduction molecule involved in the activation and regulation of a number of different metabolic pathways. This includes regulation of HIFs, cellular proliferation and differentiation pathways such as the mitogen-activated protein kinase (MAPK) pathway, cellular growth through the phosphatidylinositol 3 kinase (PI3K) pathway and cellular apoptosis through the nuclear factor- κ B (NF- κ B) pathway. Furthermore, an increase in ROS production as a result of NADPH oxidase activation upon infection has been linked to the transcription of proinflammatory cytokines¹²⁷.

Antioxidant safeguards are overwhelmed when there is mitochondrial damage or increased activation of NADPH oxidase, such as in the case of ischemic/ reperfusion injury. The accumulation of $O_2^{\cdot-}$ and/or H_2O_2 can lead to the formation of highly reactive hydroxyl free radicals (HO^{\cdot}) via a Haber-Weiss reaction, which in turn cause severe oxidative stress through lipid, protein and DNA oxidation. It is also well-established that hypoxia induces oxidative stress¹²⁸. The precise mechanisms underlying hypoxia-induced ROS production remains unknown. However, one hypothesis is that a higher ratio of NADH/NAD⁺ under these conditions in turn increases electron transport through protein complex I, thus promoting $O_2^{\cdot-}$ production¹²⁹. Additionally, unstable intermediates (e.g. ubisemiquinone) of the Q-cycle are also believed to contribute to $O_2^{\cdot-}$ production under non-phosphorylating conditions¹³⁰. This highlights the need to maintain the oxygen homeostasis within engineered tissues, as there is evidence that both hyperoxic and hypoxic environments can lead to pathologic ROS generation.

ROS modulation has also been explored in the context of anti-cancer therapies. For instance, increasing oxygen supply in tumors' necrotic core can improve radiation and photo-dynamic therapies. This relies on the conversion of newly available oxygen into ROS leading to cell death^{131,132}.

Biomaterials and devices for oxygen generation & perfusion

In the context of oxygen for tissue engineering, the lack of a robust, native vasculature in scaffolds or engineered grafts has both ramifications for both oxygen transport, and waste removal. Engineered approaches toward reproducing this oxygenating architecture have been attempted. For example, fabrication techniques are employed to mimic vasculature and introduce artificial channels (refer to Box 2). Notably, in the context of in vitro applications (where most preliminary data is gathered), the media is periodically replaced, thereby removing waste metabolites. Therefore, in these models the focus has been on controlling the delivery of oxygen via chemical or mechanical means. Examples of this include the use of chemical additives such as peroxides and perfusing bioreactors; however, more recently the discussion extends to the generation of artificial blood through oxygen-carrying perfluorocarbons. Recent advances on these strategies and release kinetics are discussed below (refer also to Box 3).

Peroxides

Many non-biological approaches for in situ oxygen delivery have been explored. Perhaps the most prominent example is the integration of inorganic peroxides. Both soluble and solid peroxides are effective at oxygen release⁷⁸. They work by directly releasing hydrogen peroxide, which chemically decomposes to form water and molecular oxygen (following the chemical decomposition reaction, $2H_2O_2 \rightarrow 2H_2O + O_2$). Their oxygen release rate varies depending on the purity and solubility of the peroxides (e.g., solubility CaO_2 : 1.65g/L vs MgO_2 : 0.86g/L), presence of antioxidants (e.g., enzyme systems), and parameters such as temperature and pH. However, under physiological conditions (pH 5-7, 37 °C) rapid decomposition upon liquid exposure (≤ 48 h), termed burst oxygen generation, can be expected⁷⁸. This is problematic for regenerative approaches that require sustained oxygen release (days-months).

Mammalian cells can decompose $2H_2O_2$ at low concentrations via *2-cys peroxiredoxin* scavenger. However, excess (>10 nM) can lead to intracellular perturbation, causing oxidative damage and triggering

apoptosis⁷⁹. Consequences include tissue damage and gas emboli (e.g., arterial emboli). In addition, tissue detriment can originate from the highly exothermic reaction (standard enthalpy of -98.2 kJ/mol) itself^{80,81}. As such, challenges for integrating peroxides into tissue engineering solutions include: (i) optimizing exposure time and concentration while balancing redox reactions; (ii) tissue selectivity; and (iii) decreasing toxicity.

To mitigate the adverse effects of burst oxygen generation, a strategic approach is to reduce peroxide exposure to water by encapsulating them within polymeric structures⁸². Synthetic polymers, such as PCL, have been explored for this purpose, achieving dissolved oxygen concentrations ranging from 5% to 29% (0.12–0.71 mmHg) with increasing peroxide loading^{83,84}. Another example is the use of in situ oxygen generation, such as by Coronel et al., where, by using calcium peroxide encapsulated in polydimethylsiloxane tablets, oxygen was raised from 0.1 mmHg to 214 mmHg, resulting in reduced anaerobic glycolysis and stimulating insulin release in pancreatic islets. From these examples, it is evident that the release rate also depends on the peroxide (e.g., molecular weight $CaCO_3$ MW= 100 g/mol), the encapsulating material properties and the morphology of the delivery vehicle (e.g., nanosized particles vs bulk hydrogels)⁸⁵. Recent research has devised parameters for oxygen release from encapsulating materials, of which the interested reader can find further discussion in Box 3.

An additional reported advantage of controlling oxygen release is the indirect reduction of ROS and free radicals. As discussed in Section 3.4., this can be enhanced by the supplementation of antioxidants such as CAT enzymes⁸⁶. This has been exploited to create bioinks for 3D-printed constructs, improving cell viability under severe hypoxia (1–5%)⁷². The use of peroxides incorporated in bio-inks has the goal of improving spatial distribution of peroxides while satisfying the need for oxygen in large 3D printed structured and further into large tissue repair^{87,88}. The introduction of oxygenating architecture within bioinks can also serve as a structural support for maturation. With these strategies to control the spatio-temporal release of oxygen through physical encapsulation, gradient-controlled oxygen release patterns could be expected soon.

BOX 2**Vascularisation and engineered oxygenating architecture**

Vascularization can be achieved by stimulating angiogenesis and activating the host microvasculature. Several engineering approaches, including (i) the modification of the chemical composition, architecture, and porosity of constructs; (ii) the addition of angiogenic growth factors such as VEGF, and (iii) cell seeding with endothelial cell (EC) progenitors, have been employed to promote the ingrowth of new blood vessels⁵. However, this process is slow (8 weeks-8 months)^{133,134}.

Alternatively, template channels or structures can be fabricated within engineered tissue scaffolds to direct the distribution of EC progenitors towards forming organized, interconnected tubular vascular networks, thus enhancing the integration of grafted ECs (and by extension, other transplanted cells) with the host¹³⁵⁻¹³⁷. Without such structures, encapsulating ECs leads to an unstable or unconnected tubular network and irregular vessel architecture. This can increase the susceptibility to early thrombosis¹³⁸ and cause delays between implantation and perfusion¹³⁹.

Techniques for building perfusable template channels include micromolding, laser degradation, 3D printing, and coaxial microfluidic spinning¹⁴⁰. For example, an aligned "cord" consisting of ECs and collagen fabricated by microtissue molding can form new capillaries along its length (1 cm). When studied in the context of hepatocyte transplantation, the survival of hepatocytes and hepatic function were significantly enhanced¹⁴¹. Despite the ease of micromolding, this technology remains limited by the design constraints on the planar substrate¹⁴². 3D bioprinting can also be implemented to generate heterogeneous tissue with multiple patterned cells, materials, and vascular channels¹⁴². However, bioinks have limitations, such as possible shear damage to cells during extrusion¹⁴³. Coaxial microfluidic spinning has created an exciting avenue for biomaterials. In this technique, hollow microfibers are formed by the flowing fluids in microfluidic devices. Yu et al. demonstrated how ECs can be encapsulated into the microfibres, driving an annular distribution of the cells along the direction of the microfibers¹⁴⁴, thus successfully constructing vessels¹⁴⁵. However, these materials are not robust and require delicate handling¹⁴⁴.

Providing a prevascularised environment for transplanted cells has other advantages in addition to oxygen delivery, as it also enables waste exchange. As such, materials utilized to preform vasculature should be permeable to water-soluble gases, metabolites and molecules¹⁴⁶.

Current challenges of preformed artificial vasculature include the demand for customized and expensive equipment or materials, such as laser-ablation microscopes, sterile environments, 3D printers, or unique bioinks¹⁴⁷. This highlights the necessity for the development of simpler, less expensive approaches to improve scalability and accessibility. Importantly, preformed vasculatures require rapidly developing interconnections to the host tissue post-implantation. Notably, on implantation of the prevascularised materials in vivo, the host models and anatomic implant locations need to be taken into consideration, as any discrepancies can illicit an inflammatory response¹³⁹.

Perfluorocarbons

Perfluorocarbons (PFCs) differ structurally from conventional alkyl chains. The abbreviation 'PFC' often refers to perfluorinated chemicals. Herein, PFCs denote only perfluorocarbons. In comparison to hydrocarbon chain (H-chains), F-chains adopt a helical arrangement (versus the planar configuration of H-Chains) and a more rigid structure (cross section of ca. 30 Å²). In PFCs (C_nF_n) the fluorine atoms surrounding the carbon backbone contribute to their hydrophobic nature, enabling them to accommodate gases, including oxygen. The solubility of oxygen in PFCs is up to one thousand times the solubility of oxygen in water, with a molecular ratio of dissolved oxygen I_{O₂}: 200 in water compared to 5_{O₂}: 1 in PFCs, such as perfluorodecalin⁸⁹.

The use of PFCs to provide oxygen in aqueous solutions was first proposed in 1966, and later as blood substitutes⁹⁰. Unlike globins, PFCs oxygen release kinetics vary linearly according to pO₂ (following Henry's law), and for blood applications, its oxygen-carrying capacity is often compared against hemoglobin. For instance, the oxygen-delivering capacity of a 1-g PFC/kg body weight dose is estimated to be around 1.5 g hemoglobin for commercial blood-substitute, Oxygent⁹¹. After decades of investigation, clinical use of PFCs as blood substitutes and as oxygen carriers for tissue engineering still faces challenges, including: (i) fabricating an optimized, stable formulation; (ii) overcoming interference with clinical oxygen measurements; and (iii) a reliable large production process.

Optimizing PFCs formulations to improve their limited miscibility has been tested by using nanoemulsions as parental formulations. Common emulsion parameters, such as droplet size (10-1000 nm), fabrication method, and surfactant choice, influence their stability and shelf-life^{92,93} (Fig. 5). Commercial oxygen delivery systems based on PFCs have included emulsions loaded with 11-40% PFCs. Efforts to optimize carriers for PFCs continue, with recent research focusing on fabricating deformable core-shell particles⁹⁴, using natural molecule-based carriers (such as albumin)⁹⁵, sequence-controlled

delivery⁹⁶ and doubly encapsulated integrated into lipid particle-hydrogel scaffolds⁹⁷.

Few toxic effects have been linked to PFCs, and often the effects are attributed to co-factor deposition (e.g., hepatic deposition >65 days) and surfactant-related side effects. Despite of this, industrial PFCs gases derived mainly from electronic manufacturing, are considered potent pollutants⁹⁸. In the context of tissue engineering and clinical use, a consideration is the interference of PFCs with pulse oximetry equipment and chemistry analytes^{99,100} which could be overcome by turbidity correction algorithms as proposed by Toffaletti et al.¹⁰¹. In addition, product quality control for PFCs must be closely regulated. Lambert et al., have recently discussed parameters for scaling up the process to use PFCs for oxygen delivery and proposed a quality by-design practice adaptation for PFC nanoemulsion optimization¹⁰². Broad adoption of these methods could improve PFC translatability.

Nanobubbles

Nanobubbles are single bubbles that are inherently unstable and have a theoretical lifespan of <1s according to Laplace pressure $\Delta P = 2(\delta/r)$, where r is the radius of the bubble and δ is the interfacial pressure¹⁰³. As a result, lowering surface tension can help to maintain their stability by reducing the interfacial pressure (e.g., by PEGylated lipids). Reducing the particle size (10-200 nm) as well as avoiding coalescence (e.g., negative charge increase: -15 to -60 mV) can also improve their oxygenating potential.

Nanobubbles can be created in biological solutions such as cell culture media via gas-liquid agitation method with oxygen discharge over 48 h (with 31% of initial nanobubbles remaining intact after this time)¹⁰⁴ but their containment via shell entrapping the gas core using lipids, proteins, and polymers can extend their lifespan (12-100 days)^{105,106}. When integrated into engineered polymeric scaffolds, nanobubbles gradually discharge oxygen (8 h-8 weeks), shifting

BOX 3**Mechanisms of oxygen delivery from secondary materials**

The encapsulation of biomolecules, peroxides, perfluorocarbons and nanobubbles into second materials that limit their release kinetics (i.e. secondary encapsulation) is common method to prolong oxygen delivery. The oxygen release is thus influenced by i) the primary material or biomolecule and ii) the secondary matrix in which the primary material or biomolecule is encapsulated. As such, the release is an interplay between the material characteristics, the biological demands (and events such as enzymatic degradation), and the encapsulating method¹⁴⁸. When a matrix is implanted or injected at time $t = 0$ where the polymer relaxation time t_r and the characteristic diffusion time T_d satisfies $(t_r) \gg T_d$, diffusion will be governed typically by Fickian diffusion mechanisms described by Fick's second law¹⁴⁹. However, other cases of mass transport have also been described (such as non-Fickian) and various mathematical models have been proposed describing release profiles (or fractional amounts released), material geometries (such as particles and slabs), or physicochemical characteristics (e.g., swelling and degradation)¹⁵⁰.

Hydrogels for tissue engineering and regenerative medicine often present a release characterized by a rapid burst at early time points ($t < 12h$) followed by a slow release, e.g. ($24h < t < 30days$), or cargo depletion. The rate of release from these hydrogels -in the absence of binding mechanisms- strongly depends on the physical characteristics between the cargo (e.g., molecular weight and charge) and the material (e.g., the mesh size ζ)^{151,152}. In contrast, high-swelling hydrogel matrices or erodible polymers associate the release of the cargo with combined diffusion and surface/ bulk degradation mechanisms. An exemption to this is when the oxygen-carrying molecule is part of the matrix itself, as exemplified by the work of Park et al., where a thiolated gelatin was crosslinked via calcium peroxide-mediated oxidative reaction¹⁵³. Similarly, Ding et al., fabricated an oxygen generating and ROS-scavenging injectable hydrogel based on the reaction between synthetic ROS-cleavable hyper-branched polymers and methacrylate hyaluronic acid¹⁵⁴.

The complexity of the release kinetics extends to the use of multi-encapsulating materials (e.g., microspheres containing peroxides encapsulated in scaffolds). In theory, the release kinetics of a system designed to constantly release oxygen can be expected to follow a zero-order kinetics (described by $C = C_0 + k_0t$ where C is the amount of oxygen released at time t , and C_0 is the initial amount of drug in the solution and k_0 the zero-order constant). However, this material design might not be parallel to the oxygen demands in biological systems that constantly fluctuate its therapeutic window.

hypoxic conditions, and have shown efficiency in maintaining cellular metabolism in vivo (8 weeks)¹⁰⁷. Integration of nanobubbles needs careful design; methods like sonication create particles of varied sizes (10-500 nm)¹⁰⁸, making it difficult to anticipate the oxygen release rate. To overcome this, fabrication techniques such as microfluidic-formed nanobubbles have emerged^{109,110}.

A limitation of encapsulated nanobubbles is that excessive encapsulation can hinder oxygen release, negating their intended purpose. Stimuli-responsive oxygen release is an emerging method to improve this. An emerging approach is to use microbubble-enhanced ultrasound to disrupt physiological barriers, such as the blood-brain barrier (BBB) for brain drug delivery¹¹¹ (Fig. 5), which works by disrupting a particle's membrane layer via cavitation, thus allowing on-demand oxygen release. For example, Wang et al., used phase transition-triggered low-intensity ultrasound nanobubbles that were filled with hemoglobin and then mixed into gelatine methacryloyl (GelMA)¹¹². The authors used a blend of GelMA-Hemoglobin nanobubbles to 3D print cardiomyocyte patches and showed increased cell survival against non-hemoglobin and non-ultrasound-stimulated structures. A challenge to overcome with this technique is that, unless there is continuous production of oxygen-carrying nanobubbles, it is a single-time point delivery mechanism. Low-intensity pulsed ultrasound to target sites and integration of biologicals to carry oxygen as a secondary oxygen release source may address this limitation in future investigations.

Oxygen perfusing and oxygen-generating devices

Oxygen-perfusing devices, such as bioreactors, function by circulating dissolved oxygen in media around scaffolds and cells, mechanically assisting in waste removal and supplying fresh nutrients and oxygen¹¹³. Bioreactors have been investigated for their utility in the maturation of large scaffolds, where the center may otherwise experience reduced pO_2 ¹¹⁴. Common bioreactor types used in tissue engineering are rotating wall vessels, spinner flasks, perfusion, and compression systems. The rate of oxygen transfer can be approximated based on the mass transfer coefficient, concentration gradient and geometry of the bioreactors. An important consideration is that in bioreactors the

oxygen transfer rate (OTR) often surpasses the oxygen uptake rate (OUR) from the cells. To satisfy an adequate system, the OTR:OUR must be 1:1 or $OTR \geq OUR$. However, this condition is challenged by the constant fluctuations in cellular OUR. Advancements in bioreactors have begun to offer real-time monitoring capabilities, integrated sensors for oxygen tension, and customized flow patterns based on advanced mathematical models to closely mimic physiological conditions¹¹⁵.

Microfluidic devices, functioning as micro-bioreactors, contribute to circulating media through microchannels mimicking capillary networks. The advent of "organ-on-a-chip" systems leverages microfluidics to create complex multi-tissue interfaces, allowing for tissue models and drug testing platforms¹¹⁶. Besides the complexity of tissues and microfabrication of advanced vasculature, microfluidics for single-cell oxygenation, tailored systems for cell culture oxygen delivery, and cellular oxygen monitoring are emerging developments¹¹⁷. Although on a much smaller scale than the large bioreactors, microfluidic devices still rely on an adequate flow rate and shear stress to provide oxygen to cell cultures, with typical flow rates varying from 0.2 to 3 $\mu\text{l}/\text{min}$ and channels widths from 1 to 200 μm .

Electronic-assisted microfluidics, or electronic integrated systems for oxygen delivery, are finding a new avenue in regenerative medicine. Electrochemical oxygen generators use electrochemical reactions to split water molecules into molecular hydrogen and oxygen, which can be delivered directly to tissue engineering constructs or fabricated as implantable oxygen devices^{118,119}. Oxygen dosage can be controlled through pulse width modulation of the electrode potential, providing various oxygen ranges (0–10 $\mu\text{mol}/\text{h}$)¹²⁰.

Oxygen monitoring and ROS detection devices, specifically designed for tissue engineering and biomedical applications, offer novel insight into oxygen demands^{121,122}. An interesting trend is dual-function sensing and delivery of oxygen. For example, a recent study showed a platform for locally producing (321.5 $\mu\text{mol}/\text{h}$) and measuring oxygen (with a range of 312.5–1625 $\mu\text{mol}/\text{h}$) in a wound area¹²³.

As the field evolves, the need for accurate measuring of oxygen is evident, with few reliable devices and assays designed for tissue engineering and biomedical applications such as micro-probes (size

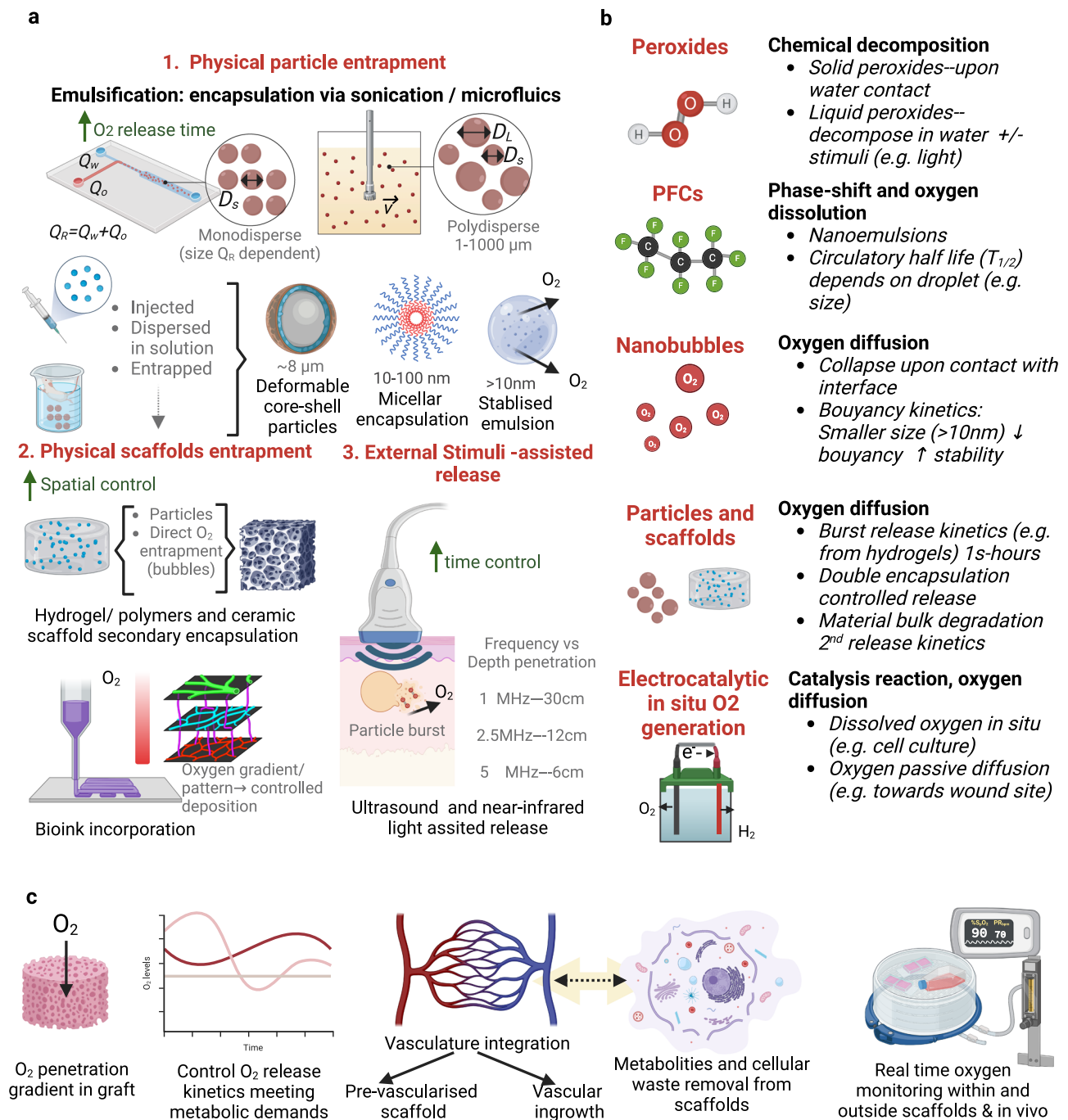


Fig. 5 | Controlled delivery strategies, oxygen release mechanisms and challenges in the field. a Oxygen-controlled release strategies include physical entrapment in emulsions of monodisperse size (e.g., microfluidic-generated) or polydisperse (particle dependent on velocity and shear of the system) generating particles such as micelles or multi-layered particles. Other strategies include

scaffold encapsulation and stimuli-dependent release. **b** Core release mechanisms for biomaterials and devices able to generate oxygen. **c** Remaining challenges in the field of oxygen tissue engineering. Figure created with BioRender.com, released under a Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International license.

50 μm–1.5 mm), luminescence optical sensors (0-500 hPa) and oxygen-reacting cellular assays (via nonspecific energy-dependent endocytosis). A limitation of these is that they are designed for in vitro applications, while in vivo non-invasive products able to test scaffold transplantation performance and bioscaffold cellular development are yet to be developed.

Concluding remarks and future directions

The intricate machinery of tissue repair is fueled by oxygen; powering cellular functions, lighting signaling pathways, and enabling structural

reorganization to achieve optimal healing or regeneration. Promising avenues have emerged in the past few years to redefine our understanding and approach to tissue regeneration mediated by oxygen delivery. Notably, hemoproteins offer well-studied oxygen binding and releasing kinetics. However, they are prone to degradation and only few investigations have created protective environments to combat this. Designing proteins with higher oxygen retention and their inclusion into novel biomaterials can provide tailored oxygen delivery. To overcome a major limitation of oxygen-generating materials, such as ROS production and subsequent oxidative stress, the use

of ROS-targeting enzymes might offer circumvention at cellular microenvironment level.

Despite the undeniable translational potential of PFCs and peroxides in medical applications, a breakthrough medical product remains to be launched. Investigations towards optimized formulations using polymeric encapsulations and receptor-binding interactions have been identified as potential facilitators to this end. The continuous research into PCFs underscores the possibility of overcoming translation barriers (such as scalability) by using new industry fabrication frameworks, thus leading to innovative medical products. 3D printing technologies can offer oxygen-containing bio-inks that promise spatio-temporal oxygen delivery into more complex scaffolds than traditional tissue engineering. However, this development must be accompanied by meeting the fluctuating oxygen demands of the cells confined within bio-inks and also be able to sustain such demands in *in vivo* scenarios. To that end, deeper investigation of the cellular oxygen demands over regenerative processes is needed. Similarly, oxygen-perfusing devices such as bioreactors and microfluidics have shown the need for adapting miniaturized and precise real-time monitoring capabilities. Future research should aim to improve the precision measurement of oxygen levels and rapid adjustment of oxygen transfer rates to closely match cellular oxygen uptake. Furthermore, the oxygen and oxygen pressure monitoring set-up should move from bulky *in vitro* systems to accurate, lightweight and minimally invasive *in vivo* monitoring (e.g. *in situ* scaffold oxygen-releasing monitoring).

Future research is expected to investigate the fabrication of dynamic materials or devices that can adapt oxygen release in response to their environment and cellular needs. With the regenerative medicine and tissue engineering field continuously pushing boundaries, numerous innovative approaches are on the horizon, holding promise for a golden era of tissue engineering via oxygen delivery systems.

References

- Mitrousis, N., Fokina, A. & Shoichet, M. S. Biomaterials for cell transplantation. *Nat. Rev. Mater.* **3**, 441–456 (2018).
- Croll, T. I. et al. Modelling oxygen diffusion and cell growth in a porous, vascularising scaffold for soft tissue engineering applications. *Chem. Eng. Sci.* **60**, 4924–4934 (2005).
- Kellner, K. et al. Determination of oxygen gradients in engineered tissue using a fluorescent sensor. *Biotechnol. Bioeng.* **80**, 73–83 (2002).
- Hosseini, M., Brown, J. & Shafiee, A. Strategies to induce blood vessel ingrowth into skin grafts and tissue-engineered substitutes. *Tissue Eng. Part C Methods* **28**, 113–126 (2022).
- Wu, X. et al. Biomimetic porous hydrogel scaffolds enabled vascular ingrowth and osteogenic differentiation for vascularized tissue-engineered bone regeneration. *Appl. Mater. Today* **27**, 101478 (2022).
- Potente, M., Gerhardt, H. & Carmeliet, P. Basic and therapeutic aspects of angiogenesis. *Cell* **146**, 873–887 (2011).
- Oh, S. H., Ward, C. L., Atala, A., Yoo, J. J. & Harrison, B. S. Oxygen generating scaffolds for enhancing engineered tissue survival. *Biomaterials* **30**, 757–762 (2009).
- Chin, K., Khattak, S., Bhatia, S. & Roberts, S. Hydrogel-perfluorocarbon composite scaffold promotes oxygen transport to immobilized cells. *Biotechnol. Prog.* **24**, 358–366 (2008).
- Ciray, H. N., Aksoy, T., Yaramanci, K., Karayaka, I. & Bahceci, M. *In vitro* culture under physiologic oxygen concentration improves blastocyst yield and quality: a prospective randomized survey on sibling oocytes. *Fertil. Steril.* **91**, 1459–1461 (2009).
- Dumoulin, J. C. et al. Effect of oxygen concentration on human *in vitro* fertilization and embryo culture*. *Hum. Reprod.* **14**, 465–469 (1999).
- Nanassy, L. et al. Comparison of 5% and ambient oxygen during days 3–5 of *in vitro* culture of human embryos. *Fertil. Steril.* **93**, 579–585 (2010).
- Mantikou, E. et al. Low oxygen concentrations for embryo culture in assisted reproductive technologies. *Hum. Reprod. Update* **19**, 209 (2013).
- Simon, M. C. & Keith, B. The role of oxygen availability in embryonic development and stem cell function. *Nat. Rev. Mol. Cell Biol.* **9**, 285–296 (2008).
- Vanderkooi, J. M., Erecińska, M. & Silver, I. A. Oxygen in mammalian tissue: methods of measurement and affinities of various reactions. *Am. J. Physiol.* **260**, C1131–1150 (1991).
- Ortiz-Prado, E., Dunn, J. F., Vasconez, J., Castillo, D. & Viscor, G. Partial pressure of oxygen in the human body: a general review. *Am. J. Blood Res.* **9**, 1–14 (2019).
- Tintu, A. N., le Noble, F. A. C. & Rouwet, E. V. Hypoxia disturbs fetal hemodynamics and growth. *Endothelium* **14**, 353–360 (2007).
- Ramani, M., Bradley, W. E., Dell’Italia, L. J. & Ambalavanan, N. Early exposure to hyperoxia or hypoxia adversely impacts cardiopulmonary development. *Am. J. Respir. Cell Mol. Biol.* **52**, 594–602 (2015).
- Srivatsa, B., Hagan, J. L., Clark, R. H. & Kupke, K. G. Oxygenation factors associated with retinopathy of prematurity in infants of extremely low birth weight. *J. Pediatr.* **247**, 46–52.e4 (2022).
- Lithopoulos, M. A. et al. Neonatal hyperoxia in mice triggers long-term cognitive deficits via impairments in cerebrovascular function and neurogenesis. *J. Clin. Investig.* **132**, e146095 (2022).
- Reily, C., Stewart, T. J., Renfrow, M. B. & Novak, J. Glycosylation in health and disease. *Nat. Rev. Nephrol.* **15**, 346–366 (2019).
- Rigby, B. J., Mitchell, T. W. & Robinson, M. S. Oxygen participation in the *in vivo* and *in vitro* aging of collagen fibres. *Biochem. Biophys. Res. Commun.* **79**, 400–405 (1977).
- Rappu, P., Salo, A., Myllyharju, J. & Heino, J. Role of prolyl hydroxylation in the molecular interactions of collagens. *Essays Biochem.* **63**, 325–335 (2019).
- Rahman, S. et al. Inability of low oxygen tension to induce chondrogenesis in human infrapatellar fat pad mesenchymal stem cells. *Front. Cell Dev. Biol.* **9**, 703038 (2021).
- Buckley, C. T., Vinardell, T. & Kelly, D. J. Oxygen tension differentially regulates the functional properties of cartilaginous tissues engineered from infrapatellar fat pad derived MSCs and articular chondrocytes. *Osteoarthritis Cartilage* **18**, 1345–1354 (2010).
- Howcroft, T. K. et al. The role of inflammation in age-related disease. *Aging* **5**, 84–93 (2013).
- Bowser, J. L., Lee, J. W., Yuan, X. & Eltzschig, H. K. The hypoxia-adenosine link during inflammation. *J. Appl. Physiol.* **123**, 1303–1320 (2017).
- Triner, D. & Shah, Y. M. Hypoxia-inducible factors: a central link between inflammation and cancer. *J. Clin. Investig.* **126**, 3689–3698 (2016).
- Lun, J., Zhang, H., Guo, J., Yu, M. & Fang, J. Hypoxia inducible factor prolyl hydroxylases in inflammatory bowel disease. *Front. Pharmacol.* **14**, 1045997 (2023).
- Dorrello, N. V. et al. Functional vascularized lung grafts for lung bioengineering. *Sci. Adv.* **3**, e1700521 (2017).
- Levenberg, S. et al. Engineering vascularized skeletal muscle tissue. *Nat. Biotechnol.* **23**, 879–884 (2005).
- O’Connell, C. D. et al. Within or without you? A perspective comparing *in situ* and *ex situ* tissue engineering strategies for articular cartilage repair. *Adv. Healthc. Mater.* **11**, 2201305 (2022).
- Ngo, J. P. et al. Vascular geometry and oxygen diffusion in the vicinity of artery-vein pairs in the kidney. *Am. J. Physiol.-Ren. Physiol.* **307**, F1111–F1122 (2014).
- Androjna, C., Gatica, J. E., Belovich, J. M. & Derwin, K. A. Oxygen diffusion through natural extracellular matrices: implications for

- estimating "critical thickness" values in tendon tissue engineering. *Tissue Eng. Part A* **14**, 559–569 (2008).
34. Mendez-Romero, O., Ricardez-García, C., Castañeda-Tamez, P., Chiquete-Félix, N. & Uribe-Carvajal, S. Thriving in oxygen while preventing ROS overproduction: no two systems are created equal. *Front. Physiol.* **13**, 874321 (2022).
 35. Cho, C. H. et al. Oxygen uptake rates and liver-specific functions of hepatocyte and 3T3 fibroblast co-cultures. *Biotechnol. Bioeng.* **97**, 188–199 (2007).
 36. Carroll, S. F., Buckley, C. T. & Kelly, D. J. Measuring and modeling oxygen transport and consumption in 3D hydrogels containing chondrocytes and stem cells of different tissue origins. *Front. Bioeng. Biotechnol.* **9**, 591126 (2021).
 37. Magliaro, C. et al. Oxygen consumption characteristics in 3D constructs depend on cell density. *Front. Bioeng. Biotechnol.* **7**, 251 (2019).
 38. Li, S. et al. Preconditioning in neuroprotection: from hypoxia to ischemia. *Prog. Neurobiol.* **157**, 79–91 (2017).
 39. Hsia, C. C. Respiratory function of hemoglobin. *N. Engl. J. Med.* **338**, 239–248 (1998).
 40. Liu, Y., Zhao, X., Zhao, C., Zhang, H. & Zhao, Y. Responsive porous microcarriers with controllable oxygen delivery for wound healing. *Small* **15**, 1901254 (2019).
 41. Zhao, Y. et al. All-in-one bioactive properties of photothermal nanofibers for accelerating diabetic wound healing. *Biomaterials* **295**, 122029 (2023).
 42. Amberson, W. R. et al. On the use of ringer-locke solutions containing hemoglobin as a substitute for normal blood in mammals. *J. Cell. Comp. Physiol.* **5**, 359–382 (1934).
 43. Buehler, P. W., D'Agnillo, F. & Schaer, D. J. Hemoglobin-based oxygen carriers: from mechanisms of toxicity and clearance to rational drug design. *Trends Mol. Med.* **16**, 447–457 (2010).
 44. Bunn, H. F., Esham, W. T. & Bull, R. W. The renal handling of hemoglobin. *J. Exp. Med.* **129**, 909–924 (1969).
 45. Estep, T. N. *Blood Substitutes and Oxygen Biotherapeutics*, 287–292 (Springer International Publishing, 2022).
 46. Hunt, S. D. & Elg, F. Clinical effectiveness of hemoglobin spray (Granulox®) as adjunctive therapy in the treatment of chronic diabetic foot ulcers. *Diabet. Foot Ankle* **7**, 33101 (2016).
 47. Ito, Y. et al. Neuroprotective effects of combination therapy of regional cold perfusion and hemoglobin-based oxygen carrier administration on rat transient cerebral ischemia. *Brain Res.* **1746**, 147012 (2020).
 48. Gekka, M. et al. Novel hemoglobin-based oxygen carrier bound with albumin shows neuroprotection with possible antioxidant effects. *Stroke* **49**, 1960–1968 (2018).
 49. Wang, J. et al. Polynitroxylated PEGylated hemoglobin protects pig brain neocortical gray and white matter after traumatic brain injury and hemorrhagic shock. *Front. Med. Technol.* **5**, 1074643 (2023).
 50. Chen, L., Yang, Z. & Liu, H. Hemoglobin-based oxygen carriers: where are we now in 2023? *Medicina* **59**, 396 (2023).
 51. Jansman, M. M. T. et al. Hemoglobin-based oxygen carriers camouflaged with membranes extracted from red blood cells: Optimization and assessment of functionality. *Biomater. Adv.* **134**, 112691 (2022).
 52. Toma, V. A. et al. Comparative in vivo effects of hemoglobin-based oxygen carriers (HBOC) with varying prooxidant and physiological reactivity. *PLoS ONE* **11**, e0153909 (2016).
 53. Fontes, P. et al. Liver preservation with machine perfusion and a newly developed cell-free oxygen carrier solution under subnormothermic conditions. *Am. J. Transplant.* **15**, 381–394 (2015).
 54. Bian, Y. & Chang, T. M. S. A novel nanobiotherapeutic poly-[hemoglobin-superoxide dismutase-catalase-carbonic anhydrase] with no cardiac toxicity for the resuscitation of a rat model with 90 minutes of sustained severe hemorrhagic shock with loss of 2/3 blood volume. *Artif. Cells Nanomed. Biotechnol.* **43**, 1–9 (2015).
 55. Guo, C. & Chang, T. M. S. Long term safety and immunological effects of a nanobiotherapeutic, bovine poly-[hemoglobin-catalase-superoxide dismutase-carbonic anhydrase], after four weekly 5% blood volume top-loading followed by a challenge of 30% exchange transfusion. *Artif. Cells Nanomed. Biotechnol.* **46**, 1349–1363 (2018).
 56. Guo, C., Gynn, M. & Chang, T. M. S. Extraction of superoxide dismutase, catalase, and carbonic anhydrase from stroma-free red blood cell hemolysate for the preparation of the nanobiotechnological complex of polyhemoglobin-superoxide dismutase-catalase-carbonic anhydrase. *Artif. Cells Nanomed. Biotechnol.* **43**, 157–162 (2015).
 57. Bian, Y., Rong, Z. & Chang, T. M. S. Polyhemoglobin-superoxide dismutase-catalase-carbonic anhydrase: a novel biotechnology-based blood substitute that transports both oxygen and carbon dioxide and also acts as an antioxidant. *Artif. Cells Blood Substit. Biotechnol.* **39**, 127–136 (2011).
 58. Dasmeh, P. & Kepp, K. P. Bridging the gap between chemistry, physiology, and evolution: quantifying the functionality of sperm whale myoglobin mutants. *Comp. Biochem. Physiol. Part A: Mol. Integr. Physiol.* **161**, 9–17 (2012).
 59. Balcerak, B. et al. A broad diversity in oxygen affinity to haemoglobin. *Sci. Rep.* **10**, 16920 (2020).
 60. Gros, G., Wittenberg, B. A. & Jue, T. Myoglobin's old and new clothes: from molecular structure to function in living cells. *J. Exp. Biol.* **213**, 2713–2725 (2010).
 61. Burke, M. et al. Regulation of scaffold cell adhesion using artificial membrane binding proteins. *Macromol. Biosci.* **17**, 1600523 (2017).
 62. Castro-Forero, A., Jiménez, D., López-Garriga, J. & Torres-Lugo, M. Immobilization of myoglobin from horse skeletal muscle in hydrophilic polymer networks. *J. Appl. Polym. Sci.* **107**, 881–890 (2008).
 63. Wang, Y. et al. Hydrogel oxygen reservoirs increase functional integration of neural stem cell grafts by meeting metabolic demands. *Nat. Commun.* **14**, 457 (2023).
 64. Helmerhorst, H. J. F. et al. Metrics of arterial hyperoxia and associated outcomes in critical care*. *Crit. Care Med.* **45**, 187–195 (2017).
 65. Armstrong, J. P. K. et al. Artificial membrane-binding proteins stimulate oxygenation of stem cells during engineering of large cartilage tissue. *Nat. Commun.* **6**, 7405 (2015).
 66. Gunther, M. R., Kelman, D. J., Corbett, J. T. & Mason, R. P. Self-peroxidation of metmyoglobin results in formation of an oxygen-reactive tryptophan-centered radical. *J. Biol. Chem.* **270**, 16075–16081 (1995).
 67. Burmester, T., Weich, B., Reinhardt, S. & Hankeln, T. A vertebrate globin expressed in the brain. *Nature* **407**, 520–523 (2000).
 68. Ascenzi, P. et al. Neuroglobin: from structure to function in health and disease. *Mol. Asp. Med.* **52**, 1–48 (2016).
 69. Watanabe, S., Takahashi, N., Uchida, H. & Wakasugi, K. Human neuroglobin functions as an oxidative stress-responsive sensor for neuroprotection. *J. Biol. Chem.* **287**, 30128–30138 (2012).
 70. Sun, Y. et al. Neuroglobin protects the brain from experimental stroke in vivo. *Proc. Natl Acad. Sci. USA* **100**, 3497–3500 (2003).
 71. Khan, A. A. et al. Neuroglobin-overexpressing transgenic mice are resistant to cerebral and myocardial ischemia. *Proc. Natl Acad. Sci. USA* **103**, 17944–17948 (2006).
 72. Willemsen, N. G. A. et al. Enzyme mediated alleviation of peroxide toxicity in self oxygenating biomaterials. *Adv. Healthc. Mater.* **11**, 2102697 (2022).
 73. Markhard, A. L., McCoy, J. G., To, T.-L. & Mootha, V. K. A genetically encoded system for oxygen generation in living cells. *Proc. Natl Acad. Sci. USA* **119**, e2207955119 (2022).
 74. Komatsu, T. *Hemoglobin-Based Oxygen Carriers as Red Cell Substitutes and Oxygen Therapeutics*, 339–348 (Springer Berlin Heidelberg, 2013).

75. Komatsu, T., Ohmichi, N., Zunszain, P. A., Curry, S. & Tsuchida, E. Dioxygenation of human serum albumin having a prosthetic heme group in a tailor-made heme pocket. *J. Am. Chem. Soc.* **126**, 14304–14305 (2004).
76. Stapleton, L. M. et al. Microfluidic encapsulation of photosynthetic cyanobacteria in hydrogel microparticles augments oxygen delivery to rescue ischemic myocardium. *J. Biosci. Bioeng.* **135**, 493–499 (2023).
77. Hwangbo, H. et al. Photosynthetic cyanobacteria can clearly induce efficient muscle tissue regeneration of bioprinted cell-constructs. *Adv. Funct. Mater.* **33**, 2209157 (2023).
78. Rastinfard, A., Dalisson, B. & Barralet, J. Aqueous decomposition behavior of solid peroxides: effect of pH and buffer composition on oxygen and hydrogen peroxide formation. *Acta Biomater.* **145**, 390–402 (2022).
79. Huang, B. K. & Sikes, H. D. Quantifying intracellular hydrogen peroxide perturbations in terms of concentration. *Redox Biol.* **2**, 955–962 (2014).
80. Krautschick, I., Krugmann, J. & Neuenfeld, M. The effect of peroxides on the vascular endothelium of isolated pig aorta in vitro. *Exp. Toxicol. Pathol.* **47**, 51–61 (1995).
81. Steg, H. et al. Control of oxygen release from peroxides using polymers. *J. Mater. Sci. Mater. Med.* **26**, 207 (2015).
82. Aleemardani, M., Solouk, A., Akbari, S., Dehghan, M. M. & Moeini, M. Silk-derived oxygen-generating electrospun patches for enhancing tissue regeneration: Investigation of calcium peroxide role and its effects on controlled oxygen delivery. *Materialia* **14**, 100877 (2020).
83. Suvarnapathaki, S., Nguyen, A., Goulopoulos, A. & Camci-Unal, G. Oxygen-generating scaffolds for cardiac tissue engineering applications. *ACS Biomater. Sci. Eng.* **9**, 409–426 (2023).
84. Peng, Z. et al. 3D printed polycaprolactone/beta-tricalcium phosphate/magnesium peroxide oxygen releasing scaffold enhances osteogenesis and implanted BMSCs survival in repairing the large bone defect. *J. Mater. Chem. B* **9**, 5698–5710 (2021).
85. Khorshidi, S., Karkhaneh, A. & Bonakdar, S. Fabrication of amine-decorated nonspherical microparticles with calcium peroxide cargo for controlled release of oxygen. *J. Biomed. Mater. Res. Part A* **108**, 136–147 (2020).
86. Shiekh, P. A., Singh, A. & Kumar, A. Oxygen-releasing antioxidant cryogel scaffolds with sustained oxygen delivery for tissue engineering applications. *ACS Appl. Mater. Interfaces* **10**, 18458–18469 (2018).
87. Lu, Z., Jiang, X., Chen, M., Feng, L. & Kang, Y. J. An oxygen-releasing device to improve the survival of mesenchymal stem cells in tissue engineering. *Biofabrication* **11**, 045012 (2019).
88. Erdem, A. et al. 3D bioprinting of oxygenated cell-laden gelatin methacryloyl constructs. *Adv. Healthc. Mater.* **9**, 1901794 (2020).
89. Jägers, J., Wrobeln, A. & Ferenz, K. B. Perfluorocarbon-based oxygen carriers: from physics to physiology. *Pflug. Arch.* **473**, 139–150 (2021).
90. Winslow, R. M. *Blood Substitutes*. 5–16. (ed. Winslow, R. M.) (Academic Press, 2006).
91. Riess, J. G. & Krafft, M. P. *Blood Substitutes*. p. 259–275 (ed. Winslow, R. M.) (Academic Press, 2006).
92. Boyd, J., Parkinson, C. & Sherman, P. Factors affecting emulsion stability, and the HLB concept. *J. Colloid Interface Sci.* **41**, 359–370 (1972).
93. Caballero-Aguilar, L. M. et al. Microencapsulation of growth factors by microfluidic system. *MethodsX* **8**, 101324 (2021).
94. Fu, X. et al. Bioinspired perfluorocarbon-based oxygen carriers with concave shape and deformable shell. *Adv. Mater. Technol.* **7**, 2100573 (2022).
95. Hester, S., Ferenz, K. B., Eitner, S. & Langer, K. Development of a lyophilization process for long-term storage of albumin-based perfluorodecalin-filled artificial oxygen carriers. *Pharmaceutics* **13**, 584 (2021).
96. Ma, P. X. & Choi, J. W. Biodegradable polymer scaffolds with well-defined interconnected spherical pore network. *Tissue Eng.* **7**, 23–33 (2001).
97. Lin, X. et al. Perfluorocarbon-encapsulated porphyrin-lipid nanoparticles as a photoactive pyroptosis inducer for cancer therapy. *J. Biomed. Nanotechnol.* **19**, 28–35 (2023).
98. Sovacool, B. K., Griffiths, S., Kim, J. & Bazilian, M. Climate change and industrial F-gases: a critical and systematic review of developments, sociotechnical systems and policy options for reducing synthetic greenhouse gas emissions. *Renew. Sustain. Energy Rev.* **141**, 110759 (2021).
99. Cohn, C. S. & Cushing, M. M. Oxygen therapeutics: perfluorocarbons and blood substitute safety. *Crit. Care Clin.* **25**, 399–414 (2009).
100. Arnaud, F., Sanders, K., Sieckmann, D. & Moon-Massat, P. In vitro alteration of hematological parameters and blood viscosity by the perfluorocarbon: Oxycyte. *Int. J. Hematol.* **103**, 584–591 (2016).
101. Toffaletti, J. G. & Wildermann, R. F. Use of turbidity-correction algorithm eliminates the effect of perflubron emulsion on CO-oximeter results. *Clin. Chem.* **46**, 136–137 (2000).
102. Lambert, E. & Janjic, J. M. Quality by design approach identifies critical parameters driving oxygen delivery performance in vitro for perfluorocarbon based artificial oxygen carriers. *Sci. Rep.* **11**, 5569 (2021).
103. Epstein, P. S. & Plesset, M. S. On the stability of gas bubbles in liquid gas solutions. *J. Chem. Phys.* **18**, 1505–1509 (2004).
104. Heo, H. et al. Proliferative effects of nanobubbles on fibroblasts. *Biomed. Eng. Lett.* **12**, 393–400 (2022).
105. Kheir, J. N. et al. Bulk manufacture of concentrated oxygen gas-filled microparticles for intravenous oxygen delivery. *Adv. Healthc. Mater.* **2**, 1131–1141 (2013).
106. Swanson, E. J. & Borden, M. A. Injectable oxygen delivery based on protein-shelled microbubbles. *Nano LIFE* **01**, 215–218 (2010).
107. Farris, A. L. et al. 3D-printed oxygen-releasing scaffolds improve bone regeneration in mice. *Biomaterials* **280**, 121318 (2022).
108. Nirmalkar, N., Pacek, A. W. & Barigou, M. On the existence and stability of bulk nanobubbles. *Langmuir* **34**, 10964–10973 (2018).
109. Vutha, A. K. et al. A microfluidic device for real-time on-demand intravenous oxygen delivery. *Proc. Natl Acad. Sci. USA* **119**, e2115276119 (2022).
110. Paknahad, A. A., Zalloum, I. O., Karshafian, R., Kolios, M. C. & Tsai, S. S. Microfluidic nanobubbles: observations of a sudden contraction of microbubbles into nanobubbles. *Soft Matter* **19**, 5142–5149 (2023).
111. McDannold, N., Zhang, Y. & Vykhodtseva, N. The effects of oxygen on ultrasound-induced blood-brain barrier disruption in mice. *Ultrasound Med. Biol.* **43**, 469–475 (2017).
112. Wang, F. et al. Ultrasound-triggered drug delivery for glioma therapy through gambogic acid-loaded nanobubble-microbubble complexes. *Biomed. Pharmacother.* **150**, 113042 (2022).
113. Henstock, J. R. & El Haj, A. J. *Mechanobiology*, p. 275–296 (John Wiley & Sons, Ltd, 2017).
114. Vukasovic, A. et al. Bioreactor-manufactured cartilage grafts repair acute and chronic osteochondral defects in large animal studies. *Cell Prolif.* **52**, e12653 (2019).
115. Kaufman, M. H., Torgeson, J. & Stegen, J. C. Metabolic multi-reactor: practical considerations for using simple oxygen sensing optodes for high-throughput batch reactor metabolism experiments. *PLoS ONE* **18**, e0284256 (2023).
116. Luan, H. et al. Complex 3D microfluidic architectures formed by mechanically guided compressive buckling. *Sci. Adv.* **7**, eabj3686 (2021).

117. Ameri, A. R., Imanparast, A., Passandideh-Fard, M. & Mousavi Shaegh, S. A. A whole-thermoplastic microfluidic chip with integrated on-chip micropump, bioreactor and oxygenator for cell culture applications. *Anal. Chim. Acta* **1221**, 340093 (2022).
118. Islam, S. et al. Totally implantable oxygen generator (TIOG) for hypoxia and hypoxemia. *IEEE Trans. Biomed. Eng.* **70**, 1380–1388 (2023).
119. Scianmarello, N. et al. Oxygen generation by electrolysis to treat retinal ischemia. In *2016 IEEE 29th International Conference on Micro Electro Mechanical Systems (MEMS)*, 399–402 (2016).
120. Maharbiz, M., Holtz, W., Sharifzadeh, S., Keasling, J. & Howe, R. A microfabricated electrochemical oxygen generator for high-density cell culture arrays. *J. Microelectromech. Syst.* **12**, 590–599 (2003).
121. Safaee, M. M., Gravely, M. & Roxbury, D. A wearable optical microfibrous biomaterial with encapsulated nanosensors enables wireless monitoring of oxidative stress. *Adv. Funct. Mater.* **31**, 2006254 (2021).
122. Sonmezoglu, S., Fineman, J. R., Maltepe, E. & Maharbiz, M. M. Monitoring deep-tissue oxygenation with a millimeter-scale ultrasonic implant. *Nat. Biotechnol.* **39**, 855–864 (2021).
123. Ochoa, M. et al. Integrated sensing and delivery of oxygen for next-generation smart wound dressings. *Microsyst. Nanoeng.* **6**, 1–16 (2020).
124. Weidemann, A. & Johnson, R. S. Biology of HIF-1 α . *Cell Death Differ.* **15**, 621–627 (2008).
125. Li, X. et al. Targeting mitochondrial reactive oxygen species as novel therapy for inflammatory diseases and cancers. *J. Hematol. Oncol.* **6**, 19 (2013).
126. Suh, Y.-A. et al. Cell transformation by the superoxide-generating oxidase Mox1. *Nature* **401**, 79–82 (1999).
127. Checa, J. & Aran, J. M. Reactive oxygen species: drivers of physiological and pathological processes. *J. Inflamm. Res.* **13**, 1057–1073 (2020).
128. Granger, D. N. & Kvietys, P. R. Reperfusion injury and reactive oxygen species: the evolution of a concept. *Redox Biol.* **6**, 524–551 (2015).
129. Mayevsky, A. Brain NADH redox state monitored in vivo by fiber optic surface fluorometry. *Brain Res. Rev.* **7**, 49–68 (1984).
130. Turrens, J. F., Alexandre, A. & Lehninger, A. L. Ubisemiquinone is the electron donor for superoxide formation by complex III of heart mitochondria. *Arch. Biochem. Biophys.* **237**, 408–414 (1985).
131. Cheng, Y. et al. Perfluorocarbon nanoparticles enhance reactive oxygen levels and tumour growth inhibition in photodynamic therapy. *Nat. Commun.* **6**, 8785 (2015).
132. Yang, Z. et al. Targeted myoglobin delivery as a strategy for enhancing the sensitivity of hypoxic cancer cells to radiation. *iScience* **23**, 101158 (2020).
133. Rouwkema, J. & Khademhosseini, A. Vascularization and angiogenesis in tissue engineering: beyond creating static networks. *Trends Biotechnol.* **34**, 733–745 (2016).
134. Wang, Z. et al. Rapid regeneration of a neoartery with elastic lamellae. *Adv. Mater.* **34**, 2205614 (2022).
135. Niklason, L. E. & Lawson, J. H. Bioengineered human blood vessels. *Science* **370**, eaaw8682 (2020).
136. Linville, R. M. et al. Human iPSC-derived blood-brain barrier microvessels: validation of barrier function and endothelial cell behavior. *Biomaterials* **190–191**, 24–37 (2019).
137. Kinstlinger, I. S. et al. Perfusion and endothelialization of engineered tissues with patterned vascular networks. *Nat. Protoc.* **16**, 3089–3113 (2021).
138. White, S. M. et al. Longitudinal in vivo imaging to assess blood flow and oxygenation in implantable engineered tissues. *Tissue Eng. Part C Methods* **18**, 697–709 (2012).
139. Brady, E. L. et al. Engineered tissue vascularization and engraftment depends on host model. *Sci. Rep.* **13**, 1973 (2023).
140. O'Connor, C., Brady, E., Zheng, Y., Moore, E. & Stevens, K. R. Engineering the multiscale complexity of vascular networks. *Nat. Rev. Mater.* **7**, 702–716 (2022).
141. Baranski, J. D. et al. Geometric control of vascular networks to enhance engineered tissue integration and function. *Proc. Natl Acad. Sci. USA* **110**, 7586–7591 (2013).
142. Unagolla, J. M. & Jayasuriya, A. C. Hydrogel-based 3D bioprinting: a comprehensive review on cell-laden hydrogels, bioink formulations, and future perspectives. *Appl. Mater. Today* **18**, 100479 (2020).
143. Jain, P., Kathuria, H. & Dubey, N. Advances in 3D bioprinting of tissues/organs for regenerative medicine and in-vitro models. *Biomaterials* **287**, 121639 (2022).
144. Yu, Y., Shang, L., Guo, J., Wang, J. & Zhao, Y. Design of capillary microfluidics for spinning cell-laden microfibers. *Nat. Protoc.* **13**, 2557–2579 (2018).
145. Xie, R. et al. Composable microfluidic spinning platforms for facile production of biomimetic perfusable hydrogel microtubes. *Nat. Protoc.* **16**, 937–964 (2021).
146. Grebenyuk, S. et al. Large-scale perfused tissues via synthetic 3D soft microfluidics. *Nat. Commun.* **14**, 193 (2023).
147. Chen, W. C. et al. Controlled dual delivery of fibroblast growth factor-2 and Interleukin-10 by heparin-based coacervate synergistically enhances ischemic heart repair. *Biomaterials* **72**, 138–151 (2015).
148. Caballero Aguilar, L. M., Silva, S. M. & Moulton, S. E. Growth factor delivery: defining the next generation platforms for tissue engineering. *J. Control. Release* **306**, 40–58 (2019).
149. Ritger, P. L. & Peppas, N. A. A simple equation for description of solute release I. Fickian and non-fickian release from non-swelling devices in the form of slabs, spheres, cylinders or discs. *J. Control. Release* **5**, 23–36 (1987).
150. Brazel, C. S. & Peppas, N. A. Modeling of drug release from Swellable polymers. *Eur. J. Pharm. Biopharm.* **49**, 47–58 (2000).
151. Caballero Aguilar, L., Stoddart, P. R., McArthur, S. L. & Moulton, S. E. Polycaprolactone porous template facilitates modulated release of molecules from alginate hydrogels. *React. Funct. Polym.* **133**, 29–36 (2018).
152. Soltani Dehnavi, S. et al. Molecular camouflage by a context-specific hydrogel as the key to unlock the potential of viral vector gene therapy. *Chem. Eng. J.* **477**, 146857 (2023).
153. Park, S. & Park, K. M. Hyperbaric oxygen-generating hydrogels. *Biomaterials* **182**, 234–244 (2018).
154. Ding, J. et al. A reactive oxygen species scavenging and o₂ generating injectable hydrogel for myocardial infarction treatment in vivo. *Small* **16**, 2005038 (2020).

Acknowledgements

This work was supported by the the Australian Research Council Future Fellowship project FT230100220 (D.N.) and St. Vincent's Hospital Melbourne Research Endowment Fund project 94519 (L.C.A.).

Author contributions

Formulation of the concept and structure: L.M.C.A. and D.R.N.; main writing and editing of the manuscript: E.Z. and L.M.C.A.; figure generation and editing: L.M.C.A., E.Z., O.S., and C.O.; subsection writing, manuscript reviewing and editing: Y.W., C.J., O.S., S.D., C.O., B.F., S.E.M., R.W., C.P., and D.R.N.

Competing interests

The authors declare no competing interests.

Additional information

Correspondence and requests for materials should be addressed to David R. Nisbet or Lilith M. Caballero-Aguilar.

Peer review information *Nature Communications* thanks Nic Leipzig, and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

Reprints and permissions information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2024