

Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Sukee, T;Beveridge, I;Koehler, AV;Hall, R;Gasser, RB;Jabbar, A

Title:

Phylogenetic relationships of the nematode subfamily Phascolostrongylinae from macropodid and vombatid marsupials inferred using mitochondrial protein sequence data

Date:

2021-12-01

Citation:

Sukee, T., Beveridge, I., Koehler, A. V., Hall, R., Gasser, R. B. & Jabbar, A. (2021). Phylogenetic relationships of the nematode subfamily Phascolostrongylinae from macropodid and vombatid marsupials inferred using mitochondrial protein sequence data. *Parasites and Vectors*, 14 (1), <https://doi.org/10.1186/s13071-021-05028-2>.

Persistent Link:

<https://hdl.handle.net/11343/290352>

License:

CC BY

RESEARCH

Open Access



Phylogenetic relationships of the nematode subfamily Phascolstrongylinae from macropodid and vombatid marsupials inferred using mitochondrial protein sequence data

Tanapan Sukee^{1*} , Ian Beveridge^{1,2}, Anson V. Koehler¹, Ross Hall¹, Robin B. Gasser¹ and Abdul Jabbar¹

Abstract

Background: The subfamily Phascolstrongylinae (Superfamily Strongyloidea) comprises nematodes that are parasitic in the gastrointestinal tracts of macropodid (Family Macropodidae) and vombatid (Family Vombatidae) marsupials. Currently, nine genera and 20 species have been attributed to the subfamily Phascolstrongylinae. Previous studies using sequence data sets for the internal transcribed spacers (ITS) of nuclear ribosomal DNA showed conflicting topologies between the Phascolstrongylinae and related subfamilies. Therefore, the aim of this study was to validate the phylogenetic relationships within the Phascolstrongylinae and its relationship with the families Chabertiidae and Strongylidae using mitochondrial amino acid sequences.

Methods: The sequences of all 12 mitochondrial protein-coding genes were obtained by next-generation sequencing of individual adult nematodes ($n = 8$) representing members of the Phascolstrongylinae. These sequences were conceptually translated and the phylogenetic relationships within the Phascolstrongylinae and its relationship with the families Chabertiidae and Strongylidae were inferred from aligned, concatenated amino acid sequence data sets.

Results: Within the Phascolstrongylinae, the wombat-specific genera grouped separately from the genera occurring in macropods. Two of the phascolstrongyline tribes were monophyletic, including Phascolstrongylinea and Hypodontinea, whereas the tribe Macropostrongyloidea was paraphyletic. The tribe Phascolstrongylinea occurring in wombats was closely related to *Oesophagostomum* spp., also from the family Chabertiidae, which formed a sister relationship with the Phascolstrongylinae.

Conclusion: The current phylogenetic relationship within the subfamily Phascolstrongylinae supports findings from a previous study based on ITS sequence data. This study contributes also to the understanding of the phylogenetic position of the subfamily Phascolstrongylinae within the Chabertiidae. Future studies investigating the relationships between the Phascolstrongylinae and Cloacininae from macropodid marsupials may advance our knowledge of the phylogeny of strongyloid nematodes in marsupials.

Keywords: Phascolstrongylinae, Macropodid marsupials, Vombatid marsupials, Mitochondrial DNA

*Correspondence: tsukee@student.unimelb.edu.au

¹ Department of Veterinary Biosciences, Melbourne Veterinary School, The University of Melbourne, Victoria, Australia

Full list of author information is available at the end of the article



© The Author(s) 2021. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

Background

The Phascolostromylinae is a subfamily of strongyloid nematode (Nematoda: Strongyloidea) belonging to the family Chabertiidae. The subfamily comprising nine genera and 20 species, parasitises macropodid (Family Macropodidae) and vombatid (Family Vombatidae) marsupials. Most of the phascolostromyline genera occur within the intestines of their hosts. However, a few exceptions are found in the stomachs of their hosts, including *Paramacropostrongylus* from grey kangaroos (*Macropus* spp.) and *Wallabicola* from swamp wallabies (*Wallabia bicolor*) [1].

Genera of the Phascolostromylinae are currently subdivided into three tribes (Phascolostromylinea, Hypodontinea and Macropostrongyloidinea) based primarily on the features of the buccal capsule [2]. The tribe Phascolostromylinea, characterised by leaf crown elements surrounding the buccal capsule, consists of *Phascolostromylus* and *Oesophagostomoides* and occurs exclusively in wombats. Hypodontinea comprises genera with globular buccal capsules, namely *Hypodontus*, *Macropicola* and *Corollostrongylus*. The genera of the Macropostrongyloidinea, including *Macropostrongyloides*, *Paramacropostrongylus*, *Torquenema* and *Wallabicola*, possess cylindrical buccal capsules surrounded by teeth or denticles [2]. The morphological classification of the Phascolostromylinae has been uncertain due to extensive variation in oral morphology that can be challenging to observe [1, 2]. Therefore, molecular markers, including the first and second region of internal transcribed spacers (ITS-1 and ITS-2) of ribosomal DNA (rDNA), have been utilised for specific identification purposes, detection of genetic variation and phylogenetic analyses within the Phascolostromylinae [3–9]. In a recent study, two species (*Macropostrongyloides dissimilis* and *Paramacropostrongylus toraliformis*) were found to be divergent from their congeners using ITS markers [9]. Subsequently, morphological examination of these nematodes led to the description of two new genera, namely *Wallabicola dissimilis* (formerly *M. dissimilis*) from the swamp wallaby, *Wallabicola bicolor* and *Torquenema toraliforme* (formerly *P. toraliformis*) from the eastern grey kangaroo *Macropus giganteus* [10].

Although ITS sequence data can be useful for inferring phylogenetic relationships at the species and genus level, this has not been the case at higher taxonomic levels. Phylogenetic analyses within the superfamily Strongyloidea determined using the ITS-2 marker showed conflicting topologies and low nodal support for the relationships between the Phascolostromylinae, Oesophagostominae and Chabertiinae [11]. Amino acid sequences derived from the mitochondrial protein-coding genes have been used to validate phylogenetic relationships

within *Hypodontus* [12] and *Macropostrongyloides* [13] of the Phascolostromylinae. These studies validated previous phylogenetic hypotheses based on ITS sequence data with improved nodal support in the phylogenetic analyses.

Therefore, in the present study we assessed the phylogenetic relationships within the subfamily Phascolostromylinae utilising the amino acid sequence data sets derived from the mitochondrial protein-coding genes. We also assessed the phylogenetic relationship between genera of the Phascolostromylinae with other subfamilies of the Chabertiidae and with the Strongyloidea. Published mitochondrial protein sequences of genera from these families were included in the phylogenetic analyses.

Methods

Sample collection and DNA extraction

Adult male and female nematodes used in the current study (Table 1) were obtained from the frozen parasite collection at the School of Veterinary Science, The University of Melbourne. The nematodes were collected from the gastrointestinal tracts of carcasses of hosts from commercial cullings or vehicle collisions (State-issued permits: Victorian Department of Sustainability and Environment 90-053, 93-016, 10000434, 100003649; Queensland Department of Environment and Heritage Protection WA00006125). The samples had been either frozen at -80°C or preserved in 70% ethanol and then frozen at -80°C as individuals or pools. If preserved in ethanol, worms were rehydrated in distilled water prior to DNA isolation. For morphological identification, the anterior and posterior extremities of each nematode were removed with a scalpel blade, cleared with lactophenol and identified. They were subsequently deposited in the helminthological collection of the South Australia Museum, Adelaide, as voucher specimens (Registration numbers Australian Helminthological Collection [AHC] 36783, 49028, 49037, 49035, 49052, 49051, 49055, 49108). The mid-sections were used for the DNA isolation.

Genomic DNA was extracted from individual nematodes using the QiaAmp Micro Kit (Qiagen, Hilden, Germany) following the manufacturer's protocol for extracting DNA from tissues. For initial molecular identification, the ITS-1 and ITS-2 sequences were determined for each individual using an established PCR-based sequencing method [9]. Prior to sequencing, the quantity and quality of the DNA were determined using the 2200 TapeStation (Agilent Technologies, Santa Clara, CA, USA).

Sequencing and gene annotation

Illumina TruSeq indexed libraries were prepared using sheared DNA following the manufacturer's protocol

Table 1 Species of Phascolostromylinae included in the current study and reference sequences obtained from GenBank database

Family/subfamily	Species ^a	Host	Site within host ^b	Collection locality ^c	GenBank accession no.	References
Subfamily Phascolostromylinae	<i>Oesophagostomoides giltneri</i> (P)	<i>Vombatus ursinus</i>	LI	Flowerdale, Vic, Australia	OK111101	This study
	<i>Oesophagostomoides longispicularis</i> (P)	<i>Vombatus ursinus</i>	LI	Gippsland, Vic, Australia	OK111102	This study
	<i>Oesophagostomoides stirtoni</i> (P)	<i>Lasiorhinus latifrons</i>	LI	Swan Reach, SA, Australia	OK111103	This study
	<i>Phascolostromylus turleyi</i> (P)	<i>Vombatus ursinus</i>	LI	Flowerdale, Vic, Australia	OK111104	This study
	<i>Paramacropostrongylus iugalis</i> (M)	<i>Macropus giganteus</i>	S	Miles, Qld, Australia	OK111105	This study
	<i>Paramacropostrongylus iugalis</i> (M)	<i>Macropus giganteus</i>	S	Charters Towers, Qld, Australia	OK111106	This study
	<i>Paramacropostrongylus typicus</i> (M)	<i>Macropus fuliginosus</i>	LI	Nyngan, NSW, Australia	OK111107	This study
	<i>Torquenema toraliforme</i> (M)	<i>Macropus giganteus</i>	LI	Research, Vic, Australia	OK111108	This study
	<i>Macropostrongyloides mawsonae</i> (M)	<i>Macropus giganteus</i>	LI	Heathcote, Vic, Australia	MW309873	[13]
	<i>Macropostrongyloides baylisi</i> (M)	<i>Osphranter robustus</i>	LI	Cloncurry, Qld, Australia	MW309874	[13]
	<i>Macropostrongyloides yamagutii</i> (M)	<i>Macropus fuliginosus</i>	LI	Hattah Lakes, Vic, Australia	MW309875	[13]
	<i>Macropostrongyloides spearei</i> (M)	<i>Osphranter robustus</i>	LI	Kalgoorlie, WA, Australia	MW309876	[13]
	<i>Macropostrongyloides phascolomys</i> (M)	<i>Vombatus ursinus</i>	LI	Flowerdale, Vic, Australia	MW309877	[13]
	<i>Macropostrongyloides woodi</i> (M)	<i>Osphranter rufus</i>	LI	Kalgoorlie, WA, Australia	MW309878	[13]
	<i>Wallabicola dissimilis</i> (M)	<i>Wallabia bicolor</i>	S	Kamarooka, Vic, Australia	MW309879	[13]
	<i>Hypodontus macropi</i> (H)	<i>Wallabia bicolor</i>	SI + LI	Hall's Gap, Vic, Australia	KF361317	[12]
	<i>Hypodontus macropi</i> (H)	<i>Thylogale billardieri</i>	SI + LI	Launceston, Tas, Australia	KF361318	[12]
	<i>Hypodontus macropi</i> (H)	<i>Macropus robustus</i>	SI + LI	Barcaldine, Qld, Australia	KF361319	[12]
	<i>Macropicola ocydromi</i> (H)	<i>Macropus fuliginosus</i>	LI	Waroon, WA, Australia	KF361320	[12]
Subfamily Oesophagostominae	<i>Oesophagostomum dentatum</i>	<i>Sus scrofa domestica</i>	LI	Chongqing, China	FM161882	[16]
	<i>Oesophagostomum quadrispinulatum</i>	<i>Sus scrofa domestica</i>	LI	Chongqing, China	FM161883	[16]
	<i>Oesophagostomum dentatum</i>	<i>Sus scrofa domestica</i>	LI	Werribee, Vic, Australia	GQ888716	[15]
	<i>Oesophagostomum asperum</i>	<i>Capra hircus</i>	LI	Shaanxi Province, China	KC715826	[17]
	<i>Oesophagostomum columbianum</i>	<i>Ovis aries</i>	LI	Heilongjiang Province, China	KC715827	[17]
Subfamily Chabertiinae	<i>Chabertia ovina</i>	<i>Ovis aries</i>	LI	Werribee, Vic, Australia	GQ888721	[15]
	<i>Chabertia ovina</i>	<i>Capra hircus</i>	LI	Shaanxi Province, China	KF660604	[18]
	<i>Chabertia ershowi</i>	<i>Bos grunniens</i>	LI	Qinghai Province, China	KF660603	[18]
Family Strongylidae	<i>Cylicodontophorus bicornatus</i>	<i>Equus caballus</i>	LI	Heilongjiang Province, China	MH551241	[19]
	<i>Strongylus vulgaris</i>	<i>Equus caballus</i>	LI	Vic, Australia	GQ888717	[15]
Family Syngamidae	<i>Syngamus trachea</i>	<i>Gymnorhina tibicen</i>	T	Vic, Australia	GQ888718	[15]

^a H, Tribe Hypodontinae; M, tribe Macropostrongyloidea; P, tribe Phascolostromylinae^b LI, Large intestine; S, stomach; SI, small intestine; T, trachea^c NSW, New South Wales; Qld, Queensland; SA, South Australia; Tas, Tasmania; Vic, Victoria; WA, Western Australia

(Illumina Inc., San Diego, CA, USA). Briefly, the steps included: (i) end-repair and A-tailing of the 3' ends; (ii) ligation of the adaptors; (iii) enrichment of the libraries and purification of the enriched library using Ampure Beads (Beckman Coulter, Brea, CA, USA). The libraries were quantified using the 2200 TapeStation, pooled and sequenced on the Illumina MiSeq platform using the 300 cycle v3 reagent kit (2×150 paired-end reads). Illumina library preparation and sequencing were carried out at the Walter and Eliza Hall Institute (WEHI) Genomics Facility, Melbourne, Victoria, Australia.

Raw sequence data in the FASTQ format were filtered for quality in Trimmomatic v.0.38 [14] prior to de novo assembly employing the program Spades v3.13.0 under default parameters. For each assembly, the 12 protein-coding genes of the mitochondrial genome were identified by local sequence alignment (6 reading frames) using the amino acid sequence inferred from corresponding genes of reference mitochondrial genomes, using an established workflow system [15]. The reference mitochondrial genomes used were selected based on sequence similarities using Basic Local Alignment Search Tool (BLAST) for sequence analysis [20]. The mitochondrial genome sequence of *Hypodontus macropi* (NC023083) was used as a reference for *Paramacropostrongylus*, *Torquenema* and *Wallabicola*, and that of *Oesophagostomum dentatum* (NC013817) was used as the reference for *Oesophagostomoides* and *Phascolostrongylus*. The nucleotide sequences of the 12 protein-coding genes of each species included in this study were deposited in the GenBank database under the accession numbers OK111101–OK111108.

Sequence comparison and phylogenetic analyses

The nucleotide and amino acid sequences of the 12 mitochondrial protein-coding genes were aligned separately using CLUSTAL W [21] and MUSCLE [22] followed by a concatenation of the alignments using MEGA software version X [23]. The alignment included previously published mitochondrial protein-coding gene sequences of species of Phascolostrongylinae, Chabertiinae, Oesophagostominae, Strongylidae and Syngamidae (Table 1). Pairwise comparisons were calculated for the nucleotide and amino acid sequences using MEGA software. The nucleotide diversity across each of the protein-coding genes was determined using sliding window analyses (SWAN) carried out in the DnaSP v.5 program [24] using a sliding window of 100 bp and 25-bp steps.

Phylogenetic analysis was conducted using concatenated and aligned amino acid sequences derived from all 12 mitochondrial protein-coding genes. The phylogenetic analysis was conducted using Bayesian inference (BI) in MrBayes v.3.2.7 [25]. The optimal partitioning

schemes and substitution model for the BI analyses were determined using PartitionFinder 2 [26] for amino acids, with a model selection set to the Akaike information criterion and greedy search algorithm. According to PartitionFinder, the amino acid sequence alignment was partitioned into eight subsets consisting of subset 1 (cytochrome *c* oxidase [*cox*] subunit 1-2), subset 2 (*cox3*, nicotinamide adenine dinucleotide hydrogen dehydrogenase [*nad*] subunit 4), subset 3 (*nad5*), subset 4 (*nad6*, *nad2*), subset 5 (*nad4L*, *nad1*), subset 6 (adenosine triphosphate synthase subunit 6 [*atp6*]), subset 7 (cytochrome *b* [*cob*]) subset 8 (*cox3*). The Mtmam (Mitochondrial Mammalia) model was the evolutionary model for all subsets except subset 6 (general reversible Markov model for amino acid substitution of mitochondrial proteins [Mtrev] model). The BI analysis was conducted with four chains each of 10 million Markov chain Monte Carlo iterations, sampling every 1000th generation for four independent runs. Convergence was determined by the average standard deviation of split frequencies of >0.01 , with the potential scale reduction factor approaching 1. The first 25% of the sampled trees were discarded as burnin and the Bayesian consensus tree was constructed from the remaining trees. The BI trees were visualised in FigTree v.1.4.4 [27]. *Syngamus trachea* (GQ888718) belonging to the family Syngamidae was used as the outgroup.

Results

Nucleotide and amino acid sequence comparisons

The SWAN revealed that the nucleotide diversity across the alignment of 12 concatenated mitochondrial protein-coding genes ranged from 0.084 to 0.290 (Fig. 1). The 5' prime end of the *nad5* gene exhibited the highest level of nucleotide diversity, whereas *cox1* was the most conserved gene.

Pairwise amino acid sequence differences among species of Phascolostrongylinae ranged between 0.60% (between two specimens of *Paramacropostrongylus iugalis*) and 10.6% (between *Phascolostrongylus turleyi* and *Macropostrongyloides baylisi*) (Table 2). The genus *Wallabicola* was most similar to *Paramacropostrongylus typicus*, with 5.9% sequence variation, compared to 8.6% variation from *Macropostrongyloides* in which it was formerly placed. The amino acid sequence of *Torquenema* was most similar to that of *Paramacropostrongylus typicus* (6.5% sequence variation), the genus to which it was formerly assigned. Amino acid sequence variation between the wombat-specific genera *Phascolostrongylus* and *Oesophagostomoides* spp. was small (4.7–5.0%), with *Ph. turleyi* and *Oesophagostomoides longspicularis* sharing the greatest sequence similarities. Interestingly, the amino acid sequences of *Phascolostrongylus*

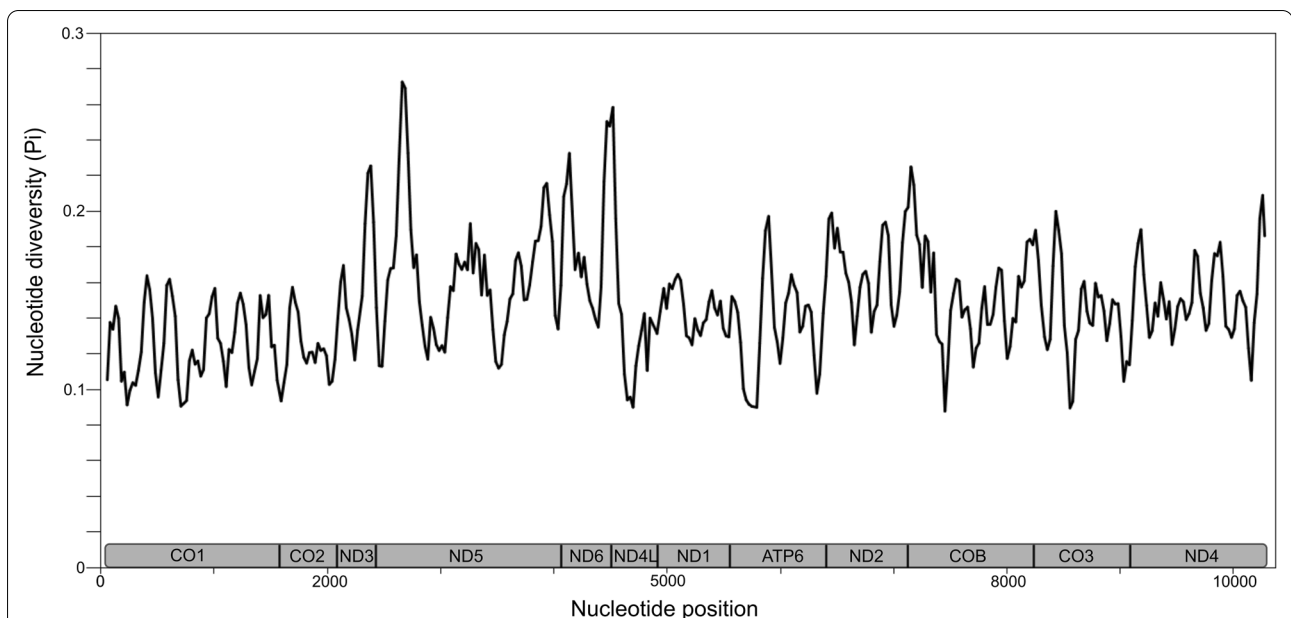


Fig. 1 Nucleotide diversity (Pi) across 12 concatenated mitochondrial protein-coding genes (y-axis) of eight genera of Phascolostromylinae, two genera of Strongylidae and one genus each of Oesophagostominae, Chabertiinae and Syngamidae. Nucleotide diversity was calculated in DnaSP version 6 software using a window of 100 bp and 25-bp steps. The nucleotide position (base pairs) is indicated on the x-axis next to the boundaries between mitochondrial protein-coding genes. CO1 – 3 = Cytochrome c oxidase subunit 1 – 3; ND1 – 6, 4L = Nicotinamide adenine dinucleotide hydrogen dehydrogenase subunits 1 – 6 and 4L; ATP6 = Adenosine triphosphate synthase subunit 6; COB = Cytochrome B

and *Oesophagostomoides* spp. shared greater similarities with two species from the subfamily Oesophagostominae, *Oe. dentatum* and *Oesophagostomum quadrispinulatum* (6.8–7.3% sequence variation) compared to genera from the same subfamily (9.1–10.6% sequence variation) (Table 2).

Phylogenetic analyses

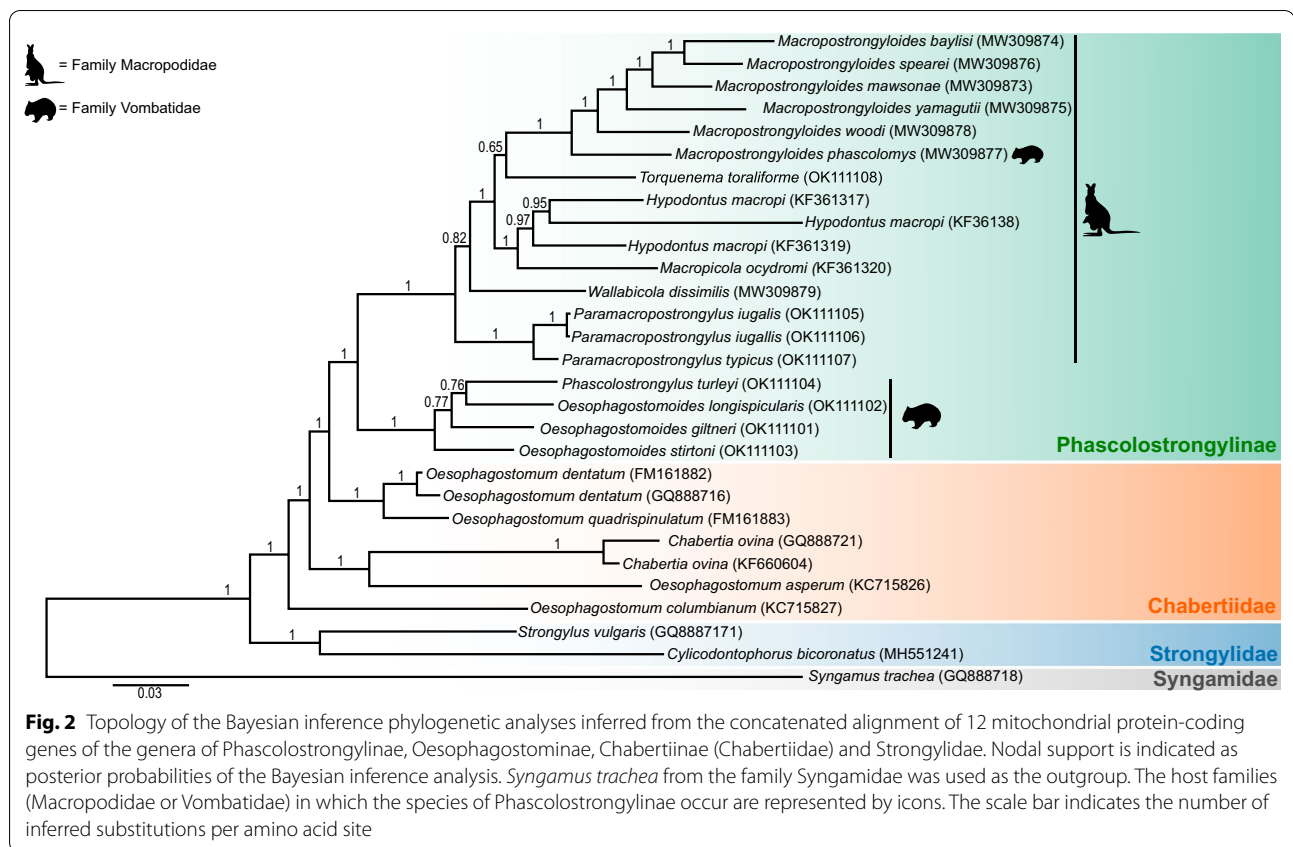
The BI analysis showed strong support for the clustering of the genera of Phascolostromylinae (Fig. 2). The tree topology showed that the genera of the Phascolostromylinae distributed between two clades, with the larger clade comprising members of the tribes Macropostromylinae and Hypodontinae from macropodid marsupials and the smaller clade comprising the tribe Phascolostromylinae from vombatid marsupials (Fig. 2). The tribe Phascolostromylinae comprising the wombat-specific genera *Phascolostromylus* and *Oesophagostomoides* formed a strongly supported monophyletic assemblage (posterior probability [pp] = 1), sister to the species occurring in macropodid marsupials and *Macropostromyloides phascolomys* from the common wombat. The clade comprising *Hypodontus* and *Macropicola* belonging to the tribe Hypodontinae was monophyletic, with strong nodal support (pp = 1). The tribe Macropostromyloidea was

paraphyletic. *Macropostromyloides* was placed as a sister group to the tribe Hypodontinae, and *Wallabicola dissimilis* from the stomach of the swamp wallaby was placed on a branch external to the clade comprising *Hypodontus* and *Macropostromyloides*. The position of *T. toraliforme*, sister to *Macropostromyloides* lacked nodal support (pp = 0.65). *Paramacropostromylus typicus* and *P. iugalis* from grey kangaroos grouped together with strong nodal support (pp = 1).

Within the tribe Oesophagostominae, *Oesophagostomum dentatum* and *Oesophagostomum quadrispinulatum* formed a strongly supported (pp = 1) clade sister to the subfamily Phascolostromylinae, with the exclusion of *Oe. asperum* and *Oe. columbianum*. *Oesophagostomum asperum* clustered with *Chabertia ovina* and *C. erschowi* (tribe Chabertiinae) whilst *Oe. columbianum* was placed on an external branch. The genera *Strongylus vulgaris* and *Cyclicodontophorus bicoronatus* are representative of the family Strongylidae that is sister to the subfamilies Phascolostromylinae, Oesophagostominae and Chabertiinae, all of which belong to the family Chabertiidae. Overall there were six genera comprising at least two species included in the study, four of these were monophyletic (*Macropostromyloides*, *Hypodontus*, *Paramacropostromylus* and *Chabertia*) and two were paraphyletic (*Oesophagostomoides* and *Oesophagostomum*).

Table 2 Pairwise difference (%) of the nucleotide (top) and amino acid (bottom) sequences derived from the 12 concatenated mitochondrial protein-coding genes among species of the Phascolostromylinae, Oesophagostominae and Chabertiinae

Species	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
1 <i>P. iugalis</i> (OK111105)	–	13.50	13.57	14.97	14.04	14.64	14.92	14.69	14.78	15.16	14.57	14.30	15.45	15.61	15.90
2 <i>T. toraliforme</i> (OK111108)	6.90	–	13.80	14.47	13.97	13.76	15.71	15.48	14.95	15.41	14.95	14.67	15.95	15.74	16.14
3 <i>W. dissimilis</i> (MW309879)	6.04	6.73	–	15.35	15.02	14.60	15.58	15.83	15.53	16.04	15.23	15.18	16.17	16.02	16.27
4 <i>M. baylisi</i> (MW309874)	8.50	8.12	8.68	–	15.32	15.13	16.36	16.60	16.50	16.29	15.98	15.58	16.50	16.27	16.97
5 <i>H. macropi</i> (KF361319)	6.73	6.49	6.49	7.94	–	13.73	15.80	16.17	15.58	16.05	15.22	15.74	16.28	15.83	16.24
6 <i>Ma. ocydromi</i> (KF361320)	7.41	7.14	7.20	7.94	6.07	–	15.54	15.94	15.62	16.28	15.63	16.08	16.16	15.63	16.34
7 <i>O. gilneri</i> (OK111101)	8.68	8.98	8.56	10.22	9.33	9.57	–	12.68	12.78	12.61	14.80	14.87	15.94	15.39	16.41
8 <i>O. longispicularis</i> (OK111102)	8.27	8.44	8.44	10.22	9.07	9.63	4.89	–	12.68	12.68	14.81	14.69	15.84	15.30	16.47
9 <i>O. stirtoni</i> (OK111103)	8.24	8.39	8.47	10.22	8.86	9.39	4.95	5.01	–	12.75	14.58	14.45	15.56	14.90	15.72
10 <i>Ph. turleyi</i> (OK111104)	9.07	9.24	8.95	10.64	9.45	9.96	5.01	4.72	4.74	–	14.72	15.04	16.12	15.64	16.30
11 <i>Oe. dentatum</i> (GQ888716)	8.50	8.74	8.36	10.22	8.15	9.33	7.08	6.90	6.76	7.32	–	12.10	15.39	4.79	15.85
12 <i>Oe. quadrispinulatum</i> (FM161883)	8.09	8.71	8.27	10.10	8.36	9.33	7.29	6.99	6.96	7.41	3.44	–	14.63	14.51	15.69
13 <i>Oe. asperum</i> (KC715826)	11.02	11.17	10.61	12.65	11.23	11.59	10.19	10.43	10.43	11.17	10.04	9.39	–	15.07	15.37
14 <i>Oe. columbianum</i> (KC715827)	11.05	11.26	10.73	12.27	11.14	11.32	10.25	10.13	10.04	10.34	8.98	8.80	11.85	–	15.48
15 <i>C. ovina</i> (GQ888721)	11.47	11.59	11.23	13.07	11.38	11.91	10.73	11.02	10.49	11.35	9.87	9.90	11.69	11.50	–



Discussion

The current study utilised the amino acid sequences derived from the mitochondrial protein-coding genes to assess the phylogenetic relationships of the subfamily Phascolostromylinae. The tree topology showed that the nine genera currently placed in the subfamily Phascolostromylinae clustered together with strong nodal support. Overall, the use of mitochondrial amino acid sequence data sets resulted in well-supported relationships among most but not all taxa included in the phylogenetic analyses. There was good support for the monophyly of the morphologically defined tribes Phascolostromylinae and Hypodontinae but not Macropostromyloidea, consistent with the findings from a previous study using ITS sequence data sets [9].

Contrary to the previous phylogeny based on ITS sequences [9], the BI analysis in the current study showed strong nodal support for the clade containing the genera *Macropicola* and *Hypodontus*. This relationship corresponded with the tribe Hypodontinae erected by Beveridge [2]. The classification of *Hypodontus* and *Macropicola* has been debated in the past as their morphology shares little resemblance to that of other strongyloid nematodes infecting marsupials. *Hypodontus* was initially placed in the family Ancylostomatidae as it

resembled hookworms, due to its ventrally bent anterior extremity and the presence of two cutting plates within the buccal capsule [28]. Subsequently, Inglis et al. [29] transferred *Hypodontus* to the subfamily Globocephaloidinae within the Trichostrongyloidea without a clear explanation. However, it was moved back to the Strongyloidea and reassigned as a hookworm in the subfamily Uncinariinae by Durette-Desset et al. [30], having formerly been placed in the hookworm subfamily Bunostomatinae by Skryabin et al. [31]. The genus *Macropicola*, also in the tribe Hypodontinae, was initially classified as a hookworm in the subfamily Globocephalinae based on its globular buccal capsule and three oesophageal teeth [32]. However, Lichtenfels [33] argued that both *Hypodontus* and *Macropicola* belonged to the subfamily Strongylinae in the Strongylidae based on the globular buccal capsule, oesophageal teeth, the Y-shaped female ovejector and a pre-anal vulva. Finally, in the most recent revision, Beveridge [2] assigned *Hypodontus* and *Macropicola* to the tribe Hypodontinae, along with *Corollostrongylus* within the subfamily Phascolostromylinae, based on their large globular or subglobular buccal capsules that are either straight or bent dorsally or ventrally. This morphological grouping was supported by the mitochondrial protein sequence data in the current study and in a previous

study on hookworms which showed the exclusion of *Hypodontus* and *Macropicola* from the Strongylinae and Ancylostomatoidea [34]. Although *Hypodontus* closely resembles hookworms, molecular data suggest that their morphological resemblances could be a result of convergent evolution. Three genotypes of *Hypodontus macropi*, each from a different host (*Osphranter robustus*, *Thylagale billardieri* and *Wallabia bicolor*) were included in the analyses. The tree topology showed that the specimens of *Hypodontus* from each host are distinct, with the specimen from the Tasmanian pademelon (*T. billardieri*) being the most divergent. This finding is consistent with a previous phylogenetic analysis also based on the mitochondrial protein sequence data [12], in addition to ITS-based studies [3, 6]. In the absence of consistent morphological differences between the distinct genotypes, *H. macropi* remains a cryptic species complex.

Analyses of both the current mitochondrial and previous ITS data sets placed *Torquenema* external to the clade comprising *Macropostrongyloides*, but with low branch support. Although *Torquenema* can be clearly distinguished from *Macropostrongyloides* by its occurrence in the stomach of its host and its prominent cervical collar, they share some common features, including a Y-shaped ovejector and small peri-oral denticles [10, 35]. Mitochondrial and ITS data support the separation of *Torquenema* from its previous position within *Paramacropostrongylus*. However, its current phylogenetic position close to *Macropostrongyloides* is uncertain due to the lack of nodal support.

The current phylogenetic analyses placed *W. dissimilis* (formerly *Macropostrongyloides dissimilis*) external to the clade comprising the genus *Macropostrongyloides* and the tribe Hypodontinae. This finding differs from that of a previous study using ITS sequence data which showed a strongly supported grouping of *W. dissimilis* and *Paramacropostrongylus* spp., both occurring within the stomach of macropodid hosts [9]. However, the current topology based on mitochondrial protein sequence data lacks nodal support (pp=0.82). Therefore, additional data, including a larger number of representatives of this species and potentially a different genetic marker, are required to validate the phylogenetic position of *Wallabicola*.

The current and previous study based on ITS sequence data showed strong support for the monophyletic grouping of the genera within tribe Phascolostrongylinea from vombatid marsupials. The topologies of the current and previous phylogenetic analyses based on ITS sequence data showed the grouping of *Ph. turleyi* and *O. longispicularis* with *O. stirtoni* on a sister branch [9]. However, nodal support was higher in the ITS study [9]. Although *Oesophagostomoides* appears to be paraphyletic

due to the position of *Phascolostrongylus*, there are no morphological features to support this apparent paraphyly. These two genera are considered valid and clearly distinguishable morphologically. The buccal capsules of *Phascolostrongylus* are shallower with thicker walls compared to *Oesophagostomoides* and have a greater number of external leaf crown elements [36]. *Oesophagostomoides longispicularis* and *Ph. turleyi* often occur at a high prevalence and together within the same host, whereas *O. giltneri* has been less commonly encountered in the common wombat [36].

The phylogenetic analysis showed strong support for the sister relationship between the genera of the subfamily Phascolostrongylinea with *Oe. dentatum* and *Oe. quadrispinulatum* (subfamily Oesophagostominae) which was not evident in a previous study based on ITS-2 sequence data [11]. The genus *Oesophagostomum* occurs in the caecum of ungulates, rodents and primates [37]. There are several subgenera within *Oesophagostomum*, some of which corresponded to those included in the current phylogenetic analyses. The separation of *Oe. asperum* from *Oe. columbianum* in the phylogenetic tree corresponded with their different subgenera, *Hysteracrum* and *Proteracrum*, respectively, whereas the clade closest to the subfamily Phascolostrongylinea, comprising *Oe. dentatum* and *Oe. quadrispinulatum*, both belong to the subgenus *Oesophagostomum*. The genus *Chabertia* appears to be monophyletic in the current analyses, although *C. ovina* from a sheep in Australia (GQ888271) was slightly divergent from the one from a goat from China (KF660603), suggesting intraspecific genetic variation within this species. The position of *Chabertia*, nested among the Oesophagostominae, is consistent with the topology of a previous phylogenetic tree based on the analysis of ITS-2 sequence data [38], suggesting that the current classifications of the Chabertiinae and Oesophagostominae may require further investigation.

The genera *Oesophagostomoides* and *Oesophagostomum* were considered synonyms by Popova [39] and Yamaguti [40] based on their morphological similarities. The key differential feature between *Oesophagostomoides* and *Oesophagostomum* are the Y-shaped ovejectors in the former genus compared with J-shaped ovejectors in the latter and the shape of their cervical papillae, which are bipartite in the former but conical in the latter [36]. The sister relationship between *Oesophagostomum* and subfamily Phascolostrongylinea found in the current study raises the question of whether the Oesophagostominae could have been the predecessors of the Strongyloidea in marsupials. However, this proposal was deemed improbable, since the most likely host, rodents, arrived in Australia after the expansive radiation of the marsupials which was believed to coincide with the radiation of their

parasitic nematodes [41]. However, in a study using 18S nuclear ribosomal gene sequence data [42], *Cyclodontostomum purvursi* from Australian rodents grouped closely to *Chabertia ovina*. Further speculation on the association between the Oesophagostominae and Phascolostromylinea is beyond the scope of this study and would require additional mitochondrial sequence datasets of other *Oesophagostomum* spp. and *Cyclodontostomum*, both of which are currently unavailable.

Although the current phylogenetic analyses suggest the monophyly of the Phascolostromylineae, not all species were included. These species were *Corollostrongylus hypsiprymmodontis* (tribe Hypodontinae) from the musky-rat kangaroo *Hypsiprymmodon moschatus* [43], *Macropostrongyloides dendrolagi* (tribe Macropostrongyloidea) from tree kangaroos, *Dendrolagus* spp. [44], *Macropostrongyloides lasiorhini* (tribe Macropostrongyloidea) from the southern hairy-nosed wombat *Lasiorhinus latifrons* and *Oesophagostomoides eppingensis* (tribe Phascolostromylineae) from the northern hairy-nosed wombat *Lasiorhinus krefftii* [45]. Each of these species occurs in rare or endangered hosts and, therefore, specimens for molecular analyses were not readily available through opportunistic sampling.

Overall, the current study demonstrated that phylogenetic inference based on amino acid sequences of mitochondrial protein-coding genes generated strong nodal support. However, the conflicting evidence between the mitochondrial- and ITS-based analyses in the case of *Wallabicola* highlights the importance of using multiple genetic markers to independently validate phylogenetic hypotheses. Furthermore, multiple samples from the same host populations should ideally be included in such analyses. However, this can be challenging due to limitations in collecting sufficient samples from wildlife hosts. Sliding window analyses of the mitochondrial protein-coding genes in current and previous studies suggest that there are variable regions flanked by conserved regions in genes such as *nad1* and *nad5*. PCR-coupled sequencing of such potentially phylogenetically informative regions could reduce the cost and amount of DNA required compared to sequencing mitochondrial genomes.

Conclusions

Utilising the amino acid sequences translated from the mitochondrial protein-coding genes, this study provided evidence for the grouping of the currently recognised genera of the Phascolostromylineae. Findings from the current study contribute to our understanding of the phylogenetic position of the subfamily Phascolostromylineae within the Chabertiidae. This study suggests that the amino acid sequence data could be used to assess the relationship within the subfamily Cloacininae which has

yet to be resolved [46]. The inclusion of genera of the subfamily Cloacininae from macropodid marsupials in future studies could further elucidate the phylogenetic relationships among the strongyloid nematodes from Australian marsupials.

Abbreviations

AHC: Australian Helminthological Collection; atp6: Adenosine triphosphate synthase subunit 6; BI: Bayesian inference; BLAST: Basic Local Alignment Search Tool; cob: Cytochrome *b*; cox1-3: Cytochrome *c* oxidase subunit 1-3; ITS-1: Internal transcribed spacer region 1; ITS-2: Internal transcribed spacer region 2; Mtmam: Mitochondrial Mammalia; Mtrev: General reversible Markov model for amino acid substitution of mitochondrial proteins; nad1-6, 4L: Nicotinamide adenine dinucleotide hydrogen dehydrogenase subunits 1-6 and 4L; MUSCLE: Multiple Sequence Comparison by Log-Expectation program; pp: Posterior probability; rDNA: Ribosomal DNA; SWAN: Sliding window analyses; WEHI: Walter and Eliza Hall Institute.

Acknowledgements

We wish to thank Stephen Wilcox for providing technical assistance with Illumina sequencing.

Authors' contributions

Conceptualisation: TS and AJ; methodology: TS, AJ, AVK, RBG and RH; software: RH; validation: TS, AJ, AVK, RBG and RH; formal analysis: TS, AVK and RH; investigation: TS; resources: AJ, RBG, TS and AVK; data curation: TS; writing—original draft preparation: TS; writing—review and editing: TS, AJ, IB, AVK, RBG and RH; visualisation: TS; supervision: AJ, IB, AVK, RBG and RH; project administration: AJ, RBG, IB, AVK; funding acquisition: AJ, TS, IB, RBG and AVK. All authors read and approved the final manuscript.

Funding

This study was financially supported by the Australian Biological Resources Study (Grant Numbers RF217-06 and CBG18-07). TS is a grateful recipient of the Australian Government Research Training Scholarship through the University of Melbourne. Further funding was provided by the University of Melbourne Research Initiatives Fund to AVK and an Australian Research Council Grant to AVK and RBG (LP160101299).

Availability of data and materials

All data generated or analysed during this study are included in this article. The sequence data generated in this study are available in the NCBI database under accession numbers OK111101–OK111108. Molecular vouchers specimens have been deposited in the South Australian Museum under the registration numbers AHC 36783, 49028, 49037, 49035, 49,052, 49,051, 49,055, 49,108.

Declarations

Ethics approval and consent to participate

Specimens were collected under the following state-issued permits: Victorian Department of Sustainability and Environment 90-053, 93-016, 10000434, 100003649; Queensland Department of Environment and Heritage Protection WA 00006125.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Veterinary Biosciences, Melbourne Veterinary School, The University of Melbourne, Victoria, Australia. ²South Australian Museum, Adelaide, South Australia, Australia.

Received: 5 August 2021 Accepted: 18 September 2021

Published online: 09 October 2021

References

- Beveridge I, Mawson PM. A taxonomic revision of the genera *Macropostrongyloides* Yamaguti and *Paramacropostrongylus* Johnston & Mawson (Nematoda: Trichonematidae) from Australian marsupials. *Aust J Zool.* 1978;26:763–87.
- Beveridge I. The systematic status of Australian Strongyloidea (Nematoda). *Bull Mus Natl Hist Nat Paris.* 1987;9:107–26.
- Chilton NB, Gasser RB, Beveridge I. Differences in a ribosomal DNA sequence of morphologically indistinguishable species within the *Hypodontus macropi* complex (Nematoda: Strongyloidea). *Int J Parasitol.* 1995;25:647–51.
- Chilton NB, Huby-Chilton F, Beveridge I, Smales LR, Gasser RB, Andrews RH. Phylogenetic relationships of species within the tribe Labiostrongylinae (Nematoda: Cloacinidae) from Australian marsupials based on ribosomal DNA spacer sequence data. *Parasitol Int.* 2011;60:381–7.
- Chilton NB, Jabbar A, Huby-Chilton F, Jex A, Gasser RB, Beveridge I. Genetic variation within the *Hypodontus macropi* (Nematoda: Strongyloidea) complex from macropodid marsupial hosts in Australia. *Electrophoresis.* 2012;33:3544–54.
- Gasser RB, Zhu X, Beveridge I, Chilton N. Mutation scanning analysis of sequence heterogeneity in the second internal transcribed spacer (rDNA) within some members of the *Hypodontus macropi* (Nematoda: Strongyloidea) complex. *Electrophoresis.* 2001;22:1076–85.
- Huby-Chilton F, Beveridge I, Gasser RB, Chilton NB. Single-strand conformation polymorphism analysis of genetic variation in *Labiostrongylus longispicularis* from kangaroos. *Electrophoresis.* 2001;22:1925–9.
- Sukee T, Beveridge I, Chilton NB, Jabbar A. Genetic variation within the genus *Macropostrongyloides* (Nematoda: Strongyloidea) from Australian macropodid and vombatid marsupials. *Parasitology.* 2019;146:1673–82.
- Sukee T, Beveridge I, Sabir AJ, Jabbar A. Phylogenetic relationships within the nematode subfamily Phascolostrongylinae (Nematoda: Strongyloidea) from Australian macropodid and vombatid marsupials. *Microorganisms.* 2021;9:1–13.
- Sukee T, Beveridge I, Jabbar A. *Torquenema* n. g., *Wallabicolina* n. g., and *Macropostrongyloides phascolomys* n. sp.: new genera and a new species of nematode (Strongyloidea: Phascolostrongylinae) parasitic in Australian macropodid and vombatid marsupials. *Animals.* 2021;11:175.
- Chilton NB, Gasser RB, Beveridge I. Phylogenetic relationships of Australian strongyloid nematodes inferred from ribosomal DNA sequence data. *Int J Parasitol.* 1997;27:1481–94.
- Jabbar A, Beveridge I, Mohandas N, Chilton NB, Littlewood DTJ, Jex AR, et al. Analyses of mitochondrial amino acid sequence datasets support the proposal that specimens of *Hypodontus macropi* from three species of macropodid hosts represent distinct species. *BMC Evol Biol.* 2013;13:259.
- Sukee T, Koehler AV, Hall R, Beveridge I, Gasser RB, Jabbar A. Phylogenetic analysis of mitogenomic data sets resolves the relationship of seven *Macropostrongyloides* species from Australian macropodid and vombatid marsupials. *Pathogens.* 2020;9:1–11.
- Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics.* 2014;30:2114–20.
- Jex AR, Hall RS, Littlewood DTJ, Gasser RB. An integrated pipeline for next-generation sequencing and annotation of mitochondrial genomes. *Nucleic Acids Res.* 2010;38:522–33.
- Lin RQ, Liu GH, Hu M, Song HQ, Wu XY, Li MW, et al. *Oesophagostomum dentatum* and *Oesophagostomum quadrispinulatum*: characterization of the complete mitochondrial genome sequences of the two pig nodule worms. *Exp Parasitol.* 2012;131:1–7.
- Zhao GH, Hu B, Cheng WY, Jia YQ, Li HM, Yu SK, et al. The complete mitochondrial genomes of *Oesophagostomum asperum* and *Oesophagostomum columbianum* in small ruminants. *Infect Genet Evol.* 2013;19:205–11.
- Liu GH, Zhao L, Song HQ, Zhao GH, Cai JZ, Zhao Q, et al. *Chabertia erschowi* (Nematoda) is a distinct species based on nuclear ribosomal DNA sequences and mitochondrial DNA sequences. *Parasit Vectors.* 2014;7:44.
- Gao Y, Wang XX, Ma XX, Zhang ZH, Lan Z, Qiu YY, et al. Characterization of the complete mitochondrial genomes of *Coronocylus labiatus* and *Cylicodontophorus bicoronatus*: Comparison with Strongylidae species and phylogenetic implication. *Vet Parasitol.* 2021;290:109359.
- Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. Basic local alignment search tool. *J Mol Biol.* 1990;215:403–10.
- Thompson JD, Higgins DG, Gibson TJ. CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* 1994;22:4673–80.
- Edgar RC. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 2004;32:1792–7.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. MEGA X: molecular evolutionary genetics analysis across computing platforms. *Mol Biol Evol.* 2018;35:1547.
- Librado P, Rozas J. DnaSP v5: a software for comprehensive analysis of DNA polymorphism data. *Bioinformatics.* 2009;25:1451–2.
- Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Höhna S, et al. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol.* 2012;61:539–42.
- Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B. PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. *Mol Biol Evol.* 2017;34:772–3.
- Rambaut A. FigTree v1. 4.4. <http://tree.bio.ed.ac.uk/software/figtree/> 2009. Accessed 14 July 2021.
- Mönnig HO. *Hypodontus macropi* n. gen., n. sp., a hookworm of the kangaroo. Union of South Africa 15th Report, Director Vet Services. 1929;1:303–6.
- Inglis WG. The geographical and evolutionary relationships of Australian trichostrongyloid parasites and their hosts. *Zool J Linn Soc.* 1968;47:327–47.
- Durette-Desset M-C, Denke MA, Murua R. Présence chez un Rongeur du Chili d'un Nématode Inglamidinae (sub fam nov) appartenant aux Amidostomatidae, famille connue des Mammifères d'Australie. *Ann Parasitol Hum Comp.* 1976;51:453–60.
- Skryabin KI, Schulz RS, Popova TI, Boev SN, Delyamure SL. Key to parasitic nematodes, volume 3—Strongylata. Leiden: Brill Publishers; 1992.
- Mawson PM. *Macropicola ocydromi* n.g., n.sp. (Nematoda: Strongylidae) from a Western Australian kangaroo. *Trans R Soc S Aust.* 1978;102:113–5.
- Lichtenfels JR. A conventional approach to a new classification of the Strongyloidea, nematode parasites of mammals. *Am Zool.* 1979;19:1185–94.
- Haynes BT, Marcus AD, Higgins DP, Gongora J, Gray R, Šlapeta J. Unexpected absence of genetic separation of a highly diverse population of hookworms from geographically isolated hosts. *Infect Genet Evol.* 2014;28:192–200.
- Sukee T, Beveridge I, Jabbar A. New species of *Macropostrongyloides* Yamaguti, 1961 (Nematoda: Strongylida) and the redescription of *Ma baylisi* (Wood, 1930) from Australian macropodid marsupials. *Syst Parasitol.* 2020;97:267–84.
- Beveridge I. A taxonomic revision of the genera *Phascolostrongylus* Canavan, and *Oesophagostomoides* Schwartz (Nematoda: Trichonematidae) from wombats. *Aust J Zool.* 1978;26:585–602.
- Lichtenfels JR. CIH keys to the nematode parasites of vertebrates. No. 7. Keys to the genera of the superfamily Strongyloidea. Farnham Royal: Commonwealth Agriculture Bureaux; 1980.
- Newton LA, Chilton NB, Beveridge I, Gasser RB. Systematic relationships of some members of the genera *Oesophagostomum* and *Chabertia* (Nematoda: Chabertiidae) based on ribosomal DNA sequence data. *Int J Parasitol.* 1998;28:1781–9.
- Popova TI. Principles of nematology, Strongyloidea of animals and man. Trichonematidae. In: Skryabin, editor. Essentials of nematology, vol. 7. Moscow: Izdatelstvo Akademii Nauk SSSR; 1958. p. 424.
- Yamaguti S. Systema Helminthum. Vol. 111. The nematodes of vertebrates. New York: Interscience Publishers; 1961.
- Spratt DM, Beveridge I. Helminth parasites of Australasian monotremes and marsupials. *Zootaxa.* 2016;4123:1–198.
- Chilton NB, Huby-Chilton F, Gasser RB, Beveridge I. The evolutionary origins of nematodes within the order Strongylida are related to predilection sites within hosts. *Mol Phylogenet Evol.* 2006;40:118–28.

43. Beveridge I. *Corollostrongylus hypsiprymnodontis* gen et sp. n. (Nematoda: Strongylidae) from the rat-kangaroo, *Hypsiprymnodon moschatus* (Marsupialia). *J Parasitol.* 1978;64:657–60.
44. Beveridge I. *Macropostrongyloides dendrolagi* n. sp. and *Mbaisoneima coronatum* n.g., n. sp., two new species of nematodes (Strongyloidea: Cloacinidae) from tree-kangaroos, *Dendrolagus* spp. (Marsupialia: Macropodidae) from Irian Jaya. *Indonesia Syst Parasitol.* 1997;38:25–31.
45. Smales LR. A new species of *Oesophagostomoides* (Nematoda: Cloacinidae) from the northern hairy-nosed wombat *Lasiorhinus krefftii* with a key to species of the genus. *J Parasitol.* 1994;80:638–43.
46. Beveridge I, Jabbar A, Koehler A, Sukee T. A morphological and molecular phylogenetic analysis of relationships between genera of the nematode sub-family Cloacininae (Stossich) (Strongyloidea: Chabertiidae) parasitic in kangaroos, wallabies and rat-kangaroos (Marsupialia: Macropodoidea). *Zootaxa.* 2020;4851:271–88.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

