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Cross-talk between tumors at anatomically distinct sites

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34 **Running title:** Cross-talk between metastatic tumors

35 **List of abbreviations:**

36 TME – Tumor microenvironment

37 PLR – paraneoplastic leukemoid reaction

38 GM-CSF/G-CSF – granulocyte-macrophage/granulocyte colony stimulating factor

39 IL – interleukin

40 TNF – Tumor necrosis factor

41 MDSC – myeloid derived suppressor cell

42 VEGF – vascular endothelial growth factor

43 DC – dendritic cell

44 EV – extracellular vesicle

45 EGFR – epithelial growth factor receptor

46 TCR – T cell receptor

47 NK – natural killer

48 Treg – regulatory T cell

49 FLT3L – FMS-like tyrosine kinase receptor ligand

50 SC - subcutaneous

51 **Abstract**

52 Cancer tissue is not homogenous, and individual metastases at different anatomical locations can
53 differ from the primary tumor and from one another in both their morphology and cellular
54 composition, even within an individual patient. Tumors are composed of cancer cells and a range of

55 other cell types, which, together with a variety of secreted molecules, collectively comprise the tumor
56 microenvironment. Cells of the tumor microenvironment can communicate with each other and with
57 distant tissues in a form of molecular cross-talk to influence their growth and function. Cross-talk
58 between cancer cells and local immune cells is well described, and can lead to the induction of local
59 immunosuppression. Recently, it has become apparent that tumors located remotely from each other,
60 can engage in cross-talk that can influence their responsiveness to various therapies, including
61 immunotherapy. In this article, we review studies that describe how tumors systemically communicate
62 with distant tissues through motile cells, extracellular vesicles and secreted molecules that can affect
63 their function. In addition, we summarize evidence from mouse studies and the clinic that indicate an
64 ability of some tumors to influence the progression and therapeutic responses of other tumors in
65 different anatomical locations.

66 **Introduction**

67 Tumors in humans often spread to multiple different sites within one patient. All tissues of
68 the body are connected through lymphatic and vascular systems, and systemic immune
69 responses against tumors have been shown to impact distal tissues and tumors. Tumor cells
70 and their associated cells, collectively known as the tumor microenvironment (TME), in one
71 location can have a significant influence on the TME of distal sites. Various factors secreted
72 from tumors, such as growth factors and tumor-derived extracellular vesicles (EVs), have
73 been observed in the circulation (**Figure 1A and B**). Immune cell populations in peripheral
74 blood and lymphoid organs are influenced by tumor cell derived factors, leading to systemic
75 changes within the host (**Figure 1D**).

76
77 The impact of tumors on non-cancerous distal tissues is largely demonstrated in studies of the
78 pre-metastatic niche, where secreted factors from a primary tumor create an environment
79 within the pre-metastatic tissue conducive to tumor cell growth (**Figure 1C**). In addition, the
80 term “abscopal effect” that was first used by Mole in 1953 in radiotherapy describes a
81 phenomenon where local treatment of one tumor leads to regression of a distant tumor [1].
82 This suggests it is possible that tumors in different metastatic sites can influence each other
83 during tumor development, progression and therapy. However, very few studies have
84 investigated how tumors in different metastatic sites might impact each other. In this article,
85 we review studies that consider the existing evidence on the concept of cross-talk between
86 tumors in different anatomical locations (**Figure 1E**).

87

88 **Systemic changes mediated by tumor cells**

89 ***Circulating factors from tumors***

90 Tumor cells secrete various molecules including cytokines and growth factors that maintain
91 the local microenvironment. These tumor molecules can also be released into the circulation
92 where they can influence distant tissues. A clinical example of this comes from
93 paraneoplastic leukemoid reaction (PLR) which describes a variety of hematological clinical
94 disorders due to a non-hematolymphoid tumor. PLR has been reported in patients with a
95 variety of cancers, including lung cancers, melanoma, mesothelioma, head and neck cancer
96 and cancers of other origins [2]. The most commonly secreted cytokine by these tumors
97 include granulocyte colony stimulating factor (G-CSF), granulocyte-macrophage colony
98 stimulating factor (GM-CSF), IL-1, IL-6 and TNF- α . In fact, GM-CSF is expressed by a
99 number of tumor types constitutively, with one study reporting 23 of 75 human tumor lines
100 producing GM-CSF [3]. These cytokines stimulate bone marrow myelopoiesis and inhibit
101 myeloid cell differentiation that leads to accumulation of immature myeloid cells, which are
102 called myeloid-derived suppressor cells (MDSCs) that suppress effector cell responses.

103
104 Another common tumor-secreted protein which has systemic effects is vascular-endothelial
105 growth factor (VEGF). Increased plasma levels of VEGF in cancer patients correlates with
106 poorer prognosis [4] and is thought to impact on dendritic cell (DC) differentiation [5].
107 Furthermore, arginase activity is higher in peripheral blood of gastric cancer patients and
108 could potentially be involved in suppressing functions of circulating immune cells [6]. In
109 each of these examples, tumor growth can result in an altered serum proteome which, can
110 impact on distal tissues and alter the normal physiology of the host. Of course, many other
111 proteins are secreted by tumor cells, but have no function in malignancy or transformation of
112 distal tissues. Examples include elevated serum AFP (hepatocellular cancer), HCG
113 (choriocarcinoma), PSA (prostate cancer), CEA (colorectal cancer), CA 125 (ovarian cancer)
114 and CA 15-3 (breast cancer). Many of these proteins have been used for decades as
115 biomarkers of tumor size indicating both tumor progression and response to therapy [7].

116

117 ***Extracellular vesicles***

118 EVs are small membrane-encapsulated particles, derived from cells, which contain DNA,
119 RNA and proteins representative of the host cells. EVs describe both exosomes and
120 microvesicles which can mediate communication between cells, through their content, to
121 modulate both the local and distal microenvironments. Exosomes are released from the cell

122 by fusion of multivesicular endosomes with the plasma membrane whereas microvesicles are
123 formed by budding off the plasma membrane [8]. Tumor-derived or tumor-associated EVs
124 can exchange molecules from malignant or non-malignant cells within the TME to other cells
125 throughout the body [9]. This communication can impact on signaling between the tumor and
126 host, and can influence many aspects of tumorigenesis. Expression of ligands such as PD-L1,
127 which can engage with receptors expressed by T cells to impair their function, have been
128 identified on EVs. High levels of EV PD-L1 associate with clinical non-responders to anti-
129 PD-1 therapy in metastatic melanoma [10].

130

131 EVs can be found in the blood and lymph through which they travel to distant tissues. A
132 number of studies have demonstrated the influence of EVs on the pre-metastatic niche.
133 Tumor-derived EVs have been shown to reprogram fibroblasts in pre-metastatic sites to
134 generate an acidic microenvironment inhibitory for anti-tumor immune responses [11-13].
135 Epidermal growth factor receptor (EGFR)-expressing EVs secreted by gastric cancer cells
136 enhance liver metastasis through induction of hepatocyte growth factor signaling in liver
137 stromal cells [14]. Melanoma-derived EVs have been shown to induce vascular leakiness,
138 allowing for increased accessibility for tumor cells into pre-metastatic sites [15]. In addition,
139 tumor-derived EVs can also suppress monocyte maturation, leading to increased MDSCs,
140 effectively increasing systemic tumor-mediated immunosuppression [16]. Thus, tumor
141 derived EVs represent a form of communication from tumors to distal tissues that may
142 potentially impact on distal TMEs and immune responses in other tumors.

143

144 ***Circulating cells***

145 Various studies have focused on changes in the peripheral blood of cancer patients and pre-
146 clinical cancer models. One study generated a novel system to match interactions across
147 tissues to link pathways within primary tumors with changes in the systemic response in 173
148 breast cancer patients [17]. This study revealed that expression of genes in circulating blood
149 cells are tightly linked to the genes in the primary tumors, and such gene expression patterns
150 vary with breast cancer subtypes. Therefore, the RNA profiles of tumors can be indicative of
151 systemic immune responses. A separate study of breast cancer patients identified changes in
152 the peripheral blood immune cell profile between breast patients with or without lymph node
153 metastasis [18].

154

155 Indication of a systemic T cell response against cancer is evident from studies where T cells
156 from peripheral blood of cancer patients can destroy tumor cells *in vitro* [19]. In addition, the
157 presence of MDSCs in peripheral blood is associated with increased disease burden and
158 negative prognosis in multiple solid tumors, presumably by infiltrating tumor sites, pre-
159 metastatic niches and with their potential to inhibit circulating T cells [20, 21]. In ovarian
160 cancer patients, the diversity in T cell receptor (TCR) clones within the circulation decreased
161 during disease progression [22]. The diversity of peripheral TCR clones is decreased in lung
162 and ovarian cancer patients compared to healthy controls, indicating that the presence of
163 negatively impacts on T cells systemically[22, 23]. Intriguingly, during treatment with
164 chemotherapy, the circulating T cell receptor (TCR) clonality is altered in various cancers,
165 indicating a potential enhanced immune response that can be identified systemically in these
166 cases [24, 25]. In breast cancer patients, increased diversity of the circulating TCR repertoire
167 of CD8⁺ T cells was associated with increased response to chemotherapy[25].Furthermore, a
168 study in melanoma found that the combinatorial diversity evenness in the TCR repertoire
169 pretreatment can be predictive of checkpoint blockade efficacy [26].

170

171 A recent study investigated systemic immune responses in experimental mouse models.
172 Spitzer *et al.* found multiple changes within lymphoid tissue and within the circulation of
173 mice receiving effective immunotherapies during the priming and rejection phase of tumors
174 [27]. In peripheral blood, there were increases in NK cells, DCs, activated B cells, naïve
175 CD4⁺ T cells, CD90^{hi} T-helper cells and CD8⁺ T cells expressing Ly6C and CD44, observed
176 during the priming phase. During the rejection phase, there were substantial decreases in
177 immune cell frequencies in the circulation likely denoting migration of immune cells into the
178 tissues. However, many cell populations in the circulation at this time retained expression of
179 Ki67, indicating potential ongoing expansion and maintenance of these cells.

180

181 Thus, changes in the TME and immunity against a tumor is not just confined to the tumor,
182 and/or the tumor-draining lymph nodes, but can cause systemic immune dysfunction. In this
183 light, a study of osteosarcoma found that T cells in the spleen of tumor bearing mice showed
184 an exhaustion status [28]. In addition, there was an increase in the percent populations of
185 Tregs and MDSCs in the spleens of the tumor bearing mice. In this model, treatment with
186 anti-PD-L1 decreased T cell exhaustion and increased monocyte maturation. These data
187 suggest that cancer induced systemic immune dysregulation should be considered and
188 targeted during cancer treatment.

189

190 ***Seed and soil hypothesis***

191 In 1889 Stephen Paget proposed the seed and soil hypothesis, where metastatic spread
192 depends on cross-talk between the cancer cells (the ‘seed’) and the organ microenvironment
193 (the ‘soil’)[29]. This hypothesis has been built upon with clinical observations and murine
194 models of cancer in which certain tumor types tend to metastasize to specific organs
195 independent of vascularization, rate of blood flow or the number of tumor cells delivered.

196

197 Experimental data to support this hypothesis first came from a study of B16 murine
198 melanoma cells which grow in the lungs when injected intravenously. It had been assumed
199 that the explanation for this is trivial – that the melanoma nodules develop in the lungs
200 because they lodge in the first capillary bed they encounter. However, B16 cells also grew in
201 lung tissue, but not in renal tissue, that had been implanted intra-muscularly demonstrating
202 that the tissue microenvironment is important in determining the site of metastasis [30].
203 Comparison of B16 and K-1735 melanoma cell lines injected into the internal carotid artery
204 revealed differences in brain metastasis patterns, as they produced only meningeal or
205 parenchymal growths, respectively [31]. This indicates different regions and
206 microenvironments, even within the same organ, may support the seeding and growth of
207 different tumor cell types.

208

209 Furthermore, establishment of a ‘pre-metastatic niche’ has now been identified as a precursor
210 for metastasis. The pre-metastatic niche is defined as aberrant immune cells and extracellular
211 matrix that initiate the transformation of a healthy microenvironment to one supportive of
212 tumor cell growth. The first-described pre-metastatic niche found a role for VEGFR1⁺
213 hematopoietic progenitor cells that formed cellular clusters in pre-metastatic sites before
214 tumor cell arrival [32]. Blocking VEGFR1 function or removing these cells ablated the
215 cellular clusters and decreased tumor metastasis. Various proteins such as fibronectin,
216 collagen IV, tenascin, periostin and certain chemokines and cytokines can be aberrantly
217 expressed by tumor-influenced stromal cells in distal tissues [33-37]. EVs from pancreatic
218 cancer can establish pre-metastatic niches in the liver and promote liver metastasis [38].
219 Tumor derived EVs can also be taken up by Kupffer cells of the liver, which produce TGFβ
220 and fibronectin to recruit bone marrow derived cells (BMDCs) that support metastasis [39].

221

222 Cells attracted to the pre-metastatic niche include MDSCs, neutrophils, macrophages and
223 regulatory T cells (Tregs) [33, 36, 40-45]. For example, a murine study of breast cancer
224 demonstrated that granulocytic MDSCs were important for establishing a pre-metastatic
225 niche [46]. Thus, it is well established that signals from the primary tumor can influence
226 distal tissues and impact healthy organ microenvironments. It could be expected that tumors
227 in different primary and metastatic sites can also influence each other in a similar manner.

228

229 *Abscopal effects of localized radiation*

230 A well-documented example of one tumor impacting on a distal tumor in metastatic cancer
231 patients is the abscopal effect of local radiation - in rare cases, localized radiation therapy of
232 one tumor is followed by regression of distant untreated tumor/s in an immune-dependent
233 manner. The abscopal effect has been observed in melanoma [47], renal cell [48], breast [49],
234 hepatocellular [50], non-small cell lung and thymic cancers [51]. Between 1969 and 2014,
235 there were an estimated 46 case reports of the abscopal effect from radiation therapy [52].

236

237 The involvement of the immune system in abscopal responses has been demonstrated in
238 mouse models. In a mouse fibrosarcoma model, the dose needed for 50% tumor control in T
239 cell-deficient mice was 64.5 Gray compared with 30 Gray in immunocompetent mice,
240 demonstrating the key role of the immune system [53]. In a bilateral mouse model of breast
241 cancer, irradiation in one flank along with systemic delivery of FMS-like tyrosine kinase
242 receptor 3 ligand (FLT3L) resulted in a T cell dependent delay in growth of both irradiated
243 and non-irradiated tumors [54]. Another study using the Lewis lung carcinoma model
244 observed control of lung metastases when footpad tumors were irradiated and the mice
245 received systemic Flt3L [55].

246

247 Since the initial observations of abscopal responses, combined immunotherapy and radiation
248 has increased the reports of abscopal responses. In a clinical trial combining radiotherapy
249 with GM-CSF in patients with solid cancer, abscopal responses were observed in 30% of
250 patients [51]. In melanoma, α PD-1 or α CTLA4/ α PD-L1 in combination with radiotherapy
251 also increased abscopal responses [56, 57].

252

253 Mechanistic studies on combined radio-immunotherapy have shown that cross-presentation
254 by BATF3-dependent DCs, CD8 T cells and type I interferons are required for abscopal
255 responses [58-61]. Radiation therapy is thought to enhance the release of tumor associated

256 antigens and promote local promotion of inflammatory responses, especially stimulation of
257 the innate immune system, which consequently leads to CD8⁺ T cell priming by BATF3-
258 dependent DCs and T cell circulation and homing to tumor sites [62, 63]. In summary, tumors
259 in distal sites can influence each other, in the context of radiation therapy, whereby tumor
260 destruction by radiation leads to enhanced anti-tumor immune responses that become
261 systemic.

262
263 Abscopal responses to other localized therapies have been observed when they are combined
264 with PD-1/PD-L1 inhibitors [27]. In the MMTV-PyMT mouse model, which produce
265 spontaneous tumors in multiple mammary glands, only those tumors that were directly
266 injected with a tumor-binding antibody therapy regressed. However, when combined with
267 systemic anti-PD-L1, intra-tumoral injection resulted in regression of distal tumors, even
268 though anti-PD-L1 alone did not bring about systemic immune responses. Another study,
269 which utilized vascular-targeted photodynamic therapy in primary murine Renca kidney
270 tumors, resulted in systemic immune responses against lung metastases, when administered
271 alongside anti-PD-1/PD-L1 [64].

272 273 **Cross-talk between tumors in different anatomical sites**

274 *Evidence from experimental models*

275 As discussed so far, complex interactions occur between the tumor and host, and these
276 interactions can have systemic consequences. Thus, it is reasonable to hypothesize that
277 metastatic tumors in different anatomical sites are not completely independent entities and
278 that connectivity between tumors could impact on tumor growth and response to therapy. The
279 limited number of studies that have investigated this hypothesis to date are discussed in detail
280 in the following section.

281
282 Studies into how tumors can affect each other's growth and response to immunity and
283 therapy go back many years. In 1905, a study by Paul Ehrlich found that engrafted tumors
284 exhibit a negative effect on each other. In this study, earlier implanted tumors inhibited the
285 growth of tumors injected later and the inhibitory effect increased with the size of the first
286 tumor [65]. Studies in anti-angiogenic therapies have observed a decrease in the growth of
287 metastatic tumors by angiogenesis inhibitors, only when the primary tumor is present. The
288 serum and urine of tumor bearing mice contained angiostatin which inhibits endothelial cell

289 proliferation and was determined to be a systemic mediator of the effects in metastatic tumors
290 in this study [66].

291

292 Another study used mathematical modelling to investigate dynamic behavior and interactions
293 between tumor and immune cells at various metastatic sites, with or without therapy [67].
294 Simulation of T cell recruitment in lung and breast tumors was assessed in isolation, and
295 upon metastatic seeding after 200 days, aimed at analyzing T cell redistribution and immune
296 cell population dynamics between the two sites, and to model the effect of surgery or
297 radiation. This mathematical modelling revealed that a proportion of the T cells activated in
298 the breast were recruited to the metastatic site. A larger fraction of circulating immune cells
299 traffic through the pulmonary circulation and therefore have a greater chance of extravasation
300 into the lung. The authors hypothesized that this would result in increased primary tumor
301 growth, given the relocation of tumor reactive T cells to the lungs and estimated a 30%
302 reduction in lung tumor size when a secondary breast tumor is seeded. Thus, this study
303 provides a simulation and mathematical model for interconnectivity between existing tumor
304 sites, which is based on assumptions from redistribution of T cells and systemic immunity in
305 various tumor studies and other mathematical models of tumor growth.

306

307 A study by our laboratory was the first to demonstrate the influence of one tumor on
308 immunotherapy responses to tumor located in a distant anatomical site [68]. Using the Renca
309 kidney cancer model, in which subcutaneous (SC) tumors were responsive and orthotopic
310 kidney tumors resistant to a triple antibody (trimAb) therapy, this study assessed therapy
311 responses in mice simultaneously implanted with SC and kidney tumors. When mice had
312 only contralateral SC tumors, responses to trimAb therapy were sustained. However, when
313 mice had one SC tumor and one kidney tumor, growth of the SC tumor was accelerated with
314 therapy, indicating that the Renca kidney tumor was inhibiting the response of the SC tumor.

315

316 In exploring a possible mechanism, we determined that M2 macrophages, which are
317 commonly found in Renca kidney tumors increased in number in SC tumors when a kidney
318 tumor was also present. The increase in M2 macrophages was accompanied by a decrease in
319 T and NK cell numbers and cytotoxicity within the SC tumors. Furthermore, CCL2 blockade
320 or systemic macrophage depletion using Clodrolip improved therapy responses in SC tumors
321 when kidney tumors were present. As there were no soluble factors identified to mediate the
322 suppressive effects, we hypothesized that M2 macrophages generated within the suppressive

323 kidney tumor could traffic to the SC tumor, where they suppressed immune responses and
324 decreased the therapeutic response.

325

326 A more recent study investigated the efficacy of dual checkpoint blockade (α PD-1/ α CTLA4)
327 on melanoma brain metastases [69]. Taggart *et al* found that an extracranial tumor (SC, in
328 their model) had to be present for α PD-1/ α CTLA4 to be efficacious against intra-cranial
329 tumors. Mice implanted only with intra-cranial B16 melanoma cells responded poorly to
330 α PD-1/ α CTLA4, whereas those bearing both intra-cranial and SC tumors had enhanced
331 survival and decreased growth of both extra- and intra- cranial tumors. When an extracranial
332 tumor was present, intra-cranial tumors had enhanced infiltration of immune cells and
333 particularly, CD8⁺ T cells, bone marrow-derived macrophages and brain microglia. The
334 beneficial effect of an extracranial tumor was abolished when mice were depleted of CD8⁺ T
335 cells and NK cells. Furthermore, CD44⁺CD62L⁻ effector CD8⁺ T cells were increased in the
336 peripheral blood of mice with extracranial disease and transfer of these cells from spleens of
337 α PD-1/ α CTLA4 treated mice demonstrated enhanced homing into intracranial tumors
338 compared with the same cell population from spleens of treatment-naive mice. The authors
339 also demonstrated that this enhanced homing may be due to enhanced T cell trafficking
340 determinants on intracranial blood vessels when extracranial tumors were present.

341

342 Interestingly, further evidence in support of immune cross-talk between tumors can be found
343 in studies utilizing neoadjuvant therapy [70]. In this approach, mice bearing both primary
344 breast tumors and lung metastases were treated with immunotherapy prior to surgical removal
345 of the primary. Mice undergoing neoadjuvant therapy survived longer, with decreased
346 primary recurrence and progression of lung metastases, compared with mice undergoing
347 surgery followed by adjuvant immunotherapy. The presence of the primary tumor during
348 immunotherapy enabled a more pronounced tumor-specific T cell response that subsequently
349 impacted on tumor relapse and metastatic formation.

350

351 ***Evidence from human cancers***

352 There is increasing evidence for tumors in different metastatic sites influencing each other in
353 human cancers. Evaluation of 371 patients with metastatic melanoma treated with high-dose
354 IL-2 revealed that cutaneous and subcutaneous melanoma metastases were more responsive
355 than visceral metastases [71]. Interestingly, patients with both cutaneous/subcutaneous and
356 visceral metastases had an overall response rate of 17%, whereas patients with only

357 cutaneous/subcutaneous metastases had on overall response rate of 50%, supporting the
358 notion of visceral metastases having a negative impact on therapy response.

359

360 Data presented at the 2017 American Society of Clinical Oncology meeting reported that
361 metastasis to the liver in melanoma patients was associated with reduced α PD-1 response
362 [72]. The authors recapitulated these findings in the B16 melanoma mouse model, in that
363 mice with liver and SC tumors were unable to achieve SC tumor clearance upon treatment
364 with α PD-1, whereas 30% of mice with SC tumors alone achieved tumor rejection.
365 Interestingly, the presence of liver metastases from an unrelated MC38 tumor line did not
366 impact on SC tumor rejection, suggesting a tumor- or antigen-specific process. Although
367 neither study controlled for the possibility that visceral/liver metastases signify more
368 aggressive or progressive disease, these studies suggest that in humans, metastatic tumors
369 situated in specific organs can have an influence on the overall response to therapy.

370

371 Recent results from human trials of immune checkpoint blockade in the neoadjuvant setting
372 support the observations in mice described above [73, 74]. Patients with melanoma or
373 glioblastoma undergoing neoadjuvant checkpoint blockade had better outcomes than patients
374 receiving adjuvant checkpoint blockade, with increased time to progression and increased
375 overall survival. Enhanced systemic immune responses were observed in some patients
376 receiving neoadjuvant therapy, supporting the interpretation that, despite a larger tumor
377 burden, the presence of the primary tumor during checkpoint blockade led to an increased T
378 cell response against the remaining unresectable malignant disease.

379

380 ***Concluding remarks***

381 Given the complexity of cancers, with metastases to various organs, and the ability of tissues
382 to communicate across very distinct anatomical sites, it is not surprising that tumors might
383 also affect each other's progression and responses to therapy. Tumors at different sites can
384 differ in their immune composition, and the immune system is made up of a diverse variety
385 of motile cells and secreted molecules that have the potential to move throughout the body. In
386 addition, other cell types within tumors, including malignant cells, can also produce a variety
387 of molecules and particles, many of which presumably remain to be identified, that may
388 influence other tissues and tumors remotely.

389

390 In this review, we have described studies supporting the concept of cross-talk between
391 tumors, which can impact on the growth of metastases in various sites in the body and their
392 response to various therapies, including immunotherapy (**Figure 1**). In some instances, cross-
393 talk can clearly lead to a reduced response to therapy at certain sites, but there are also
394 instances where tumors can become more responsive to therapy. With further advancement of
395 this field, it may be possible to identify tumors that, based on their histological origin and
396 anatomical site, might have a positive or a negative impact on the response to therapy. In at
397 least some limited instances, it may be feasible to consider excising tumors at certain specific
398 sites, or subjecting them to localized radiotherapy, in order to enhance the patient's overall
399 response and prolong survival.

400

401 **Figure Legend**

402

403 **Figure 1: Mechanisms of cross-talk of tumors with distal tumors and tissues.** The
404 presence of a tumor in one location results in systemic changes and can impact on distal
405 tumors in other locations. **A)** Tumor-derived EVs and tumor-secreted proteins from
406 malignant and non-malignant cells within the tumor microenvironment (TME), along with
407 cells from the TME, can be released into the circulatory and lymphatic systems. **B)** The
408 consequent changes in the molecular and cellular composition systemically have been
409 detected in cancer patients compared to healthy individuals, and patients before, during and
410 after treatment with various anti-cancer therapies. This highlights that events happening
411 within the TME have an impact on the entire host. **C)** and **D)** Distant healthy tissues are
412 impacted by secreted factors from the primary tumor which can disrupt the extracellular
413 matrix and promote local immunosuppression in order to create the pre-metastatic niche (C)
414 and impact on the composition and production of immune cells within lymphoid organs (D).
415 For example, GM-CSF secreted by tumor cells can increase production of myeloid derived
416 suppressor cells (MDSCs) from the bone marrow. **E)** The impact of one tumor on other distal
417 tumors has recently been investigated in a limited number of studies. It seems that tumors can
418 exhibit both positive and negative effects on each other, especially in the context of
419 immunotherapy responses. For example, murine studies indicate that the presence of a
420 primary tumor can enhance the generation of tumor specific T cell responses in metastatic
421 tumors upon treatment with immunotherapies. Other studies suggest that immunosuppressive
422 cells, such as M2 macrophages, that are generated in one tumor can traffic to, infiltrate and
423 exhibit local immunosuppression in distal tumors thus dampening immunotherapy responses.

424

425 **References**

- 426 1. Mole, R. H. (1953) Whole body irradiation; radiobiology or medicine?, *Br J Radiol.* **26**, 234-41.
- 427 2. Abukhiran, I. A., Jasser, J. & Syrbu, S. (2020) Paraneoplastic leukemoid reactions induced by
428 cytokine-secreting tumours, *J Clin Pathol.*
- 429 3. Bronte, V., Chappell, D. B., Apolloni, E., Cabrelle, A., Wang, M., Hwu, P. & Restifo, N. P. (1999)
430 Unopposed production of granulocyte-macrophage colony-stimulating factor by tumors inhibits
431 CD8+ T cell responses by dysregulating antigen-presenting cell maturation, *J Immunol.* **162**, 5728-37.
- 432 4. Toi, M., Kondo, S., Suzuki, H., Yamamoto, Y., Inada, K., Imazawa, T., Taniguchi, T. & Tominaga, T.
433 (1996) Quantitative analysis of vascular endothelial growth factor in primary breast cancer, *Cancer.*
434 **77**, 1101-6.
- 435 5. Gabrilovich, D. I., Chen, H. L., Girgis, K. R., Cunningham, H. T., Meny, G. M., Nadaf, S., Kavanaugh,
436 D. & Carbone, D. P. (1996) Production of vascular endothelial growth factor by human tumors
437 inhibits the functional maturation of dendritic cells, *Nat Med.* **2**, 1096-103.
- 438 6. Wu, C. W., Chi, C. W., Lin, E. C., Lui, W. Y., P'Eng F, K. & Wang, S. R. (1994) Serum arginase level in
439 patients with gastric cancer, *J Clin Gastroenterol.* **18**, 84-5.
- 440 7. Duffy, M. J. (2001) Clinical uses of tumor markers: a critical review, *Crit Rev Clin Lab Sci.* **38**, 225-
441 62.
- 442 8. Raposo, G. & Stoorvogel, W. (2013) Extracellular vesicles: exosomes, microvesicles, and friends, *J*
443 *Cell Biol.* **200**, 373-83.
- 444 9. Tung, K. H., Ernstoff, M. S., Allen, C. & Shu, S. (2019) A Review of Exosomes and their Role in The
445 Tumor Microenvironment and Host-Tumor "Macroenvironment", *J Immunol Sci.* **3**, 4-8.
- 446 10. Chen, G., Huang, A. C., Zhang, W., Zhang, G., Wu, M., Xu, W., Yu, Z., Yang, J., Wang, B., Sun, H.,
447 Xia, H., Man, Q., Zhong, W., Antelo, L. F., Wu, B., Xiong, X., Liu, X., Guan, L., Li, T., Liu, S., Yang, R., Lu,
448 Y., Dong, L., McGettigan, S., Somasundaram, R., Radhakrishnan, R., Mills, G., Lu, Y., Kim, J., Chen, Y.
449 H., Dong, H., Zhao, Y., Karakousis, G. C., Mitchell, T. C., Schuchter, L. M., Herlyn, M., Wherry, E. J., Xu,
450 X. & Guo, W. (2018) Exosomal PD-L1 contributes to immunosuppression and is associated with anti-
451 PD-1 response, *Nature.* **560**, 382-386.
- 452 11. Shu, S., Yang, Y., Allen, C. L., Maguire, O., Minderman, H., Sen, A., Ciesielski, M. J., Collins, K. A.,
453 Bush, P. J., Singh, P., Wang, X., Morgan, M., Qu, J., Bankert, R. B., Whiteside, T. L., Wu, Y. & Ernstoff,
454 M. S. (2018) Metabolic reprogramming of stromal fibroblasts by melanoma exosome microRNA
455 favours a pre-metastatic microenvironment, *Sci Rep.* **8**, 12905.

- 456 12. Asgharzadeh, M. R., Barar, J., Pourseif, M. M., Eskandani, M., Jafari Niya, M., Mashayekhi, M. R.
457 & Omid, Y. (2017) Molecular machineries of pH dysregulation in tumor microenvironment: potential
458 targets for cancer therapy, *Bioimpacts*. **7**, 115-133.
- 459 13. Bellone, M., Calcinotto, A., Filipazzi, P., De Milito, A., Fais, S. & Rivoltini, L. (2013) The acidity of
460 the tumor microenvironment is a mechanism of immune escape that can be overcome by proton
461 pump inhibitors, *Oncoimmunology*. **2**, e22058.
- 462 14. Zhang, H., Deng, T., Liu, R., Bai, M., Zhou, L., Wang, X., Li, S., Wang, X., Yang, H., Li, J., Ning, T.,
463 Huang, D., Li, H., Zhang, L., Ying, G. & Ba, Y. (2017) Exosome-delivered EGFR regulates liver
464 microenvironment to promote gastric cancer liver metastasis, *Nat Commun*. **8**, 15016.
- 465 15. Peinado, H., Aleckovic, M., Lavotshkin, S., Matei, I., Costa-Silva, B., Moreno-Bueno, G., Hergueta-
466 Redondo, M., Williams, C., Garcia-Santos, G., Ghajar, C., Nitadori-Hoshino, A., Hoffman, C., Badal, K.,
467 Garcia, B. A., Callahan, M. K., Yuan, J., Martins, V. R., Skog, J., Kaplan, R. N., Brady, M. S., Wolchok, J.
468 D., Chapman, P. B., Kang, Y., Bromberg, J. & Lyden, D. (2012) Melanoma exosomes educate bone
469 marrow progenitor cells toward a pro-metastatic phenotype through MET, *Nat Med*. **18**, 883-91.
- 470 16. Domenis, R., Cesselli, D., Toffoletto, B., Bourkoura, E., Caponnetto, F., Manini, I., Beltrami, A. P.,
471 Ius, T., Skrap, M., Di Loreto, C. & Gri, G. (2017) Systemic T Cells Immunosuppression of Glioma Stem
472 Cell-Derived Exosomes Is Mediated by Monocytic Myeloid-Derived Suppressor Cells, *PLoS One*. **12**,
473 e0169932.
- 474 17. Dumeaux, V., Fjukstad, B., Fjosne, H. E., Frantzen, J. O., Holmen, M. M., Rodegerdts, E.,
475 Schlichting, E., Borresen-Dale, A. L., Bongo, L. A., Lund, E. & Hallett, M. (2017) Interactions between
476 the tumor and the blood systemic response of breast cancer patients, *PLoS Comput Biol*. **13**,
477 e1005680.
- 478 18. Zuckerman, N. S., Yu, H., Simons, D. L., Bhattacharya, N., Carcamo-Cavazos, V., Yan, N., Dirbas, F.
479 M., Johnson, D. L., Schwartz, E. J. & Lee, P. P. (2013) Altered local and systemic immune profiles
480 underlie lymph node metastasis in breast cancer patients, *Int J Cancer*. **132**, 2537-47.
- 481 19. Slingluff, C. L., Jr., Cox, A. L., Stover, J. M., Jr., Moore, M. M., Hunt, D. F. & Engelhard, V. H. (1994)
482 Cytotoxic T-lymphocyte response to autologous human squamous cell cancer of the lung: epitope
483 reconstitution with peptides extracted from HLA-Aw68, *Cancer Res*. **54**, 2731-7.
- 484 20. Veglia, F., Perego, M. & Gabrilovich, D. (2018) Myeloid-derived suppressor cells coming of age,
485 *Nat Immunol*. **19**, 108-119.
- 486 21. Zhang, S., Ma, X., Zhu, C., Liu, L., Wang, G. & Yuan, X. (2016) The Role of Myeloid-Derived
487 Suppressor Cells in Patients with Solid Tumors: A Meta-Analysis, *PLoS One*. **11**, e0164514.

- 488 22. Cui, J. H., Lin, K. R., Yuan, S. H., Jin, Y. B., Chen, X. P., Su, X. K., Jiang, J., Pan, Y. M., Mao, S. L.,
489 Mao, X. F. & Luo, W. (2018) TCR Repertoire as a Novel Indicator for Immune Monitoring and
490 Prognosis Assessment of Patients With Cervical Cancer, *Front Immunol.* **9**, 2729.
- 491 23. Liang, N., Chen, S., Yi, Y., Guan, Y.-F., Xia, X. & Yi, X. (2019) Characteristics of T-cell receptor
492 repertoire between lung cancer patients and healthy people, *Journal of Clinical Oncology.* **37**,
493 e20728-e20728.
- 494 24. Zhang, C., Palashati, H., Tan, Q., Ku, W., Miao, Y., Xiong, H. & Lu, Z. (2018) Immediate and
495 substantial evolution of T-cell repertoire in peripheral blood and tumor microenvironment of
496 patients with esophageal squamous cell carcinoma treated with preoperative chemotherapy,
497 *Carcinogenesis.* **39**, 1389-1398.
- 498 25. Lin, K. R., Pang, D. M., Jin, Y. B., Hu, Q., Pan, Y. M., Cui, J. H., Chen, X. P., Lin, Y. X., Mao, X. F.,
499 Duan, H. B. & Luo, W. (2018) Circulating CD8(+) T-cell repertoires reveal the biological characteristics
500 of tumors and clinical responses to chemotherapy in breast cancer patients, *Cancer Immunol*
501 *Immunother.* **67**, 1743-1752.
- 502 26. Hogan, S. A., Courtier, A., Cheng, P. F., Jaberg-Bentele, N. F., Goldinger, S. M., Manuel, M., Perez,
503 S., Plantier, N., Mouret, J. F., Nguyen-Kim, T. D. L., Raaijmakers, M. I. G., Kvistborg, P., Pasqual, N.,
504 Haanen, J., Dummer, R. & Levesque, M. P. (2019) Peripheral Blood TCR Repertoire Profiling May
505 Facilitate Patient Stratification for Immunotherapy against Melanoma, *Cancer Immunol Res.* **7**, 77-
506 85.
- 507 27. Spitzer, M. H., Carmi, Y., Reticker-Flynn, N. E., Kwek, S. S., Madhireddy, D., Martins, M. M.,
508 Gherardini, P. F., Prestwood, T. R., Chabon, J., Bendall, S. C., Fong, L., Nolan, G. P. & Engleman, E. G.
509 (2017) Systemic Immunity Is Required for Effective Cancer Immunotherapy, *Cell.* **168**, 487-502 e15.
- 510 28. Markel, J. E., Noore, J., Emery, E. J., Bobnar, H. J., Kleinerman, E. S. & Lindsey, B. A. (2018) Using
511 the Spleen as an In Vivo Systemic Immune Barometer Alongside Osteosarcoma Disease Progression
512 and Immunotherapy with alpha-PD-L1, *Sarcoma.* **2018**, 8694397.
- 513 29. Paget, S. (1989) The distribution of secondary growths in cancer of the breast. 1889, *Cancer*
514 *Metastasis Rev.* **8**, 98-101.
- 515 30. Hart, I. R. & Fidler, I. J. (1980) Role of organ selectivity in the determination of metastatic
516 patterns of B16 melanoma, *Cancer Res.* **40**, 2281-7.
- 517 31. Schackert, G. & Fidler, I. J. (1988) Site-specific metastasis of mouse melanomas and a
518 fibrosarcoma in the brain or meninges of syngeneic animals, *Cancer Res.* **48**, 3478-84.
- 519 32. Kaplan, R. N., Riba, R. D., Zacharoulis, S., Bramley, A. H., Vincent, L., Costa, C., MacDonald, D. D.,
520 Jin, D. K., Shido, K., Kerns, S. A., Zhu, Z., Hicklin, D., Wu, Y., Port, J. L., Altorki, N., Port, E. R., Ruggero,

521 D., Shmelkov, S. V., Jensen, K. K., Rafii, S. & Lyden, D. (2005) VEGFR1-positive haematopoietic bone
522 marrow progenitors initiate the pre-metastatic niche, *Nature*. **438**, 820-7.

523 33. Liu, Y., Gu, Y., Han, Y., Zhang, Q., Jiang, Z., Zhang, X., Huang, B., Xu, X., Zheng, J. & Cao, X. (2016)
524 Tumor Exosomal RNAs Promote Lung Pre-metastatic Niche Formation by Activating Alveolar
525 Epithelial TLR3 to Recruit Neutrophils, *Cancer Cell*. **30**, 243-256.

526 34. Oskarsson, T. (2013) Extracellular matrix components in breast cancer progression and
527 metastasis, *Breast*. **22 Suppl 2**, S66-72.

528 35. Barkan, D., Green, J. E. & Chambers, A. F. (2010) Extracellular matrix: a gatekeeper in the
529 transition from dormancy to metastatic growth, *Eur J Cancer*. **46**, 1181-8.

530 36. Erler, J. T., Bennewith, K. L., Cox, T. R., Lang, G., Bird, D., Koong, A., Le, Q. T. & Giaccia, A. J.
531 (2009) Hypoxia-induced lysyl oxidase is a critical mediator of bone marrow cell recruitment to form
532 the premetastatic niche, *Cancer Cell*. **15**, 35-44.

533 37. Hiratsuka, S., Watanabe, A., Aburatani, H. & Maru, Y. (2006) Tumour-mediated upregulation of
534 chemoattractants and recruitment of myeloid cells predetermines lung metastasis, *Nat Cell Biol*. **8**,
535 1369-75.

536 38. Costa-Silva, B., Aiello, N. M., Ocean, A. J., Singh, S., Zhang, H., Thakur, B. K., Becker, A., Hoshino,
537 A., Mark, M. T., Molina, H., Xiang, J., Zhang, T., Theilen, T. M., Garcia-Santos, G., Williams, C., Ararso,
538 Y., Huang, Y., Rodrigues, G., Shen, T. L., Labori, K. J., Lothe, I. M., Kure, E. H., Hernandez, J., Dousot,
539 A., Ebbesen, S. H., Grandgenett, P. M., Hollingsworth, M. A., Jain, M., Mallya, K., Batra, S. K.,
540 Jarnagin, W. R., Schwartz, R. E., Matei, I., Peinado, H., Stanger, B. Z., Bromberg, J. & Lyden, D. (2015)
541 Pancreatic cancer exosomes initiate pre-metastatic niche formation in the liver, *Nat Cell Biol*. **17**,
542 816-26.

543 39. Hoshino, A., Costa-Silva, B., Shen, T. L., Rodrigues, G., Hashimoto, A., Tesic Mark, M., Molina, H.,
544 Kohsaka, S., Di Giannatale, A., Ceder, S., Singh, S., Williams, C., Sopol, N., Uryu, K., Pharmed, L., King,
545 T., Bojmar, L., Davies, A. E., Ararso, Y., Zhang, T., Zhang, H., Hernandez, J., Weiss, J. M., Dumont-Cole,
546 V. D., Kramer, K., Wexler, L. H., Narendran, A., Schwartz, G. K., Healey, J. H., Sandstrom, P., Labori, K.
547 J., Kure, E. H., Grandgenett, P. M., Hollingsworth, M. A., de Sousa, M., Kaur, S., Jain, M., Mallya, K.,
548 Batra, S. K., Jarnagin, W. R., Brady, M. S., Fodstad, O., Muller, V., Pantel, K., Minn, A. J., Bissell, M. J.,
549 Garcia, B. A., Kang, Y., Rajasekhar, V. K., Ghajar, C. M., Matei, I., Peinado, H., Bromberg, J. & Lyden,
550 D. (2015) Tumour exosome integrins determine organotropic metastasis, *Nature*. **527**, 329-35.

551 40. Giles, A. J., Reid, C. M., Evans, J. D., Murgai, M., Vicioso, Y., Highfill, S. L., Kasai, M., Vahdat, L.,
552 Mackall, C. L., Lyden, D., Wexler, L. & Kaplan, R. N. (2016) Activation of Hematopoietic
553 Stem/Progenitor Cells Promotes Immunosuppression Within the Pre-metastatic Niche, *Cancer Res*.
554 **76**, 1335-47.

- 555 41. Chafe, S. C., Lou, Y., Sceneay, J., Vallejo, M., Hamilton, M. J., McDonald, P. C., Bennewith, K. L.,
556 Moller, A. & Dedhar, S. (2015) Carbonic anhydrase IX promotes myeloid-derived suppressor cell
557 mobilization and establishment of a metastatic niche by stimulating G-CSF production, *Cancer Res.*
558 **75**, 996-1008.
- 559 42. Sceneay, J., Chow, M. T., Chen, A., Halse, H. M., Wong, C. S., Andrews, D. M., Sloan, E. K., Parker,
560 B. S., Bowtell, D. D., Smyth, M. J. & Moller, A. (2012) Primary tumor hypoxia recruits
561 CD11b+/Ly6C^{med}/Ly6G⁺ immune suppressor cells and compromises NK cell cytotoxicity in the
562 premetastatic niche, *Cancer Res.* **72**, 3906-11.
- 563 43. Qian, B. Z., Li, J., Zhang, H., Kitamura, T., Zhang, J., Campion, L. R., Kaiser, E. A., Snyder, L. A. &
564 Pollard, J. W. (2011) CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis,
565 *Nature.* **475**, 222-5.
- 566 44. Kowanetz, M., Wu, X., Lee, J., Tan, M., Hagenbeek, T., Qu, X., Yu, L., Ross, J., Korsisaari, N., Cao,
567 T., Bou-Reslan, H., Kallop, D., Weimer, R., Ludlam, M. J., Kaminker, J. S., Modrusan, Z., van Bruggen,
568 N., Peale, F. V., Carano, R., Meng, Y. G. & Ferrara, N. (2010) Granulocyte-colony stimulating factor
569 promotes lung metastasis through mobilization of Ly6G+Ly6C+ granulocytes, *Proc Natl Acad Sci U S*
570 *A.* **107**, 21248-55.
- 571 45. Olkhanud, P. B., Baatar, D., Bodogai, M., Hakim, F., Gress, R., Anderson, R. L., Deng, J., Xu, M.,
572 Briest, S. & Biragyn, A. (2009) Breast cancer lung metastasis requires expression of chemokine
573 receptor CCR4 and regulatory T cells, *Cancer Res.* **69**, 5996-6004.
- 574 46. Ouzounova, M., Lee, E., Piranlioglu, R., El Andaloussi, A., Kolhe, R., Demirci, M. F., Marasco, D.,
575 Asm, I., Chadli, A., Hassan, K. A., Thangaraju, M., Zhou, G., Arbab, A. S., Cowell, J. K. & Korkaya, H.
576 (2017) Monocytic and granulocytic myeloid derived suppressor cells differentially regulate
577 spatiotemporal tumour plasticity during metastatic cascade, *Nat Commun.* **8**, 14979.
- 578 47. Postow, M. A., Callahan, M. K., Barker, C. A., Yamada, Y., Yuan, J., Kitano, S., Mu, Z., Rasalan, T.,
579 Adamow, M., Ritter, E., Sedrak, C., Jungbluth, A. A., Chua, R., Yang, A. S., Roman, R. A., Rosner, S.,
580 Benson, B., Allison, J. P., Lesokhin, A. M., Gnjatic, S. & Wolchok, J. D. (2012) Immunologic correlates
581 of the abscopal effect in a patient with melanoma, *N Engl J Med.* **366**, 925-31.
- 582 48. Wersall, P. J., Blomgren, H., Pisa, P., Lax, I., Kalkner, K. M. & Svedman, C. (2006) Regression of
583 non-irradiated metastases after extracranial stereotactic radiotherapy in metastatic renal cell
584 carcinoma, *Acta Oncol.* **45**, 493-7.
- 585 49. Hu, Z. I., McArthur, H. L. & Ho, A. Y. (2017) The Abscopal Effect of Radiation Therapy: What Is It
586 and How Can We Use It in Breast Cancer?, *Curr Breast Cancer Rep.* **9**, 45-51.

- 587 50. Ohba, K., Omagari, K., Nakamura, T., Ikuno, N., Saeki, S., Matsuo, I., Kinoshita, H., Masuda, J.,
588 Hazama, H., Sakamoto, I. & Kohno, S. (1998) Abscopal regression of hepatocellular carcinoma after
589 radiotherapy for bone metastasis, *Gut*. **43**, 575-7.
- 590 51. Golden, E. B., Chhabra, A., Chachoua, A., Adams, S., Donach, M., Fenton-Kerimian, M., Friedman,
591 K., Ponzio, F., Babb, J. S., Goldberg, J., Demaria, S. & Formenti, S. C. (2015) Local radiotherapy and
592 granulocyte-macrophage colony-stimulating factor to generate abscopal responses in patients with
593 metastatic solid tumours: a proof-of-principle trial, *Lancet Oncol*. **16**, 795-803.
- 594 52. Abuodeh, Y., Venkat, P. & Kim, S. (2016) Systematic review of case reports on the abscopal
595 effect, *Curr Probl Cancer*. **40**, 25-37.
- 596 53. Stone, H. B., Peters, L. J. & Milas, L. (1979) Effect of host immune capability on radiocurability
597 and subsequent transplantability of a murine fibrosarcoma, *J Natl Cancer Inst*. **63**, 1229-35.
- 598 54. Demaria, S., Ng, B., Devitt, M. L., Babb, J. S., Kawashima, N., Liebes, L. & Formenti, S. C. (2004)
599 Ionizing radiation inhibition of distant untreated tumors (abscopal effect) is immune mediated, *Int J*
600 *Radiat Oncol Biol Phys*. **58**, 862-70.
- 601 55. Chakravarty, P. K., Alfieri, A., Thomas, E. K., Beri, V., Tanaka, K. E., Vikram, B. & Guha, C. (1999)
602 Flt3-ligand administration after radiation therapy prolongs survival in a murine model of metastatic
603 lung cancer, *Cancer Res*. **59**, 6028-32.
- 604 56. Ribeiro Gomes, J., Schmerling, R. A., Haddad, C. K., Racy, D. J., Ferrigno, R., Gil, E., Zanuncio, P. &
605 Buzaid, A. C. (2016) Analysis of the Abscopal Effect With Anti-PD1 Therapy in Patients With
606 Metastatic Solid Tumors, *J Immunother*. **39**, 367-372.
- 607 57. Twyman-Saint Victor, C., Rech, A. J., Maity, A., Rengan, R., Pauken, K. E., Stelekati, E., Benci, J. L.,
608 Xu, B., Dada, H., Odorizzi, P. M., Herati, R. S., Mansfield, K. D., Patsch, D., Amaravadi, R. K.,
609 Schuchter, L. M., Ishwaran, H., Mick, R., Pryma, D. A., Xu, X., Feldman, M. D., Gangadhar, T. C., Hahn,
610 S. M., Wherry, E. J., Vonderheide, R. H. & Minn, A. J. (2015) Radiation and dual checkpoint blockade
611 activate non-redundant immune mechanisms in cancer, *Nature*. **520**, 373-7.
- 612 58. Grass, G. D., Krishna, N. & Kim, S. (2016) The immune mechanisms of abscopal effect in radiation
613 therapy, *Curr Probl Cancer*. **40**, 10-24.
- 614 59. Vatner, R. E., Cooper, B. T., Vanpouille-Box, C., Demaria, S. & Formenti, S. C. (2014)
615 Combinations of immunotherapy and radiation in cancer therapy, *Front Oncol*. **4**, 325.
- 616 60. Mittal, D., Gubin, M. M., Schreiber, R. D. & Smyth, M. J. (2014) New insights into cancer
617 immunoediting and its three component phases--elimination, equilibrium and escape, *Curr Opin*
618 *Immunol*. **27**, 16-25.
- 619 61. Lynch, T. J., Bondarenko, I., Luft, A., Serwatowski, P., Barlesi, F., Chacko, R., Sebastian, M., Neal,
620 J., Lu, H., Cuillerot, J. M. & Reck, M. (2012) Ipilimumab in combination with paclitaxel and

621 carboplatin as first-line treatment in stage IIIB/IV non-small-cell lung cancer: results from a
622 randomized, double-blind, multicenter phase II study, *J Clin Oncol.* **30**, 2046-54.

623 62. Rodriguez-Ruiz, M. E., Vanpouille-Box, C., Melero, I., Formenti, S. C. & Demaria, S. (2018)
624 Immunological Mechanisms Responsible for Radiation-Induced Abscopal Effect, *Trends Immunol.* **39**,
625 644-655.

626 63. Ngwa, W., Irabor, O. C., Schoenfeld, J. D., Hesser, J., Demaria, S. & Formenti, S. C. (2018) Using
627 immunotherapy to boost the abscopal effect, *Nat Rev Cancer.* **18**, 313-322.

628 64. O'Shaughnessy, M. J., Murray, K. S., La Rosa, S. P., Budhu, S., Merghoub, T., Somma, A.,
629 Monette, S., Kim, K., Corradi, R. B., Scherz, A. & Coleman, J. A. (2018) Systemic Antitumor Immunity
630 by PD-1/PD-L1 Inhibition Is Potentiated by Vascular-Targeted Photodynamic Therapy of Primary
631 Tumors, *Clin Cancer Res.* **24**, 592-599.

632 65. Ehrlich, P. & Apolant, H. (1905) *Beobachtungen über maligne Mäusetumoren*, L. Schumacher.

633 66. O'Reilly, M. S., Holmgren, L., Shing, Y., Chen, C., Rosenthal, R. A., Moses, M., Lane, W. S., Cao, Y.,
634 Sage, E. H. & Folkman, J. (1994) Angiostatin: a novel angiogenesis inhibitor that mediates the
635 suppression of metastases by a Lewis lung carcinoma, *Cell.* **79**, 315-28.

636 67. Walker, R., Poleszczuk, J., Pilon-Thomas, S., Kim, S., Anderson, A., Czerniecki, B. J., Harrison, L. B.,
637 Moros, E. G. & Enderling, H. (2018) Immune interconnectivity of anatomically distant tumors as a
638 potential mediator of systemic responses to local therapy, *Sci Rep.* **8**, 9474.

639 68. Devaud, C., John, L. B., Westwood, J. A., Yong, C. S., Beavis, P. A., Schwendener, R. A., Darcy, P.
640 K. & Kershaw, M. H. (2015) Cross-talk between tumors can affect responses to therapy,
641 *Oncoimmunology.* **4**, e975572.

642 69. Taggart, D., Andreou, T., Scott, K. J., Williams, J., Rippaus, N., Brownlie, R. J., Ilett, E. J., Salmond,
643 R. J., Melcher, A. & Loriger, M. (2018) Anti-PD-1/anti-CTLA-4 efficacy in melanoma brain metastases
644 depends on extracranial disease and augmentation of CD8(+) T cell trafficking, *Proc Natl Acad Sci U S*
645 *A.* **115**, E1540-E1549.

646 70. Liu, J., Blake, S. J., Yong, M. C., Harjunpaa, H., Ngiow, S. F., Takeda, K., Young, A., O'Donnell, J. S.,
647 Allen, S., Smyth, M. J. & Teng, M. W. (2016) Improved Efficacy of Neoadjuvant Compared to
648 Adjuvant Immunotherapy to Eradicate Metastatic Disease, *Cancer Discov.* **6**, 1382-1399.

649 71. Chang, E. & Rosenberg, S. A. (2001) Patients with melanoma metastases at cutaneous and
650 subcutaneous sites are highly susceptible to interleukin-2-based therapy, *J Immunother.* **24**, 88-90.

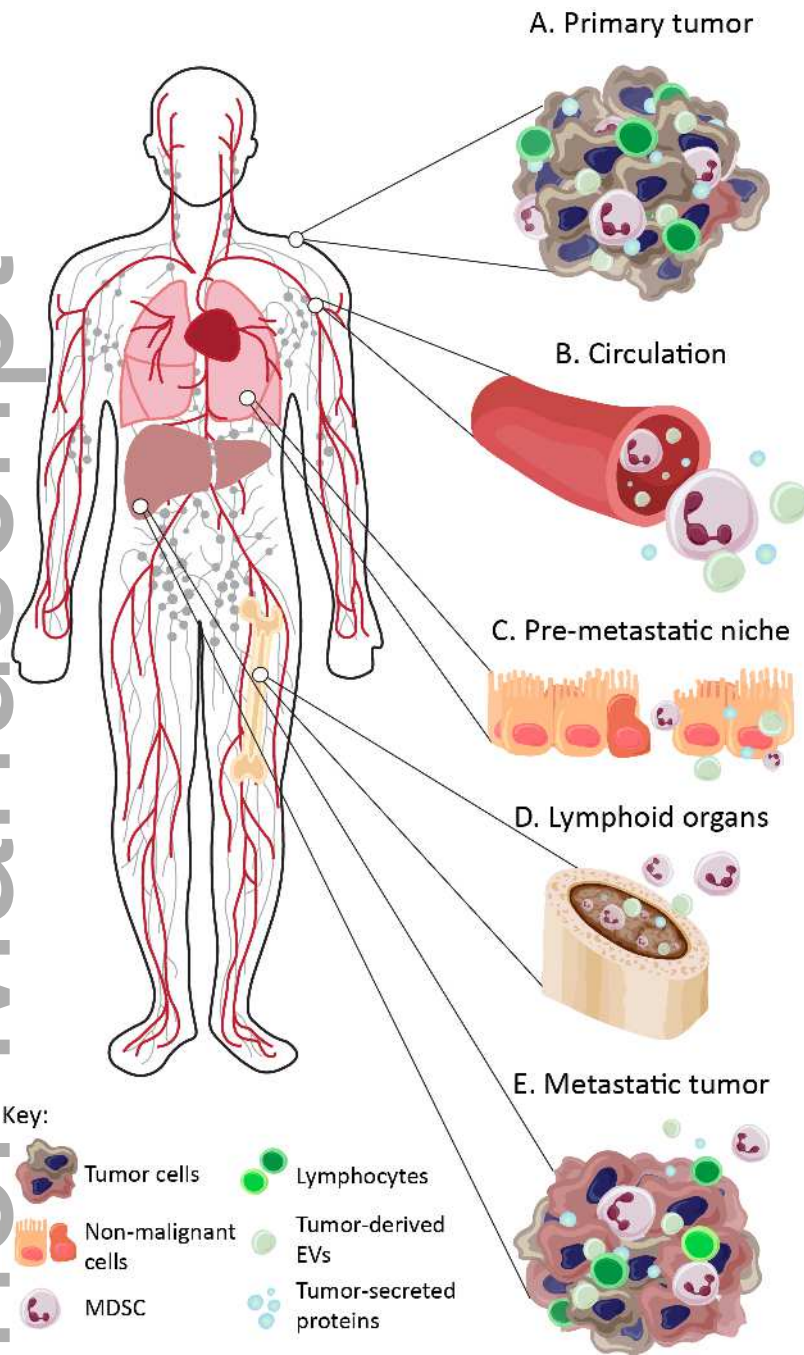
651 72. Lee, J. C.-C., Tsai, K. K., Algazi, A. P., Rosenblum, M., Bluestone, J. & Daud, A. (2017) Relationship
652 between liver metastases and PD-1 blockade in melanoma in, American Society of Clinical Oncology,

653 73. Amaria, R. N., Reddy, S. M., Tawbi, H. A., Davies, M. A., Ross, M. I., Glitza, I. C., Cormier, J. N.,
654 Lewis, C., Hwu, W. J., Hanna, E., Diab, A., Wong, M. K., Royal, R., Gross, N., Weber, R., Lai, S. Y.,

655 Ehlers, R., Blando, J., Milton, D. R., Woodman, S., Kageyama, R., Wells, D. K., Hwu, P., Patel, S. P.,
656 Lucci, A., Hessel, A., Lee, J. E., Gershenwald, J., Simpson, L., Burton, E. M., Posada, L., Haydu, L.,
657 Wang, L., Zhang, S., Lazar, A. J., Hudgens, C. W., Gopalakrishnan, V., Reuben, A., Andrews, M. C.,
658 Spencer, C. N., Prieto, V., Sharma, P., Allison, J., Tetzlaff, M. T. & Wargo, J. A. (2018) Neoadjuvant
659 immune checkpoint blockade in high-risk resectable melanoma, *Nat Med.* **24**, 1649-1654.
660 74. Cloughesy, T. F., Mochizuki, A. Y., Orpilla, J. R., Hugo, W., Lee, A. H., Davidson, T. B., Wang, A. C.,
661 Ellingson, B. M., Rytlewski, J. A., Sanders, C. M., Kawaguchi, E. S., Du, L., Li, G., Yong, W. H., Gaffey, S.
662 C., Cohen, A. L., Mellingerhoff, I. K., Lee, E. Q., Reardon, D. A., O'Brien, B. J., Butowski, N. A.,
663 Nghiemphu, P. L., Clarke, J. L., Arrillaga-Romany, I. C., Colman, H., Kaley, T. J., de Groot, J. F., Liao, L.
664 M., Wen, P. Y. & Prins, R. M. (2019) Neoadjuvant anti-PD-1 immunotherapy promotes a survival
665 benefit with intratumoral and systemic immune responses in recurrent glioblastoma, *Nat Med.* **25**,
666 477-486.

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