

Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Imamura, F;Fretts, AM;Marklund, M;Ardisson Korat, AV;Yang, WS;Lankinen, M;Qureshi, W;Helmer, C;Chen, TA;Virtanen, JK;Wong, K;Bassett, JK;Murphy, R;Tintle, N;Yu, C;Brouwer, IA;Chien, KL;Chen, YY;Wood, AC;del Gobbo, LC;Djousse, L;Geleijnse, JM;Giles, GG;de Goede, J;Gudnason, V;Harris, WS;Hodge, A;Hu, F;Koulman, A;Laakso, M;Lind, L;Lin, HJ;McKnight, B;Rajaobelina, K;Riserus, U;Robinson, JG;Samieri, C;Senn, M;Siscovick, DS;Soedamah-Muthu, SS;Sotoodehnia, N;Sun, Q;Tsai, MY;Tuomainen, TP;Uusitupa, M;Wagenknecht, LE;Wareham, NJ;Wu, JHY;Micha, R;Lemaitre, RN;Mozaffarian, D;Forouhi, NG

Title:

Fatty acids in the de novo lipogenesis pathway and incidence of type 2 diabetes: A pooled analysis of prospective cohort studies

Date:

2020-06-01

Citation:

Imamura, F., Fretts, A. M., Marklund, M., Ardisson Korat, A. V., Yang, W. S., Lankinen, M., Qureshi, W., Helmer, C., Chen, T. A., Virtanen, J. K., Wong, K., Bassett, J. K., Murphy, R., Tintle, N., Yu, C. I., Brouwer, I. A., Chien, K. L., Chen, Y. Y., Wood, A. C., ... Forouhi, N. G. (2020). Fatty acids in the de novo lipogenesis pathway and incidence of type 2 diabetes: A pooled analysis of prospective cohort studies. *Plos Medicine*, 17 (6), <https://doi.org/10.1371/journal.pmed.1003102>.

Persistent Link:

<https://hdl.handle.net/11343/304456>

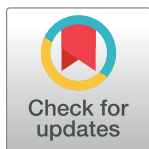
License:

[CC BY](#)

RESEARCH ARTICLE

Fatty acids in the de novo lipogenesis pathway and incidence of type 2 diabetes: A pooled analysis of prospective cohort studies

Fumiaki Imamura^{1*}, Amanda M. Fretts², Matti Marklund^{3,4,5}, Andres V. Ardisson Korat^{6,7}, Wei-Sin Yang⁸, Maria Lankinen⁹, Waqas Qureshi¹⁰, Catherine Helmer¹¹, Tzu-An Chen¹², Jyrki K. Virtanen⁹, Kerry Wong¹³, Julie K. Bassett¹³, Rachel Murphy¹⁴, Nathan Tintle¹⁵, Chaoyu Ian Yu¹⁶, Ingeborg A. Brouwer¹⁷, Kuo-Liong Chien⁸, Yun-yu Chen^{8,18}, Alexis C. Wood¹², Liana C. del Gobbo¹⁹, Luc Djousse²⁰, Johanna M. Geleijnse²¹, Graham G. Giles^{13,22,23}, Janette de Goede²¹, Vilmondur Gudnason²⁴, William S. Harris^{25,26}, Allison Hodge^{13,22}, Frank Hu^{6,7}, InterAct Consortium¹, Albert Koulman^{1,27,28,29}, Markku Laakso^{30,31}, Lars Lind³², Hung-Ju Lin³³, Barbara McKnight¹⁶, Kalina Rajaobelina¹¹, Ulf Riserus³, Jennifer G. Robinson³⁴, Cecilia Samieri¹¹, Mackenzie Senn¹², David S. Siscovick³⁵, Sabita S. Soedamah-Muthu^{21,36,37}, Nona Sotoodehnia³⁸, Qi Sun^{6,7}, Michael Y. Tsai³⁹, Tomi-Pekka Tuomainen⁹, Matti Uusitupa⁹, Lynne E. Wagenknecht⁴⁰, Nick J. Wareham¹, Jason H. Y. Wu⁴, Renata Micha⁵, Rozenn N. Lemaitre³⁸, Dariush Mozaffarian⁵, Nita G. Forouhi¹



OPEN ACCESS

Citation: Imamura F, Fretts AM, Marklund M, Ardisson Korat AV, Yang W-S, Lankinen M, et al. (2020) Fatty acids in the de novo lipogenesis pathway and incidence of type 2 diabetes: A pooled analysis of prospective cohort studies. *PLoS Med* 17(6): e1003102. <https://doi.org/10.1371/journal.pmed.1003102>

Academic Editor: Xu Lin, Hangzhou Institute for Advanced Study, CHINA

Received: October 10, 2019

Accepted: April 28, 2020

Published: June 12, 2020

Copyright: © 2020 Imamura et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: The institutional IRB approvals and data sharing agreements for the participating cohorts allowed us to share cohort results. Individual participant data are owned by individual participating cohorts and are available to researchers consented from participating cohorts. For further queries or requests, please contact force@tufts.edu. Further details are available at the FORCE website: <http://force.nutrition.tufts.edu/>.

1 MRC Epidemiology Unit, University of Cambridge, Cambridge, United Kingdom, **2** Cardiovascular Health Research Unit, Department of Epidemiology, University of Washington, Seattle, Washington, United States of America, **3** Department of Public Health and Caring Sciences, Clinical Nutrition and Metabolism, Uppsala University, Uppsala, Sweden, **4** The George Institute for Global Health, the Faculty of Medicine, University of New South Wales, Sydney, Australia, **5** Friedman School of Nutrition Science and Policy, Tufts University, Boston, Massachusetts, United States of America, **6** Department of Nutrition and Epidemiology, Harvard T.H. Chan School of Public Health, Boston, Massachusetts, United States of America, **7** Channing Division of Network Medicine, Department of Medicine, Brigham and Women’s Hospital and Harvard Medical School, Boston, Massachusetts, United States of America, **8** Institute of Epidemiology and Preventive Medicine, College of Public Health, National Taiwan University, Taipei City, the Republic of China, **9** Institute of Public Health and Clinical Nutrition, University of Eastern Finland, Kuopio, Finland, **10** Section of Cardiovascular Medicine, Department of Internal Medicine, Wake Forest University School of Medicine, Winston-Salem, North Carolina, United States of America, **11** INSERM, UMR 1219, Bordeaux Population Health Research Center, University of Bordeaux, Bordeaux, France, **12** USDA/ARS Children’s Nutrition Research Center, Department of Pediatrics, Baylor College of Medicine, Houston, Texas, United States of America, **13** Cancer Epidemiology Division, Cancer Council Victoria, Melbourne, Australia, **14** Centre of Excellence in Cancer Prevention, School of Population & Public Health, Faculty of Medicine, The University of British Columbia, Vancouver, Canada, **15** Department of Mathematics and Statistics, Dordt University, Sioux Center, Iowa, United States of America, **16** Department of Biostatistics, University of Washington School of Public Health, Seattle, Washington, United States of America, **17** Department of Health Sciences, Faculty of Science, Vrije Universiteit Amsterdam, Amsterdam Public Health Research Institute, Amsterdam, the Netherlands, **18** Division of Cardiology, Department of Medicine, Taipei Veterans General Hospital, Taipei City, the Republic of China, **19** Department of Medicine, Division of Cardiovascular Medicine, Stanford University School of Medicine, Stanford, California, United States of America, **20** Divisions of Aging, Department of Medicine, Brigham and Women’s Hospital and Harvard Medical School, Boston, Massachusetts, United States of America, **21** Division of Human Nutrition and Health, Wageningen University, Wageningen, the Netherlands, **22** Centre for Epidemiology and Biostatistics, The University of Melbourne, Parkville, Australia, **23** Precision Medicine, School of Clinical Sciences at Monash Health, Monash University, Clayton, Australia, **24** Icelandic Heart Association Research Institute, Kopavogur, Iceland, **25** Department of Internal Medicine, Sanford School of Medicine, University of South Dakota, Sioux Falls, South Dakota, United States of America, **26** OmegaQuant Analytics, Sioux Falls, South Dakota, United States of America, **27** National Institute for Health Research Biomedical Research Centres Core Nutritional Biomarker Laboratory, University of Cambridge, Addenbrooke’s Hospital, Cambridge, United Kingdom, **28** National Institute for Health Research Biomedical Research Centres Core Metabolomics and Lipidomics Laboratory, University of Cambridge, Addenbrooke’s Hospital, Cambridge, United Kingdom, **29** Medical Research Council Elsie Widdowson Laboratory, Cambridge, United Kingdom, **30** Institute of Clinical Medicine, Internal Medicine, University of Eastern Finland, Kuopio, Finland, **31** Department of Medicine, Kuopio University Hospital, Kuopio, Finland,

Funding: No specific funding was received for this study. This manuscript does not reflect opinions or conclusions of any funding agency. No funding bodies had any role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. FI, NGF, and NJW were funded by the United Kingdom Medical Research Council Epidemiology Unit core grant (MC_UU_12015/5 and MC_UU_12015/1); NJW, NGF and AK acknowledge National Institute for Health Research (NIHR) Biomedical Research Centre Cambridge (IS-BRC-1215-20014); NJW is an NIHR Senior Investigator; IAB, by the European Union (FP7 and Horizon2020) and the Dutch Scientific Organization (ZonMW); CS, by the Foundation Plan Alzheimer; and AAK, by the NIH training grant (3T32DK007703 and T32CA009001). Alpha Omega Cohort was funded by the Netherlands Heart Foundation (grant 2000T401), the NIH (NIH/ National Heart, Lung, and Blood Institute [NHLBI], and ODS, grant R01HL-076200), and Unilever R&D, Vlaardingen (margarine production and distribution). The authors acknowledge Kamalita Pertiwi, Wageningen University, the Netherlands, for analyses of data from the Alpha Omega Cohort. Age, Gene/Environment Susceptibility Study Reykjavik was funded by Office of Dietary Supplements, the United States National Institute of Health (NIH) contract N01-AG012100, the National Institute of Aging (NIA) Intramural Research Program, Hjartavernd (the Icelandic Heart Association), and the Althingi (the Icelandic Parliament). Additional support was provided by the Michael Smith Foundation for Medical Research (#17644) and the Canadian Cancer Society (grant #704735). Chin-Shan Community Cardiovascular Study was funded by Ministry of Science and Technology and National Taiwan University, Taiwan (MOST 103-2314-B-002-135-MY3, NSC 100-2314-B-002-113-MY3, NTUH 105-S3120, NTUH 106-S3453). The Cardiovascular Health Study (CHS) was supported by contracts HHSN268201200036C, HHSN268200800007C, HHSN268201800001C, N01HC55222, N01HC85079, N01HC85080, N01HC85081, N01HC85082, N01HC85083, N01HC85086, and grants R01-HL-085710, U01HL080295 and U01HL130114 from the NHLBI, with additional contribution from the National Institute of Neurological Disorders and Stroke (NINDS). Additional support was provided by R01AG023629 from NIA. The Framingham Heart Study was supported by the NHLBI in collaboration with Boston University (Contract No. N01-HC-25195). The Health Professionals Follow-up Study was funded by UM1 CA167552, R01 HL35464, AA11181, HL35464, CA55075, HL60712, and P30

32 Department of Medical Sciences, Uppsala University, Uppsala, Sweden, 33 Department of Internal Medicine, National Taiwan University Hospital, Taipei City, the Republic of China, 34 Preventive Intervention Center, Departments of Epidemiology, the University of Iowa College of Public Health, Iowa City, Iowa, United States of America, 35 The New York Academy of Medicine, New York, New York, United States of America, 36 Center of Research on Psychological and Somatic disorders, Department of Medical and Clinical Psychology, Tilburg University, Tilburg, the Netherlands, 37 Institute for Food, Nutrition and Health, University of Reading, Reading, United Kingdom, 38 Cardiovascular Health Research Unit, Department of Medicine, University of Washington, Seattle, Washington, United States of America, 39 Department of Laboratory Medicine and Pathology, University of Minnesota, Minneapolis, Minnesota, United States of America, 40 Public Health Sciences, Wake Forest School of Medicine, Winston-Salem, North Carolina, United States of America

* fumiaki.imamura@mrc-epid.cam.ac.uk

Abstract

Background

De novo lipogenesis (DNL) is the primary metabolic pathway synthesizing fatty acids from carbohydrates, protein, or alcohol. Our aim was to examine associations of in vivo levels of selected fatty acids (16:0, 16:1n7, 18:0, 18:1n9) in DNL with incidence of type 2 diabetes (T2D).

Methods and findings

Seventeen cohorts from 12 countries (7 from Europe, 7 from the United States, 1 from Australia, 1 from Taiwan; baseline years = 1970–1973 to 2006–2010) conducted harmonized individual-level analyses of associations of DNL-related fatty acids with incident T2D. In total, we evaluated 65,225 participants (mean ages = 52.3–75.5 years; % women = 20.4%–62.3% in 12 cohorts recruiting both sexes) and 15,383 incident cases of T2D over the 9-year follow-up on average. Cohort-specific association of each of 16:0, 16:1n7, 18:0, and 18:1n9 with incident T2D was estimated, adjusted for demographic factors, socioeconomic characteristics, alcohol, smoking, physical activity, dyslipidemia, hypertension, menopausal status, and adiposity. Cohort-specific associations were meta-analyzed with an inverse-variance-weighted approach. Each of the 4 fatty acids positively related to incident T2D. Relative risks (RRs) per cohort-specific range between midpoints of the top and bottom quintiles of fatty acid concentrations were 1.53 (1.41–1.66; $p < 0.001$) for 16:0, 1.40 (1.33–1.48; $p < 0.001$) for 16:1n7, 1.14 (1.05–1.22; $p = 0.001$) for 18:0, and 1.16 (1.07–1.25; $p < 0.001$) for 18:1n9. Heterogeneity was seen across cohorts ($I^2 = 51.1\%–73.1\%$ for each fatty acid) but not explained by lipid fractions and global geographical regions. Further adjusted for triglycerides (and 16:0 when appropriate) to evaluate associations independent of overall DNL, the associations remained significant for 16:0, 16:1n7, and 18:0 but were attenuated for 18:1n9 (RR = 1.03, 95% confidence interval (CI) = 0.94–1.13). These findings had limitations in potential reverse causation and residual confounding by imprecisely measured or unmeasured factors.

Conclusions

Concentrations of fatty acids in the DNL were positively associated with T2D incidence. Our findings support further work to investigate a possible role of DNL and individual fatty acids in the development of T2D.

DK46200 from the NIH. The Insulin Resistance Atherosclerosis Study was funded by grants U01-HL-47892, U01-HL-47902, DK-29867, R01-58329, and DK-079888 from NHLBI and grant M01-RR-43 from the NIH. The InterAct project was funded by the EU FP6 programme (grant no. LSHM_CT_2006_037197). The Kuopio Ischaemic Heart Disease Risk Factor Study was supported mainly by the funding from the Academy of Finland to Jukka T. Salonen. The Melbourne Collaborative Cohort Study was funded by VicHealth and Cancer Council Victoria; and by grants 209057 and 126403 from Australia's National Health and Medical Research Council and by infrastructure provided by Cancer Council Victoria. Multi-ethnic Study of Atherosclerosis was funded by contracts HHSN268201500003I, N01-HC-95159, N01-HC-95160, N01-HC-95161, N01-HC-95162, N01-HC-95163, N01-HC-95164, N01-HC-95165, N01-HC-95166, N01-HC-95167, N01-HC-95168 and N01-HC-95169 from the NHLBI and by grants UL1-TR-000040 and UL1-TR-001079 from NCRR. The Metabolic Syndrome in Men Study was funded by the grants from the European Union, the Academy of Finland, and the Juselius Foundation. The Nurses' Health Study was funded by the NIH (CA186107, CA87969, CA49449, HL34594, HL35464, CA167552, HL60712, and HL088521). Prospective Investigation of the Vasculature in Uppsala Seniors was supported by Uppsala University Hospital and the Swedish Research Council for Health, Working Life and Welfare. The Tree City Study was conducted under a partnership agreement between the Institut National de la Sante et de la Recherche Medicale, the University Bordeaux 2 Victor Segalen, and Sanofi; and funded by the Fondation pour la Recherche Medicale, the Caisse Nationale Maladie des Travailleurs Salaries, Direction Generale de la Sante, MGEN, Institut de la Longevite, Conseils Regionaux d'Aquitaine et Bourgogne, Fondation de France, Ministry of Research—Institut National de la Sante and de la Recherche Medicale Programme Cohortes, grant COGINUT ANR-06-PNRA-005 from the Agence Nationale de la Recherche, grant FCS 2009-2012 from the Fondation Plan Alzheimer, and the Caisse Nationale pour la Solidarite et l'Autonomie. The Uppsala Longitudinal Studies of Adult Men 50 and 70 were funded by the Swedish Research Council for Health, Working Life and Welfare, Uppsala City Council, Swedish Research Council, and Swedish Diabetes Foundation (UR). The Women's Health Initiative was funded by the NHLBI, NIH, U.S. Department of Health and Human Services through contracts HHSN268201600018C, HHSN268201600001C, HHSN268201600002C, HHSN268201600003C, and

Author summary

Why was this study done?

- De novo lipogenesis (DNL) is a metabolic pathway involved in the endogenous synthesis of specific fatty acids, such as 16:0, 16:1n7, 18:0, and 18:1n9, and it is linked to the pathophysiology of cardiometabolic diseases, including type 2 diabetes (T2D).
- Circulating or tissue concentrations of these fatty acids have been investigated for the associations with T2D incidence in epidemiological research. However, published studies reported inconsistent associations inconsistently and were subject to publication bias.
- Summary evidence is not available to date for the associations between these fatty acids and T2D incidence. An integration of available cohort studies would increase statistical power and allow assessment of generalizability, standardization of analytical strategies, and evidence synthesis with the potential publication bias minimized.

What did the researchers do and find?

- As a part of the Fatty Acids and Outcomes Research Consortium (FORCE), we conducted new individual-participant data analyses of 17 cohort studies of a total of 65,225 adults free of T2D at baseline, among whom 15,383 developed incident T2D over up to 20 years of follow-up.
- The cohort studies analyzed the associations between fatty acids (16:0, 16:1n7, 18:0, and 18:1n9) and the risk of developing T2D with standardized analytic strategy.
- In pooled analyses, each of the fatty acids was positively associated with a higher risk of developing T2D. The associations were independent of major risk factors for T2D, such as age, sex, race/ethnicity, socioeconomic characteristics, smoking status, physical activity, and obesity.

What do these findings mean?

- The findings provide the first summary evidence to date for the positive relationships of concentrations of the DNL-related fatty acids with a risk of T2D, indicating the strong relevance of DNL and its determinants to the development of T2D.
- These fatty acids potentially reflect the status of DNL activity, which may be stimulated or suppressed by a combination of carbohydrate intake, alcohol intake, polyunsaturated fatty acid intake, and other lifestyle and clinical factors. Therefore, the current findings indicate the need for investigation into determinants and consequences of elevated concentrations of these fatty acids.
- Despite several advantages of our individual-level data analysis in this pooling project, the results cannot establish whether elevated concentrations of these fatty acids caused the development of T2D or whether underlying peripheral or hepatic insulin resistance, for example, may elevate both the fatty acid concentrations and the risk of T2D independently.

HHSN268201600004C. A full listing of each of CHS, MESA, and WHI investigators can be found at <http://www.CHS-NHLBI.org>, <http://www.mesa-nhlbi.org>, and <http://www.whi.org/researchers/>, respectively. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: I have read the journal's policy and the authors of this manuscript have the following competing interests: JYW and RM report research support from Unilever for other projects on fatty acid biomarkers. LCDG reported receiving ad hoc consulting fees from the Life Sciences Research Organization. CH reported receiving fees for a conference from Novartis. IAB reported involvement in a research project partly funded by Unilever. JGR received research grants from Amarin and Astra-Zeneca. DM reported receiving ad hoc honoraria from Bunge, Pollock Institute, and Quaker Oats; ad hoc consulting for Foodminds, Life Sciences Research Organization, Nutrition Impact, Amarin, AstraZeneca, Winston, and Strawn LLP; membership in Unilever North America Scientific Advisory Board; and chapter royalties from UpToDate. SSSM reported receiving an international award and unrestricted grants for meta-analysis work on dairy foods and cardiometabolic diseases from Global and Dutch Dairy Associations. Other authors do not have any conflict of interest to declare.

Abbreviations: AOC, Alpha Omega Cohort; BMI, body mass index; CCCC, Chin-Shan Community Cardiovascular Cohort Study; CHS, Cardiovascular Health Study; CI, confidence interval; DNL, de novo lipogenesis; FORCE, Fatty Acids and Outcomes Research Consortium; MCCC, Melbourne Collaborative Cohort Study; MESA, Multi-Ethnic Study of Atherosclerosis; PRISMA, Preferred Reporting Items for Systematic Reviews and Meta-Analyses; RR, relative risk; SCD, stearoyl-coenzyme A desaturase 1 gene; T2D, type 2 diabetes; WHIMS, Women's Health Initiative Memory Study.

Introduction

De novo lipogenesis (DNL) is a metabolic pathway for the endogenous synthesis of triglycerides and other lipids from dietary starch, sugar, and protein [1,2]. Palmitic acid (16:0) is the major fatty acid product of DNL and can be elongated to stearic acid (18:0) and desaturated to form palmitoleic acid (16:1n7) and from stearic acid to oleic acid (18:1n9). Tissue levels of these fatty acids have been previously reported to show associations with insulin resistance and to be higher among adults with type 2 diabetes (T2D) than healthy adults [3].

Experimental studies have supported causal detrimental effects of 16:0 on inflammatory responses and pancreatic function, whereas protective effects of 16:1n7 and 18:1n9 on pancreatic function have been suggested [4–7]. In addition, greater DNL activity has been reported to be driven by lifestyle habits such as excessive consumption of carbohydrates or alcohol and lower physical activity [8–10], although the relative contributions of different lifestyle habits influencing DNL remain undefined. Investigation into how fatty acids in the DNL pathway relate to incident T2D may provide important etiological knowledge and stimulate future work on modifiable risk factors and preventive treatments.

Individual studies have examined the associations between circulating DNL-related fatty acids and incident T2D, showing mixed associations [11–18]. For instance, higher concentrations of 16:0 were associated with a higher incidence of T2D in several studies but not in others [14,18]. Similarly inconsistent findings were observed for 16:1n7, 18:0, and 18:1n9. To our knowledge, no prior studies have comprehensively brought together available evidence relating these fatty acids to incidence of T2D or investigated potential factors underlying the heterogeneous findings. Varied findings to date could reflect unstable results from some relatively small-scale studies (e.g., N cases < 200 in many cohorts), different lipid fractions evaluated across studies, and differences in demographics and analytic approaches. Also, there is little evidence whether the fatty acids in the DNL pathway may have a pathophysiological role independent of the overall DNL activity or triglycerides, one of the end products of the DNL. Therefore, to better characterize the prospective associations of fatty acids in the DNL pathway with incidence of T2D, we conducted de novo pooled individual-level analysis using harmonized methods across 17 studies in the global Fatty Acids and Outcomes Research Consortium (FORCE).

Methods

Cohorts and study variables

FORCE was initially formed from the Cohorts for Heart and Aging Research in Genomic Epidemiology consortium. FORCE is an ongoing consortium project to study relationships of fatty acid biomarkers with health outcomes (<http://force.nutrition.tufts.edu/>) [19–21]. The current project included 17 prospective studies (cohorts and nested case-control or case-cohort studies). These studies agreed to participate after confirming the inclusion criteria met: recruitment of adults aged 18 years or over and without prevalent diabetes at the time of fatty acid assessment; available data of circulating or adipose 16:0, 16:1n7, 18:0, and 18:1n9; and ascertainment of incident T2D (S1 Text). Other cohorts participated in FORCE for other projects [19–21] but did not contribute to this study because incident T2D were not ascertained. All cohorts had obtained approval from each institutional review board and written informed consent from participants. This study is reported as per the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guideline (S1 Checklist).

A standardized analysis protocol was developed, approved by the FORCE investigators, and provided to each participating cohort (S2 Text). The protocol prespecified the inclusion

criteria mentioned above, as well as the exposures (DNL-related fatty acids), standardized covariates, effect modifiers, incident T2D, and statistical methods. Following this protocol we developed centrally, each cohort performed new individual-participant data analysis. Cohort-specific results were recorded in a standardized electronic form and centrally compiled and meta-analyzed. The data underlying the results presented in the study are available for researchers who meet criteria of each participating cohort.

[S1 Text](#) includes information on participating cohorts, study participants, and methods for fatty acid measurement and ascertainment of incident T2D. Briefly, each cohort isolated fatty acid molecules from one or more lipid compartments including erythrocyte phospholipids, plasma phospholipids, plasma cholesteryl esters, plasma triglycerides, total plasma or serum, or adipose tissue. Then, *in vivo* fatty acid concentrations were measured with gas chromatography. Concentrations of each fatty acid were quantified as a percent of total fatty acids in the lipid fraction.

Incident T2D was ascertained on the basis of one or more criteria including fasting glucose ≥ 7.0 mmol/L; glucose 11.1 mmol/L from 2-hour oral glucose tolerance test; new use of oral antidiabetic medication; or concentration of HbA1c $\geq 6.5\%$ ([S1 Text](#)). The Melbourne Collaborative Cohort Study (MCCS) [14] and Alpha Omega Cohort (AOC) [22] ascertained incident T2D based on self-reported physician diagnosis, use of antidiabetic medication, or both. The EPIC-InterAct Study ascertained incident T2D by adjudicating self-reported T2D diagnosis or verifying diagnosis in disease registries [17].

Statistical analysis in individual studies

Individual-participant data analyses were prespecified and documented in the protocol, with the primary exposure variables being 16:0, 16:1n7, 18:0, and 18:1n9. We examined Pearson correlation coefficients between these fatty acids within each lipid fraction. To assess associations of interest, Cox proportional hazard regression was modeled to time-to-event data, with sampling weights applied in EPIC-InterAct with a case-cohort design [17]. Each cohort calculated follow-up time from time of fatty acid measurement to either date of incident T2D, death from any cause, or loss to follow-up, or censoring at end of follow-up, whichever available and occurred first. In the 2 cohorts (AOC and MCCS) without individuals' person-time data [14,22], logistic regression was used as the most efficient approach to obtaining estimates of interest from the 2 cohorts. The fatty acid variables were evaluated as a continuous linear variable in a unit of the study-specific interquintile range (the difference between the midpoints of the top and bottom quintiles) and, in a separate model, as categorical indicator variables (quintile categories, with the lowest quintile as the reference). We used an interquintile range and quintile categories in continuous and categorical approaches, respectively, because two approaches allowed estimation of the associations over the same exposure range and improvement of comparability between the two approaches.

Covariates for statistical adjustment were prespecified, including their categorization (e.g., continuous, quintiles, etc.). Each participating study prespecified the use of some study-specific covariates (e.g., the number of categories for education status), depending on availability. The primary model included field site, age, sex, race/ethnicity, occupation, education, smoking status, physical activity, alcohol consumption, prevalent hypertension (self-reported or treated), prevalent dyslipidemia (self-reported or treated), prevalent heart disease, and self-reported health status. The second model further adjusted for adiposity measures (body mass index [BMI] and waist circumference). For the mechanistic investigation, the third model further adjusted for circulating 16:0 (for analysis of 16:1n7, 18:0, and 18:1n9) and triglycerides to assess whether associations of 16:1n7, 18:0, and 18:1n9 with incident T2D would be independent of 16:0 and triglycerides and for analysis of 16:0, of triglycerides.

We assessed study-specific measures of interaction by age, sex, BMI, and race/ethnicity using the second model that adjusted for potential confounders including the adiposity measures. Each fatty acid, these prespecified potential effect modifiers, and their relevant cross-product terms and variance-covariance measures were analyzed to evaluate the potential interaction within each cohort.

Pooled analyses

Study-specific regression coefficients, either log hazard ratios or log odds ratios, and standard errors were meta-analyzed with an inverse-variance weighted method to estimate summary relative risks (RRs) and confidence intervals (CIs). Heterogeneity in results between studies was quantified as I^2 [23]. A few cohorts included fatty acid measures in more than one lipid fraction. To avoid double-counting estimates from such cohorts, we prespecified primary use of estimates of phospholipid (plasma or erythrocyte) fatty acids. These lipid fractions were most commonly used among participating cohorts and generally reflect longer-term exposure than the other compartments except for adipose tissue [24]. In secondary analyses, estimates in each different lipid fraction were also meta-analyzed separately, using each available cohort with measurements in that lipid fraction.

To test interactions by age, sex, BMI, and race/ethnicity, cohort-specific coefficients of cross-product terms were meta-analyzed. Because we considered the tests for interactions as exploratory, we applied correction for multiple testing as $\alpha_{\text{two-tailed}} = 0.0031$ (0.05/4 fatty acids/4 potential effect modifiers). If an interaction was statistically significant, stratum-specific associations were estimated by using regression coefficients and variance-covariance matrices and then pooled using meta-analysis. We fitted meta-regression models and stratified meta-analyses to investigate potential sources of heterogeneity due to study-specific characteristics. Factors examined included lipid fraction, geographical region (Europe/Australia, United States, Asia), and prevalence of dyslipidemia. To further explore sources of heterogeneity, we evaluated the following factors post hoc: prevalence of hypertension, mean triglyceride concentrations, fasting status, availability of time-to-event data, and mean years of follow-up. As a sensitivity analysis, we conducted random-effects meta-analysis and meta-analysis after converting odds ratios to risk ratios in AOC and MCCS [25]. Meta-analyses were performed using Stata 14.2 (StataCorp, College Station, Texas) with $\alpha_{\text{two-tailed}} = 0.05$, unless specified otherwise.

Results

Population characteristics

Among 17 participating cohorts, mean age ranged from 52.3 to 76.0 years (Table 1). Three cohorts recruited men only, two cohorts recruited women only, and the others recruited both (%women = 20.4%–62.3%). Study-specific mean BMIs ranged from 25.2 to 28.4 kg/m², except for the Chin-Shan Community Cardiovascular Cohort Study (CCCC) in Taiwan (mean BMI = 23.2 kg/m²). Most studies recruited participants of European descent predominantly. Participants of non-European descent were recruited in the Multi-Ethnic Study of Atherosclerosis (MESA; 71.6% nonwhite), the Women's Health Initiative Memory Study (WHIMS; 11.6% nonwhite), the Cardiovascular Health Study (CHS; 11.0% nonwhite), and the CCCC (100% East Asian).

The concentrations of the selected fatty acids in the DNL pathway varied by lipid compartment (Fig 1). Concentrations of 16:0 ranged from 15% to 35% of total fatty acids in most lipid compartments except for cholesteryl esters (10% to 13%). Concentrations of 16:1n7 were less than 1.0% when measured in phospholipids (plasma or red blood cell membrane) and 1.0% to 9.0% when measured in the other compartments. In phospholipids, average concentrations of

Table 1. Baseline characteristics of 17 studies of the pooling analysis of fatty acids on DNL pathway and incident T2D: FORCE^a.

Study	Country	Study design	Baseline year(s)	Follow-up years, median	N adults (N cases)	Age, mean y	Sex, % women	BMI, mean kg/m ²	Triglycerides, mmol/L	Biomarker fraction
CHS	United States	Cohort	1992	10.6	3,179 (284)	75.1	61.5	26.4	1.57	PL
MESA	United States	Cohort	2000–2002	9.3	2,252 (309)	61.0	53.9	27.6	1.49	PL
IRAS	United States	Cohort	1992–1997	5.3	719 (146)	55.1	55.8	28.4	1.53	Total plasma
FHS	United States	Cohort	2005–2008	5.8	2,209 (98)	64.4	57.2	27.8	1.26	RBC PL
WHIMS	United States	Cohort	1996	11.0	6,510 (502)	70.1	100	28.1	1.56	RBC PL
NHS	United States	Cohort	1990	16.9	1,760 (177)	60.4	100	25.3	N/A	RBC PL, total plasma
HPFS	United States	Cohort	1994	11.1	1,519 (112)	64.1	0	25.8	N/A	RBC PL, total plasma
EPIC-InterAct ^b	8 European countries	Case cohort	1993–1997	12.3	27,296 (12,132)	52.3	62.3	26.0	1.35	PL
AGESR	Iceland	Cohort	2002–2006	5.2	753 (28)	75.5	59.5	27.0	1.14	PL
Three C	France	Cohort	1999–2000	8.0	565 (39)	76.0	64.3	25.0	1.28	RBC PL
AOC	Netherlands	Cohort	2002–2006	2.5	1,741 (201)	68.9	20.4	27.4	1.83	RBC PL, CE
ULSAM	Sweden	Cohort	1970–1973	21.4	2009 (396)	54.4	0	25.2	1.77	Adipose tissue
PIVUS	Sweden	Cohort	2001–2004	10.0	879 (67)	72.5	51.0	26.7	1.24	PL, CE
KIHD	Finland	Cohort	1998–2001	10.3	1,543 (205)	62.7	52.7	27.6	1.23	Total serum
METSIM	Finland	Cohort	2006–2010	5.5	1,302 (71)	57.3	0	26.4	1.35	PL, CE, TG
MCCS	Australia	Case cohort	1990–1994	4.0	6,151 (490)	56.3	53.9	27.0	1.27	PL
CCCC	Taiwan	Cohort	1992–1993	6.0	1,838 (128)	58.7	40.0	23.2	1.29	Total plasma

^aBaseline characteristics at the time of fatty acid biomarker measurement

^bThe EPIC-InterAct Study provided pooled estimates from across 8 European countries: Denmark, France, Germany, Italy, the Netherlands Spain, Sweden, and the United Kingdom.

Abbreviations: AGESR: Age, Genes, Environment Susceptibility Study (Reykjavik); AOC, Alpha Omega Cohort; CCCC, Chin-Shan Community Cardiovascular Cohort Study; CE, cholesteryl esters; CHS, Cardiovascular Health Study; FHS, Framingham Heart Study; FORCE, Fatty Acids and Outcomes Research Consortium; HPFS, Health Professionals' Follow-up Study; KIHD, Kuopio Ischaemic Heart Disease; MCCS, Melbourne Collaborative Cohort Study; MESA, Multi-Ethnic Study of Atherosclerosis; METSIM, Metabolic Syndrome in Men Study; NHS, Nurses' Health Study; PIVUS, Prospective Investigation of the Vasculature in Uppsala Seniors; PL, phospholipids; RBC, red blood cells; TG, triglycerides; Three C, Three City Study; ULSAM, Uppsala Longitudinal Study of Adult Men; WHIMS, Women's Health Initiative Memory Study

<https://doi.org/10.1371/journal.pmed.1003102.t001>

18:0 were 11.0% to 16.3% and consistently higher than those of 18:1n9. In other lipid compartments, conversely, concentrations of 18:0 were much lower than those of 18:1n9.

Correlations between fatty acids also varied by lipid compartment (S1 Table). For example, in phospholipids, 16:0 positively correlated with 16:1n7 (weighted-average $r = 0.47$) and 18:1n9 ($r = 0.23$) but negatively with 18:0 ($r = -0.63$). By contrast, in cholesteryl ester, triglycerides, and adipose tissue, the correlation between 16:0 and 18:0 was positive ($r = 0.39, 0.39,$ and 0.53 , respectively).

Prospective associations with incident T2D

In pooled analyses for each of the 4 fatty acids evaluating a total of 65,225 participants and 15,383 incident T2D cases, significant positive associations were identified, whether before (S1 Fig) or after adjustment (Fig 2) for adiposity measures. For example, RRs (95% CI) per the cohort-specific midpoints of the top and bottom quintiles for 16:0 were 1.63 (1.50–1.76) and 1.53 (1.41–1.66) with and without adjustment for adiposity, respectively ($p < 0.001$ for each). For 16:1n7, 18:0, and 18:1n9, similar or weaker significantly positive associations were observed.

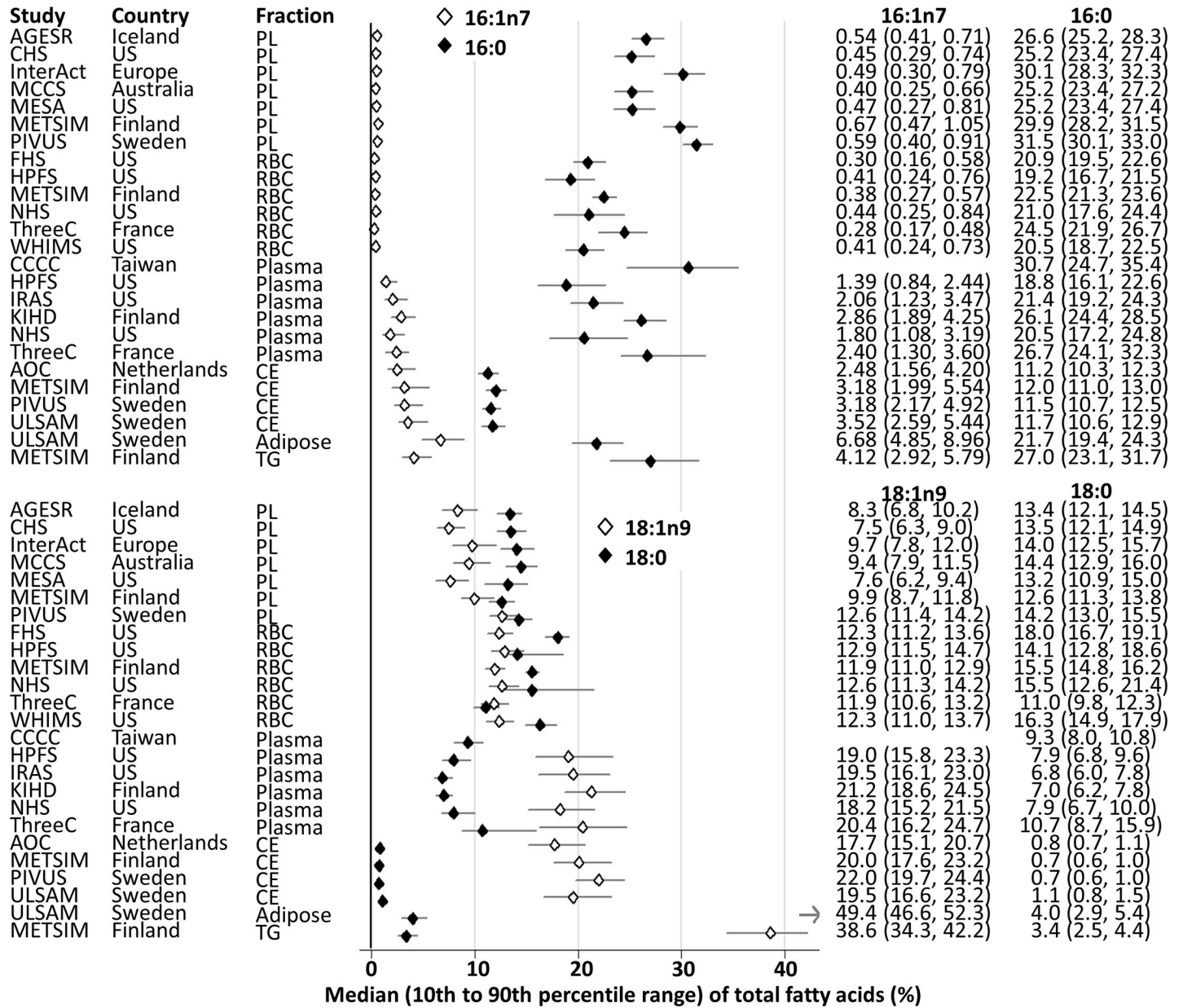


Fig 1. Proportions of fatty acids in the DNL pathway. Plots represent median (diamond) and the range of 10th to 90th percentiles (horizontal bar). See Table 1 for cohort names. CE, cholesteryl ester; DNL, de novo lipogenesis; PL, phospholipid; RBC, red blood cell; US, United States.

<https://doi.org/10.1371/journal.pmed.1003102.g001>

Further adjusting for triglycerides, the association of 16:0 was modestly attenuated, with RR (95% CI) of 1.36 (1.24–1.50; $p < 0.001$; S2 Fig). After adjustment for triglycerides and 16:0, associations of 16:1n7 and 18:0 with T2D incidence were attenuated but still evident with RRs (95% CI) of 1.17 (1.11–1.24; $p < 0.001$) and 1.16 (1.06–1.27; $p = 0.001$), respectively, whereas the association of 18:1n9 with T2D risk was attenuated to the null (RR = 1.03, 95% CI 0.94–1.13, $p = 0.40$). Findings were similar when each fatty acid was evaluated categorically (Fig 3).

Heterogeneity was seen in these pooled analyses, with I^2 ranging from 52.1% to 73.1% (Fig 2). The between-study heterogeneity was not associated with the global region or lipid fraction (S2 Table, S3–S6 Figs for 16:0, 16:1n7, 18:0, and 18:1n9, respectively). Among post hoc meta-

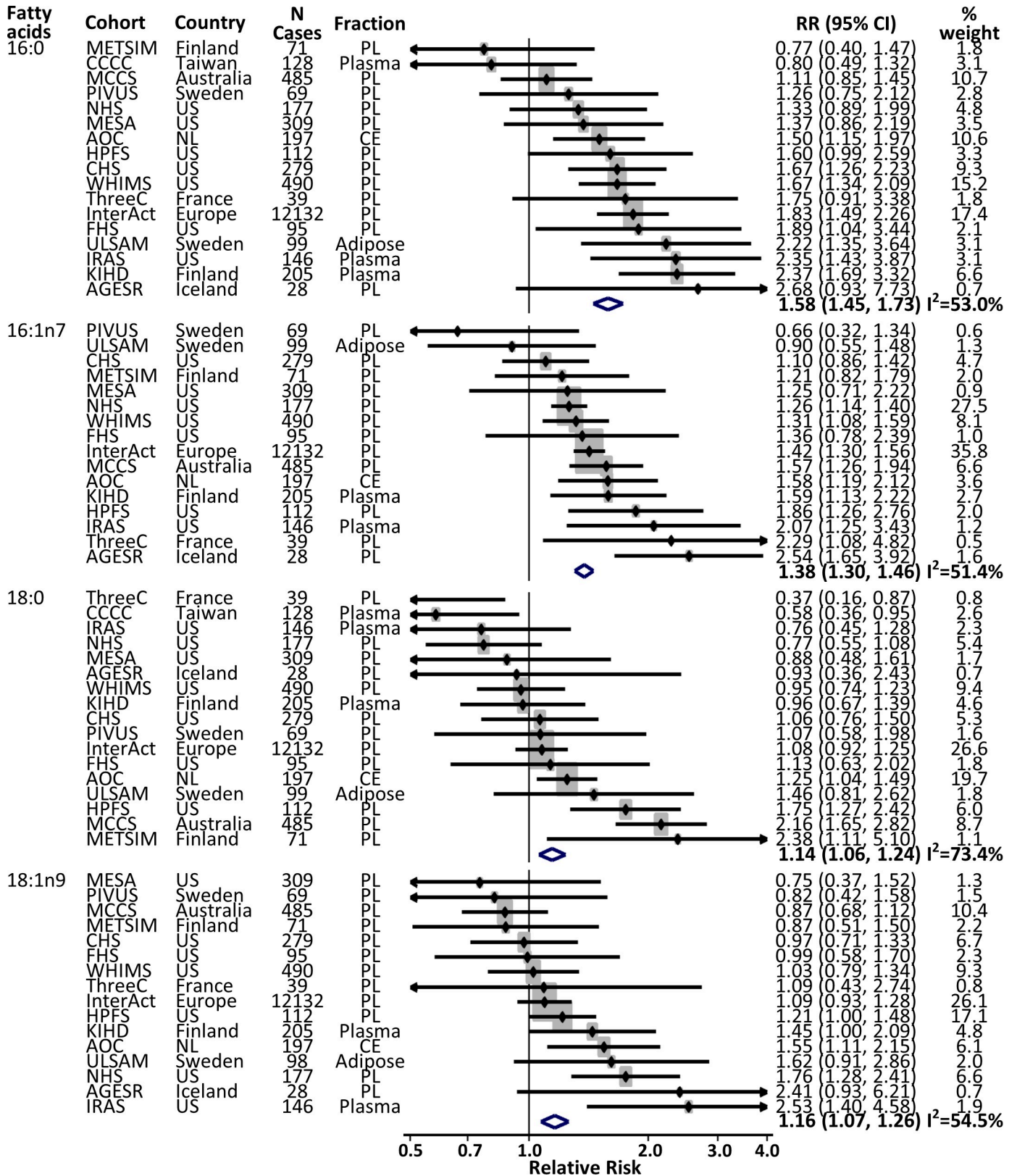


Fig 2. Associations of fatty acids in the DNL pathway with the risk of developing T2D. RRs and 95% CIs are presented in the scale per study-specific range from the midpoints of the first and fifth quintile groups (i.e., 10th to 90th percentiles): dots from individual studies and diamonds as summary estimates meta-

analyzed. The sizes of the squares of point estimates represent relative contributions of each cohort to each summary estimate (% weight). Each cohort-specific association was assessed with multivariable-adjusted regression controlling for field site (if appropriate), sex, age, race/ethnicity, socioeconomic characteristics (education, occupation), smoking status, alcohol consumption, physical activity, family history of diabetes, dyslipidaemia, hypertension, menopausal status (women), prevalent coronary heart disease, BMI, and waist circumference. Results remained similar in the other models (S1 Fig and S2 Fig), except for 18:1n9, which showed no significant result in the most adjusted model ($p = 0.69$, S2 Fig). CE, cholesteryl esters; CI, confidence interval; DNL, de novo lipogenesis; PL, phospholipids; RR, relative risk; T2D, type 2 diabetes.

<https://doi.org/10.1371/journal.pmed.1003102.g002>

regression analyses, average follow-up years explained heterogeneity of the association of 16:1n7 with T2D risk (S2 Table). I^2 estimates were 52.1% and 36.8% before and after controlling for follow-up years in meta-regression, respectively. Further stratification of cohorts into those with <10 years and those with ≥ 10 years of mean follow-up showed RRs (95% CI) of 1.64 (1.43–1.87) and 1.33 (1.25–1.41), respectively. Significant interactions were also not identified by age, sex, race, or BMI, except for sex and 18:1n9 ($p = 0.002$; S7 Fig). In exploratory meta-analysis including cohorts with both sexes and cohorts recruiting only men or women, sex-specific RRs (95% CI) for 18:1n9 were 1.17 (1.05–1.30; $p = 0.005$) for men and 1.10 (0.98–1.24; $p = 0.10$) for women.

Discussion

In this pooling project using harmonized, de novo individual-participant analyses from 17 prospective cohorts across 12 countries, biomarker concentrations of 16:0, 16:1n7, 18:0, and 18:1n9 were associated with higher risk of T2D. Associations appeared strongest for the 16-carbon saturated and monounsaturated fatty acids, followed by the 18-carbon fatty acids, and were independent of measures of adiposity. The relationships appeared partly confounded or mediated by circulating levels of blood triglycerides, a marker of DNL, although independent associations with T2D remained evident for 16:0, 16:1n7, and 18:0. Statistical heterogeneity between cohorts was largely not explained by age, sex, lipid compartment, or world region. These novel findings across 17 global cohorts suggest that the pathophysiological process of developing T2D is linked to activity of the DNL pathway and/or these circulating fatty acids.

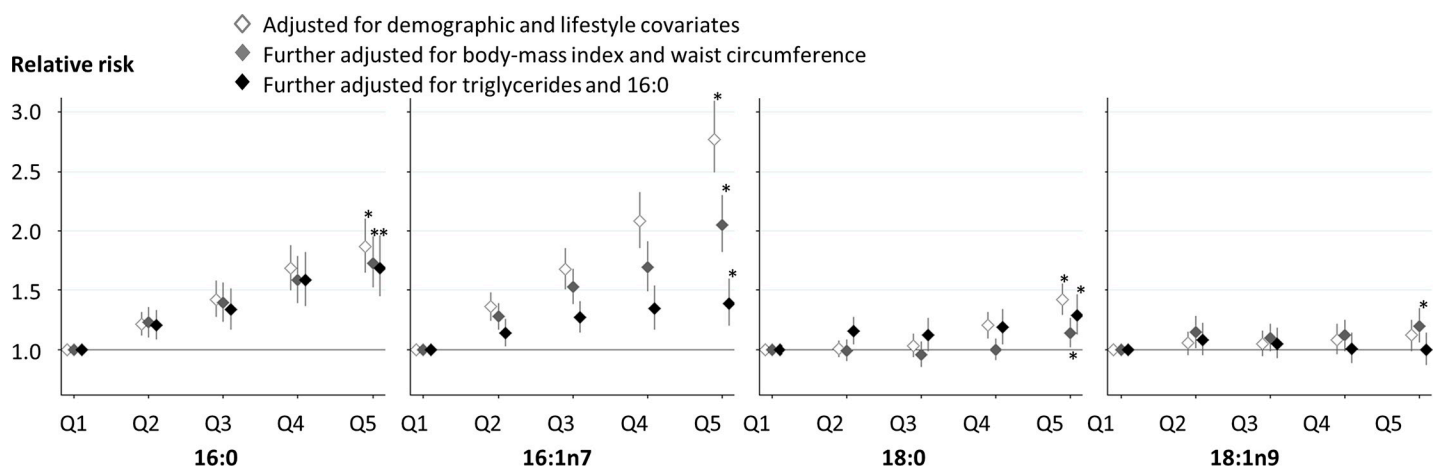


Fig 3. Associations of fatty acids in the DNL pathway with the incidence of T2D. Cohort-specific measures of associations across the quintile groups were pooled with inverse-variance-weighted meta-analysis. In each cohort, 3 different models were fitted: the first, adjusting for study field (if available), sex, age, smoking status, alcohol consumption, socioeconomic status, physical activity, dyslipidaemia, hypertension, and menopausal status (only for women); the second, adjusting for BMI and waist circumference; and the third, adjusting for triglycerides and 16:0 (for 16:1n7, 18:0 and 18:1n9) as the main products of DNL. A trend across quintiles of each fatty acid was tested with meta-analysis of cohort-specific regression coefficients of an ordinal variable of each fatty acid. The association with an asterisk showed $p < 0.001$ except for the second results for 18:0 ($p = 0.0158$) and for 18:1n9 ($p = 0.0162$). BMI, body mass index; DNL, de novo lipogenesis; T2D, type 2 diabetes.

<https://doi.org/10.1371/journal.pmed.1003102.g003>

Experimental evidence provides biological plausibility to support these findings; 16:0, the major product of DNL, appears to exert a direct toxic effect on pancreatic cells, activating membrane-bound toll-like receptor 4 and promoting pro-inflammatory responses [26], leading to impaired insulin secretion capacity [4–6]. In cells expressing insulin receptors, 16:0-ceramides attenuate insulin sensitivity by antagonizing the insulin-receptor signaling cascade and impairing endoplasmic reticulum function [27]. DNL also elevates levels of diacylglycerols, which inhibit insulin signaling and impair insulin sensitivity in skeletal muscle [28]. These mechanistic effects support our current findings of a robust positive association between in vivo 16:0 concentrations and incidence of T2D.

16:1n7 positively correlated with 16:0, and mutual adjustment partly but not fully attenuated the association between 16:1n7 and T2D. In rodents, blocking the expression of stearoyl-coenzyme A desaturase 1 gene (SCD), a rate-limiting enzyme for synthesis of 16:1n7 from 16:0, protected against insulin resistance [29]. Our results support the need for future mechanistic investigations of whether 16:1n7 and 16:0 as well as hepatic DNL and SCD activity have overlapping or partly independent roles in the pathogenesis of T2D.

The major modifiable factors that influence circulating levels of 18:0 and 18:1n9 are not well characterized. Lipid compartment-specific analyses of these fatty acids in total plasma/serum versus phospholipids suggested potentially varying associations with T2D, although the availability of cohorts to confirm such heterogeneity was limited. Further work is needed to clarify the determinants, roles, and effects on metabolic risk of 18:0 and 18:1n9 in different lipid fractions, including the potential relevance of DNL versus dietary intakes of these fatty acids.

A number of lifestyle and dietary factors may regulate DNL. Consumption of starch and sugars high in glycemic load are likely to promote DNL by increasing insulin and/or activating the carbohydrate-response pathway in the liver [1,8,9,30–32]. Certain dietary factors, such as coffee and omega-6 polyunsaturated fat, appear to suppress DNL [8,32–35] and are associated with lower incidence of T2D [20,36–38]. Other modifiable factors that may influence DNL include sleeping behavior and meal frequency [39]. Dietary intakes of saturated and monounsaturated fatty acids directly influence 16:0, 16:1n7, 18:0, and 18:1n9 [38,40,41], but it remains unclear whether these effects are similar or even smaller than the influence of endogenous synthesis and metabolism in long-term settings [42–44]. For example, limited evidence from Swedish cohorts suggests the negative or null association of carbohydrate intake with concentrations of DNL-related fatty acids in adipose tissue and phospholipids and highlights a role of saturated fat or alcohol as determining DNL fatty acids [45,46]. Further research should address this uncertainty of dietary carbohydrates and saturated fat in terms of each impact on circulating concentrations of DNL-related fatty acids, genetic activity of *SCD*, and also the accumulation of hepatic fat [35,41]. Further overall mechanistic evidence is crucial to help interpret the current dietary evidence: in contrast to our observed associations based on in vivo circulating biomarkers, dietary monounsaturated fat improves several markers of glucose-insulin homeostasis in randomized feeding trials, but dietary saturated fat has neutral effects compared with dietary carbohydrates [47].

Our analysis has several strengths. We collaboratively pooled new, standardized participant-level analyses across multiple cohorts in various global regions, improving a statistical power from a large number of studies. Our consortium approach should be robust against the potential publication bias. The standardized approaches to defining the populations, exposures, outcomes, and multivariable-adjusted analyses minimized bias and heterogeneity by method.

Our study also has limitations. The diagnosis of T2D could be missed or misclassified in some participants. However, most cohorts operated regular study visits and measurements needed for T2D ascertainment, reducing potential measurement error in the outcome ascertainment, free from bias due to health consciousness leading to T2D screening. Additionally,

any outcome misclassification would occur at random across fatty acid measures. As another limitation, we cannot rule out reverse causation that unmeasured diabetes pathophysiology may result in dysregulation of lipolysis, elevate DNL-related fatty acids, and further elevate incident T2D via separate causal pathways. The 16:1n7-T2D association significantly varied by follow-up duration, and this finding could be by chance or regression dilution but may indicate the possibility of reverse causation. This finding only for 16:1n7 may also have reflected a unique role of SCD in the development of T2D. The limitation, nonetheless, indicates the importance of the DNL pathway as a strong noncausal indicator, a causal determinant of T2D risk, or both.

We analyzed fatty acid concentrations and study covariates measured at baseline only. Those measurement errors, temporary variations over time, and unmeasured confounding factors, such as dietary correlates with carbohydrates, fat, and alcohol, could potentially bias our findings in either direction. Of note, the exposure duration represented with a single fatty acid measurement is unclear and likely to vary by laboratory, tissue, fraction type, and cohort settings. A single measure may reflect approximately 3 to 4 weeks of a habitual diet, e.g., according to published kinetic studies on essential fatty acids [24,48]. A 6-month high-carbohydrate diet resulted in higher DNL-related fatty acids concentrations in phospholipids than a 6-month high-fat diet [9]. Isotope-labeling studies related fatty acids to hepatic DNL activity and showed variable responses of specific fatty acids to a diet [49]. However, those observations tend to have been limited in size (e.g., approximately 20 adults in detailed assessments) and represent an acute dietary effect on hepatic DNL (a single meal to a few days), not representing a long-term effect or fatty acids exchanged between circulating lipids and cells [49,50]. Additionally, measures of the DNL-related fatty acids were reproducible over years (correlation coefficients = 0.3–0.7 over 5–18 years) in population-based cohorts [51,52]. Therefore, while temporality of the DNL activity is not clear with our exposure assessment, the DNL-related fatty acids we evaluated are likely to have reflected a “usual” or habitual lifestyle and metabolic status over years.

Statistical between-study heterogeneity was evident but not explained by measured characteristics except for the associations of 16:1n7 with T2D risk systematically varying by follow-up years. A limited number of cohorts investigated certain lipid compartments such as triglycerides and adipose tissue, and laboratory settings were not standardized between cohorts. In addition, some of the observed heterogeneity in this current study could reflect variation in lifestyle factors across the 17 studies. We did not identify significant heterogeneity by race/ethnicity, but the number of participants of non-European descent was relatively limited. The inclusion of only a few cohorts of non-European descent and unknown sources of heterogeneity of the observed associations limit the generalizability of our findings. To better understand the generalizability of our findings and to understand sources of heterogeneity, research in different populations with varying dietary practices is required.

In summary, the current FORCE consortium study including 17 prospective cohorts identified significant associations of higher concentrations of fatty acids related to DNL, especially 16-carbon fatty acids, in relation to incidence of T2D. These findings highlight the potential importance of DNL and these individual fatty acids in the development of T2D, and the need for further investigations on how lifestyle behavioral factors and potential interventions may influence levels of these fatty acids and DNL.

Supporting information

S1 PRISMA Checklist. The PRISMA guideline checklist. PRISMA, Preferred Reporting Items for Systematic Reviews and Meta-Analyses. (DOCX)

S1 Table. Correlations between fatty acids in the DNL pathway. DNL, de novo lipogenesis. (DOCX)

S2 Table. Exploratory analyses of the associations of fatty acids in the DNL pathway with incident T2D. DNL, de novo lipogenesis; T2D, type 2 diabetes. (DOCX)

S1 Fig. Prospective associations of fatty acids in the DNL pathway with the risk of T2D mellitus. DNL, de novo lipogenesis; T2D, type 2 diabetes. (TIF)

S2 Fig. Associations of fatty acids in the DNL pathway with the risk of T2D mellitus after adjustment for 16:0 (for 16:1n7, 18:0, 18:1n9) and triglycerides. DNL, de novo lipogenesis; T2D, type 2 diabetes. (TIF)

S3 Fig. Associations of palmitic acid (16:0), one of the fatty acids in the DNL pathway, with the risk of T2D mellitus: pooled analysis stratified by lipid fraction. DNL, de novo lipogenesis; T2D, type 2 diabetes. (TIF)

S4 Fig. Associations of palmitoleic acid (16:1n7), one of the fatty acids in the DNL pathway, with the risk of T2D mellitus: pooled analysis stratified by lipid fraction. DNL, de novo lipogenesis; T2D, type 2 diabetes. (TIF)

S5 Fig. Associations of stearic acid (18:0), one of the fatty acids in the DNL pathway, with the risk of T2D mellitus: pooled analysis stratified by lipid fraction. DNL, de novo lipogenesis; T2D, type 2 diabetes. (TIF)

S6 Fig. Associations of stearic acid (18:1n9), one of the fatty acids in the DNL pathway, with the risk of T2D mellitus: pooled analysis stratified by lipid fraction. DNL, de novo lipogenesis; T2D, type 2 diabetes. (TIF)

S7 Fig. Forest plots of $\exp(\beta)$ of statistical interaction terms by age, sex, and BMI for an association of each of the fatty acids (16:0, 16:1n7, 18:0, and 18:1n9) related to DNL with incidence of T2D. BMI, body mass index; DNL, de novo lipogenesis; T2D, type 2 diabetes. (TIF)

S1 Text. Characteristics and references of prospective cohorts evaluating associations between fatty acids related to the DNL pathway and the risk of developing T2D. DNL, de novo lipogenesis; T2D, type 2 diabetes. (DOCX)

S2 Text. Study protocol. (DOCX)

Acknowledgments

The lead author affirms that the manuscript is an honest, accurate, and transparent account of the study being reported, that no important aspects of the study have been omitted, and that any discrepancies from the study as planned have been explained.

Author Contributions

Conceptualization: Fumiaki Imamura, Amanda M. Fretts, Matti Marklund, David S. Siscovick, Jason H. Y. Wu, Renata Micha, Rozenn N. Lemaitre, Dariush Mozaffarian.

Data curation: Fumiaki Imamura, Amanda M. Fretts, Matti Marklund.

Formal analysis: Fumiaki Imamura, Andres V. Ardisson Korat, Wei-Sin Yang, Maria Lankinen, Waqas Qureshi, Tzu-An Chen, Jyrki K. Virtanen, Kerry Wong, Julie K. Bassett, Rachel Murphy, Nathan Tintle, Chaoyu Ian Yu, Alexis C. Wood, Janette de Goede, Barbara McKnight, Kalina Rajaobelina, Sabita S. Soedamah-Muthu, Dariush Mozaffarian.

Funding acquisition: Catherine Helmer, Ingeborg A. Brouwer, Kuo-Liong Chien, Johanna M. Geleijnse, Graham G. Giles, Vilmundur Gudnason, William S. Harris, Allison Hodge, Frank Hu, Albert Koulman, Markku Laakso, Lars Lind, Hung-Ju Lin, Ulf Riserus, Jennifer G. Robinson, David S. Siscovick, Nona Sotoodehnia, Qi Sun, Michael Y. Tsai, Tomi-Pekka Tuomainen, Matti Uusitupa, Lynne E. Wagenknecht, Nick J. Wareham, Rozenn N. Lemaitre, Dariush Mozaffarian, Nita G. Forouhi.

Investigation: Fumiaki Imamura, Amanda M. Fretts, Matti Marklund, Andres V. Ardisson Korat, Jason H. Y. Wu, Dariush Mozaffarian, Nita G. Forouhi.

Methodology: Fumiaki Imamura, Amanda M. Fretts, Matti Marklund, Andres V. Ardisson Korat, Albert Koulman, Barbara McKnight, Rozenn N. Lemaitre, Dariush Mozaffarian, Nita G. Forouhi.

Project administration: Fumiaki Imamura, Amanda M. Fretts, Matti Marklund, Andres V. Ardisson Korat, Liana C. del Gobbo, Luc Djousse, Johanna M. Geleijnse, Barbara McKnight, David S. Siscovick, Nona Sotoodehnia, Jason H. Y. Wu, Renata Micha, Rozenn N. Lemaitre, Dariush Mozaffarian, Nita G. Forouhi.

Resources: Fumiaki Imamura, Amanda M. Fretts, Catherine Helmer, Ingeborg A. Brouwer, Kuo-Liong Chien, Yun-yu Chen, Alexis C. Wood, Johanna M. Geleijnse, Graham G. Giles, Janette de Goede, Vilmundur Gudnason, William S. Harris, Allison Hodge, Frank Hu, Albert Koulman, Markku Laakso, Lars Lind, Hung-Ju Lin, Jennifer G. Robinson, Cecilia Samieri, Mackenzie Senn, Sabita S. Soedamah-Muthu, Qi Sun, Michael Y. Tsai, Tomi-Pekka Tuomainen, Matti Uusitupa, Lynne E. Wagenknecht, Nick J. Wareham, Jason H. Y. Wu, Rozenn N. Lemaitre, Dariush Mozaffarian, Nita G. Forouhi.

Software: Fumiaki Imamura.

Supervision: Fumiaki Imamura, Julie K. Bassett, Alexis C. Wood, Johanna M. Geleijnse, Graham G. Giles, Vilmundur Gudnason, William S. Harris, Allison Hodge, Barbara McKnight, Ulf Riserus, David S. Siscovick, Nona Sotoodehnia, Qi Sun, Jason H. Y. Wu, Rozenn N. Lemaitre, Dariush Mozaffarian, Nita G. Forouhi.

Visualization: Fumiaki Imamura.

Writing – original draft: Fumiaki Imamura, Nita G. Forouhi.

Writing – review & editing: Fumiaki Imamura, Amanda M. Fretts, Matti Marklund, Andres V. Ardisson Korat, Wei-Sin Yang, Maria Lankinen, Waqas Qureshi, Catherine Helmer, Tzu-An Chen, Jyrki K. Virtanen, Kerry Wong, Julie K. Bassett, Rachel Murphy, Nathan Tintle, Chaoyu Ian Yu, Ingeborg A. Brouwer, Kuo-Liong Chien, Yun-yu Chen, Alexis C. Wood, Liana C. del Gobbo, Luc Djousse, Johanna M. Geleijnse, Graham G. Giles, Janette de Goede, Vilmundur Gudnason, William S. Harris, Allison Hodge, Frank Hu, Albert Koulman, Markku Laakso, Lars Lind, Hung-Ju Lin, Barbara McKnight, Kalina Rajaobelina,

Ulf Riserus, Jennifer G. Robinson, Cecilia Samieri, Mackenzie Senn, David S. Siscovick, Sabita S. Soedamah-Muthu, Nona Sotoodehnia, Qi Sun, Michael Y. Tsai, Tomi-Pekka Tuomainen, Matti Uusitupa, Lynne E. Wagenknecht, Nick J. Wareham, Jason H. Y. Wu, Renata Micha, Rozenn N. Lemaitre, Dariush Mozaffarian, Nita G. Forouhi.

References

1. Ameer F, Scanduzzi L, Hasnain S, Kalbacher H, Zaidi N. De novo lipogenesis in health and disease. *Metabolism*. 2014; 63(7):895–902. <https://doi.org/10.1016/j.metabol.2014.04.003> PMID: 24814684
2. Lodhi IJ, Wei X, Semenkovich CF. Lipoexpediency: de novo lipogenesis as a metabolic signal transmitter. *Trends Endocrinol Metab*. 2011; 22(1):1–8. <https://doi.org/10.1016/j.tem.2010.09.002> PMID: 20889351
3. Borkman M, Storlien LH, Pan DA, Jenkins AB, Chisholm DJ, Campbell L V. The relation between insulin sensitivity and the fatty-acid composition of skeletal-muscle phospholipids. *New Engl J Med*. 1993; 328(4):238–44. <https://doi.org/10.1056/NEJM199301283280404> PMID: 8418404
4. Maedler K, Spinass GA, Dyntar D, Moritz W, Kaiser N, Donath MY. Distinct effects of saturated and monounsaturated fatty acids on beta-cell turnover and function. *Diabetes*. 2001; 50(1):69–76. <https://doi.org/10.2337/diabetes.50.1.69> PMID: 11147797
5. Maedler K, Oberholzer J, Bucher P, Spinass GA, Donath MY. Monounsaturated Fatty Acids Prevent the Deleterious Effects of Palmitate and High Glucose on Human Pancreatic -Cell Turnover and Function. *Diabetes*. 2003; 52(3):726–33. <https://doi.org/10.2337/diabetes.52.3.726> PMID: 12606514
6. Shi H, Kokoeva M V, Inouye K, Tzamelis I, Yin H, Flier JS. TLR4 links innate immunity and fatty acid-induced insulin resistance. *J Clin Invest*. 2006; 116(11):3015–25. <https://doi.org/10.1172/JCI28898> PMID: 17053832
7. Cnop M, Hannaert JC, Hoorens A, Eizirik DL, Pipeleers DG. Inverse relationship between cytotoxicity of free fatty acids in pancreatic islet cells and cellular triglyceride accumulation. *Diabetes*. 2001; 50(8):1771–7. <https://doi.org/10.2337/diabetes.50.8.1771> PMID: 11473037
8. Luukkonen PK, S Sädevirta S, Zhou Y, Kayser B, Ali A, Ahonen L, et al. Saturated fat is more metabolically harmful for the human liver than unsaturated fat or simple sugars. *Diabetes Care*, 2018; 41:1732–1739 <https://doi.org/10.2337/dc18-0071> PMID: 29844096
9. King IB, Lemaitre RN, Kestin M. Effect of a low-fat diet on fatty acid composition in red cells, plasma phospholipids, and cholesterol esters: investigation of a biomarker of total fat intake. *Am J Clin Nutr*. 2006; 83(2):227–36. <https://doi.org/10.1093/ajcn/83.2.227> PMID: 16469979
10. Louvet A, Mathurin P. Alcoholic liver disease: mechanisms of injury and targeted treatment. *Nat Rev Gastroenterol Hepatol*. 2015; 12(4):231–42. <https://doi.org/10.1038/nrgastro.2015.35> PMID: 25782093
11. Krachler B, Norberg M, Eriksson JW, Hallmans G, Johansson I, Vessby B, et al. Fatty acid profile of the erythrocyte membrane preceding development of Type 2 diabetes mellitus. *Nutr Metab Cardiovasc Dis*. 2008; 18(7):503–10. <https://doi.org/10.1016/j.numecd.2007.04.005> PMID: 18042359
12. Patel PS, Sharp SJ, Jansen E, Luben RN, Khaw K-T, Wareham NJ, et al. Fatty acids measured in plasma and erythrocyte-membrane phospholipids and derived by food-frequency questionnaire and the risk of new-onset type 2 diabetes: a pilot study in the European Prospective Investigation into Cancer and Nutrition (EPIC)-Norfolk. *Am J Clin Nutr*. 2010; 92(5):1214–22. <https://doi.org/10.3945/ajcn.2010.29182> PMID: 20861175
13. Kröger J, Zietemann V, Enzenbach C, Weikert C, Jansen EH, Döring F, et al. Erythrocyte membrane phospholipid fatty acids, desaturase activity, and dietary fatty acids in relation to risk of type 2 diabetes in the European Prospective Investigation into Cancer and Nutrition (EPIC)-Potsdam Study. *Am J Clin Nutr*. 2011; 93(1):127–42. <https://doi.org/10.3945/ajcn.110.005447> PMID: 20980488
14. Hodge AM, English DR, O’Dea K, Sinclair AJ, Makrides M, Gibson RA, et al. Plasma phospholipid and dietary fatty acids as predictors of type 2 diabetes: interpreting the role of linoleic acid. *Am J Clin Nutr*. 2007; 86(1):189–97. <https://doi.org/10.1093/ajcn/86.1.189> PMID: 17616780
15. Wang L, Folsom AR, Zheng Z-JJ, Pankow JS, Eckfeldt JH. Plasma fatty acid composition and incidence of diabetes in middle-aged adults: the Atherosclerosis Risk in Communities (ARIC) Study. *Am J Clin Nutr*. 2003; 78(1):91–8. <https://doi.org/10.1093/ajcn/78.1.91> PMID: 12816776
16. Harris WS, Luo J, Pottala J V., Margolis KL, Espeland MA, Robinson JG. Red Blood Cell Fatty Acids and Incident Diabetes Mellitus in the Women’s Health Initiative Memory Study. *PLoS ONE*. 2016; 11(2): e0147894. <https://doi.org/10.1371/journal.pone.0147894> PMID: 26881936
17. Forouhi NG, Koulman A, Sharp SJ, Imamura F, Kröger J, Schulze MB, et al. Differences in the prospective association between individual plasma phospholipid saturated fatty acids and incident type 2

- diabetes: the EPIC-InterAct case-cohort study. *Lancet Diabetes Endocrinol.* 2014; 2(10):810–8. [https://doi.org/10.1016/S2213-8587\(14\)70146-9](https://doi.org/10.1016/S2213-8587(14)70146-9) PMID: 25107467
18. Lankinen M a., Stančáková A, Uusitupa M, Ågren J, Pihlajamäki J, Kuusisto J, et al. Plasma fatty acids as predictors of glycaemia and type 2 diabetes. *Diabetologia.* 2015; 58(11):2533–44. <https://doi.org/10.1007/s00125-015-3730-5> PMID: 26277381
 19. Del Gobbo LC, Imamura F, Aslibekyan S, Marklund M, Virtanen JK, Wennberg M, et al. ω -3 Polyunsaturated Fatty Acid Biomarkers and Coronary Heart Disease. *JAMA Intern Med.* 2016; 176(8):1155. <https://doi.org/10.1001/jamainternmed.2016.2925> PMID: 27357102
 20. Wu JHY, Marklund M, Imamura F, Tintle N, Ardisson Korat A V, de Goede J, et al. Omega-6 fatty acid biomarkers and incident type 2 diabetes: pooled analysis of individual-level data for 39 740 adults from 20 prospective cohort studies. *Lancet Diabetes Endocrinol.* 2017; 5(12):965–74. [https://doi.org/10.1016/S2213-8587\(17\)30307-8](https://doi.org/10.1016/S2213-8587(17)30307-8) PMID: 29032079
 21. Imamura F, Fretts A, Marklund M, Ardisson Korat A V, Yang W-S, Lankinen M, et al. Fatty acid biomarkers of dairy fat consumption and incidence of type 2 diabetes: a pooled analysis of prospective cohort studies. *PLoS Med.* 2018; 15(10):e1002670. <https://doi.org/10.1371/journal.pmed.1002670> PMID: 30303968
 22. Kromhout D, Giltay EJ, Geleijnse JM, Alpha Omega Trial Group. n-3 fatty acids and cardiovascular events after myocardial infarction. *New Engl J Med.* 2010; 363(21):2015–26. <https://doi.org/10.1056/NEJMoa1003603> PMID: 20929341
 23. Higgins JP, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Stat Med.* 2002; 21(11):1539–58. <https://doi.org/10.1002/sim.1186> PMID: 12111919
 24. Katan MB, Deslypere JP, van Birgelen AP, Penders M, Zegwaard M. Kinetics of the incorporation of dietary fatty acids into serum cholesteryl esters, erythrocyte membranes, and adipose tissue: an 18-month controlled study. *J Lipid Res.* 1997; 38(10):2012–22. <https://doi.org/10.3945/ajcn.111.021907> PMID: 9374124
 25. Zhang J, Yu KF. What's the relative risk? A method of correcting the odds ratio in cohort studies of common outcomes. *JAMA.* 1998; 280(19):1690. <https://doi.org/10.1001/jama.280.19.1690> PMID: 9832001
 26. Cao H, Gerhold K, Mayers JR, Wiest MM, Watkins SM, Hotamisligil GS. Identification of a lipokine, a lipid hormone linking adipose tissue to systemic metabolism. *Cell.* 2008; 134(6):933–44. <https://doi.org/10.1016/j.cell.2008.07.048> PMID: 18805087
 27. Hla T, Kolesnick R. C16:0-Ceramide Signals Insulin Resistance. *Cell Metab.* 2014; 20(5):703–5. <https://doi.org/10.1016/j.cmet.2014.10.017> PMID: 25440051
 28. Erion DM, Shulman GI. Diacylglycerol-mediated insulin resistance. *Nat Med.* 2010; 16(4):400–2. <https://doi.org/10.1038/nm0410-400> PMID: 20376053
 29. Brown JM, Chung S, Sawyer JK, Degirolamo C, Alger HM, Nguyen T, et al. Inhibition of Stearoyl-Coenzyme A Desaturase 1 Dissociates Insulin Resistance and Obesity From Atherosclerosis. *Circulation.* 2008; 118(14):1467–75. <https://doi.org/10.1161/CIRCULATIONAHA.108.793182> PMID: 18794388
 30. Stanhope KL. Role of fructose-containing sugars in the epidemics of obesity and metabolic syndrome. *Annu Rev Med.* 2012; 63:329–43. <https://doi.org/10.1146/annurev-med-042010-113026> PMID: 22034869
 31. Livesey G, Taylor R, Livesey H, Liu S. Is there a dose-response relation of dietary glycemic load to risk of type 2 diabetes? Meta-analysis of prospective cohort studies. *Am J Clin Nutr.* 2013; 97(3):584–96. <https://doi.org/10.3945/ajcn.112.041467> PMID: 23364021
 32. Eissing L, Scherer T, Tödter K, Knippschild U, Greve JW, Buurman WA, et al. De novo lipogenesis in human fat and liver is linked to ChREBP- β and metabolic health. *Nature Comm.* 2013; 4:1528. <https://doi.org/10.1038/ncomms2537> PMID: 23443556
 33. Wu JHY, Lemaitre RN, Imamura F, King IB, Song X, Spiegelman D, et al. Fatty acids in the de novo lipogenesis pathway and risk of coronary heart disease: the Cardiovascular Health Study. *Am J Clin Nutr.* 2011; 94(2):431–8. <https://doi.org/10.3945/ajcn.111.012054> PMID: 21697077
 34. Jump DB. N-3 polyunsaturated fatty acid regulation of hepatic gene transcription. *Curr Opin Lipidol.* 2008; 19(3):242–7. <https://doi.org/10.1097/MOL.0b013e3282ffaf6a> PMID: 18460914
 35. Rosqvist F, Kullberg J, Ståhlman M, Cedernaes J, Heurling K, Johansson H-E, et al. Overeating Saturated Fat Promotes Fatty Liver and Ceramides Compared With Polyunsaturated Fat: A Randomized Trial. *J Clin Endocrinol Metab.* 2019; 104(12):6207–19. <https://doi.org/10.1210/je.2019-00160> PMID: 31369090
 36. Ding M, Bhupathiraju SN, Chen M, van Dam RM, Hu FB. Caffeinated and decaffeinated coffee consumption and risk of type 2 diabetes: a systematic review and a dose-response meta-analysis. *Diabetes Care.* 2014; 37(2):569–86. <https://doi.org/10.2337/dc13-1203> PMID: 24459154

37. Jacobs S, Kroger J, Floegel A, Boeing H, Drogan D, Pischon T, et al. Evaluation of various biomarkers as potential mediators of the association between coffee consumption and incident type 2 diabetes in the EPIC-Potsdam Study. *Am J Clin Nutr*. 2014; 100(3):891–900. <https://doi.org/10.3945/ajcn.113.080317> PMID: 25057154
38. Forouhi NG, Imamura F, Sharp SJ, Koulman A, Schulze MB, Zheng J, et al. Association of Plasma Phospholipid n-3 and n-6 Polyunsaturated Fatty Acids with Type 2 Diabetes: The EPIC-InterAct Case-Cohort Study. *PLoS Med*. 2016; 13(7):e1002094. <https://doi.org/10.1371/journal.pmed.1002094> PMID: 27434045
39. Wu JHY, Lemaitre RN, Manichaikul A, Guan W, Tanaka T, Foy M, et al. Genome-wide association study identifies novel loci associated with concentrations of four plasma phospholipid fatty acids in the de novo lipogenesis pathway: results from the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium. *Circulation Cardiovasc Genet*. 2013; 6(2):171–83. <https://doi.org/10.1161/CIRCGENETICS.112.964619> PMID: 23362303
40. Hodson L, Skeaff CM, Fielding BA. Fatty acid composition of adipose tissue and blood in humans and its use as a biomarker of dietary intake. *Prog Lipid Res*. 2008; 47(5):348–80. <https://doi.org/10.1016/j.plipres.2008.03.003> PMID: 18435934
41. Hodson L, Rosqvist F, Parry SA. The influence of dietary fatty acids on liver fat content and metabolism. *Proc Nutr Soc*. 2020; 79(1):30–41. <https://doi.org/10.1017/S0029665119000569> PMID: 30942685
42. Song X, Huang Y, Neuhauser ML, Tinker LF, Vitolins MZ, Prentice RL, et al. Dietary long-chain fatty acids and carbohydrate biomarker evaluation in a controlled feeding study in participants from the Women's Health Initiative cohort. *Am J Clin Nutr*. 2017; 105:1272–82. <https://doi.org/10.3945/ajcn.117.153072> PMID: 28446501
43. Forsythe CE, Phinney SD, Fernandez ML, Quann EE, Wood RJ, Bibus DM, et al. Comparison of Low Fat and Low Carbohydrate Diets on Circulating Fatty Acid Composition and Markers of Inflammation. *Lipids*. 2008; 43(1):65–77. <https://doi.org/10.1007/s11745-007-3132-7> PMID: 18046594
44. Hodson L, Fielding BA. Stearoyl-CoA desaturase: rogue or innocent bystander? *Prog Lipid Res*. 2013; 52(1):15–42. <https://doi.org/10.1016/j.plipres.2012.08.002> PMID: 23000367
45. Iggman D, Årnlöv J, Cederholm T, Risérus U. Association of Adipose Tissue Fatty Acids With Cardiovascular and All-Cause Mortality in Elderly Men. *JAMA Cardiol*. 2016; 1(7):745–53. <https://doi.org/10.1001/jamacardio.2016.2259> PMID: 27541681
46. Alsharari ZD, Leander K, Sjögren P, Carlsson A, Cederholm T, de Faire U, et al. Association between carbohydrate intake and fatty acids in the de novo lipogenic pathway in serum phospholipids and adipose tissue in a population of Swedish men. *Eur J Nutr*. 2019 <https://doi.org/10.1007/s00394-019-02058-6> PMID: 31350637
47. Imamura F, Micha R, Wu JHY, de Oliveira Otto MC, Otite FO, Abioye AI, et al. Effects of Saturated Fat, Polyunsaturated Fat, Monounsaturated Fat, and Carbohydrate on Glucose-Insulin Homeostasis: A Systematic Review and Meta-analysis of Randomised Controlled Feeding Trials. *PLoS Med*. 2016; 13(7):e1002087. <https://doi.org/10.1371/journal.pmed.1002087> PMID: 27434027
48. Skeaff CM, Hodson L, McKenzie JE. Dietary-Induced Changes in Fatty Acid Composition of Human Plasma, Platelet, and Erythrocyte Lipids Follow a Similar Time Course. *J Nutr*. 2006; 136(3):565–9. <https://doi.org/10.1093/jn/136.3.565> PMID: 16484525
49. Rosqvist F, McNeil CA, Pramfalk C, Parry SA, Low WS, Cornfield T, et al. Fasting hepatic de novo lipogenesis is not reliably assessed using circulating fatty acid markers. *Am J Clin Nutr*. 2019; 109(2):260–8. <https://doi.org/10.1093/ajcn/nqy304> PMID: 30721918
50. Diraison F, Pachiadi C, Beylot M. Measuring lipogenesis and cholesterol synthesis in humans with deuterated water: use of simple gas chromatographic/mass spectrometric techniques. *J Mass Spectrometry*. 1997; 32(1):81–6. [https://doi.org/10.1002/\(SICI\)1096-9888\(199701\)32:1<81::AID-JMS454>3.0.CO;2-2](https://doi.org/10.1002/(SICI)1096-9888(199701)32:1<81::AID-JMS454>3.0.CO;2-2) PMID: 9008871
51. Lai HTM, de Oliveira Otto MC, Lee Y, Wu JHY, Song X, King IB, et al. Serial Plasma Phospholipid Fatty Acids in the De Novo Lipogenesis Pathway and Total Mortality, Cause-Specific Mortality, and Cardiovascular Diseases in the Cardiovascular Health Study. *J Am Heart Assoc*. 2019; 8(22). <https://doi.org/10.1161/JAHA.119.012881> PMID: 31711385
52. Zheng J-S, Imamura F, Sharp SJ, Koulman A, Griffin JL, Mulligan AA, et al. Changes in plasma phospholipid fatty acid profiles over 13 years and correlates of change: European Prospective Investigation into Cancer and Nutrition-Norfolk Study. *Am J Clin Nutr*. 2019; 109(6):1527–34. <https://doi.org/10.1093/ajcn/nqz030> PMID: 30997506