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Title:

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Date:

2010-10-15

Citation:

Sugawara, K., Kizaki, K., Herath, C. B., Hasegawa, Y. & Hashizume, K. (2010). Transforming growth factor beta family expression at the bovine feto-maternal interface. *Reproductive Biology and Endocrinology*, 8 (1), <https://doi.org/10.1186/1477-7827-8-120>.

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RESEARCH

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Transforming growth factor beta family expression at the bovine feto-maternal interface

Kumiko Sugawara^{2,1†}, Keiichiro Kizaki^{1†}, Chandana B Herath^{3,4}, Yoshihisa Hasegawa⁵, Kazuyoshi Hashizume^{1*}

Abstract

Background: Endometrial remodelling is necessary for implantation in all mammalian species. The TGF beta super-family plays a crucial role in this event in humans and mice. However, the role of TGF beta super-family members during implantation is still unclear in ruminants. In the present study, the spacio-temporal expression of TGF beta super-family members including activin was explored in bovine trophoblasts and endometrial tissue during the peri-implantation period in order to elucidate whether it is essential for promoting cell proliferation at the implantation site.

Methods: Gene expression in the fetal membrane and endometrium of the gravid and non-gravid horn around Day 35 of gestation were analyzed with a custom-made oligo-microarray in cattle. The expression of activin and its related genes was also analyzed with quantitative RT-PCR. Activin-like activity in trophoblastic tissue and BT-1 cells was examined using a fibroblast cell proliferation test and Western blotting.

Results: The expression of various TGF beta super-family related genes including activin was detected in trophoblasts and the endometrium in cattle. The most intensive activin expression was found in the gravid horn endometrium, and rather intense expression was detected in the non-gravid trophoblastic tissue. Extracts from the fetal membrane including trophoblasts and purified activin both stimulated fibroblast proliferation effectively, and activin was immunologically detected in BT-1 cells, which have trophoblastic features.

Conclusions: Specific expression of the activin gene (gene name: inhibin beta A) was found in the gravid horn endometrium during peri-implantation. An activin-like molecule, which was derived from the endometrium and trophoblasts, stimulated the proliferation of fibroblast cells. These results suggested that as in other species, the activity of TGF beta super-family members including activin-like molecules plays a pivotal role in endometrial remodelling, which is an essential process in implantation and placentogenesis during the peri-implantation period in cattle.

Background

Implantation involves a complex process and various kinds of factors, during which time the fetal-maternal interface develops into the placenta. Trophoblast cells and maternal endometrial cells adhere to each other and produce the feto-maternal interface, which represents the beginning of implantation and endometrial remodeling. However, the cells from the fetus and mother only fuse and develop into the placentome in a limited area, namely the caruncle (CAR) on the endometrium in

cattle [1]. Stromal cells proliferate in the CAR during peri-implantation, which corresponds to the proliferative activity of trophoblasts. In cattle, the rapid proliferation and differentiation of the early embryo and/or the conceptus begins in the gravid horn, which contains the fetus, and the conceptus grows with morphological changes such as the development of spherical, ovoid, filamentous, and elongated membranes, the latter of which then spreads to the other side of the uterine horn (non-gravid horn) [2-5]. In particular, endometrial and fetal trophoblastic cells markedly proliferate at the CAR, where the placentome develops as the feto-maternal interface including the epithelium and stroma. Various molecules derived from trophoblastic and endometrial cells including cytokines, growth factors, and hormones

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[6] may play an important role at this site in cattle, and the fusion of trophoblast and endometrial epithelial cells starts around 20 days of gestation [7-9]. Although these phenomena have been morphologically confirmed, the detailed mechanisms behind them are still unclear in ruminants. Various growth factors, such as heparin-binding epidermal growth factor (HB-EGF) and transforming growth factors (TGFs), promote crucial events during the peri-implantation period in humans and mice [10-14]. These growth factors may affect the initiation of endometrial proliferation during implantation in these species.

TGF β superfamily members including activin and some growth factors participate in this critical event in mice and humans [15-17] but little information is available about this process in ruminants. A microarray study suggested that activin (gene name: inhibin β A) coordinates endometrial remodelling in cattle [18]. In other species, there is evidence that activin is involved in endometrial remodelling [15]. Activin, which was originally identified as a peptide growth factor obtained from the ovarian follicular fluid [19], is a disulphide-linked dimeric glycoprotein composed of two subunits in various combinations, β A and β A (homodimer), β A and β B (heterodimer), and β B and β B (homodimer), which were named activin A (gene symbol: INH β A), activin AB (INH β AB), and activin B (INH β B), respectively. Both the β A and β B subunits can also be combined with the α -subunit to form inhibin-A ($\alpha\beta$ A) and -B ($\alpha\beta$ B), respectively [20]. Follistatin, which is related to activin and inhibin, and can be purified from ovarian follicular fluid, binds to activin with high affinity and neutralizes activin bioactivity [21]. Although the TGF β super-family members regulate the cell proliferation in various kinds of cells via complex mechanisms, they may stimulate endometrial cell activity and function [22,23]. In humans, activin plays a stimulatory role in decidualization and endometrial cell invasion [12,24,25].

The aims of this study are to confirm the expression of TGF β super-family members during gestation, and to explore the factors that promote cell proliferation at the implantation site and the possible role of activin, which is a TGF β super-family member, in trophoblasts and the endometrium during the peri-implantation period.

Methods

Tissue Preparation

The following tissues were used to obtain total RNA from Japanese Black cows: the fetal membrane (FM) and endometrium (END) of the gravid horn (GH) and/or the non-gravid horn (NGH) were collected separately. Some tissues such as the FM were only collected for microarray analysis, in which it was difficult to identify the cotyledonary (COT) and intercotyledonary (ICOT) sections

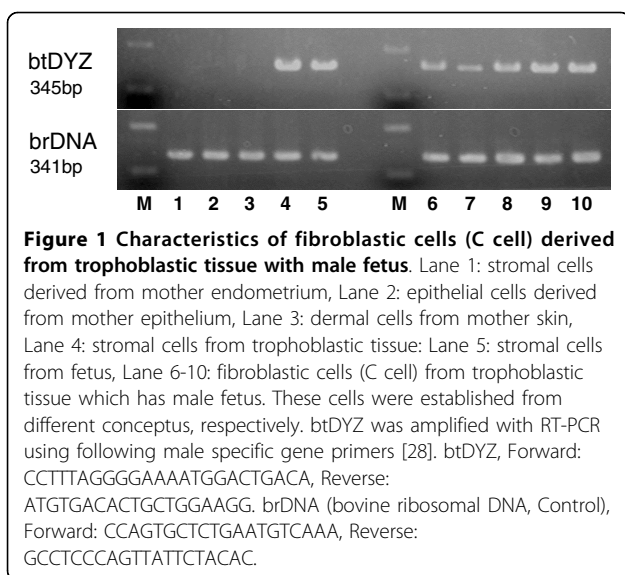
during the early period of implantation. After day 50 of gestation COT, ICOT, caruncular (CAR) and intercaruncular (ICAR) endometrium were separately collected. Endometrial tissues for gene expression analysis were collected from cows on day 7-14 of the estrous cycle ($n = 3$). The main tissue collection schedules are summarized in Table 1. The day of artificial insemination was designated as Day 0 of gestation. All tissues used for the gene, biological, and biochemical analyses were snap frozen in liquid nitrogen and stored at -80°C until RNA and protein extraction. All animal experiments and procedures were approved by the Animal Experimental Committee of Iwate University and National Institute of Agrobiological Sciences.

Bovine endometrial (E cells) or cotyledonary (C cells) fibroblast cells were derived from the endometrium and cotyledon, respectively, of Japanese Black cattle that were reared at the National Institute of Agrobiological Sciences [26,27] and used for the proliferation assay. In brief, small pieces of tissue were subjected to explant culture, and the cells that grew around the explanted tissue were collected and passaged at least three times to generate a fibroblast cell population, and then less than 10 passaged cells were used as fibroblast cells after immunohistochemical characterization with vimentin and cytokeratin, as described previously [27]. C cell was confirmed as a cell derived from cotyledonary tissue as shown in Figure 1 using Y-chromosome specific gene [28]. The bovine trophoblast cell line (BT-1), which was established in our laboratory [29], and bovine skin fibroblasts (SKIN) were prepared and used as a control in the total RNA expression and protein extraction analyses. Primary bovine SKIN cells were established from dermis tissue derived from the bovine shoulder. After all hair had been removed and the tissue had been rinsed in 70% ethanol, the tissue was washed in phosphate

Table 1 Tissue collection schedule for gene expression analysis

	Cyclic	GH	NGH	GH	NGH
a: For microarray and quantitative RT-PCR (number of animals)					
Day of gestation	END	FM	FM	END	END
35-37*	3	3	2	3	3
b: For quantitative RT-PCR (number of animals)					
Day of gestation	FM	COT	ICOT	CAR	ICAR
27-34	6	-	-	5	5
48-64		2	5	8	9
93-150		4	6	7	9
240-270		3	4	5	7

GH: gravid horn, NGH: non-gravid horn, FM: fetal membrane, END: endometrium, COT: cotyledonary fetal membrane, ICOT: intercotyledonary fetal membrane, CAR: caruncular tissue of the endometrium, ICAR: intercaruncular tissue of the endometrium. *Data was shown in text and tables as Day 35.



buffered saline (PBS) containing 100 IU/mL of penicillin and 100 µg/mL of streptomycin several times. It was then minced thoroughly with scissors and placed on plastic Petri dishes, before being carefully filled with DMEM/F-12 medium supplemented with 10% FBS. These cells were then cultured in Dulbecco's modified Eagle's medium/F-12 medium (DMEM/F-12, Sigma, Saint Louis, MI, USA) containing 100 IU/mL of penicillin and 100 µg/mL of streptomycin (Sigma), supplemented with 10% fetal bovine serum (FBS, HANA-NESCO, Tokyo, Japan), at 37°C in an atmosphere of 5% CO₂. The confluent fibroblast cells were scale passaged using 0.25% trypsin-EDTA (Sigma) and plated on flasks. The BT-1 cells were cultured according to the previously described method [30].

Protein extraction and purification

Frozen FM (Day 90 of gestation) was thawed in chilled PBS (containing penicillin and streptomycin) before being minced. After five minutes of PBS supplementation, they were centrifuged at 300 ×g for 10 minutes at 4°C, and the supernatant was removed. These steps were repeated until all blood cell factors had been removed from the supernatant. Following the addition of PBS, the minced tissues were homogenized using a polytron homogenizer (ULTRA TURRAX T25, IKA Works, NC, USA) and centrifuged at 44,000 ×g for 10 minutes at 4°C. The supernatant was then stored at -30°C until the proliferation assay.

The protein extraction from SKIN and BT-1 cells was performed using the following procedures: the SKIN cells were washed twice with serum-free medium (DMEM/F12) in close to confluent conditions and then incubated with medium (serum-free) at 37°C for two

hours. After being washed twice, the cells were scraped using a cell scraper in PBS, and the resultant cell suspension was centrifuged at 1,000 ×g for 10 minutes at room temperature. The pellet was then re-suspended in two volumes of PBS, before being homogenized and centrifuged as in the FM procedure. The supernatant was used for protein analysis. The BT-1 cells were washed three times with serum-free medium and incubated in serum-free medium at 37°C for two hours to eliminate contaminating factors from the FBS. Then, all cells were cultured in 2 mL of serum-free medium for 48 h, collected, and used for protein analysis. The conditioned medium was collected after centrifugation at 3,000 ×g for 10 minutes. The pellet was then homogenized, and the supernatant was collected as mentioned for the SKIN cell procedure. To concentrate the conditioned medium, 2 mL of medium were mixed with 8 mL of cold acetone. After being incubated overnight at -30°C, the sample was centrifuged at 10,000 ×g for 10 minutes, and the pellet was suspended in 100 µL of PBS and used for further analysis.

An activin-like substance was extracted from the BT-1 cells using sulfated Cellulofine (Seikagaku biobusiness, Tokyo, Japan), which is an affinity chromatography reagent that concentrates TGFβ related molecules [31], according to the manufacturer's instructions. In brief, after the Cellulofine extract had been washed five times with PBS, 100 µg of BT-1 extract and 100 µL of Cellulofine were mixed and incubated for 2 hours at 4°C on the rotator. The mixtures were then washed three times with PBS, and 50 µg of PBS with 1 M NaCl were added. After 1 h incubation at 4°C, they were centrifuged at 10,000 ×g for 5 min at 4°C, and the supernatants were collected.

Proliferation assay

The E and C cells were plated at 1 × 10⁵ cells/dish and incubated in DMEM/F12 medium containing 10% FBS at 37°C in an atmosphere of 5% CO₂. The cells were then washed twice with PBS and incubated in DMEM/F12 medium without FBS for 24 h at 37°C. The cells extracts (protein concentrations ranging from 1~100 µg/mL) and growth factors (as shown below) were diluted to each concentration with serum-free medium, and the cells were incubated with them for 48 h at 37°C. After the cells had been dissociated with 0.25% trypsin solution, suspended in medium, and mixed with 0.5% Trypan blue reagent (Wako Pure Chemical Industries, Osaka, Japan; 1:1), the total number of cells was counted using Thoma's hemocytometer. All assays were performed in triplicate. All values are presented as the mean ± SD.

Activin A, activin AB, and follistatin, which were purified by Y.H. [32], were used for cell proliferation analysis

with 1~1000 ng/mL. Other growth factors were purchased from commercial companies: FGF2 (0.1~100 ng/mL, WAKO 067-02831), HB-EGF (0.1~100 ng/mL, WAKO 084-08281), and TGFβ1 (0.01~10 ng/mL, WAKO 204-14291) were from Wako chemical co.

Microarray analysis

A customized bovine oligonucleotide microarray with 11,000 unique genes (GPL9136) was used to detect the genes expressed in the fetal membrane and endometrial tissues in the gravid and non-gravid uterine horn. The oligo-microarray produced by Agilent Technologies (Santa Clara, CA, USA) was used in this study. Sixty-mer nucleotide probes for the customized microarray were synthesized on a glass slide. The annotated bovine oligonucleotide array represented 10263 sequences, 4466 of which were known bovine genes, 5697 were unknown sequences and possible candidates for novel bovine genes, and 100 were internal references. All total RNA and mRNA tested including those examined by RT-PCR had their quality and quantity confirmed using either a Bioanalyzer (Agilent) or Nanodrop (Nanodrop Co.). We performed one-color microarray analysis, and fluorescence-labelled (Cy3) cRNA probes were prepared from 400 ng of total RNA of each sample using a Low RNA Input Linear Amplification Kit (Agilent Technologies). Labeled cRNA probes (600 ng each) were then hybridized and washed using the Gene Expression Hybridization Kit and Gene Expression Wash Pack Kit (Agilent Technologies), according to the manufacturer's instructions. After being washed, the arrays were scanned using an Agilent Microarray Scanner (Agilent Technologies), and Feature Extraction ver. 9.1 (Agilent Technologies) was used for image analysis and data extraction. The microarray data from each sample were imported into GeneSpring 7.3 (Agilent Technologies) for further data characterization. The GEO accession number is as GSE23981 <http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE23981>, and Platforms are GPL9136 and GPL10134.

Real-time RT-PCR(QPCR)

The total RNA was isolated from the fetal and endometrial tissues, BT-1 and SKIN cells, respectively used for microarray or RT-PCR analysis using TRIzol Reagent (Invitrogen, Carlsbad, CA, USA), according to the manufacturer's instructions. After verifying the quality of the RNA, one µg of total RNA was used for reverse transcription. The total RNA was then reverse transcribed into cDNA for 120 min at 37°C by MultiScribe™ reverse transcriptase (Perkin-Elmer Applied Biosystems, Foster City, CA) with a random primer dNTP mixture, MgCl₂, and an RNase inhibitor.

The gene expression levels of fibroblast growth factor 2 (FGF2), follistatin (FST), heparin-binding epidermal

growth factor-like growth factor (HBEGF), inhibin alpha (INHA), activin beta A (INHβA), activin beta B (INHβB), and transforming growth factor beta 1 (TGFβ1) were quantitatively confirmed by real-time RT-PCR analysis (QPCR) using the SYBR Green assay. Primer pairs were designed using the Primer Express 3.0 software program (Applied Biosystems) and are listed in Table 2. The thermal cycling conditions included the initial incubation of the samples at 50°C for two minutes and 95°C for ten minutes, followed by forty cycles of 95°C for fifteen seconds and 60°C for one minute. The cycle-threshold values (C_T) indicate the quantity of the target gene in each sample. The relative difference in the initial amount of each mRNA species (or cDNA) was determined by comparing the C_T values. The standard curves for each gene were generated by serially diluting plasmids containing FGF2, FST, HBEGF, INHA, INHβA, INHβB, TGFβ1, or GAPDH cDNA to quantify the mRNA concentrations produced. The expression ratio of each gene to GAPDH mRNA was calculated to adjust for any variation in the RT-PCR reaction. All values are presented as the mean ± SD. The expression of the inhibin βA gene during gestation was examined by another QPCR procedure. Briefly, one microgram of total RNA was reverse transcribed into cDNA in a total volume of 20 µL. QPCR was performed using SYBR Green fluorescence (Applied Biosystems, Foster City, CA, USA) to quantify inhibin βA gene expression. Bovine glyceraldehyde-3-phosphate dehydrogenase (GAPDH) was used as endogenous reference gene. The reaction mixture contained the cDNA template (1 µL), 0.4 µM each primer, SYBR Green PCR master mix (Applied Biosystems), and nuclease free water. The thermal cycling conditions included 2 min at 50 C and 10 min at 95 C. Thermal cycling proceeded with 40 cycles

Table 2 Sequences of the oligonucleotide primers used for real-time RT-PCR

Gene name	Strand	Sequence
FGF2	Forward	5'-CCAGTTGGTATGTGGCACTGA-3'
	Reverse	5'-GGTCCTGTTTTGGGTCCAAGT-3'
FST	Forward	5'-GCGTGCTGCTGGAAGTGAA-3'
	Reverse	5'-CGGTGCTCTCCGAAATGGA-3'
HBEGF	Forward	5'-AGCTCCGGTTCACCACT-3'
	Reverse	5'-ATGGCACCTCTCTCCGTGAT-3'
INHA	Forward	5'-AGGAGGGCCTCTTCACGTATG-3'
	Reverse	5'-TCCAGTCTGTGTGGAACCA-3'
INHβA	Forward	5'-GGCCACCCTCCCAAGG-3'
	Reverse	5'-TTAACGGCCTCCACCATCTC-3'
INHβB	Forward	5'-CCGGGTCCGCCTGTACTT-3'
	Reverse	5'-GCCTGCACCACAAACAGTT-3'
TGFβ1	Forward	5'-TGAGCCAGAGCGGACTACT-3'
	Reverse	5'-TGCCGTATCCACCATAGCA-3'

of 95 C for 15 sec and 60 C for 1 min. The primers were designed using the primer express software version 1.0, according to the manufacturer's instructions (Applied Biosystems). The primer sequences are given below together with their respective GenBank accession numbers within brackets. GAPDH (U85042): forward, 5'-aaggccatcaccatcttcca-3'; reverse, 5'-ccactacatactcagcac-cagcat-3'. The primers were commercially synthesized (Espec Oligo Service, Japan), and QPCR detection was performed using an ABI PRISM 7700 sequence detector and SDS software version 1.7.

Western blot analysis

To analyze activin-like protein expression, BT-1 cell extracts (100 µg) and supernatants (200 µL) were applied to Western blotting, as described previously with some modifications [29]. In brief, these proteins were added to sample buffer and boiled for 3 min, before being applied to an SDS polyacrylamide slab gel according to the method of Laemmli [33] using a Mini Protean III apparatus (Millipore corporation, BioRad, MA, USA). The proteins were electroblotted onto a polyvinylidene difluoride membrane (Immobilon-P, Millipore) using transblot apparatus (transblot SD cell, BioRad). After being blocked with 5% ECL blocking agent (GE Healthcare, NJ, USA), the membrane was incubated with anti-activin βA diluted 1000-fold overnight at 4°C, followed by incubation with alkaline phosphatase-conjugated antibody (Sigma) for 1 h at room temperature. For the absorption experiment, 1 µg of primary antibody was incubated with 8 µg of purified activin A for 1 hr, before they were incubated with the membrane. Anti-activin βA antibody and purified activin A were prepared by Dr Hasegawa of Kitasato University [34]. Immunopositive bands were visualized using an Alkaline Phosphatase Conjugate Substrate Kit (BioRad).

Statistical Analysis

The cell proliferation and QPCR data were analyzed by JMP software (SAS Institute Inc, Cary NC, USA) with one way ANOVA followed by the Dunnett's multiple comparison test. P-value of < 0.05 was considered significant.

Results

Gene expression levels of TGFβ super-family members in the fetal membrane and endometrium during implantation

Around Day 35 of gestation, no placental villi (chorionic villi), which were identified as spots with fine capillaries, were found in the FM in the NGH, and less than 30 villi were macromorphologically confirmed in the FM in the GH (data not shown). Microarray analysis showed that the expression levels of various trophoblast

specific genes were higher in the END of the GH than those in the END of the NGH; namely *CSH1*, *PRP*, *PAG*, *AIF*, and *Muc-1*. In FM, activin-related genes like *INHA*, *INHβA*, *INHβB*, *FST*, *TGFβ1*, and *HB-EGF* were detectable in both the GH and the NGH; however, *FGF2* was not detectable (Tables 3, 4). These data were confirmed by QPCR (Figure 2). There were no significant differences in the expression levels of these genes between the FM in the GH and NGH. No difference was seen in the endometrial *INHA* and *FST* expression levels between the GH and NGH, and quite high expression levels were detected during the estrous cycle compared to those observed during gestation although the difference was not significant. *INHβA* expression in the GH.END was much higher than those in other tissues (over 0.2 vs. less than 0.05). In the GH, the expression levels of *INHβA*, *INHβB*, *FGF2*, and *IGF1* in the END were significantly higher than those in the FM ($p < 0.05$).

When *INHβA* expression throughout gestation was analyzed by QPCR, higher expression levels were detected in the COT during early gestation and in the ICOT during the middle period of gestation. Similar

Table 3 Gene expression in the fetal membrane on Day 35 of gestation

Gene	Gravid (n = 3)	Non-gravid (n = 2)	Ratio Gravid/Non-gravid
EGF	0.03 ± 0.03 ^a	0.01	2.52
EGFR	1.68 ± 0.66	1.57	1.07
FGF2	0.01 ± 0.00	0.01	0.76
FST	84.74 ± 77.11	85.60	0.99
IGF1	0.02 ± 0.02	0.01	0.35
INHA	6.95 ± 2.37	6.10	1.14
INHBA	3.29 ± 2.84	248.39	0.01
INHBB	0.03 ± 0.03	0.15	0.19
TGFB1	1.41 ± 0.55	1.54	0.91
BMP3b	0.02	0.03	0.6
BMP4	1.41 ± 0.76	2.08	0.7
BMP15	0.01 ± 0.00	0.02	0.5
ACVR1	0.84 ± 0.13	1.02	0.8
ACVR2	0.90 ± 0.01	0.87	1.0
ACVR2B	0.48 ± 0.04	0.61	0.8
TGFBRI	1.02 ± 0.75	0.24	4.3
BMPRI1A	0.10 ± 0.01	0.17	0.6
BMPRI1B	0.04 ± 0.03	0.20	0.2
BMPRI2	0.33 ± 0.13	0.37	0.9
SMAD2	7.30 ± 0.90	8.05	0.9
CSH1	184.28 ± 16.47	2.71	68.0
PAG1	72.83 ± 15.22	0.59	124.0
PRP1	13.55 ± 4.70	0.37	37.0
IFNT	0.04 ± 0.05	0.24	0.2

^aMean ± SD.

Table 4 Gene expression in the endometrium on Day 35 of gestation

Gene	Gravid (n = 3)	Non-gravid (n = 3)	Ratio Gravid/Non-gravid
EGF	0.60 ± 0.95 ^a	0.06 ± 0.04	10.58
EGFR	0.60 ± 0.35	0.78 ± 0.41	0.76
FGF2	0.05 ± 0.07	0.13 ± 0.01	0.39
FST	21.57 ± 10.07	11.35 ± 6.47	1.90
IGF1	0.07 ± 0.09	0.17 ± 0.20	0.40
INHA	0.96 ± 0.32	1.01 ± 0.48	0.96
INHBA	536.20 ± 314.75	31.78 ± 40.56	16.87
INHBB	0.14 ± 0.11	0.03 ± 0.01	5.55
TGFB1	0.51 ± 0.31	0.76 ± 0.26	0.67
BMP3b	0.01	0.01	1.0
BMP4	0.98 ± 0.49	0.87 ± 0.34	1.1
BMP15	0.15 ± 0.24	0.08 ± 0.11	2.0
ACVR1	0.83 ± 0.28	1.02 ± 0.14	0.8
ACVR2	0.13 ± 0.10	0.15 ± 0.01	0.8
ACVR2B	0.31 ± 0.16	0.42 ± 0.20	0.7
TGFB1	0.13 ± 0.10	0.08 ± 0.07	1.6
BMP1A	0.08 ± 0.07	0.32 ± 0.14	0.3
BMP1B	0.01 ± 0.00	0.01 ± 0.00	1.0
BMP2	0.29 ± 0.15	0.27 ± 0.09	1.1
SMAD2	4.93 ± 1.98	4.34 ± 0.65	1.1

^aMean ± SD.

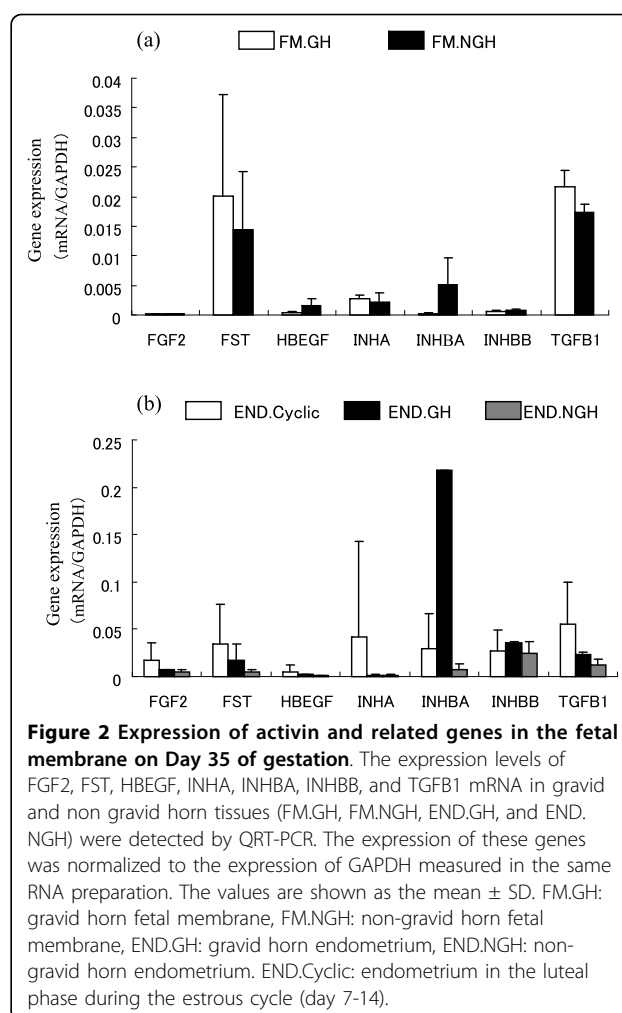
expression profiles were confirmed in the CAR and ICAR. However, two to three times higher expression intensities were detected in area from the END to the FM (Figures 3, 4).

Expression of TGFβ super-family members in BT-1 cells

BT-1 cells possess some trophoblastic cell features derived from in vitro fertilized ova [30], so we used them as a representative source replacing in vivo trophoblastic cells from the extra-embryonic membrane. First, the expression of various growth factors/cytokines was examined by QPCR. As a result, BT-1 cells were found to express *FGF2*, *HBEGF*, *INHβA*, *INHβB*, *FST*, and *TGFB1*, all of which were also detected in SKIN cells, which were used as the control (general somatic cells) (Figure 5). There were significant differences between the expression levels of these molecules in BT-1 and SKIN cells (except for in *INHA* expression) ($P < 0.05$).

Effects of trophoblast extracts on cell proliferation

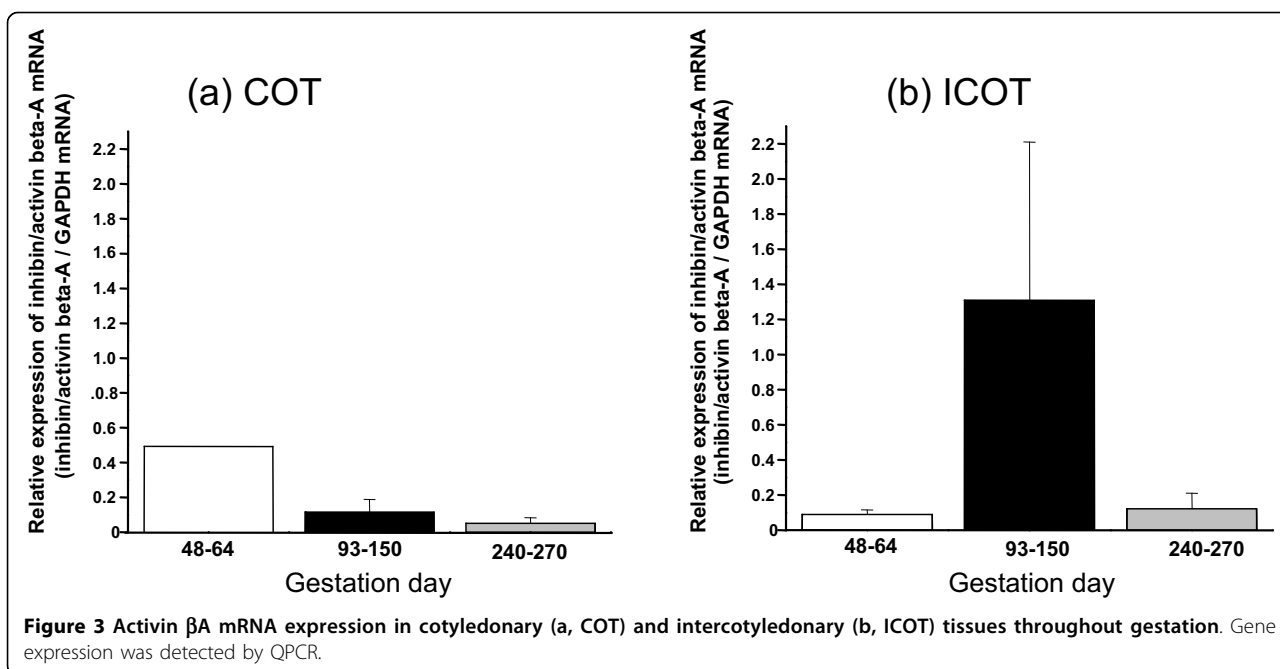
To examine whether trophoblastic factors stimulate endometrial cell proliferation, extracts containing 1-100 µg/mL proteins from the fetal membrane, BT-1, and SKIN cells were applied to in vitro cell cultures of two cell lines with different origins, E as bovine endometrial fibroblast cells and C as bovine cotyledonary fibroblast cells. The trophoblast and BT-1 extracts stimulated cell



proliferation in both E and C cells (Figure 6a and 6b, respectively); however, the SKIN cell extract had no effect. Cell proliferation analysis using C cells showed that the extracts from trophoblasts and BT-1 cells enhanced cell growth.

Activin protein expression in trophoblast cells

We next examined whether activin protein is expressed in trophoblastic cells using BT-1 cells as a model. Although activin-βA protein was found in BT-1 extract and culture supernatant by Western blot analysis, only small amounts of the appropriate sized proteins were detected in BT-1 cells and culture media (Figure 7). The activin-βA subunit was found in BT-1 culture supernatant as a 13 kDa protein but many nonspecific bands including a 26 kDa protein were also found in the extract (Figure 7a). The excess application of semi-purified activin to anti-activin sera showed the simultaneous disappearance of both the 13 and 26 kDa proteins in BT-1 cells.

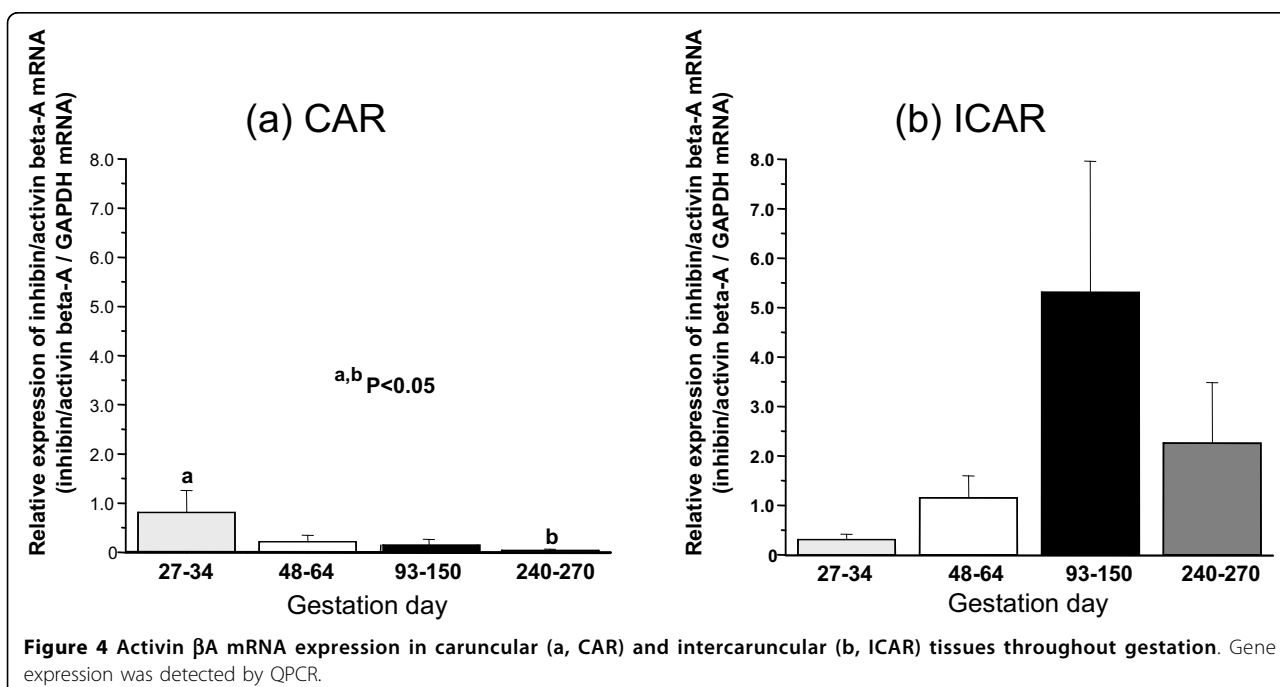


Cell proliferative activity of growth factors

To confirm whether activin displays proliferative activity in the endometrium, we applied various growth factors including activin-A and -AB to E and C fibroblast cell preparations. Although activin-A and -AB, FGF2, in and TGF β stimulated the proliferation of both cell lines, follistatin had no effect on either cell type. HB-EGF stimulated E cell growth (Figure 8).

Discussion

Implantation is a complex process involving various factors and biochemical and molecular modifications: cell-to-cell contact between cells with different origins, cell invasion and/or fusion depending on species, endometrial cell proliferation, and matrix reorganization. In the ruminant endometrium, feto-maternal events mainly occur in a limited area named the caruncle [1,35].



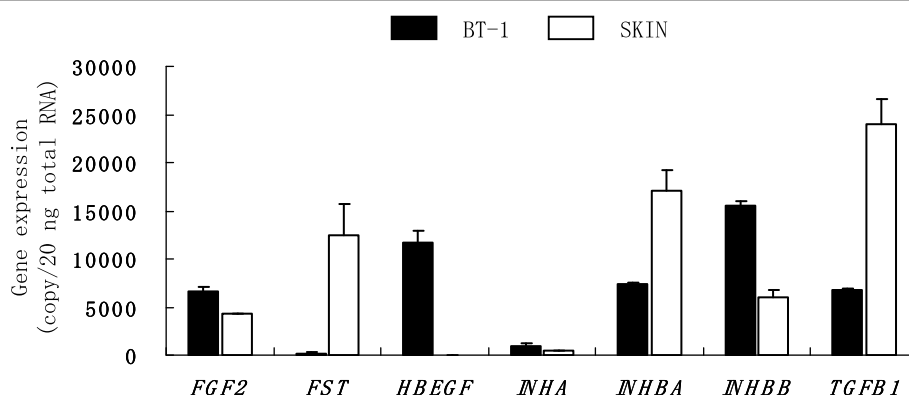


Figure 5 Expression of activin and related genes in trophoblastic cells (BT-1). Gene expression was detected by QPCR in BT-1 and SKIN cells. The expression of these genes is shown as copy number per 20 ng total RNA. The values are shown as mean $\pm$ SD. BT-1: a trophoblastic cell line established by Shimada et al. (2001), SKIN: cells derived from bovine skin as a control.

However, how and why cell-cell contact and active cell proliferation are initiated in the caruncle in ruminants rather than in the intercaruncle remain answered. During implantation, the embryo is stimulated by maternal factors, uterine milk protein and serpin [36,37], which ensure a suitable environment for fetal development. At

the same time, endometrial functions, and especially those in the caruncle in bovines, are regulated by fetal-side factors, for example interferon-tau, which stimulates endometrial prostaglandins and the ovarian corpus luteum, which functions as a factor for the recognition of gestation [38-42]. In the present study, we explored

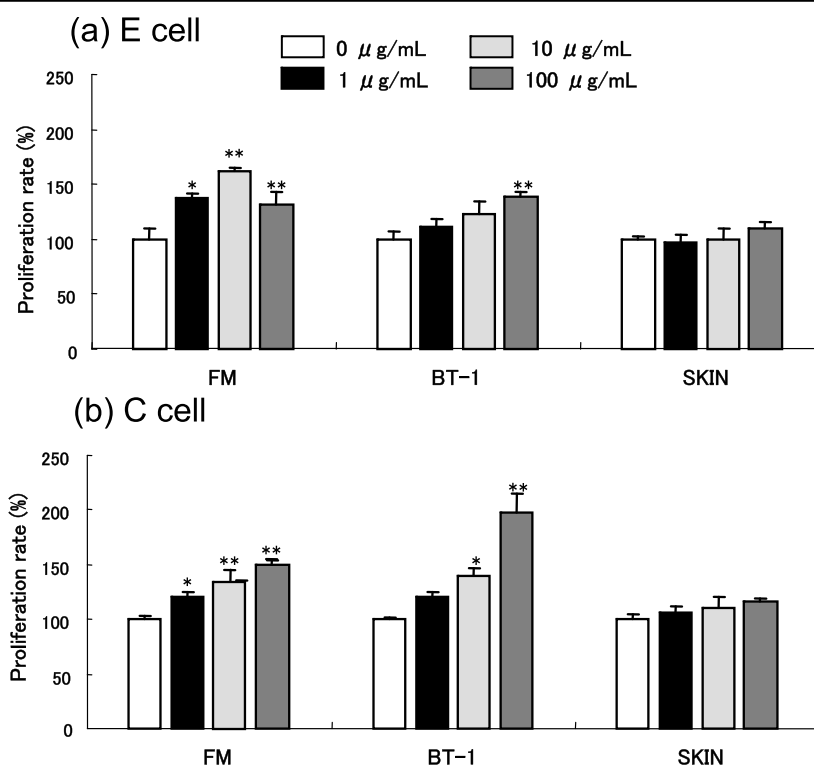
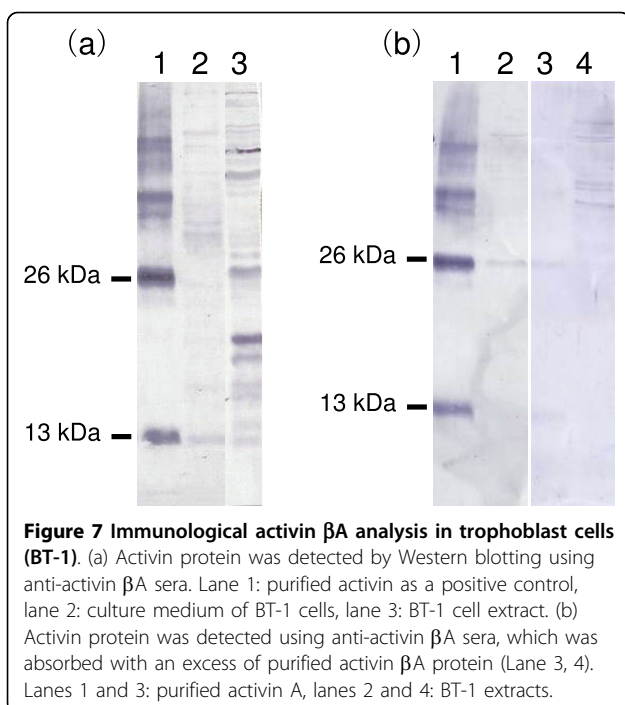


Figure 6 Effects of trophoblastic tissue extracts on the proliferation of fibroblast cells. EM: trophoblast tissue extracts with protein concentrations ranging from 1~100 μ g/mL, BT-1: cell extracts with the same protein concentrations as the EM, SKIN: cell extracts from the skin. These extracts were applied to endometrial fibroblast cells (E) and cotyledonary fibroblast cells (C). The values (mean $\pm$ SD) are expressed as percentages relative to the value obtained for the relevant untreated group (control). Dunnett's multiple comparison test was carried out to test the differences between the control and the treatment groups: *P < 0.05, **P < 0.01.



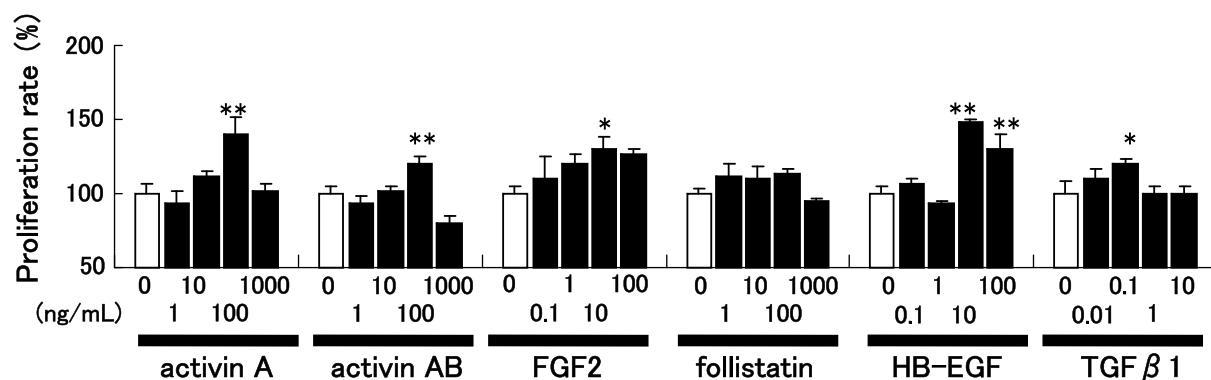
the possible role of activin, a member of the TGF β super-family, at the feto-maternal interface. As in other species such as humans and mice, activin plays a crucial role in endometrial remodelling, but there is little information regarding the role of the TGF β super-family in ruminants [15,43]. Although the structure of the bovine placenta is different from those of humans and mice; namely, the former is a non-invasive placenta but the latter is an invasive placenta, endometrial remodelling is still necessary for placentogenesis in bovines [44]. Discordant feto-maternal organization causes embryo loss. The caruncular and intercaruncular area play different roles in the bovine endometrium, and their gene expression patterns have been identified [45,46].

Extracts from the extra-embryonic membrane stimulated cell proliferation in an in vitro fibroblast cell culture, and BT-1 cells, which are derived from bovine blastocysts and have trophoblastic features, also stimulated cell proliferation effectively (Figure 4). These data suggest that trophoblastic cells contain a factor that stimulates bovine endometrial cell proliferation in the caruncle. Although trophoblast tissue expresses many factors including vascular factors (VEGF and their receptors, TGF, IGF, PIGF, etc. [47]), the most important factor remains unknown. Microarray and QPCR confirmed that various molecules including *FGF2*, *INH β A*, *INH β B*, *FST*, *HBEGF*, and *TGF β* were detectable in the END; however, most of them were highly expressed in the END.GH. A comparison between the gene expression patterns in gravid and non-gravid horn

trophoblasts showed that in the fetal membrane specific molecules such as placental lactogen (PL), prolactin-related protein (PRP), and pregnancy-associated glycoprotein (PAG) were expressed more in gravid horn trophoblasts. These results agreed with those of a previous report [8] and imply that members of the TGF β super-family including activin play roles in endometrial receptivity in ruminants. As in the gravid horn, the fusion of trophoblast cells and the endometrial epithelium starts earlier than in the non-gravid horn; namely, it may be evidence of the initiation of implantation [8,43].

Activin was detected in bovine blastocysts as well as in human and mouse blastocysts [11,13,48,49]. It stimulates the development of the embryo; i.e., cell proliferation of the blastocyst and attachment in humans and mice [48,50,51]. Placental villi produce activin A and promote hormone production, and these hormones direct the formation of the trophoblastic syncytium [52]. Although endometrial activin production is known to occur in these species [26], there is no information regarding its function in the bovine caruncular endometrium. TGF β super-family genes including activin genes (*INH β A* and *INH β B*) may play crucial roles in the proliferation of various cell types such as endometrial, immune, stem, ovarian and tumor cells [53-55]. In the present study, high *INH β A* expression was found in the gravid endometrium, and our findings suggest that activin plays a similar role as a regulator of endometrial remodelling in humans even though bovine endometrial cells are not involved in decidualization during implantation [17,52]. Endometrial remodelling is necessary for bovine implantation [56,57], and a comparison between gravid and non-gravid endometrial molecular expression implied that the initiation of implantation and cell proliferation in the bovine caruncular endometrium is regulated by fetal-side trophoblasts. However, there are no reports regarding fetal-side stimulation. We hypothesized that trophoblastic factors participate in rapid spatiotemporal endometrial cell proliferation during bovine implantation. Using BT-1 cells, which possess trophoblastic features, we attempted to identify activin and detected various molecules immunologically (Figure 5). Extracts from fetal trophoblastic tissues and BT-1 cells stimulated cell proliferation. These findings suggest that fetal-side tissues stimulate endometrial remodelling and development during implantation. Activin activity is regulated by various factors including FST, and FST expression is higher than that of activin genes in bovine trophoblastic tissues [20,55]. Therefore, activin may not be active in fetal trophoblasts at this time. Although activin is not the only factor that stimulates cell proliferation in the bovine endometrium, it may play an important role in endometrial remodelling and development [52]. The detailed roles of activin in bovine

(a) E cell



(b) C cell

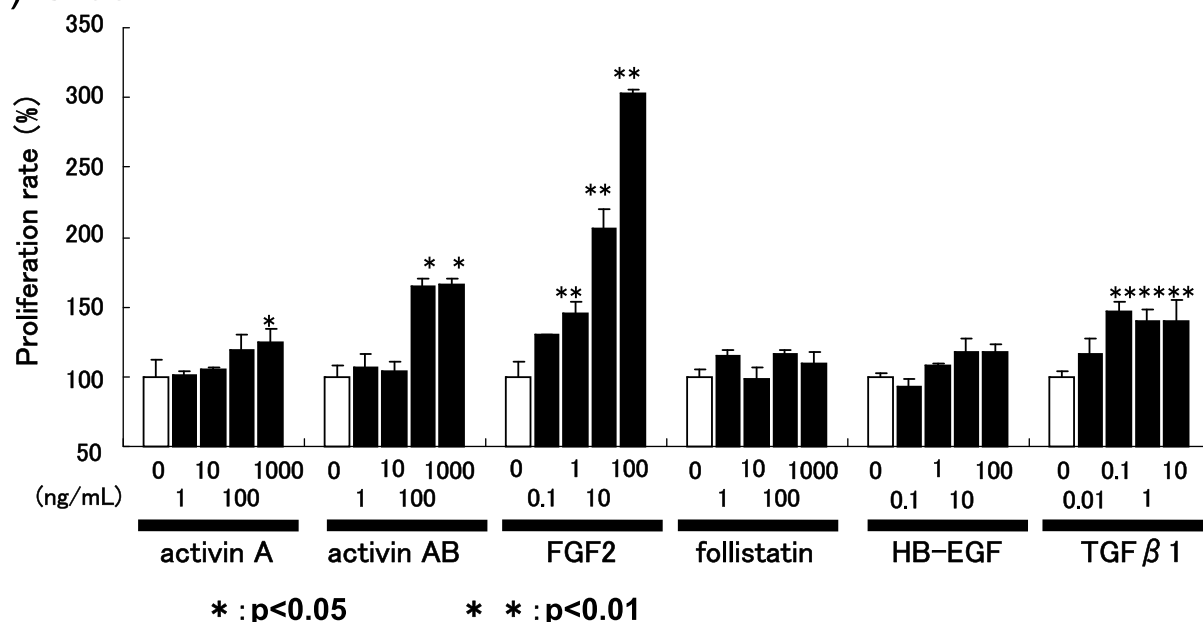


Figure 8 Cell proliferation analysis of activin and related growth factors. Purified activin A, activin AB, and follistatin were applied to E and C cells at concentrations ranging from 1 to 1000 ng/mL. FGF2 and HB-EGF were used at concentrations ranging from 0.1 to 100 ng/mL. TGFβ1 was used at concentrations ranging from 0.01 to 10 ng/mL. The values (mean ± SD) are expressed as percentages relative to the value obtained for the relevant untreated group (control). Dunnett's multiple comparison test was carried out to test the differences between the control and treatment groups: *P < 0.05, **P < 0.01.

endometrial remodelling and implantation require further study.

In the present study, other molecules were found to be expressed in the fetal and endometrial tissues, (FGF2, HB-EGF, and TGFβ1), but their biological roles are still obscure. Almost no expression of FGF2 was detected in the trophoblastic tissues but it did demonstrate proliferative activity in cell culture. Endometrial FGF2 may be important for endometrial remodelling and implantation and may be related to interferon tau expression [58,59].

In addition, HB-EGF may play a specific role during implantation on both the maternal and fetal sides [12,60,61]; however, this requires further study.

Conclusions

TGFβ super-family genes are specifically expressed in the bovine endometrial and trophoblastic tissues in a specific spacio-temporal manner. *INHβA* (activin βA) was found to be highly expressed in the gravid horn endometrium, but its expression was low in trophoblasts. Several factors

derived from trophoblastic tissues stimulated the proliferation of fibroblast cells, and activin may participate in endometrial proliferation. These results suggest that TGF β super-family members including activin, which are produced by trophoblasts and the endometrium, play a crucial role in endometrial remodeling during the peri-implantation period in cattle. In addition, bovine trophoblastic tissues contain various other cytokine/growth factors that may be important for placental growth during bovine placentation.

Acknowledgements

This research was supported by a grant from the Research Project for Utilizing Advanced Technologies (05-1770) from the Ministry of Agriculture, Forestry, and Fisheries of Japan and grants (Kiban-kenkyu C 17580284; Kiban-kenkyu B 20380159) from the Ministry of Education, Culture, Sport, Science, and Technology of Japan. We thank Dr. Nobuhiko Yamauchi, Kyushu University for supply information of bovine endometrial and cotyledonary fibroblast cells. We thank Drs. Toru Takahashi, Misa Hosoe and Koichi Ushizawa for help to collect samples and performed microarray.

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Authors' contributions

KS and KK participated in the design of study, carried out the experiments and prepared the manuscript. CBH performed QPCR for TGF β super-family during gestation. YH purified Activin, Inhibin and raised their antibody. KH participated in the design and coordination of study and helped to draft the manuscript. All authors read and approved the final manuscript for publication.

Competing interests

The authors declare that they have no competing interests.

Received: 5 March 2010 Accepted: 15 October 2010

Published: 15 October 2010

References

- Hashizume K, Ushizawa K, Patel OV, Kizaki K, Imai K, Yamada O, Nakano H, Takahashi T: Gene expression and maintenance of pregnancy in bovine: roles of trophoblastic binucleate cell-specific molecules. *Reprod Fertil Dev* 2007, **19**:79-90.
- King GJ, Atkinson BA, Robertson HA: Development of the bovine placentome during the second month of gestation. *J Reprod Fertil* 1979, **55**:173-180.
- Spencer TE, Burghardt RC, Johnson GA, Bazer FW: Conceptus signals for establishment and maintenance of pregnancy. *Anim Reprod Sci* 2004, **82**:537-550.
- Blomberg L, Hashizume K, Viebahn C: Blastocyst elongation, trophoblastic differentiation, and embryonic pattern formation. *Reproduction* 2008, **135**:181-195.
- Bazer FW, Spencer TE, Johnson GA, Burghardt RC, Wu G: Comparative aspects of implantation. *Reproduction* 2009, **138**:195-209.
- Dimitriadis E, White CA, Jones RL, Salamonsen LA: Cytokines, chemokines and growth factors in endometrium related to implantation. *Hum Reprod Update* 2005, **11**:613-630.
- King GJ, Atkinson BA, Robertson HA: Development of the bovine placentome from days 20 to 29 of gestation. *J Reprod Fertil* 1980, **59**:95-100.
- Yamada O, Todoroki J, Kizaki K, Takahashi T, Imai K, Patel OV, Schuler LA, Hashizume K: Expression of prolactin-related protein I at the fetomaternal interface during the implantation period in cows. *Reproduction* 2002, **124**:427-437.
- Maddox-Hyttel P, Alexopoulos NI, Vajta G, Lewis I, Rogers P, Cann L, Callesen H, Tveden-Nyborg P, Trounson A: Immunohistochemical and ultrastructural characterization of the initial post-hatching development of bovine embryos. *Reproduction* 2003, **125**:607-623.
- Albano RM, Groome N, Smith JC: Activins are expressed in preimplantation mouse embryos and in ES and EC cells and are regulated on their differentiation. *Development* 1993, **117**:711-723.
- Lu RZ, Matsuyama S, Nishihara M, Takahashi M: Developmental expression of activin/inhibin beta A, beta B, and alpha subunits, and activin receptor-11B genes in preimplantation mouse embryos. *Biol Reprod* 1993, **49**:1163-1169.
- Das SK, Wang XN, Paria BC, Damm D, Abraham JA, Klagsbrun M, Andrews GK, Dey SK: Heparin-binding EGF-like growth factor gene is induced in the mouse uterus temporally by the blastocyst solely at the site of its apposition: a possible ligand for interaction with blastocyst EGF-receptor in implantation. *Development* 1994, **120**:1071-1083.
- He ZY, Liu HC, Mele CA, Barmat L, Veeck LL, Davis O, Rosenwaks Z: Expression of inhibin/activin subunits and their receptors and binding proteins in human preimplantation embryos. *J Assist Reprod Genet* 1999, **16**:73-80.
- Chow JF, Lee KF, Chan ST, Yeung WS: Quantification of transforming growth factor beta1 (TGFbeta1) mRNA expression in mouse preimplantation embryos and determination of TGFbeta receptor (type I and type II) expression in mouse embryos and reproductive tract. *Mol Hum Reprod* 2001, **7**:1047-1056.
- Munson L, Wilhite A, Boltz VF, Wilkinson JE: Transforming growth factor beta in bovine placentas. *Biol Reprod* 1996, **55**:748-755.
- Jones RL, Stoikos C, Findlay JK, Salamonsen LA: TGF-beta superfamily expression and actions in the endometrium and placenta. *Reproduction* 2006, **132**:217-232.
- Stoikos CJ, Harrison CA, Salamonsen LA, Dimitriadis E: A distinct cohort of the TGFbeta superfamily members expressed in human endometrium regulate decidualization. *Hum Reprod* 2008, **23**:1447-1456.
- Ishiwata H, Katsuma S, Kizaki K, Patel OV, Nakano H, Takahashi T, Imai K, Hirasawa A, Shiojima S, Ikawa H, Suzuki Y, Tsujimoto G, Izaika Y, Todoroki J, Hashizume K: Characterization of gene expression profiles in early bovine pregnancy using a custom cDNA microarray. *Mol Reprod Dev* 2003, **65**:9-18.
- Ling N, Ying SY, Ueno N, Shimasaki S, Esch F, Hotta M, Guillemin R: A homodimer of the beta-subunits of inhibin A stimulates the secretion of pituitary follicle stimulating hormone. *Biochem Biophys Res Commun* 1986, **138**:1129-1137.
- Shimasaki S, Moore RK, Otsuka F, Erickson GF: The bone morphogenetic protein system in mammalian reproduction. *Endocr Rev* 2004, **25**:72-101.
- Silva JR, van den Hurk R, van Tol HT, Roelen BA, Figueiredo JR: Gene expression and protein localisation for activin-A, follistatin and activin receptors in goat ovaries. *J Endocrinol* 2004, **183**:405-415.
- Yamashita S, Maeshima A, Kojima I, Nojima Y: Activin A is a potent activator of renal interstitial fibroblasts. *J Am Soc Nephrol* 2004, **15**:91-101.
- Fumagalli M, Musso T, Vermi W, Scutera S, Daniele R, Alotto D, Cambieri I, Ostorero A, Gentili F, Caposio P, Zucca M, Sozzani S, Stella M, Castagnoli C: Imbalance between activin A and follistatin drives postburn hypertrophic scar formation in human skin. *Exp Dermatol* 2007, **16**:600-610.
- Caniggia I, Lye SJ, Cross JC: Activin is a local regulator of human cytotrophoblast cell differentiation. *Endocrinology* 1997, **138**:3976-3986.
- Jones RL, Kaitu'u-Lino TJ, Nie G, Sanchez-Partida LG, Findlay JK, Salamonsen LA: Complex expression patterns support potential roles for maternally derived activins in the establishment of pregnancy in mouse. *Reproduction* 2006, **132**:799-810.
- Yamauchi N, Takezawa T, Kizaki K, Herath CB, Hashizume K: Proliferative potential of endometrial stromal cells, and endometrial and placental expression of cyclin in the bovine. *J Reprod Dev* 2003, **49**:553-560.

27. Yamauchi N, Yamada O, Takahashi T, Imai K, Sato T, Ito A, Hashizume K: **A three-dimensional cell culture model for bovine endometrium: regeneration of a multicellular spheroid using ascorbate.** *Placenta* 2003, **24**:258-269.
28. Kadokawa H, Minezawa M, Yamamoto Y, Takahashi M, Shimada K, Takahashi H, Kariya T: **Freemartinism among singleton bovine females born from multiple embryo transfer.** *Theriogenology* 1995, **44**:295-306.
29. Shimada A, Nakano H, Takahashi T, Imai K, Hashizume K: **Isolation and characterization of a bovine blastocyst-derived trophoblastic cell line, BT-1: development of a culture system in the absence of feeder cell.** *Placenta* 2001, **22**:652-662.
30. Hashizume K, Shimada A, Nakano H, Takahashi T: **Bovine trophoblast cell culture systems: a technique to culture bovine trophoblast cells without feeder cells.** *Methods Mol Med* 2006, **121**:179-188.
31. Wada W, Maeshima A, Zhang Y-Q, Hasegawa Y, Kuwano H, Kojima I: **Assessment of the function of the bC-subunit of activin in cultured hepatocytes.** *Am J Physiol Endocrinol Metab* 2004, **287**:E247-E254.
32. Hasegawa Y, Eto Y, Ibuki Y, Sugino H: **Activin as autocrine and paracrine factor in the ovary.** *Horm Res.* 1994, **41**(1):55-62.
33. Laemmli UK: **Cleavage of structural proteins during the assembly of the head of bacteriophage T4.** *Nature* 1970, **227**:680-685.
34. Kaneko H, Noguchi J, Kikuchi K, Todoroki J, Hasegawa Y: **Alterations in peripheral concentrations of inhibin A in cattle studied using a time-resolved immunofluorometric assay: relationship with estradiol and follicle-stimulating hormone in various reproductive conditions.** *Biol Reprod* 2002, **67**:38-45.
35. Klisch K, Pfarrer C, Schuler G, Hoffmann B, Leiser R: **Tripolar acytokinetic mitosis and formation of feto-maternal syncytia in the bovine placenta: different modes of the generation of multinuclear cells.** *Anat Embryol (Berl)* 1999, **200**:229-237.
36. Ing NH, Francis H, McDonnell JJ, Amann JF, Roberts RM: **Progesterone induction of the uterine milk proteins: major secretory proteins of sheep endometrium.** *Biol Reprod* 1989, **41**:643-654.
37. Stewart MD, Johnson GA, Gray CA, Burghardt RC, Schuler LA, Joyce MM, Bazer FW, Spencer TE: **Prolactin receptor and uterine milk protein expression in the ovine endometrium during the estrous cycle and pregnancy.** *Biol Reprod* 2000, **62**:1779-1789.
38. Majewska M, Woclawek-Potocka I, Bah MM, Hapunik J, Piotrowska KK, Tasaki Y, Acosta TJ, Okuda K, Skarzynski DJ: **Is interleukin-1alpha a luteotrophic or luteolytic agent in cattle?** *Reproduction* 2010, **139**:665-672.
39. Roberts RM, Cross JC, Farin CE, Hansen TR, Klemann SW, Imakawa K: **Interferons at the placental interface.** *J Reprod Fertil* 1990, **41**:63-74.
40. Flint AP, Parkinson TJ, Stewart HJ, Vallet JL, Lamming GE: **Molecular biology of trophoblast interferons and studies of their effects in vivo.** *J Reprod Fertil* 1991, **43**:13-25.
41. Martal J, Chène N, Camous S, Huynh L, Lantier F, Hermier P, L'Haridon R, Charpigny G, Charlier M, Chaouat G: **Recent developments and potentialities for reducing embryo mortality in ruminants: the role of IFN-tau and other cytokines in early pregnancy.** *Reprod Fertil Dev* 1997, **9**:355-380.
42. Cooke FNT, Pennington KA, Yang Q, Ealy AD: **Several fibroblast growth factors are expressed during pre-attachment bovine conceptus development and regulate interferon-tau expression from trophoblast.** *Reproduction* 2009, **137**:259-269.
43. Ravelich SR, Shelling AN, Wells DN, Peterson AJ, Lee RS, Ramachandran A, Keelan JA: **Expression of TGF-beta1, TGF-beta2, TGF-beta3 and the receptors TGF-betaRI and TGF-betaRII in placentomes of artificially inseminated and nuclear transfer derived bovine pregnancies.** *Placenta* 2006, **27**:307-316.
44. Uekita T, Yamanouchi K, Sato H, Tojo H, Seiki M, Tachi C: **Expression and localization of matrix metalloproteinases (MT1-MMP, MMP-2) and tissue inhibitor of metalloproteinase-2 (TIMP-2) during synepitheliochorial placentation of goats (*Capra hircus*).** *Placenta* 2004, **25**:810-819.
45. Mansouri-Attia N, Aubert J, Reinaud P, Giraud-Delville C, Taghouti G, Galio L, Everts RE, Degrelle S, Richard C, Hue I, Yang X, Tian XC, Lewin HA, Renard JP, Sandra O: **Gene expression profiles of bovine caruncular and intercaruncular endometrium at implantation.** *Physiol Genomics* 2009, **39**:14-27.
46. Berg DK, van Leeuwen J, Beaumont S, Berg M, Pfeffer PL: **Embryo loss in cattle between Days 7 and 16 of pregnancy.** *Theriogenology* 2010, **73**:250-260.
47. Ushizawa K, Herath CB, Kaneyama K, Shiojima S, Hirasawa A, Takahashi T, Imai K, Ochiai K, Tokunaga T, Tsunoda Y, Tsujimoto G, Hashizume K: **cDNA microarray analysis of bovine embryo gene expression profiles during the pre-implantation period.** *Reprod Biol Endocrinol* 2004, **2**:77.
48. Yoshioka K, Suzuki C, Iwamura S: **Activin A and follistatin regulate developmental competence of In vitro-produced bovine embryos.** *Biol Reprod* 1998, **59**:1017-1022.
49. Patel OV, Bettgowda A, Ireland JJ, Coussens PM, Lonergan P, Smith GW: **Functional genomics studies of oocyte competence: evidence that reduced transcript abundance for follistatin is associated with poor developmental competence of bovine oocytes.** *Reproduction* 2007, **133**:95-106.
50. Orimo T, Taga M, Matsui H, Minaguchi H: **The effect of activin-A on the development of mouse preimplantation embryos in vitro.** *J Assist Reprod Genet* 1996, **13**:669-674.
51. Mtango NR, Varisanga MD, Dong YJ, Rajamahendran R, Suzuki T: **Growth factors and growth hormone enhance in vitro embryo production and post-thaw survival of vitrified bovine blastocysts.** *Theriogenology* 2003, **59**:1393-402.
52. Jones RL, Findlay JK, Farnworth PG, Robertson DM, Wallace E, Salamonsen LA: **Activin A and inhibin A differentially regulate human uterine matrix metalloproteinases: potential interactions during decidualization and trophoblast invasion.** *Endocrinology* 2006, **147**:724-732.
53. Seder CW, Hartojo W, Lin L, Silvers AL, Wang Z, Thomas DG, Giordano TJ, Chen G, Chang AC, Orringer MB, Beer DG: **Upregulated INHBA expression may promote cell proliferation and is associated with poor survival in lung adenocarcinoma.** *Neoplasia* 2009, **11**:388-396.
54. Sulzbacher S, Schroeder IS, Truong TT, Wobus AM: **Activin A-induced differentiation of embryonic stem cells into endoderm and pancreatic progenitors-the influence of differentiation factors and culture conditions.** *Stem Cell Rev* 2009, **5**:159-173.
55. Xia Y, Schneyer AL: **The biology of activin: recent advances in structure, regulation and function.** *J Endocrinol* 2009, **202**:1-12.
56. Hashizume K: **Analysis of uteroplacental-specific molecules and their functions during implantation and placentation in the bovine.** *J Reprod Dev* 2007, **53**:1-11.
57. Kizaki K, Ushizawa K, Takahashi T, Yamada O, Todoroki J, Sato T, Ito A, Hashizume K: **Gelatinase (MMP-2 and -9) expression profiles during gestation in the bovine endometrium.** *Reprod Biol Endocrinol* 2008, **6**:66.
58. Michael DD, Alvarez IM, Ocón OM, Powell AM, Talbot NC, Johnson SE, Ealy AD: **Fibroblast growth factor-2 is expressed by the bovine uterus and stimulates interferon-tau production in bovine trophoblast.** *Endocrinology* 2006, **147**:3571-3579.
59. Ocón-Grove OM, Cooke FN, Alvarez IM, Johnson SE, Ott TL, Ealy AD: **Ovine endometrial expression of fibroblast growth factor (FGF) 2 and conceptus expression of FGF receptors during early pregnancy.** *Domest Anim Endocrinol* 2008, **34**:135-145.
60. Lessey BA, Gui Y, Apparao KB, Young SL, Mulholland J: **Regulated expression of heparin-binding EGF-like growth factor (HB-EGF) in the human endometrium: a potential paracrine role during implantation.** *Mol Reprod Dev* 2002, **62**:446-455.
61. Jessmon P, Leach RE, Armant DR: **Diverse functions of HBEGF during pregnancy.** *Mol Reprod Dev* 2009, **76**:1116-1127.

doi:10.1186/1477-7827-8-120

Cite this article as: Sugawara *et al.*: Transforming growth factor beta family expression at the bovine feto-maternal interface. *Reproductive Biology and Endocrinology* 2010 **8**:120.