



Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Jordan, R;Prober, SM;Hoffmann, AA;Dillon, SK

Title:

Combined analyses of phenotype, genotype and climate implicate local adaptation as a driver of diversity in eucalyptus microcarpa (grey box)

Date:

2020-05-01

Citation:

Jordan, R., Prober, S. M., Hoffmann, A. A. & Dillon, S. K. (2020). Combined analyses of phenotype, genotype and climate implicate local adaptation as a driver of diversity in eucalyptus microcarpa (grey box). *Forests*, 11 (5), <https://doi.org/10.3390/F11050495>.

Persistent Link:

<https://hdl.handle.net/11343/274506>

License:

CC BY

Article

Combined Analyses of Phenotype, Genotype and Climate Implicate Local Adaptation as a Driver of Diversity in *Eucalyptus microcarpa* (Grey Box)

Rebecca Jordan ^{1,2,*}, Suzanne M. Prober ³, Ary A. Hoffmann ¹  and Shannon K. Dillon ⁴

¹ Bio21 Institute, School of BioSciences, University of Melbourne, 30 Flemington Rd, Parkville, VIC 3010, Australia; ary@unimelb.edu.au

² CSIRO Land & Water, 15 College Rd, Sandy Bay, TAS 7005, Australia

³ CSIRO Land & Water, 147 Underwood Ave, Floreat, WA 6014, Australia; suzanne.prober@csiro.au

⁴ CSIRO Agriculture & Food, Building 801, Clunies Ross Street, Black Mountain, ACT 2601, Australia; shannon.dillon@csiro.au

* Correspondence: rebecca.jordan@csiro.au; Tel.: +61-3-6237-5641

Received: 30 March 2020; Accepted: 22 April 2020; Published: 28 April 2020



Abstract: Trees are a keystone species in many ecosystems and a critical component of ecological restoration. Understanding their capacity to respond to climate change is essential for conserving biodiversity and determining appropriate restoration seed sources. Patterns of local adaptation to climate between populations within a species can inform such conservation decisions and are often investigated from either a quantitative trait or molecular genetic basis. Here, we present findings from a combined analysis of phenotype (quantitative genetic analysis), genotype (single nucleotide polymorphism (SNP) trait associations), and climate associations. We draw on the strength of this combined approach to investigate pre-existing climate adaptation and its genetic basis in *Eucalyptus microcarpa* (Grey box), an important tree for ecological restoration in south-eastern Australia. Phenotypic data from a 26-year-old provenance trial demonstrated significant genetic variation in growth and leaf traits at both the family and provenance levels. Growth traits were only associated with temperature, whilst leaf traits were associated with temperature, precipitation and aridity. Genotyping of 40 putatively adaptive SNPs from previous genome-wide analyses identified 9 SNPs associated with these traits. Drawing on previous SNP–climate association results, several associations were identified between all three comparisons of phenotype, genotype and climate. By combining phenotypic with genomic analyses, these results corroborate genomic findings and enhance understanding of climate adaptation in *E. microcarpa*. We discuss the implication of these results for conservation management and restoration under climate change.

Keywords: climate; quantitative trait; restoration; SNP-trait association; tree

1. Introduction

Understanding adaptive variation within species is a major goal for forest tree research, past and present [1]. Given their keystone role in many terrestrial systems worldwide [2], tree responses to changes in climate could have significant implications for wider ecosystems. Despite the fact that trees generally have wide climate tolerances, recent environmental changes have had notable impacts on tree populations, including canopy die-back and mortality [3–6] and shifts in distribution [7]. Understanding the potential of trees to adapt to climate change will help in the management of forests through approaches such as assisting gene flow and provenancing across climatic gradients [8,9].

At present, genomic approaches are becoming more commonly used for assessing adaptive variation in trees ([10–12]; reviewed in [13]) as well as potential genomic vulnerability to projected

change [14,15]. Genomics provides the advantage of enabling investigations of adaptation in shorter time frames and without the logistical investments required for common garden experiments [1,16]. Furthermore, genomics can identify genes and genomic regions that have responded to selection [17,18].

Nevertheless, it remains important to understand phenotypic variation to validate genomic results and provide insight into mechanistic drivers of adaptive variation. Common garden trials have been extensively used to understand adaptive variation in trees [19], demonstrating significant genetic trait variation along environmental clines in numerous traits, for example growth [20], leaf morphology [21], stress tolerance [22], physiology [23], and even plasticity [24]. Whilst trait variation can be influenced by multiple selection pressures, climate (and especially temperature, precipitation and aridity) has been identified as a key driver of adaptive variation [23–25].

Evidence for adaptation can be built by combining both genomic and phenotypic approaches, corroborating evidence of adaptation across all three components of genotype, phenotype and climate [16,26,27]. Such an approach has been useful for several tree species: for example, relationships have been established through a combined approach between outlier single nucleotide polymorphisms (SNPs), temperature and leaf traits in *Alnus glutinosa* (Black alder) [28], between aridity-associated adaptive genetic variation and leaf traits in *Eucalyptus tricarpa* (Red ironbark) [11], and between candidate genes, temperature and bud cold tolerance in *Pseudotsuga menziesii* (Douglas fir) [29]. Demonstrating phenotypic variation and linking climate-adapted traits to climate-adaptive genotypes enables validation of genomic signatures of selection and provides insights into the potential role and function of genomics in mediating phenotypes under selection [1,26,27].

Eucalypts are the dominant tree in the majority of forests and woodlands in Australia [30]. Furthermore, they are widely used in ecological restoration plantings [31]. For many eucalypt species, climate change is projected to decrease the extent of climate-suitable habitat [32]. Migration is unlikely to match rates of climate change, hindered by the long generation times of trees, altered land use and low topographic relief in Australia, the latter making the physical distances required to track current climate niches greater than in areas of high topographic relief e.g., mountains with strong elevation gradients [33,34]. Understanding current climate adaptation will be important for utilising adaptive variation to enhance evolutionary potential in both natural and restored sites for these keystone species.

Adaptive genetic variation has been identified in eucalypts through both phenotypic and genomic approaches. There is an extensive history of quantitative trait studies in eucalypts which have demonstrated genetic variation in many traits related to both abiotic and biotic factors [19], including growth and morphology [35,36], phenology [37], physiology [38], and plasticity [24,39]. Previous quantitative trait loci (QTL) mapping has identified genomic regions associated with particular traits [19]. More recently genomic analyses have revealed potential genes and genomic regions associated with climate adaptation [11,12,40]. Whilst phenotypic and genomic approaches have been used to investigate climate adaptation in eucalypts, few studies have combined them (however, see [11]).

In this study, we combined phenotypic and genomic data to infer evidence of climate adaptation in *Eucalyptus microcarpa* (Maiden) Maiden (Grey Box). *Eucalyptus microcarpa* is an important restoration species in south-eastern Australia, used to mitigate widespread habitat loss across the species' distribution due to clearing for agriculture. Previous landscape genomic analyses identified genomic signatures of climate adaptation in *E. microcarpa* [12] including genomic variants associated with temperature, precipitation and aridity. Building on these results, this study aimed to investigate evidence of variation in quantitative traits to provide a greater understanding of adaptive variation in this species. It then aimed to further support genomic results by combining genotypic, phenotypic and climate association analyses. In particular, this study asked the following questions: (1) Is there evidence of quantitative genetic variation in growth and leaf traits of *E. microcarpa*? (2) Is variation in these quantitative traits associated with climate? (3) Are putatively adaptive genomic variants associated with quantitative traits and do these associations support evidence of climate adaptation?

2. Materials and Methods

2.1. Trial Design and Trait Measurements

A provenance trial for *Eucalyptus microcarpa* was established by the Western Australian Department of Conservation and Land Management in 1988 near Collie, Western Australia (Figure 1). Though outside the species normal distribution and established for a different purpose, this trial provided an opportunity to explore trait variation in well-established, mature trees in a common environment. Temperature at the trial site was mid-range compared to original provenance locations (Table 1). The site was wetter than original provenance locations, though intrannual variability differed with wetter winters and drier summers (Table 1). Seedlings were planted using a randomised complete block design with four replicates, with families nested by provenance within a replicate. Each replicate contained 10 open-pollinated progeny from 10 mother trees (families) from each provenance, except for the Benalla provenance which had 11 families.

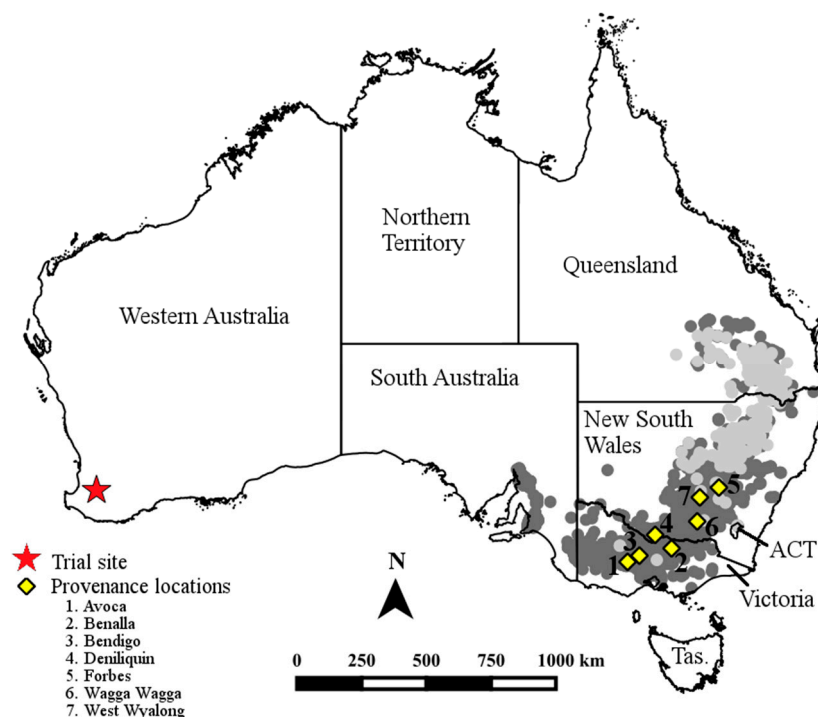


Figure 1. Map of Australia showing original provenance locations and the provenance trial site of *Eucalyptus microcarpa* (Grey box). Grey dots indicate recorded occurrences of *E. microcarpa* (dark) and *E. woollsiana* (light) respectively (data from [41]), providing an indication of the species' distribution. ACT = Australian Capital Territory. Tas. = Tasmania.

For this study, we sampled seven of the 12 planted provenances across three replicates. Provenances from Victoria and New South Wales (NSW) were sampled, avoiding potentially different species recognised after the trial was established (Queensland provenances potentially *E. woollsiana* and a NSW provenance from Gillgandra marked as *E. pilligaensis* (Narrow-leaved grey box), now considered *E. woollsiana*; [42]) and highly disjunct populations where population structure is likely to confound association analyses (South Australian provenances, disjunct from general south-eastern Australian distribution; Figure 1).

Table 1. Sample and climate data for the seven *Eucalyptus microcarpa* (Grey box) provenances measured in the provenance trial, including number of trees sampled for traits, number of trees genotyped, and climate data of original provenance locations. Climate data from [41]. See Table S1 for climate variable definitions.

Site	State	Number of Samples (<i>n</i>)				Aridity Index (Ratio) ¹		Annual (Bio12)	Precipitation (mm)			Temperature (°C)			
		DBH	Height	Leaf Traits	Genotyped	Mean Annual	Maximum Month		Summer	Winter	Driest Period (Bio14)	Wettest Period (Bio13)	Annual Mean (Bio01)	Max. Month Abs. Mean Max.	Warmest Period Max. (Bio05)
Avoca	Vic	251	65	62	58	0.740	1.797	529	103	176	6	14	13.7	43	28.8
Benalla	Vic	289	69	67	58	0.698	1.745	550	108	174	7	16	15.4	44	30.9
Bendigo	Vic	240	75	71	66	0.640	1.613	489	88	154	5	13	14.4	44	29.9
Deniliquin	NSW	250	71	64	60	0.394	0.989	374	85	109	5	10	15.6	46	31.5
Forbes	NSW	245	69	63	57	0.479	1.017	556	158	134	8	13	16.8	45	33.1
Wagga Wagga	NSW	243	73	66	61	0.487	1.189	489	110	133	7	12	16.4	45	33
West Wyalong	NSW	226	74	66	62	0.427	1.010	466	124	113	7	12	16.4	45	32.8
Total		1744	496	459	422										
Minimum						0.394	0.989	374	85	109	5	10	13.7	43	28.8
Maximum						0.740	1.797	556	158	176	8	16	16.8	46	33.1
Trial site	WA					0.796	2.445	587	42	307	2	24	15.2	45	30.6

¹ Ratio precipitation to potential evaporation (pan, free-water surface). Vic = Victoria; NSW = New South Wales; WA = Western Australia; DBH = diameter at breast height; Max. = maximum; Abs. = absolute.

Growth measurements and leaf samples were taken in October–November 2014. Diameter at breast height (DBH) and number of stems at breast height were measured on all surviving trees from the 71 families available across the seven Victorian and NSW provenances. For multi-stem trees, DBH was calculated based on the total area of all stems at breast height. Leaf samples were collected from a subset of trees for both trait measurement and genotyping. For Benalla, only 10 of the 11 families were sampled for leaf traits to remain consistent with other provenances. Approximately two randomly-selected trees per family per replicate were sampled, giving approximately six trees per family in total or 459 trees across 70 families from seven provenances sampled (Table 1). For trait measurements, 10 fully expanded, mature leaves were sampled from a single mid-point in the outer canopy. A single canopy point was used owing to the challenge of sampling from tree canopies >10 m high. To ensure this did not impact results, a second set of 10 leaves was collected from a different side of the canopy for 32 trees across families and replicates. Assessment of trait variation between canopy points for leaf area, length, weight and thickness found little difference in traits between canopy points and greater variance between individual trees than between canopy points (see Supplementary Methods, Tables S2 and S3 for more detail). This suggested most trait variation occurred between rather than within trees and thus the single canopy point was sufficient for the questions of this study. All leaves for trait measurements were stored at 4 °C during fieldwork before later being dried at 40 °C. Several additional leaves were collected and immediately dried on silica gel for later DNA extraction and genotyping.

Dried leaves for trait measurements were weighed and leaf area and length measured using a Licor (USA) LI-3000C leaf area meter and leaf thickness measured using a micrometer (Mitutoyo, Japan). Leaf weight, area, length and thickness measurements were averaged across the 10 leaves to create a single measurement per tree. Specific leaf area (SLA; area/weight) and density ([weight/area]/thickness) were calculated from average dry leaf area, weight and thickness measurements. Height was measured using a clinometer (Suunto, Finland) for the subset of trees with leaf samples and a set of additional trees (total 496; Table 1)

2.2. Phenotypic Analysis

2.2.1. Genetic Variance

To estimate the proportion of phenotypic variance due to genetic effects, family and provenance were fitted as random terms in a bivariate linear mixed model framework for each trait as per [43]:

$$y = Xb + Zu + e \quad (1)$$

where y is the vector of trait observations, b and u are vectors of fixed and random effect estimates, X and Z are incidence matrices for fixed and random model terms and e is a vector of random residual effects. Models were tested with fixed effects of Replicate and ‘Edge’—a binary variable defining a tree’s location on the outer row or column of a replicate block. ‘Edge’ accounted for potential edge effects due to inconsistent boundary tree planting and survival around the trial plots. The number of stems at breast height was included as a fixed effect for growth traits of DBH, height and size ratio. Provenance, as a random effect, was found to significantly improve models for all traits, based on likelihood-ratio tests, and was therefore retained in all models. A Wald test was used to test for significance of fixed effects, with only those fixed effects identified as significant ($p < 0.05$) retained for that trait model. Final models per trait (Table S4) were checked for linearity, homoscedasticity and normality of residuals by plotting fitted vs. residual values and a histogram of residuals.

Narrow-sense heritability (h^2) was calculated as:

$$h^2 = \frac{V_A}{V_P} = \frac{2.5\sigma_f^2}{\sigma_f^2 + \sigma_{residual}^2} \quad (2)$$

where V_A is the additive genetic variance, V_P the total phenotypic variance, σ_f^2 the family-level variance and $\sigma_{residual}^2$ the residual variance of the model. A relatedness coefficient of 1/2.5 was used to account for greater relatedness than the expected 1/4 for half-sibs commonly found within open-pollinated, half-sib families of eucalypts [44,45]. Within family relatedness estimates from SNP data in this study (see Section 2.3.1 Genotyping) suggest this is also appropriate for *E. microcarpa* (average within family relatedness coefficient = $0.32 \pm$ standard deviation (s.d.) 0.09). This estimate is an approximation of narrow-sense heritability as it includes potential maternal and dominance effects. Initial model testing for heritability estimates was performed in R v 3.2.1 [46], using the R package 'lme4' v 1.1-12 [47], before final calculations in ASReml-R (Release 3; [48]) using the pin.R function to estimate standard errors (SE; [49]).

To estimate genetic differentiation of quantitative traits between provenances, Q_{ST} was calculated from estimated variance components as per [50]:

$$Q_{ST} = \frac{\sigma_p^2}{\sigma_p^2 + 2V_A} = \frac{\sigma_p^2}{\sigma_p^2 + 2(\sigma_f^2 \times 2.5)} \quad (3)$$

where σ_p^2 is the genetic variance between populations and V_A and σ_f^2 are as defined above. Standard errors for Q_{ST} were calculated in ASReml-R using the pin.R function. Q_{ST} was not estimated for leaf density due to a lack of additive genetic variance for this trait.

To determine whether provenance-level differentiation was potentially due to selection or neutral processes, trait Q_{ST} values were compared to a distribution of neutral F_{ST} values. The expected distribution of Q_{ST} for neutral traits may be approximated by the distribution of F_{ST} for neutral genomic loci [50]. If Q_{ST} is significantly greater than the mean neutral F_{ST} , a trait may be considered significantly more differentiated than neutral expectations and potentially under diversifying selection. The distribution of neutral F_{ST} can be well approximated by a χ^2 distribution; specifically

$$\frac{(n-1)\hat{F}_{ST}}{\bar{F}_{ST}} \approx \chi^2 (n-1) \quad (4)$$

where \hat{F}_{ST} = per locus F_{ST} , \bar{F}_{ST} = mean F_{ST} and n = number of demes [51].

Individual Q_{ST} estimates (as percentages i.e., $Q_{ST} \times 100$) were deemed significantly greater than neutral expectation if the lower SE was significant ($p < 0.05$) when tested against χ^2 with $df = \bar{F}_{ST}$ (as a percentage). Mean F_{ST} was calculated from 418 'neutral' SNPs in 157 samples from the seven original provenance populations [12]; see Supplementary Methods). To avoid limitations associated with estimating a distribution of F_{ST} values from a single mean value [50], Q_{ST} estimates were also compared to the empirical F_{ST} distribution of the 418 'neutral' SNPs and similar results were obtained (see Supplementary Methods, Figure S1).

2.2.2. Genetic Trait–Trait Correlations

To explore evidence of a shared genetic basis between traits, trait-trait correlations were performed in ASReml-R to test genetic correlations between traits at both the family and provenance levels, using the mixed model framework previously described. Leaf density was not tested as no additive genetic variance was found. Models included replicate, edge and stem as fixed effects, and covariance between traits was estimated by fitting random family and provenance terms and residuals across traits in an unstructured covariance matrix, assuming heterogeneous variance estimates. Genetic correlations (r) were calculated as:

$$r_{1,2} = \frac{cov_{1,2}}{\sqrt{\sigma_1^2 \times \sigma_2^2}} \quad (5)$$

where $cov_{1,2}$ is the covariance between trait 1 and trait 2 and σ_1^2 and σ_2^2 are the variance of traits 1 and 2 respectively. A two-tailed likelihood-ratio test, testing against the null model of covariance = 0 ('diag' covariance matrix for random effects), was used to calculate the significance of genetic correlations. To account for multiple testing, q -values were calculated using the R package 'qvalue' v 1.43.0 [52] within each test level separately.

2.2.3. Climate Associations

To investigate climate as a potential selection pressure driving provenance-level trait genetic differentiation, climate associations between quantitative traits and climate of the original provenance location (Table 1) were tested using linear models in R. Ten climate variables previously used in a genomic assessment of climate adaptation in *E. microcarpa* were tested [12]; (Table S1). Briefly, climate variables were selected based on projected future hotter and drier climatic conditions in south-eastern Australia, increased extreme events, and variable precipitation patterns [53]. Climate variables related to temperature, precipitation, evaporation, moisture and aridity were reduced to a set of 10 key representative variables, minimising redundancy and correlations between variables (Table 1). Climate data for the original provenance locations were downloaded from the Atlas of Living Australia [41] (0.01 degree (~1km) gridded data). Provenance-level Best Linear Unbiased Estimates (BLUEs, equivalent to least-square means) were calculated for each trait using the R package 'lsmeans' v 2.2.3 [54] from trait specific models (Table S4) with Provenance fitted as a fixed effect.

To reduce correlations between traits, BLUEs for growth traits (DBH, height and size ratio) and leaf traits (area, length, thickness, weight, density and SLA) were reduced via separate principal component analyses (PCA) using the R package 'FactoMineR' [55]; see Table S5 for trait correlations and contributions to principal component axes). The first two axes (principal components, PC) for growth (98.4% of variation) and first three axes for leaf trait (97.9% of variation) were tested for associations with climate variables using separate linear models ('lm' function) in R. An analysis of variance (ANOVA) was performed in R ('anova' function) to calculate the p -value for the environmental component of the model. Correction for multiple testing was applied to environmental component p -values using 'qvalue' in R. To ensure associations were not due to neutral population structure, linear models were repeated including latitude and longitude of the original provenance location. This assumes a population structure pattern of isolation-by-distance, which appears to be the case for *E. microcarpa* [12,56].

2.3. Genotype-Phenotype Analysis

2.3.1. Genotyping

To look for links between potentially adaptive genomic variation and quantitative traits, a set of 40 putatively adaptive candidate SNPs were genotyped in 422 trees for which both growth and leaf trait data were collected (Table 1). Using data from genome scans, rather than candidate genes based on known gene function, has the potential to uncover new genomic regions associated with traits, without the need for *a priori* selection of potential genes of interest [57]. Candidate adaptive SNPs were chosen based on previous genomic analyses of climate adaptation in *E. microcarpa*, using reduced-representation, DArTseq SNP data [12]. Potential candidate SNPs used in this study were those identified as an F_{ST} outlier in at least one of the four previous analyses—BayeScan [58], hierarchical FDIST2 [59], FDIST2 [60,61] or Bayenv2 $X^T X$ [62], or those identified as having a strong association with at least one of the 10 climate variables tested (Bayenv2; [62,63]). In the previous analyses, environmental association analyses were only performed on SNPs identified as F_{ST} outliers. To expand the list of potential candidate SNPs for genotyping in this study, environmental associations were performed in Bayenv2 for all 4218 SNPs, using the same methodology as the previous analysis [12]. The F_{ST} outlier analyses account for population structure using various demographic models to create a 'null' distribution against which individual SNPs are compared, whilst Bayenv2 employs a covariance matrix,

generated from genomic data, to account for population structure when performing SNP–climate associations. Candidate SNPs were, therefore, those that were more differentiated than expected (F_{ST} outliers) or were associated with climate even after accounting for population structure (Bayenv2).

To determine a subset of 40 putatively adaptive candidate SNPs for genotyping, potential candidate SNPs were filtered to those with (i) a minor allele frequency of greater than 0.05 in at least six of the seven original provenance sites to ensure a high enough frequency across the dataset to enable effective SNP-trait association analyses, (ii) to SNPs within 2000 basepairs (bp) of a putative *E. grandis* gene with an *Arabidopsis thaliana* orthologue (TAIR10) based on *E. grandis* v 1.1 annotation [64] to enable exploration of potential gene function or physiological processes underlying traits, and (iii) those with an appropriate flanking sequence for DArTmp, the genotyping methodology used (see below).

An additional 25 ‘non-adaptive’ SNPs were included as a proxy for neutral population structure. Based on previous analysis, ‘non-adaptive’ SNPs were selected from those not significant in all four F_{ST} outlier tests—BayeScan ($q > 0.2$), hierarchical FDIST2 ($p > 0.1$), FDIST2 ($q > 0.2$) and $X^T X$ (outside top 10% of 3 runs), not strongly associated with any of the 10 climate variables tested (Bayenv2, Bayes Factor (BF) < 20), and with a minor allele frequency of greater than 0.05 in at least six of seven original provenance sites. Whilst SNPs deemed non-significant in F_{ST} outlier tests may still be under selection [65], this is likely to be a weaker selection and, therefore, not strongly impact the primary aim of these SNPs, being to approximate neutral structure. Despite the significantly smaller dataset, the final dataset of 65 SNPs provided a fair representation of the population structure found in the full dataset of 4218 SNPs [12]—correlation of $r = 0.69$ for provenance-level pairwise F_{ST} estimates using the 65 genotyped SNPs within the provenance trial compared to the full dataset of 4218 SNPs for the seven original natural sites (pairwise F_{ST} calculated in Arlequin v 3.5.1.2 [66]).

Flanking sequences for each SNP were extracted from genomic read data from the previous analysis [12]. Firstly, a *E. microcarpa* consensus sequence was created from genomic read data flanking the SNPs of interest by modifying the *E. grandis* v 1.1 sequence using samtools mpileup v 1.2-10 and bcftools v 1.3 [67]. *E. microcarpa* sequences for each SNP locus were then created by extracting flanking regions covered by known *E. microcarpa* reads from the “whole genome” consensus.

To genotype the SNPs, approximately 25 mm² of dried leaf material per tree and fasta sequences of SNP loci were sent to Diversity Arrays Technology Pty Ltd. for DNA extraction and subsequent SNP genotyping via DArTmp multiplex PCR. Genotypes were called from allele sequencing read counts using the maximum likelihood method described by [68] in R, using an error rate of 0.05 and only scoring genotypes with a minimum total read depth of 20. Data were filtered to SNPs and individuals with <50% missing data. Little linkage was found between SNPs (average $r^2 = 0.003 \pm$ s.d. 0.005), nor within adaptive or neutral SNP subsets ($r^2 = 0.004 \pm 0.006$ and $r^2 = 0.004 \pm 0.005$, respectively; linkage (as measured by correlation, r^2) was calculated using the ‘r’ function in PLINK v 1.90b3p [69]).

2.3.2. Genotype–Phenotype Associations

SNP-trait associations between the 40 putatively adaptive SNPs and nine scored traits were performed on 422 individuals in TASSEL v.5 [70]. To account for population structure, a PCA based on covariance was performed in TASSEL using all 65 SNPs. Overall F_{ST} , estimated via analysis of molecular variance (AMOVA) in Arlequin using all 65 SNPs, found 3% of variance between populations. Therefore, the first PC accounting for the highest amount of variance and reflecting between-population variance (PC1 = 4.1%) was retained in all analyses.

To account for relatedness between individuals, a kinship matrix was created in Coancestry v. 1.0.0.1 [71]. Coefficient of relatedness (2θ) was calculated using the triadic likelihood estimator [72] and using 100 reference individuals. This estimator can account for potential inbreeding, an important consideration given the mixed mating system of eucalypts [73]. Inbreeding (f) was calculated using the estimator of [74], which bounds inbreeding coefficient estimates between 0 and 1. Two different kinship matrices were tested. The first used individual kinship estimates, allowing for variation in the degree of relatedness between all individuals, including across families and provenances. However,

due to the small number of SNPs used in this study, relatedness estimates may be subject to large error rates. In addition, identity-by-state (genetic similarity) may be mistaken as identity-by-descent (true relatedness), especially between provenances where true sibling or cousin relationships are unlikely. To address these issues, an alternative matrix was created using the average family-level relatedness within and between families. This can reduce the effects of individual spurious estimates, especially between provenances; however, it may reduce genuine unique relationships. The average relatedness (2θ) within and between each family and the average inbreeding (f) within each family was calculated and an individual matrix created using family-level averages.

SNP-trait associations were tested in TASSEL, using a mixed linear model (MLM) based on the same underlying framework as previously described for variance component estimation (Equation (1)); with trait observations as the dependent variable (y), provenance, replicate, edge and stems as well as adaptive SNP and one PC to account for population structure as fixed effects, and kinship included as a random effect to account for relatedness between individuals. Both the raw individual kinship matrix and family-average kinship matrix were tested.

2.4. Genotype-Phenotype-Climate Analysis

To provide additional support for climate adaptation in *E. microcarpa*, and possible underlying genetic mechanisms, we compared results from the three independent pairwise associations of genotype, phenotype and climate performed in this and previous analyses [12]. Results of SNP–climate associations from the previous study were compared to trait–climate and SNP–trait associations in this study. For trait–climate comparisons, climate association results for the principal component(s) to which individual traits loaded were used (Table S5).

3. Results

3.1. Genetic Variation Within Quantitative Traits

Significant additive genetic variance suggested a genetic basis for quantitative trait variation observed in *E. microcarpa*. Family-level heritability (h^2) ranged from 0.106 to 0.373 (Table 2). Significant heritability was suggested for all traits, except leaf area, with estimates more than one standard error (SE) from 0. Additive genetic variance was not found for leaf density. Specific leaf area (SLA) and diameter at breast height (DBH) had the highest heritabilities (0.373 and 0.318, respectively), followed by leaf weight and tree height (0.230 and 0.210, respectively). Heritability estimates for other traits were below 0.2 (Table 2). Errors for heritability estimates tended to be large, potentially reflecting low replication per family.

Significant trait differentiation was found between provenances with Q_{ST} greater than one SE from 0 for all traits, excluding leaf density (Table 2). Furthermore, Q_{ST} estimates for tree height, leaf area and leaf length exceeded neutral F_{ST} (\bar{F}_{ST} 'neutral' SNPs = 0.033), suggesting these traits are potentially under divergent selection (Table 2; Figure S1). Low, neutral \bar{F}_{ST} here is in line with low differentiation identified in *E. microcarpa* using greater numbers of loci and samples [12]. Q_{ST} ranged from 0.094 to 0.483, with leaf area and leaf length having the highest Q_{ST} estimates (> 0.4), followed by leaf weight and thickness (0.224 and 0.205, respectively). For all other traits, Q_{ST} was less than 0.2. In addition to significant variation between provenances, Q_{ST} estimates also suggested greater than 50% of trait variation could be attributed to variation within provenances (Table 2, Figure S2).

3.2. Genetic Trait–Trait Correlations

Not all traits were genetically independent. Correlations in additive genetic variance suggested variation in some pairs of traits had a common heritable basis (Table 3). At the family level, all three growth traits were significantly correlated ($|r| = 0.75$ to 0.97 , $q < 0.1$), as were all six leaf traits ($|r| = 0.36$ to 0.98 , $q < 0.1$; Table 3). Between growth and leaf traits, DBH and leaf traits were independent except for leaf area ($r = -0.20$), height was correlated with all leaf traits except leaf weight ($r = -0.30$ to -0.56),

and size ratio was correlated with leaf weight and thickness ($r = -0.55$ and -0.42 respectively; Table 3). In contrast, most traits were genetically independent at the provenance level, with only five pairs of traits having significant genetic correlations ($r = 0.79$ to -0.95 , $q < 0.1$; Table 3). Genetic correlations between DBH and height ($r = 0.93$), leaf area and leaf length ($r = 0.79$), and leaf area and leaf weight ($r = 0.95$) reflected family-level results. Leaf length and SLA were also significantly correlated at the provenance level; however, contrary to family-level results, greater leaf length was correlated with higher SLA ($r = 0.93$). Although not significant at the family-level, higher leaf weight was correlated with larger DBH at the provenance level ($r = 0.92$).

Table 2. Heritability (h^2) and between provenance differentiation (Q_{ST}) for nine quantitative traits measured in *Eucalyptus microcarpa* (Grey box). SE = one standard error.

Trait	Abbreviation	Units	Family Level		Provenance Level		
			h^2	SE	Q_{ST}	SE	
Growth traits							
Diameter at breast height	DBH	cm	0.318	0.226	0.094	0.065	
Height	Height	m	0.210	0.137	0.431	0.190	*
Size Ratio	Size Ratio	height:DBH	0.106	0.080	0.165	0.162	
Leaf traits							
Leaf area	Area	cm ²	0.155	0.169	0.467	0.220	*
Leaf length	Length	cm	0.144	0.100	0.483	0.226	*
Leaf weight	Weight	g	0.235	0.102	0.224	0.145	
Leaf thickness	Thickness	mm	0.193	0.098	0.205	0.147	
Specific leaf area	SLA	mm ² mg ⁻¹	0.373	0.116	0.182	0.115	
Leaf density	Density	mg mm ⁻³	0.000	-	-	-	

* = $Q_{ST} > F_{ST}$. Lower bound SE ($Q_{ST} - SE$) significant ($p < 0.05$) against χ^2 distribution with df = mean neutral F_{ST} (3.3%).

Table 3. Family-level (upper right) and provenance-level (lower left) genetic correlations between traits in *Eucalyptus microcarpa* (Grey box). Note leaf density excluded due to no additive variance found.

	Growth Traits			Leaf Traits				
	DBH	Height	Size Ratio	Area	Length	Weight	Thickness	SLA
Growth traits								
DBH		0.88 *	-0.97 *	-0.20 *	-0.08	-0.03	-0.18	-0.15
Height	0.93 *		-0.75 *	-0.30 *	-0.56 *	0.16	-0.34 *	-0.35 *
Size Ratio	0.35	0.81		-0.35	-0.16	-0.55 *	-0.42 *	-0.15
Leaf traits								
Area	0.76	0.53	0.04		0.72 *	0.97 *	0.46 *	-0.36 *
Length	0.40	0.32	0.38	0.79 *		0.84 *	0.38 *	-0.36 *
Weight	0.92 *	0.56	-0.01	0.95 *	0.59		0.70 *	-0.77 *
Thickness	-0.37	-0.22	0.22	-0.74	-0.50	-0.76		-0.98 *
SLA	0.16	0.19	0.06	0.70	0.93 *	0.34	-0.38	

* $q < 0.1$ (accounting for multiple testing).

3.3. Climate Associations with Quantitative Trait Variation

Significant heritable genetic variation (h^2) and genetic differentiation between provenances (Q_{ST}) suggest genetic variation in phenotypes that selection can act on, and that selection has likely driven trait variation in *E. microcarpa*. An adaptive hypothesis was further supported by associations between phenotypic variation and climate among provenances, implicating local adaptation to climate as a driver of genetic variation in phenotypes.

Provenance level trait estimates (BLUEs) were reduced to two major principal component axes for growth traits and three major axes for leaf traits (Table S5). Nine of 50 potential associations between

growth or leaf trait axes and climate were significant, although only marginally after correcting for multiple testing ($p < 0.05$, $q < 0.15$; Table 4a; Table S6 for individual trait-climate associations). Stronger associations were found between climate and leaf trait axes than growth trait axes (Table 4a; Figure 2). For growth traits, the second major axis, predominately driven by increasing size ratio and to a lesser extent decreasing DBH (Table S5), was positively correlated with warmest period maximum temperature and mean annual temperature. For leaf traits, the second major axis, driven primarily by decreasing leaf density and increasing SLA and to a lesser extent increasing leaf length and thickness, was positively correlated with the three temperature variables and was negatively correlated with both aridity variables and winter precipitation. The third leaf trait axis, primarily reflecting increasing leaf weight and thickness, was negatively correlated with summer precipitation.

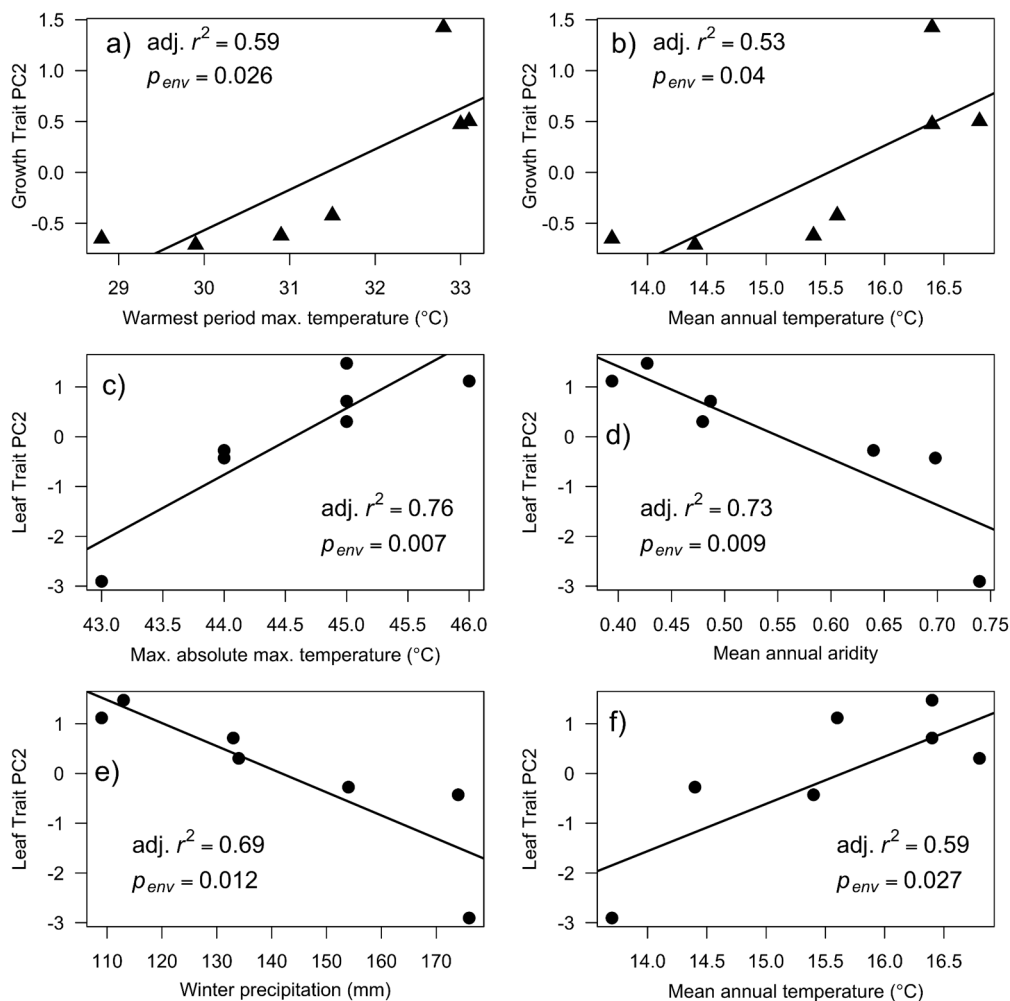


Figure 2. Example associations between climate and provenance level trait principal component (PC) axes in *Eucalyptus microcarpa* (Grey box). (a,b) Climate associations with second growth trait PC, reflecting primarily (increasing) size ratio and (decreasing) DBH. (c–f) Climate associations with second leaf trait PC, reflecting predominately (decreasing) leaf density and (increasing) SLA and to a lesser extent, (increasing) leaf thickness and leaf length. Adj. r^2 = adjusted r^2 of linear regression; p_{env} = significance of environmental component of the linear model (see Table 4). Refer to Table S5 for further details on trait contributions and correlations to PC axes.

Table 4. Top climate associations ($p_{env} < 0.05$) for leaf and growth trait principal component (PC) axes in *Eucalyptus microcarpa* (Grey box), calculated (a) without and (b) with adjustment for spatial population structure (latitude and longitude). Note p_{env} = significance of environmental component in the linear model; q_{env} = accounts for multiple testing.

Trait	Environment	(a) Trait ~ Environment					(b) Trait ~ Lat. + Long. + Environment				
		Adj. r^2	Assoc.	F_{env} [1,5]	p_{env}	q_{env}	Adj. r^2	Assoc.	F_{env} [1,3]	p_{env}	q_{env}
Growth traits											
PC2	Warmest period max. temp.	0.59	+	9.7	0.026	0.13	0.56	+	0.1	0.831	0.811
PC2	Mean annual temp.	0.53	+	7.7	0.040	0.15	0.56	−	0.1	0.835	0.811
Leaf traits											
PC2	Max. abs. mean max. temp.	0.76	+	20.1	0.007	0.13	0.69	+	7.4	0.073	0.400
PC2	Mean annual aridity	0.73	−	17.1	0.009	0.13	0.71	−	8.4	0.063	0.400
PC2	Winter precipitation	0.69	−	14.6	0.012	0.13	0.81	−	13.8	0.034	0.400
PC2	Max. aridity	0.63	−	11.1	0.021	0.13	0.58	−	4.7	0.119	0.575
PC2	Warmest period max. temp.	0.62	+	10.6	0.022	0.13	0.70	+	7.9	0.067	0.400
PC2	Mean annual temp.	0.59	+	9.5	0.027	0.13	0.74	+	9.4	0.055	0.400
PC3	Summer precipitation	0.52	−	7.6	0.040	0.15	0.26	−	2.5	0.215	0.811
PC2	Summer precipitation	−0.17	+	0.1	0.732	0.61	0.88	−	23.1	0.017	0.385
PC2	Driest period prec.	−0.17	+	0.1	0.742	0.61	0.87	−	22.7	0.018	0.385
PC2	Annual precipitation	0.13	−	1.9	0.231	0.43	0.77	−	11.4	0.043	0.400

Adj. r^2 = adjusted r^2 of linear model; Lat. = Latitude; Long. = Longitude; Assoc. = association (+ = positive; − = negative).

After accounting for possible neutral demographic effects, associations with growth traits were no longer significant (Table 4b). However, local adaptation cannot be ruled out due to strong spatial autocorrelation of the two temperature variables and latitude ($r = 0.91$ and $r = 0.90$ for mean annual temperature and warmest period maximum temperature respectively). Associations with leaf traits remained significant, although p -values were higher. Additional associations were identified when accounting for population structure—the second leaf trait axis was also associated with summer precipitation, precipitation of the driest period and annual precipitation, although these were not significant after correcting for multiple testing ($p < 0.05$, $q > 0.15$; Table 4b).

3.4. Genotype–Phenotype Associations

Associations between putatively adaptive SNPs and quantitative trait variation further support a genetic basis to trait variation, validating results of previous genomic adaptation analyses [12] and providing insights into possible genomic regulatory factors. Nine of the 40 putatively adaptive SNPs genotyped were significantly associated with at least one of the nine measured traits (Table 5). *Arabidopsis thaliana* orthologues (TAIR10) of putative *E. grandis* genes within 2000 bp of associated SNPs (Table 5; [75]) suggest trait variation in *E. microcarpa* may be influenced by genes associated with development (e.g., CHC1, [75]) and stress responses (e.g., PRK5, [76]) as well as gene regulation (e.g., MYB78, [77]). For growth traits, three significant SNP-trait associations ($p < 0.05$) were identified, each explaining approximately 0.9%–1.9% of trait variation. For leaf traits, 11 significant associations were identified (Table 5). The amount of leaf trait variance explained by an individual SNP varied from 0.9%–3%. No SNP-trait associations were significant after correction for multiple testing across SNP-trait comparisons.

3.5. Genotype–Phenotype–Climate Associations

Support for climate adaptation was strengthened by four climate-related associations that were corroborated across the three independent analyses of trait-climate, SNP-trait and, drawing on previous analyses, SNP–climate associations [12] (examples in Figure 3). In these cases, the results lend weight to a hypothesis for climate adaptation of genetic variation underlying traits in *E. microcarpa* and suggest potential candidate genes that may have been targeted by selection. It should be noted candidate genes are interpreted here as indicative only and warranting further analysis, given low linkage disequilibrium in eucalypts [78] and assumptions of sufficient similarity between *E. microcarpa* and *E. grandis* genomes and *A. thaliana* orthologues reflecting possible gene functions.

Table 5. Significant SNP-trait associations ($p < 0.05$) in *Eucalyptus microcarpa* (Grey box) and possible gene function of SNPs based on *Arabidopsis thaliana* orthologues (TAIR10) of predicted *E. grandis* genes (information from *E. grandis* v1.1 genome annotation). Mixed linear models (MLM) accounted for kinship using either a kinship matrix based on individual kinship (indv. kinship) or family-average kinship (avg. kinship).

Trait Marker	MLM (Indv. Kinship)		MLM (Avg. Kinship)		<i>Eucalyptus grandis</i> (v1.1) Gene Information (+/-2000 bp)		Best TAIR10 Gene Orthologue		
	p	r^2	p	r^2	Name	Gene Effect	Name	Symbol	Definition
DBH									
3:59841756	0.043	0.019	0.049	0.019	Eucgr.C03147	synonymous	AT3G13980.1		
Size Ratio									
10:29282238	0.023	0.012	0.015	0.014	Eucgr.J02333	synonymous	AT3G27150.1		Galactose oxidase/kelch repeat superfamily protein
10:29282238	0.023	0.012	0.015	0.014	Eucgr.J02334	upstream	AT5G14170.1	CHC1	SWIB/MDM2 domain superfamily protein
2:58822368	0.060	0.009	0.044	0.010	Eucgr.B03399	downstream	AT1G27150.1		Tetratricopeptide repeat (TPR)-like superfamily protein
Leaf area									
4:30801453	0.069	0.013	0.037	0.016	Eucgr.D01681	synonymous, intron	AT1G64660.1	ATMGL, MGL	methionine gamma-lyase
11:4085447	0.052	0.016	0.044	0.017	Eucgr.K00355	downstream	AT5G38280.1	PR5K	PR5-like receptor kinase
Leaf length									
2:63702271	0.036	0.016	0.002	0.030	Eucgr.B03985	missense	AT5G49620.1	AtMYB78, MYB78	myb domain protein 78
4:30801453	0.012	0.021	0.004	0.026	Eucgr.D01681	synonymous, intron	AT1G64660.1	ATMGL, MGL	methionine gamma-lyase
5:5009176	0.030	0.016	0.130	0.009	Eucgr.E00527	missense	AT4G23020.1		
Leaf weight									
5:4625458	0.028	0.017	0.027	0.017	Eucgr.E00491	upstream	AT3G60890.2	ZPR2	protein binding
Leaf thickness									
5:4625458	0.006	0.023	0.007	0.023	Eucgr.E00491	upstream	AT3G60890.2	ZPR2	protein binding
SLA									
5:4625458	0.024	0.018	0.017	0.020	Eucgr.E00491	upstream	AT3G60890.2	ZPR2	protein binding
11:4085447	0.020	0.021	0.015	0.022	Eucgr.K00355	downstream	AT5G38280.1	PR5K	PR5-like receptor kinase
Leaf density									
5:4625458	0.102	0.010	0.042	0.014	Eucgr.E00491	upstream	AT3G60890.2	ZPR2	protein binding
6:39617441	0.086	0.014	0.044	0.018	Eucgr.F02999	synonymous	AT1G22610.1		C2 calcium/lipid-binding plant phosphoribosyltransferase family protein

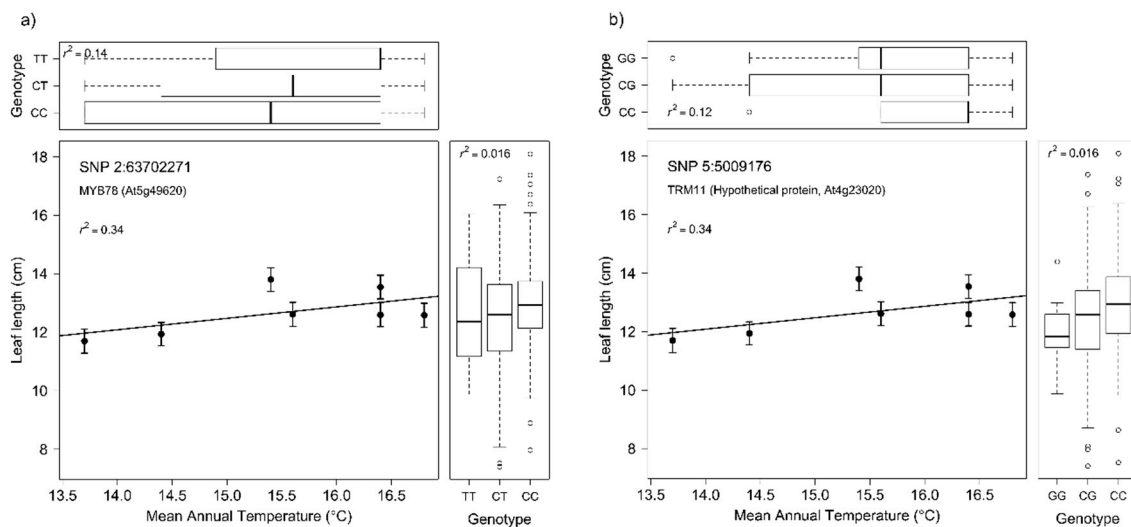


Figure 3. Examples of corroboration between three independent association analyses in *Eucalyptus microcarpa* (Grey box)—genotype (SNP; (a) SNP 2:63702271, (b) SNP 5:5009176), phenotype (leaf length) and climate (mean annual temperature). Main plot = trait (provenance level best linear unbiased estimates, BLUEs) vs. climate. Box plot (right) = SNP vs. trait (provenance BLUEs). Box plot (top) = SNP vs. climate. Error bars in main plot = 95% confidence interval. Box plot whiskers extend to $1.5 \times$ interquartile range.

Two SNPs associated with leaf length also had several climate associations in common with this trait (SNP 2:63702271, 5:5009176; Table 5; Figure 3). Leaf length correlated primarily with the first, but also the second leaf trait axis. Both leaf length, via its correlation to the second leaf trait axis and both SNPs were associated with mean annual temperature and SNP 2:63702271 was also associated with warmest period maximum temperature. In addition, the second leaf trait axis and SNP 5:5009176 were associated with aridity. Based on *A. thaliana* orthologues of predicted *E. grandis* genes, possible genes or functions that may influence climate-related variation in leaf length include an MYB domain protein (MYB78; SNP 2:63702271), and proteins of unknown function or hypothetical proteins (SNP 5:5009176; Table 5).

Results for leaf density suggest a possible link to SNP 6:39617441 (putative *A. thaliana* orthologue, C2 calcium/lipid-binding plant phosphoribosyltransferase family protein; Table 5) and temperature, with both the SNP and leaf density, via its correlation with the second leaf trait axis, associated with mean annual temperature and warmest period maximum temperature.

Finally, a potential link was found between size ratio and SNP 10:29282238, supported by both the trait, via correlation with the second growth trait axis, and the SNP associated with mean annual temperature and warmest period maximum temperature. Based on *A. thaliana* orthologues of predicted *E. grandis* genes, a possible SWIB/MDM2 domain or galactose oxidase/kelch repeat superfamily protein may be involved in temperature-related variation in size ratio (Table 5).

4. Discussion

Building on previous genomic-only assessments of adaptive variation [12], we found evidence for climate driven, local adaptation of genetic variation in growth and leaf traits of *E. microcarpa*. The identification of adaptive phenotypic variation provides support for local adaptation to climate within *E. microcarpa* and demonstrates that the genomic variation previously identified likely reflects adaptive phenotypic variation. Furthermore, the identification of both genetic variance and trait variation within provenances suggest there is variation that selection could act upon, potentially enabling adaptation in the future. Although representing a small sample of the genome as well as trait and population diversity, the combination of both phenotypic and genomic analyses bolstered evidence of

climate-related, adaptive variation in this species that warrants further investigation, especially for understanding future adaptive potential and seed sourcing under climate change.

4.1. Evidence of Genetic Variance and Climate Adaptation

In line with many ecological traits in eucalypts [19], this study detected a genetic component to variation in several traits of *E. microcarpa*, upon which selection could potentially act. Heritability estimates for growth traits in this study were similar to, or higher than, estimates seen in other eucalypts (e.g., DBH $h^2 = 0.06$ – 0.37 , height $h^2 = 0.13$; [22,35]), although estimates for leaf traits were low relative to other studies (e.g., leaf length $h^2 = 0.60$ – 0.77 ; [36,79]). Furthermore, genetic trait–trait correlations identified in this study suggest a common genetic basis, or pleiotropy, between leaf and growth traits. Genetic variance and correlation estimates in this study are likely influenced by low replication within families, increasing the standard error, as well as potential upward bias in additive variance due to maternal, dominance and epistatic effects unaccounted for [80]. Further analysis using more samples per family, as well as more test families and provenances, would greatly improve genetic variance estimates. Despite these limitations, genetic variance detected for leaf and growth traits in *E. microcarpa* here suggests potential for future evolution in response to environmental change.

Greater quantitative genetic trait differentiation between provenances than expected under a neutral model suggest selection has driven adaptive differences in a number of traits in *E. microcarpa*. This result is consistent with evidence of selection on leaf and growth traits commonly found in eucalypts as well as other tree species [19,24,35,36,81]. Whilst significant adaptive divergence in quantitative traits was detected between provenances, a large proportion (greater than 50%) of total genetic variance was attributed to variation within provenances. The presence of such standing variation further suggests a genetic source for future adaptation in these traits.

Associations between trait variation and climate suggest local adaptation in *E. microcarpa* resulting from divergent selection. This supports the results of genomic adaptation analyses in *E. microcarpa* [12] suggesting significant, climate-associated genomic differences and evidence of phenotypic variation driven by similar climatic variables in other widespread trees species [25,28,35,36]. Trait associations with maximum temperature identified here and in other studies [20,36], suggest climate extremes may be important drivers of adaptive trait differences in trees. In line with *a priori* expectations of thicker, denser leaves under higher water stress [82,83], thicker leaves in this study (second leaf trait axis) were associated with warmer, more arid home climates (consistent with [24]). However, contrary to this expectation, higher leaf density and lower SLA (second leaf trait axis) were associated with a cooler, wetter climate of origin. Plastic responses of leaf density and SLA have been found to differ between provenances and species of eucalypts [24,39,82]. Leaf density and SLA climate associations in this study may therefore reflect variation in plastic responses to environment at the trial site, especially in the case of leaf density for which no additive genetic variance was found.

One of the challenges for association studies in trees is disentangling local adaptation from neutral processes, especially where correlations between climate and population structure exist. False positives—differences due to drift rather than adaptation—can arise if population structure is not accounted for. Conversely, correcting for population structure that correlates with environment reduces the environmental effect and can result in false negatives, as shown in simulation studies of genotype–environment association analyses [65]. In this study, higher p -values when latitude and longitude were included in trait–climate associations likely reflect reduced power rather than a lack of association. Even with reduced climate effects due to correlations with spatial variables and reduced power (fewer degrees of freedom) given the small sample size (seven provenances), several associations remained marginally significant ($p = 0.034$ – 0.073 ; Table 4). Furthermore, a lack of significance after correcting for multiple testing potentially reflects a further reduction in power for this small study rather than a true lack of association. Indeed, differentiation (Q_{ST}) greater than neutral population structure supports local adaptation rather than neutral processes. Future analyses, with a greater

number of provenances will help to more clearly define those climate variables driving local adaptation in *E. microcarpa*.

4.2. Linking Genotype and Phenotype

Whilst genomics is providing advances for identifying signatures of adaptation, assessing phenotypic variation in combination with genomic data provides greater support for adaptive variation and increases understanding of potential adaptive mechanisms [16,26,84]. SNP-trait associations in this study linked previous genomic analyses [12] with quantitative analyses of climate adaptation in *E. microcarpa*, validating earlier results, providing stronger evidence of climate as a driver of local adaptation and highlighting potential genes or genic regions underlying quantitative traits.

Combining genomic and phenotypic analyses firstly demonstrated that putatively adaptive genomic markers explained, individually, a small proportion of genetic variation in quantitative traits. This, as well as multiple SNPs associating with individual traits, is in line with the highly polygenic nature of quantitative traits, and characteristic of association studies in trees [18,19,85]. Indeed, variance estimates found here are comparable to other SNP-trait association studies in trees [25,29,81]. The small number of SNP-trait associations identified and the weak significance found is likely due to the small size and power of this study, although they may also reflect the tendency of single marker analyses to miss associations due to small effect sizes [84,86]. Similarly, non-significance after correcting for multiple testing may be due to corrections being too stringent given the small study size, combined with the more subtle associations underlying highly polygenic traits. Future analyses are warranted before concrete conclusions are drawn from these results, including studies which employ more genomic markers (e.g., [18,84]), that use polygenic [85] or “omnigenic” [87] assessments of complex traits instead of single SNP associations (e.g., [88]) and that assess a wider range of traits, including phenology and physiological traits. Furthermore, additional common garden sites, in contrasting environments, would assist in separating plastic from genetic responses as well as identifying gene-by-environment interactions that were not able to be separated in this study. Lastly, multi-species analysis could provide additional evidence for genomic regions and mechanisms underlying climate adaptation (e.g., [18]).

As found in other studies [28,29], our results demonstrated how combining both genomic and phenotypic datasets can validate genomic signatures of adaptation and reveal adaptive variation and potential within a species. Phenotypic variation in this study complement and validate previous genomic variation, together demonstrating genetic variation between provenances (heritability in this study and [12]) and high variation within provenances (Q_{ST} and [56]) and thus suggesting the presence of variation upon which selection may act. Results of this study, therefore, support the combination of traditional common garden trials with modern genomic technologies as a method for investigating local adaptation and the potential for future adaptation in trees [16,26].

4.3. Conservation and Restoration under Climate Change

Incorporating evolutionary potential into conservation management will be essential for long-term sustainability in natural systems under environmental change [89]. In trees, high standing genetic variation, fecundity, gene flow, and large effective population size are likely to help enhance evolutionary potential by maintaining variation upon which selection can act [81,90]. Supporting previous findings [12,56], significant additive genetic variance and trait variation found in this study suggests high levels of standing genetic variation and therefore potential for future adaptation in *E. microcarpa*. Additionally, adaptive differences between provenances provide an opportunity for the movement of ‘pre-adapted alleles’, via gene flow or assisted migration, to help facilitate climate adaptation [90]. However, whilst additive variance is important for adaptation, it does not necessarily equate to adaptive capacity [91]. If genetic correlations identified in this study represent true genetic dependence between traits, pleiotropy may also constrain adaptive responses in one trait in favour of increased fitness in another [19]. Furthermore, additional stresses such as habitat fragmentation, reducing population size and potentially disrupting gene flow, could slow the rate of adaptation within tree populations

relative to climate change [92]. Where adaptation may not occur quickly enough, introduction of pre-adapted genetic variation through assisted migration and restoration planting may be necessary to assist evolutionary potential in natural populations [8].

As restoration seed sourcing strategies move away from 'local' and toward capturing evolutionary potential, especially pre-adapted climate-related diversity [9], understanding climate-related variation across species' distributions will help inform effective seed sourcing for long-term sustainability of populations under climate change. The results of this study, together with genomic results [12] suggest the presence of climate-associated genetic variation within *E. microcarpa* that could be utilised to enhance diversity and pre-adapted climate variation within restoration plantings and the wider landscape. In particular, temperature, aridity and summer and winter precipitation appear to be important climate drivers of adaptation in *E. microcarpa*. Sourcing seed along these climatic gradients, towards projected future climates, may therefore enhance the long-term potential of restoration sites as well as support adaptation in the wider landscape through the introduction of pre-adapted genetic variation.

5. Conclusions

Assessing phenotypic variation provides additional support and confirmation of previous genomic-based assessments of adaptive variation as well as climate as a driver of this variation. The presence of adaptive variation within and between populations suggests potential future adaptation in natural populations and may enable selective seed sourcing to enhance climate resilience in restoration plantings. However, whether the variation found is sufficient for adaptation to future climate change remains unknown. Further quantitative trait and genomic studies, especially whole-genome sequencing and polygenic assessments of adaptation, are needed to clarify the extent of adaptive variation in this species and thus potential adaptability, or vulnerability, to climate change.

Supplementary Materials: The following are available online at <http://www.mdpi.com/1999-4907/11/5/495/s1>, Supplementary Methods, Table S1: Definitions for climate variables used in climate association analysis of quantitative traits and putatively adaptive SNPs in *Eucalyptus microcarpa* (Grey box).; Table S2: Results of two sample t-tests assessing differences in leaf trait measurements between two different canopy points within a single tree (10 leaves per canopy point) for *Eucalyptus microcarpa* (Grey box).; Table S3: Mixed model estimates of variance between trees (Tree) and between two canopy points within a tree (Position) for leaf trait measurements in 32 *Eucalyptus microcarpa* (Grey box) trees, including fold difference between variance estimates.; Table S4: Mixed models for calculating narrow-sense heritability (h^2) and Q_{ST} in nine quantitative traits measured in *Eucalyptus microcarpa* (Grey box).; Table S5: Trait correlations (a) and contributions (b) to trait principal component (PC) axes in *Eucalyptus microcarpa* (Grey box).; Table S6: Climate associations for individual leaf and growth traits in *Eucalyptus microcarpa* (Grey box), calculated (a) without and (b) with adjustment for spatial population structure; Figure S1: Empirical per locus 'neutral' F_{ST} distribution (bar plot) for *Eucalyptus microcarpa* (Grey box) and Q_{ST} estimates; Figure S2: Provenance level, Best Linear Unbiased Estimates (BLUEs) for nine quantitative traits measured in *Eucalyptus microcarpa* (Grey box).

Data Availability: Raw trait data for all measured trees (FileS2_Emicrocarpa_trait_data.xlsx) and raw genotypes for all genotyped individual SNP data for the 65 SNPs used in this study (FileS3_Emicrocarpa_SNPgenotypes.vcf; FileS4_README_vcf_file.txt) are available from the CSIRO Data Access Portal: <https://doi.org/10.25919/5e9e4b5b2241e>.

Author Contributions: All authors (R.J., S.M.P., A.A.H., S.K.D.) contributed to the conceptualization and methodology of this study; R.J. undertook and supervised fieldwork, data collection and data curation; formal analysis performed by R.J. and S.K.D. with interpretation of results by all authors; R.J. wrote the original draft preparation with all authors reviewing and editing the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This study was funded by a CSIRO Julius Award (SMP), the Australian Department of Environment and Energy (SMP), Science and Industry Endowment Fund (AAH), Australian Postgraduate Award (RJ), Eucalypt Australia (RJ, AAH, SMP, SKD), Australian Flora Foundation (RJ, AAH, SMP, SKD), and Holsworth Wildlife Research Endowment (RJ).

Acknowledgments: Many thanks to Richard Mazanec for allowing use of the provenance trial he established (under then Department of Conservation and Land Management, Western Australia) and to the Department of Water (Western Australia) for access to the trial site. The authors would also like to thank Alex Arnold for fieldwork assistance, Craig Macfarlane for assistance sourcing field equipment, Nick Bell for assisting with leaf trait measurements, Washington Gapare and Milosh Ivkovich for assistance with the quantitative trait analyses,

David Bush for assistance with kinship matrices and six anonymous reviewers for constructive comments on earlier versions of this manuscript.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Neale, D.B.; Kremer, A. Forest tree genomics: Growing resources and applications. *Nat. Rev. Genet.* **2011**, *12*, 111–122. [[CrossRef](#)] [[PubMed](#)]
2. Petit, R.J.; Hampe, A. Some evolutionary consequences of being a tree. *Annu. Rev. Ecol. Evol. Syst.* **2006**, *37*, 187–214. [[CrossRef](#)]
3. Matusick, G.; Ruthrof, K.X.; Brouwers, N.C.; Dell, B.; Hardy, G.S.J. Sudden forest canopy collapse corresponding with extreme drought and heat in a mediterranean-type eucalypt forest in southwestern Australia. *Eur. J. For. Res.* **2013**, *132*, 497–510. [[CrossRef](#)]
4. Klein, T.; Cahanovitch, R.; Sprintsin, M.; Herr, N.; Schiller, G. A nation-wide analysis of tree mortality under climate change: Forest loss and its causes in Israel 1948–2017. *For. Ecol. Manag.* **2019**, *432*, 840–849. [[CrossRef](#)]
5. Andrew, M.E.; Ruthrof, K.X.; Matusick, G.; Hardy, G.E.S.J. Spatial configuration of drought disturbance and forest gap creation across environmental gradients. *PLoS ONE* **2016**, *11*, e0157154. [[CrossRef](#)]
6. Hall, A.A.G.; Gherlenda, A.N.; Hasegawa, S.; Johnson, S.N.; Cook, J.M.; Riegler, M. Anatomy of an outbreak: The biology and population dynamics of a *Cardiaspina* psyllid species in an endangered woodland ecosystem. *Agric. For. Entomol.* **2015**, *17*, 292–301. [[CrossRef](#)]
7. Wearne, L.J.; Morgan, J.W. Recent forest encroachment into subalpine grasslands near Mount Hotham, Victoria, Australia. *Arct. Antarct. Alp. Res.* **2001**, *33*, 369–377. [[CrossRef](#)]
8. Aitken, S.N.; Bemmels, J.B. Time to get moving: Assisted gene flow of forest trees. *Evol. Appl.* **2016**, *9*, 271–290. [[CrossRef](#)]
9. Prober, S.M.; Byrne, M.; McLean, E.H.; Steane, D.A.; Potts, B.M.; Vaillancourt, R.E.; Stock, W.D. Climate-adjusted provenancing: A strategy for climate-resilient ecological restoration. *Front. Ecol. Evol.* **2015**, *3*, 65. [[CrossRef](#)]
10. Mahony, C.R.; MacLachlan, I.R.; Lind, B.M.; Yoder, J.B.; Wang, T.; Aitken, S.N. Evaluating genomic data for management of local adaptation in a changing climate: A lodgepole pine case study. *Evol. Appl.* **2019**, *13*, 116–131. [[CrossRef](#)]
11. Steane, D.A.; Potts, B.M.; McLean, E.; Prober, S.M.; Stock, W.D.; Vaillancourt, R.E.; Byrne, M. Genome-wide scans detect adaptation to aridity in a widespread forest tree species. *Mol. Ecol.* **2014**, *23*, 2500–2513. [[CrossRef](#)] [[PubMed](#)]
12. Jordan, R.; Hoffmann, A.A.; Dillon, S.K.; Prober, S.M. Evidence of genomic adaptation to climate in *Eucalyptus microcarpa*: Implications for adaptive potential to projected climate change. *Mol. Ecol.* **2017**, *26*, 6002–6020. [[CrossRef](#)] [[PubMed](#)]
13. Holliday, J.A.; Aitken, S.N.; Cooke, J.E.K.; Fady, B.; González-Martínez, S.C.; Heuertz, M.; Heuertz, M.; Jaramillo-Correa, J.P.; Lexer, C.; Staton, M.; et al. Advances in ecological genomics in forest trees and applications to genetic resources conservation and breeding. *Mol. Ecol.* **2016**, *26*, 706–717. [[CrossRef](#)] [[PubMed](#)]
14. Supple, M.A.; Bragg, J.G.; Broadhurst, L.M.; Nicotra, A.B.; Byrne, M.; Andrew, R.L.; Widdup, A.; Aitken, N.C.; Borevitz, J.O. Landscape genomic prediction for restoration of a *Eucalyptus* foundation species under climate change. *eLife* **2018**, *7*, e31835. [[CrossRef](#)]
15. Ingvarsson, P.K.; Bernhardsson, C. Genome-wide signatures of environmental adaptation in European aspen (*Populus tremula*) under current and future climate conditions. *Evol. Appl.* **2019**, *13*, 132–142. [[CrossRef](#)]
16. Sork, V.L.; Aitken, S.N.; Dyer, R.J.; Eckert, A.J.; Legendre, P.; Neale, D.B. Putting the landscape into the genomics of trees: Approaches for understanding local adaptation and population responses to changing climate. *Tree Genet. Genomes* **2013**, *9*, 901–911. [[CrossRef](#)]
17. Dillon, S.; McEvoy, R.; Baldwin, D.S.; Rees, G.N.; Parsons, Y.; Southerton, S. Characterisation of adaptive genetic diversity in environmentally contrasted populations of *Eucalyptus camaldulensis* Dehnh. (River Red Gum). *PLoS ONE* **2014**, *9*, e103515. [[CrossRef](#)]

18. Yeaman, S.; Hodgins, K.A.; Lotterhos, K.E.; Suren, H.; Nadeau, S.; Degner, J.C.; Nurkowski, K.A.; Smets, P.; Wang, T.; Gray, L.K.; et al. Convergent local adaptation to climate in distantly related conifers. *Science* **2016**, *353*, 23–26. [[CrossRef](#)]
19. Kremer, A.; Potts, B.M.; Delzon, S. Genetic divergence in forest trees: Understanding the consequences of climate change. *Funct. Ecol.* **2014**, *28*, 22–36. [[CrossRef](#)]
20. McKown, A.D.; Guy, R.D.; Klápště, J.; Geraldles, A.; Friedmann, M.; Cronk, Q.C.B.; El-Kassaby, Y.A.; Mansfield, S.D.; Douglas, C.J. Geographical and environmental gradients shape phenotypic trait variation and genetic structure in *Populus trichocarpa*. *New Phytol.* **2014**, *201*, 1263–1276. [[CrossRef](#)]
21. Vitasse, Y.; Lenz, A.; Kollas, C.; Randin, C.F.; Hoch, G.; Körner, C. Genetic vs. non-genetic responses of leaf morphology and growth to elevation in temperate tree species. *Funct. Ecol.* **2014**, *28*, 243–252. [[CrossRef](#)]
22. Costa e Silva, J.; Potts, B.M.; Dutkowski, G.W. Genotype by environment interaction for growth of *Eucalyptus globulus* in Australia. *Tree Genet. Genomes* **2006**, *2*, 61–75. [[CrossRef](#)]
23. Bansal, S.; Harrington, C.A.; Gould, P.J.; St Clair, J.B. Climate-related genetic variation in drought-resistance of Douglas-fir (*Pseudotsuga menziesii*). *Glob. Chang. Biol.* **2015**, *21*, 947–958. [[CrossRef](#)] [[PubMed](#)]
24. McLean, E.H.; Prober, S.M.; Stock, W.D.; Steane, D.A.; Potts, B.M.; Vaillancourt, R.E.; Byrne, M. Plasticity of functional traits varies clinally along a rainfall gradient in *Eucalyptus tricarpa*. *Plant Cell Environ.* **2014**, *37*, 1440–1451. [[CrossRef](#)] [[PubMed](#)]
25. Holliday, J.A.; Ritland, K.; Aitken, S.N. Widespread, ecologically relevant genetic markers developed from association mapping of climate-related traits in Sitka spruce (*Picea sitchensis*). *New Phytol.* **2010**, *188*, 501–514. [[CrossRef](#)] [[PubMed](#)]
26. De Villemereuil, P.; Gaggiotti, O.E.; Mouterde, M.; Till-Bottraud, I. Common garden experiments in the genomic era: New perspectives and opportunities. *Heredity* **2016**, *116*, 249–254. [[CrossRef](#)]
27. Rellstab, C.; Gugerli, F.; Eckert, A.J.; Hancock, A.M.; Holderegger, R. A practical guide to environmental association analysis in landscape genomics. *Mol. Ecol.* **2015**, *24*, 4348–4370. [[CrossRef](#)]
28. De Kort, H.; Vandepitte, K.; Bruun, H.H.; Closset-Kopp, D.; Honnay, O.; Mergeay, J. Landscape genomics and a common garden trial reveal adaptive differentiation to temperature across Europe in the tree species *Alnus glutinosa*. *Mol. Ecol.* **2014**, *23*, 4709–4721. [[CrossRef](#)]
29. Eckert, A.J.; Bower, A.D.; Wegrzyn, J.L.; Pande, B.; Jermstad, K.D.; Krutovsky, K.V.; Clair, J.B.S.; Neale, D.B. Association genetics of coastal Douglas fir (*Pseudotsuga menziesii* var. *menziesii*, Pinaceae). I. Cold-hardiness related traits. *Genetics* **2009**, *182*, 1289–1302. [[CrossRef](#)]
30. Williams, J.E.; Brooker, I. Eucalypts: An introduction. In *Eucalypt Ecology*; Williams, J.E., Woinarski, J., Eds.; Cambridge University Press: Cambridge, UK, 1997; pp. 1–15.
31. Prober, S.M.; Potts, B.M.; Bailey, T.; Byrne, M.; Dillon, S.; Harrison, P.A.; Hoffmann, A.A.; Jordan, R.; McLean, E.H.; Steane, D.A.; et al. Climate adaptation and ecological restoration in eucalypts. *Proc. R. Soc. Victoria* **2016**, *128*, 40–53. [[CrossRef](#)]
32. Butt, N.; Pollock, L.J.; McAlpine, C.A. Eucalypts face increasing climate stress. *Ecol. Evol.* **2013**, *3*, 5011–5022. [[CrossRef](#)]
33. Hughes, L. Climate change and Australia: Key vulnerable regions. *Reg. Environ. Chang.* **2011**, *11*, 189–195. [[CrossRef](#)]
34. Aitken, S.N.; Yeaman, S.; Holliday, J.A.; Wang, T.; Curtis-McLane, S. Adaptation, migration or extirpation: Climate change outcomes for tree populations. *Evol. Appl.* **2008**, *1*, 95–111. [[CrossRef](#)] [[PubMed](#)]
35. O'Brien, E.K.; Mazanec, R.A.; Krauss, S.L. Provenance variation of ecologically important traits of forest trees: Implications for restoration. *J. Appl. Ecol.* **2007**, *44*, 583–593. [[CrossRef](#)]
36. Gauli, A.; Vaillancourt, R.E.; Bailey, T.G.; Steane, D.A.; Potts, B.M. Evidence for local climate adaptation in early-life traits of Tasmanian populations of *Eucalyptus pauciflora*. *Tree Genet. Genomes* **2015**, *11*, 104. [[CrossRef](#)]
37. Rawal, D.S.; Kasel, S.; Keatley, M.R.; Nitschke, C.R. Climatic and photoperiodic effects on flowering phenology of select eucalypts from south-eastern Australia. *Agric. For. Meteorol.* **2015**, *214*, 231–242. [[CrossRef](#)]
38. Slatyer, R. Altitudinal variation in the photosynthetic characteristics of Snow Gum, *Eucalyptus pauciflora* Sieb. ex Spreng. III Temperature response of material grown in contrasting thermal environments. *Aust. J. Plant Physiol.* **1977**, *4*, 301–312. [[CrossRef](#)]

39. Drake, J.E.; Aspinwall, M.J.; Pfautsch, S.; Rymer, P.D.; Reich, P.B.; Smith, R.A.; Crous, K.Y.; Tissue, D.T.; Ghannoum, O.; Tjoelker, M.G. The capacity to cope with climate warming declines from temperate to tropical latitudes in two widely distributed *Eucalyptus* species. *Glob. Chang. Biol.* **2015**, *21*, 459–472. [[CrossRef](#)]
40. Steane, D.A.; McLean, E.H.; Potts, B.M.; Prober, S.M.; Stock, W.D.; Stylianou, V.M.; Vaillancourt, R.E.; Byrne, M. Evidence for adaptation and acclimation in a widespread eucalypt of semi-arid Australia. *Biol. J. Linn. Soc.* **2017**, *121*, 484–500. [[CrossRef](#)]
41. Atlas of Living Australia [Internet]. Available online: <http://www.ala.org.au> (accessed on 30 January 2017).
42. Bean, A.R. Taxonomic and nomenclatural notes on the Eastern grey boxes (*Eucalyptus* ser. *Moluccanae* Chippendale, Myrtaceae) and the reinstatement of *Eucalyptus woollsiana* R.T.Baker. *Austrobaileya* **2009**, *8*, 25–34.
43. Falconer, D.S.; Mackay, T.C. *Introduction to Quantitative Genetics*, 4th ed.; Pearson: London, UK, 1996.
44. Eldridge, K.; Davidson, J.; Harwood, C.E.; van Wyk, G. *Eucalypt Domestication and Breeding*; Clarendon Press: Oxford, UK, 1993.
45. Bush, D.; Kain, D.; Matheson, C.; Kanowski, P. Marker-based adjustment of the additive relationship matrix for estimation of genetic parameters—An example using *Eucalyptus cladocalyx*. *Tree Genet. Genomes* **2011**, *7*, 23–35. [[CrossRef](#)]
46. R Core Team. *R: A Language and Environment for Statistical Computing*; R Found. Stat. Comput.: Vienna, Austria, 2018; [Internet]; Available online: <https://www.r-project.org/> (accessed on 25 July 2015).
47. Bates, D.; Mächler, M.; Bolker, B.M.; Walker, S.C. Fitting linear mixed-effects models using lme4. *J. Stat. Softw.* **2015**, *67*. [[CrossRef](#)]
48. Butler, D.; Cullis, B.; Gilmour, A.; Gogle, B. *ASReml-R Reference Manual, Release 3. Technical Series QE02001*; Queensland Department of Primary Industries: Brisbane, Australia, 2009.
49. White, I. The R Pin Function [Internet]. 2013. Available online: <http://www.homepages.ed.ac.uk/iwhite/asrem1> (accessed on 1 December 2016).
50. Whitlock, M.C. Evolutionary inferences from, Q_{ST} . *Mol. Ecol.* **2008**, *17*, 1885–1896. [[CrossRef](#)] [[PubMed](#)]
51. Lewontin, R.; Krakauer, J. Distribution of gene frequency as a test of the theory of the selective neutrality of polymorphisms. *Genetics* **1973**, *74*, 175–195.
52. Dabney, A.; Storey, J.D. Qvalue: Q-Value Estimation for False Discovery Rate Control. R Package Version 1.43.0 [Internet]. 2015. Available online: <https://bioconductor.riken.jp/packages/3.0/bioc/html/qvalue.html> (accessed on 1 December 2016).
53. Reisinger, A.; Kitching, R.L.; Chiew, F.; Hughes, L.; Newton, P.C.D.; Schuster, S.S.; Tait, A.; Whetton, P. Australasia. In *Climate Change 2014: Impacts, Adaptation, and Vulnerability. Part B: Regional Aspects. Contribution of Working Group II to the fifth Assessment Report of the Intergovernmental Panel on Climate Change*; Barros, V.R., Field, C.B., Dokken, D.J., Mastrandrea, M.D., Mach, K.J., Bilir, T.E., Chatterjee, M., Ebi, K.L., Estrada, Y.O., Genova, R.C., et al., Eds.; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2014; pp. 1371–1438.
54. Lenth, R.V. Least-squares means: The R package lsmeans. *J. Stat. Softw.* **2016**, *69*. [[CrossRef](#)]
55. Lê, S.; Josse, J.; Husson, F. FactoMineR: An R package for multivariate analysis. *J. Stat. Softw.* **2008**, *25*, 1–18. [[CrossRef](#)]
56. Jordan, R.; Dillon, S.K.; Prober, S.M.; Hoffmann, A.A. Landscape genomics reveals altered genome wide diversity within revegetated stands of *Eucalyptus microcarpa* (Grey Box). *New Phytol.* **2016**, *212*, 992–1006. [[CrossRef](#)]
57. Tiffin, P.; Ross-Ibarra, J. Advances and limits of using population genetics to understand local adaptation. *Trends Ecol. Evol.* **2014**, *29*, 673–680. [[CrossRef](#)]
58. Foll, M.; Gaggiotti, O. A genome-scan method to identify selected loci appropriate for both dominant and codominant markers: A Bayesian perspective. *Genetics* **2008**, *180*, 977–993. [[CrossRef](#)]
59. Excoffier, L.; Hofer, T.; Foll, M. Detecting loci under selection in a hierarchically structured population. *Heredity* **2009**, *103*, 285–298. [[CrossRef](#)]
60. Beaumont, M.A.; Nichols, R.A. Evaluating loci for use in the genetic analysis of population structure. *Proc. R. Soc. B Biol. Sci.* **1996**, *263*, 1619–1626.
61. Antao, T.; Lopes, A.; Lopes, R.J.; Beja-Pereira, A.; Luikart, G. LOSITAN: A workbench to detect molecular adaptation based on a F_{ST} -outlier method. *BMC Bioinform.* **2008**, *9*, 323. [[CrossRef](#)] [[PubMed](#)]

62. Günther, T.; Coop, G. Robust identification of local adaptation from allele frequencies. *Genetics* **2013**, *195*, 205–220. [[CrossRef](#)] [[PubMed](#)]
63. Coop, G.; Witonsky, D.; Di Rienzo, A.; Pritchard, J.K. Using environmental correlations to identify loci underlying local adaptation. *Genetics* **2010**, *185*, 1411–1423. [[CrossRef](#)]
64. Myburg, A.A.; Grattapaglia, D.; Tuskan, G.A.; Hellsten, U.; Hayes, R.D.; Grimwood, J.; Jenkins, J.; Lindquist, E.; Tice, H.; Bauer, D. The genome of *Eucalyptus grandis*. *Nature* **2014**, *510*, 356–362. [[CrossRef](#)]
65. Lotterhos, K.E.; Whitlock, M.C. The relative power of genome scans to detect local adaptation depends on sampling design and statistical method. *Mol. Ecol.* **2015**, *24*, 1031–1046. [[CrossRef](#)]
66. Excoffier, L.; Laval, G.; Schneider, S. Arlequin (version 3.0): An integrated software package for population genetics data analysis. *Evol. Bioinforma.* **2005**, *1*, 47–50. [[CrossRef](#)]
67. Li, H.; Handsaker, B.; Wysoker, A.; Fennell, T.; Ruan, J.; Homer, N.; Marth, G.; Abecasis, G.; Durbin, R. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* **2009**, *25*, 2078–2079. [[CrossRef](#)]
68. Blischak, P.D.; Kubatko, L.S.; Wolfe, A.D. Accounting for genotype uncertainty in the estimation of allele frequencies in autopolyploids. *Mol. Ecol. Resour.* **2016**, *16*, 742–754. [[CrossRef](#)]
69. Purcell, S.; Neale, B.; Todd-Brown, K.; Thomas, L.; Ferreira, M.A.R.; Bender, D.; Maller, J.; Sklar, P.; De Bakker, P.I.; Daly, M.J.; et al. PLINK: A tool set for whole-genome association and population-based linkage analyses. *Am. J. Hum. Genet.* **2007**, *81*, 559–575. [[CrossRef](#)]
70. Bradbury, P.J.; Zhang, Z.; Kroon, D.E.; Casstevens, T.M.; Ramdoss, Y.; Buckler, E.S. TASSEL: Software for association mapping of complex traits in diverse samples. *Bioinformatics* **2007**, *23*, 2633–2635. [[CrossRef](#)] [[PubMed](#)]
71. Wang, J. COANCESTRY: A program for simulating, estimating and analysing relatedness and inbreeding coefficients. *Mol. Ecol. Resour.* **2011**, *11*, 141–145. [[CrossRef](#)] [[PubMed](#)]
72. Wang, J. Triadic IBD coefficients and applications to estimating pairwise relatedness. *Genet. Res.* **2007**, *89*, 135–153. [[CrossRef](#)] [[PubMed](#)]
73. House, S.M. Reproductive biology of eucalypts. In *Eucalypt Ecology*; Williams, J.E., Woinarski, J., Eds.; Cambridge University Press: Cambridge, UK, 1997; pp. 30–55.
74. Lynch, M.; Ritland, K. Estimation of pairwise relatedness with molecular markers. *Genetics* **1999**, *152*, 1753–1766. [[PubMed](#)]
75. TAIR. The Arabidopsis Information Resource (TAIR) [Internet]. 2017. Available online: <https://www.arabidopsis.org> (accessed on 1 March 2017).
76. Yoon, S.-K.; Park, E.-J.; Choi, Y.-I.; Bae, E.-K.; Kim, J.-H.; Park, S.-Y.; Kang, K.S.; Lee, H. Response to drought and salt stress in leaves of poplar (*Populus alba* × *Populus glandulosa*): Expression profiling by oligonucleotide microarray analysis. *Plant Physiol. Biochem.* **2014**, *84*, 158–168. [[CrossRef](#)] [[PubMed](#)]
77. Yanhui, C.; Xiaoyuan, Y.; Kun, H.; Meihua, L.; Jigang, L.; Zhaofeng, G.; Zhiqiang, L.; Yunfei, Z.; Xiaoxiao, W.; Xiaoming, Q.; et al. The MYB transcription factor superfamily of *Arabidopsis*: Expression analysis and phylogenetic comparison with the rice MYB family. *Plant Mol. Biol.* **2006**, *60*, 107–124. [[CrossRef](#)]
78. Silva-Junior, O.B.; Grattapaglia, D. Genome-wide patterns of recombination, linkage disequilibrium and nucleotide diversity from pooled resequencing and single nucleotide polymorphism genotyping unlock the evolutionary history of *Eucalyptus grandis*. *New Phytol.* **2015**, *208*, 830–845. [[CrossRef](#)]
79. Steane, D.A.; Conod, N.; Jones, R.C.; Vaillancourt, R.E.; Potts, B.M. A comparative analysis of population structure of a forest tree, *Eucalyptus globulus* (Myrtaceae), using microsatellite markers and quantitative traits. *Tree Genet. Genomes* **2006**, *2*, 30–38. [[CrossRef](#)]
80. White, T.L.; Adams, W.T.; Neale, D.B. *Forest Genetics*; CAB International: Oxfordshire, UK, 2007.
81. Alberto, F.J.; Aitken, S.N.; Alía, R.; González-Martínez, S.C.; Hänninen, H.; Kremer, A.; Lefèvre, F.; Lenormand, T.; Yeaman, S.; Whetten, R.; et al. Potential for evolutionary responses to climate change—Evidence from tree populations. *Glob. Chang. Biol.* **2013**, *19*, 1645–1661. [[CrossRef](#)]
82. Merchant, A.; Callister, A.; Arndt, S.; Tausz, M.; Adams, M. Contrasting physiological responses of six *Eucalyptus* species to water deficit. *Ann. Bot.* **2007**, *100*, 1507–1515. [[CrossRef](#)]
83. Niinemets, Ü. Global-scale climatic controls of leaf dry mass per area, density and thickness in trees and shrubs. *Ecology* **2001**, *82*, 453–469. [[CrossRef](#)]
84. Eckert, A.J.; Wegrzyn, J.L.; Liechty, J.D.; Lee, J.M.; Cumbie, W.P.; Davis, J.M.; Goldfarb, B.; Loopstra, C.A.; Palle, S.R.; Quesada, T.; et al. The evolutionary genetics of the genes underlying phenotypic associations for loblolly pine (*Pinus taeda*, Pinaceae). *Genetics* **2013**, *195*, 1353–1372. [[CrossRef](#)] [[PubMed](#)]

85. Lind, B.M.; Menon, M.; Bolte, C.E.; Faske, T.M.; Eckert, A.J. The genomics of local adaptation in trees: Are we out of the woods yet? *Tree Genet. Genomes* **2018**, *14*, 29. [[CrossRef](#)]
86. Le Corre, V.; Kremer, A. The genetic differentiation at quantitative trait loci under local adaptation. *Mol. Ecol.* **2012**, *21*, 1548–1566. [[CrossRef](#)] [[PubMed](#)]
87. Boyle, E.A.; Li, Y.I.; Pritchard, J.K. An expanded view of complex traits: From polygenic to omnigenic. *Cell* **2017**, *169*, 1177–1186. [[CrossRef](#)] [[PubMed](#)]
88. Rajora, O.P.; Eckert, A.J.; Zinck, J.W.R. Single-locus versus multilocus patterns of local adaptation to climate in Eastern White Pine (*Pinus strobus*, Pinaceae). *PLoS ONE* **2016**, *11*, e0158691. [[CrossRef](#)] [[PubMed](#)]
89. Hoffmann, A.; Griffin, P.; Dillon, S.; Catullo, R.; Rane, R.; Byrne, M.; Jordan, R.; Oakeshott, J.; Weeks, A.; Joseph, L.; et al. A framework for incorporating evolutionary genomics into biodiversity conservation and management. *Clim. Chang. Responses* **2015**, *2*, 1. [[CrossRef](#)]
90. Kremer, A.; Ronce, O.; Robledo-Arnuncio, J.J.; Guillaume, F.; Bohrer, G.; Nathan, R.; Bridle, J.R.; Gomulkiewicz, R.; Klein, E.K.; Ritland, K.; et al. Long-distance gene flow and adaptation of forest trees to rapid climate change. *Ecol. Lett.* **2012**, *15*, 378–392. [[CrossRef](#)]
91. Hoffmann, A.A.; Sgrò, C.M.; Kristensen, T.N. Revisiting adaptive potential, population size, and conservation. *Trends Ecol. Evol.* **2017**, *32*, 506–517. [[CrossRef](#)]
92. Lienert, J. Habitat fragmentation effects on fitness of plant populations—A review. *J. Nat. Conserv.* **2004**, *12*, 53–72. [[CrossRef](#)]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).