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8 **Global coagulation assays – Proposed reference intervals for healthy controls**

9 *Reference intervals for global coagulation assays*

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27 Dear Editors,

28 Global coagulation assays (GCA) have been proposed as better surrogate measures of the  
29 haemostatic profile compared to traditional coagulation studies<sup>1</sup>. Traditional coagulation assays  
30 such as prothrombin time (PT) and activated partial thromboplastin time (APTT) measure the  
31 time to the start of clot formation, representing less than 5% of total clot formation<sup>1</sup>.  
32 Furthermore, they were developed to measure anticoagulants, with their role in assessing  
33 bleeding and thrombotic risk is unclear. GCA, on the other hand, measure the end products of  
34 the coagulation cascade such as thrombin and fibrin, and provide a more comprehensive  
35 assessment of the haemostatic profile. These assays have been shown to be potentially more  
36 useful in assessing for bleeding and thrombotic disorders<sup>1</sup>. One of the challenges of any new  
37 assay is understanding what is normal, the variations seen with standard epidemiology factors of  
38 age and gender, and developing a reference range for these parameters. We have previously  
39 reported significant age and gender differences in thromboelastography (TEG®) and calibrated  
40 automated thrombogram (CAT®) parameters in the normal population<sup>2,3</sup>. Since our previous  
41 report, we have expanded our cohort of healthy controls and here, we report the reference  
42 intervals for TEG, CAT and a fibrin generation assay, overall haemostatic potential (OHP).

43 Healthy volunteers were recruited and written informed consent obtained. Volunteers with  
44 history of thrombosis, active cardiovascular risk factors on treatment, such as hypertension,  
45 diabetes and hyperlipidaemia, as well as those on hormonal therapies, anticoagulation or  
46 antiplatelet agents were excluded. All volunteers also underwent testing with full blood count,  
47 renal and liver function and coagulation studies and were excluded if the results were abnormal.  
48 Samples were collected via venepuncture from the cubital fossa using 21G vacutainer or  
49 butterfly collection set. Whole blood collected in citrated tube was used for immediate testing  
50 with TEG® within 4 hours and additional samples were double-spun at 2500g to obtain platelet-  
51 poor plasma (PPP). The PPP was immediately stored at -80°C and tested at a later time for CAT  
52 and OHP. The study was approved by the Austin and Northern Health Human Research Ethics  
53 Committees (H2013/04977 and P5/13).

54 Thromboelastography measures the changes in the viscoelastic properties of whole blood during  
55 fibrin clot formation, propagation and dissolution and was performed using TEG® 5000

56 (Haemonetics, USA) as per manufacturer's guidelines. 1000 $\mu$ L citrated blood was mixed with  
57 40 $\mu$ L kaolin. 340 $\mu$ L of this mix was added to a cup preheated to 37°C, containing 20 $\mu$ L of 0.2M  
58 Calcium chloride. The cup oscillates around a suspended pin. As the fibrin clot forms, the  
59 torque on the wire is recorded. CAT® (Diagnostica Stago, France) determines the rate and  
60 extent of thrombin formed over 60 minutes after a tissue factor stimulus. 80 $\mu$ L of PPP was  
61 added to either 20 $\mu$ L of PPP reagent or 20 $\mu$ L of thrombin calibrator. The final mixture has a  
62 concentration of 5pM tissue factor; 4 $\mu$ M phospholipids. Coagulation is triggered with the  
63 addition of calcium chloride in a buffer with fluorogenic substrate. OHP is derived from a fibrin  
64 aggregation curve formed from repeated spectrophotometric measurements measured on the  
65 FLUOstar Optima (BMG Labtech) at 405nm. 75 $\mu$ L of thawed PPP was added to wells with  
66 75 $\mu$ L of buffer containing either (i) thrombin (0.006IU/mL) to generate the overall coagulation  
67 potential (OCP) or (ii) thrombin and tissue plasminogen activator (tPA) (600ng/mL) to generate  
68 the OHP. Figure 1 illustrates the key parameters as measured using TEG®, CAT® and OHP.

69 Statistical analysis: Reference intervals were calculated using the non-parametric approach  
70 based on the recommendations of the International Federation of Clinical Chemistry and  
71 Laboratory Medicine<sup>4</sup>. Screening for outlier exclusion was undertaken using the Reed test<sup>5</sup> or  
72 the Tukey test<sup>6</sup>. Sex-dependent differences were explored using a Mann-Whitney U test. Age-  
73 dependent differences were assessed using analysis of variance<sup>7</sup>. Where differences were found  
74 for a given analyte, sex partitioned reference intervals were calculated. As described in the  
75 Clinical and Laboratory Standards Institute (CLSI) Guidelines C28-A3<sup>4</sup>, the non-parametric  
76 rank method was used to calculate the reference interval for partitions with a sample size  $\geq$ 120  
77 participants. For analytes with partitions containing <120 participants, the robust method of  
78 Horn and Pesce was used to calculate the reference interval<sup>4</sup>. The data are presented as 2.5-  
79 97.5<sup>th</sup> percentile ranges (two-sided) with 90<sup>th</sup> percentile confidence intervals. The MedCalc  
80 software was used for statistical analysis.

81 A total of 153 healthy volunteers, aged 18 to 80 years (mean age 42 $\pm$ 17 years, 98 females), were  
82 prospectively recruited. Table1 shows gender and age-specific reference intervals for TEG®,  
83 CAT® and OHP. For TEG®, the manufacturer's reference intervals were R-time, 2-8 min; K-  
84 time, 1-3 min;  $\alpha$ -angle, 55-78°; maximum amplitude (MA), 51-69 mm; LY30 0-8%. Age did  
85 not appear to influence TEG® parameters while gender did. Figure 2 shows the age and gender  
86 differences in some of the key GCA parameters. Compared to the manufacturer's reference  
87 intervals, 25 (20.3%) of healthy controls were outside of the reference interval for R-time, 19  
88 (15.4%) for K-time, 47 (38.2%) for  $\alpha$ -angle, 18 (14.6%) for MA and 4 (3.2%) for LY30. Only  
89 58 (47.2%) controls had all parameters within the manufacturer's reference intervals. This

90 discordance is consistent to Scarpelini et al which reported a specificity of 81% for the  
91 manufacturer's "normal" values<sup>8</sup>. Of note, the manufacturer's reference interval was established  
92 using small studies of volunteers or hospitalised patients, which may limit its generalisability.

93 To the best of our knowledge, there are no manufacturer's reference interval for CAT.  
94 Previously published reference intervals for CAT either did not delineate age and gender or used  
95 a different tissue factor concentration<sup>9,10</sup>. The results from the Gutenberg Health study involving  
96 over 1000 healthy controls and included results derived from 5pM tissue factor, reported a  
97 median of 2.7 (interquartile range (IQR) 2.3, 3.0) minutes for lag time, a mean of 1322±196 for  
98 endogenous thrombin potential (ETP) and 236±52 for peak height for males, and 2.4 minutes  
99 (IQR 2.3, 2.7) for lag time, 1318±212 nM.min for ETP and 259±53 nM for peak height for  
100 females<sup>10</sup>. Interestingly, the ETP was not significantly different between genders in the  
101 Gutenberg Health Study (p=0.71) while the female controls in our study was more  
102 hypercoagulable than the males (1391±263 vs 1236±219 nM.min, p<0.001). Both studies  
103 reported higher peak height in females (our study 236±68 vs 190±54 nM, p<0.001) even after  
104 adjusting for age<sup>3,10</sup>. While the lag time was longer in the male controls in the Gutenberg Health  
105 Study (p<0.001)<sup>10</sup>, the difference was not significant in our study (3.0 (2.7, 3.7) vs 3.3 (3.0, 3.7)  
106 min, p=0.097). No age group specific ranges were reported in the Gutenberg Health Study<sup>10</sup>.

107 There are no published reference intervals for the OHP assay and previous studies have utilised  
108 different reagent concentrations<sup>11</sup>. In our study, we found significant age and gender differences  
109 in the OHP parameters. Females appear more hypercoagulable than males (Overall coagulation  
110 potential, OCP 38.0±8.9 units vs 20.9±9.5 units (p<0.001); OHP 7.9±3.1 vs 5.9±3.4 units  
111 (p<0.001); and overall fibrinolytic potential, OFP 79.3±6.2 vs 81.5±6.6 % (p=0.020). The older  
112 controls also demonstrated higher OCP (39.9±9.1 vs 32.6±9.0 units, p<0.001) and OHP  
113 (9.0±3.6 vs 6.1±2.6 units, p<0.001) values and reduced OFP (77.4±7.6 vs 81.7±4.8 %,  
114 p<0.001).

115 The proposed reference intervals derived from this study represents an adult population within  
116 Australia's metropolitan area with diverse ethnicity background (56% Caucasians, 35% South  
117 Asians and 9% South Asians). Importantly, and unique to many normal control studies, our  
118 volunteers have been specifically screened for pre-existing cardiovascular and prothrombotic  
119 risk factors. This study confirms the gender differences seen in some of the parameters, which  
120 are not quantified by the currently published TEG® reference intervals or other studies  
121 involving CAT® using this specific study protocol. Statistically significant age-dependent  
122 differences were found for lag time, OCP, OHP and OFP. However, combined with the

123 uncertainty in the partitioned reference intervals and for standardisation, an arbitrary 50 years  
124 old cut-off was considered appropriate for these analytes. In addition, we note there is some  
125 differences across the different ethnic group, particularly with the South Asian group – the MA  
126 and ETP are significantly elevated in this group compared to the rest of the group but we note  
127 that there are only 13 subjects in this group which limits how meaningful and applicable this  
128 finding may be.

129 To the best of our knowledge, this is also a first-in-kind study looking at reference intervals in  
130 the OHP assay, a fibrin generation assay. Given the discordance of our results to reference  
131 intervals provided by the TEG® manufacturer, many otherwise well healthy individuals in our  
132 population would have been identified as “abnormal” supporting the recommendations by the  
133 manufacturer to establish each institution’s local ranges. The clinical implications of these  
134 differences were not explored in this study but given that TEG® is a point-of-care device used  
135 to support transfusion in bleeding patients, it is important to raise awareness of these  
136 discrepancies to avoid potentially unnecessarily transfusion. We acknowledge that our locally  
137 determined reference intervals, however, may not be directly transferable in laboratories serving  
138 different populations or using different study protocol and trigger reagents. Laboratories should  
139 embark on establishing local reference intervals where possible, however, our reference  
140 intervals may be useful for transference checking<sup>12</sup>.

141 In conclusion, this letter reports reference intervals for three commonly used global coagulation  
142 assays, TEG®, CAT® and OHP assays based on 153 healthy volunteers of varying age, gender  
143 and ethnic background. It is important to note that the applicability and generalisability of these  
144 reference intervals are subject to the study protocol and local population characteristics.  
145 Nevertheless, our reported reference intervals will add to the literature and understanding of  
146 global coagulation assays.

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## 148 **References**

- 149 1. Lim HY, O'Malley C, Donnan G, Nandurkar H, Ho P. A review of global coagulation  
150 assays — Is there a role in thrombosis risk prediction? *Thromb Res.* 2019;179:45-55.
- 151 2. Ho P, Ng C, Rigano J, et al. Significant age, race and gender differences in global  
152 coagulation assays parameters in the normal population. *Thromb Res.* 2017;154:80-83.
- 153 3. Lim HY, Lui B, Tacey M, et al. Global coagulation assays in healthy controls: are there  
154 compensatory mechanisms within the coagulation system? *J Thromb Thrombolysis.* 2021 Feb  
155 24. Epub ahead of print.

- 156 4. Horowitz G, Altaie S, Boyd J. Defining, establishing, and verifying reference intervals  
157 in the clinical laboratory; approved guideline: EP28-A3c. Wayne, PA: CLSI; 2010.
- 158 5. Reed A, Henry R, Mason W. Influence of statistical method used on the resulting  
159 estimate of normal range. *Clin Chem.* 1971;17:275-284.
- 160 6. Tukey JW. *Exploratory data analysis.* New York: Springer; 1977.
- 161 7. Sheskin D. *Inferential statistical tests employed with two or more dependent samples*  
162 *(and related measures of association/ correlation).* New York, NY: *Handbook of Parametric and*  
163 *Nonparametric Statistical Procedures;* 2004, 4:1021-1116 pp.
- 164 8. Scarpelini S, Rhind S, Nascimento B, et al. Normal range values for  
165 thromboelastography in healthy adult volunteers. *Braz J Med Biol Res.* 2009;42:1210-17.
- 166 9. Devreese K, Walter W, Combes I, Van kerckhoven S, Hoylaerts MF. Thrombin  
167 generation in plasma of healthy adults and children: chromogenic versus fluorogenic  
168 thrombogram analysis. *Thromb Haemost.* 2007;98:600-13.
- 169 10. van Paridon P, Panova-Noeva M, van Oerle R, et al. Thrombin generation in  
170 cardiovascular disease and mortality - results from the Gutenberg Health Study. *Haematologica.*  
171 2019;105:2327-34.
- 172 11. Antovic A. The overall hemostasis potential: a laboratory tool for the investigation of  
173 global hemostasis. *Semin Thromb Hemost.* 2010;36:772-9.
- 174 12. Ress K, Koerbin G, Li L, Chesher D, Bwititi P, Horvath A. Reference intervals for  
175 venous blood gas measurement in adults. . *Clin Chem Lab Med.* 2020;59:947-54.

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## 177 **Table and figure legends**

178

179 **Table 1** Proposed reference intervals for the global coagulation assay parameters

180 **Figure 1** shows the traces of the global coagulation assays. Fig 1A represents the TEG® trace  
181 which includes parameters such as R-time (minutes) = reaction time, time from start of test to  
182 initial clot formation; K-time (minutes) = time taken to achieve a certain level of clot strength;  
183  $\alpha$ -angle ( $^{\circ}$ ) = rate of clot formation; maximum amplitude (MA, mm) = strength of fibrin clot;  
184 LY30 (%) = percentage of decrease in amplitude at 30 minutes after MA (degree of  
185 fibrinolysis). Fig 2B represents the CAT curve which includes parameters such as lag time  
186 (min) = time required to reach one-sixth of the peak height; peak height (nM); endogenous  
187 thrombin potential (ETP, nM.min) = area under the curve or amount of thrombin formation;  
188 velocity index (nM/min) = maximum slope of the initial part of the curve. Fig 1C illustrates the

189 OHP trace which includes overall coagulation potential (OCP, units) derived from the addition  
190 of thrombin only; overall haemostatic potential (OHP, units) derived from the addition of  
191 thrombin and tissue plasminogen activator, and overall fibrinolytic potential (OFP, %) which is  
192 calculated using  $((OCP-OHP/OCP) \times 100\%)$ .

193 **Figure 2** Boxplots showing age and gender differences in the key parameters for TEG®, CAT®  
194 and OHP. (The box component indicates the 25<sup>th</sup> and 75<sup>th</sup> percentiles (interquartile range) with  
195 the median indicated by the line within the box. The whiskers indicate the upper and lower  
196 adjacent values and extreme values outside these adjacent values are indicated by dots.)

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**Table 1** Proposed reference intervals for the global coagulation assay parameters

|   | Manufacturers' reference intervals | Age group (years) | Male                |                    |                    | Female              |                    |                    |
|---|------------------------------------|-------------------|---------------------|--------------------|--------------------|---------------------|--------------------|--------------------|
|   |                                    |                   | Reference intervals | Lower limit 95% CI | Upper limit 95% CI | Reference intervals | Lower limit 95% CI | Upper limit 95% CI |
| <b>Number of volunteers</b>                           |                                    |                   | <b>55</b>           |                    |                    | <b>97</b>           |                    |                    |
| <b>APTT</b>   | 25.0-38.0                          | 20-<80            | <b>22.1-36.6</b>    | 20.8-23.5          | 35.0-38.1          | <b>21.0-34.2</b>    | 20.0-22.0          | 33.0-35.3          |
| <b>PT (sec)</b>                                       | 11.0-17.0                          | 20-<80            | <b>10.3-15.3</b>    | 9.8-10.8           | 14.8-15.7          | <b>9.3-11.9</b>     | 9.1-9.6            | 11.7-12.2          |
| <b>Thromboelastography (TEG®) (n=123)</b>             |                                    |                   |                     |                    |                    |                     |                    |                    |
| <b>Number of volunteers</b>                           |                                    |                   | <b>43</b>           |                    |                    | <b>75</b>           |                    |                    |
| <b>R time (min)</b>                                   | 2.0-8.0                            | 20-<80            | <b>2.3-11.0</b>     | 2.6-3.8            | 12.1-17.4          | <b>2.9-9.3</b>      | 2.4-3.5            | 8.7-9.8            |
| <b>K time (min)</b>                                   | 1.0-3.0                            |                   | <b>1.3-3.8</b>      | 1.0-1.5            | 3.5-4.0            | <b>0.7-3.2</b>      | 0.4-0.9            | 2.9-3.5            |
| <b>a-angle (deg)</b>                                  | 55.0-78.0                          |                   | <b>36.1-75.2</b>    | 31.5-41.2          | 69.8-79.3          | <b>36.5-83.3</b>    | 31.9-41.7          | 79.5-86.3          |
| <b>Maximum amplitude (mm)</b>                         | 51.0-69.0                          |                   | <b>46.7-69.2</b>    | 44.0-49.7          | 66.9-71.6          | <b>49.2-74.2</b>    | 47.1-51.5          | 71.6-76.6          |
| <b>LY 30 (%)</b>                                      | 0.0-8.0                            |                   | <b>0.0-4.9</b>      | 0                  | 3.6-6.1            | <b>0.0-8.6</b>      | 0                  | 6.7-16.7           |
| <b>Calibrated automated thrombogram (CAT) (n=153)</b> |                                    |                   |                     |                    |                    |                     |                    |                    |
| <b>Number of volunteers</b>                           |                                    |                   | <b>55</b>           |                    |                    | <b>98</b>           |                    |                    |
| <b>Lag time (min)</b>                                 | Not available                      | 20-<50            | <b>1.8-4.2*</b>     |                    |                    | 1.7-2.0             | 4.0-4.4            |                    |
|   |                                    | 50-<80            | <b>1.7-5.3*</b>     |                    |                    | 1.3-2.2             | 4.8-5.7            |                    |
| <b>ETP (nM.min)</b>                                   |                                    | 20-<80            | <b>789-1678</b>     | 708-878            | 1580-1766          | <b>861-1912</b>     | 787-932            | 1833-1985          |

|   |               |        |                  |           |             |                   |           |             |
|---|---------------|--------|------------------|-----------|-------------|-------------------|-----------|-------------|
| <b>Velocity Index (nM/min)</b>  |               |        | <b>16.2-147</b>  | 13.1-20.8 | 120.8-177.7 | <b>24.6-215.6</b> | 20.7-30.0 | 184.7-248.6 |
| <b>Thrombin Peak (nM)</b>   |               |        | <b>80-299</b>    | 62-100    | 278-318     | <b>101-373</b>    | 82-121    | 355-391     |
|   |               |        |                  |           |             |                   |           |             |
| <b>Overall haemostatic potential assay (OHP) (n=144)</b>  |               |        |                  |           |             |                   |           |             |
| <b>Number of volunteers</b>   |               |        | <b>33</b>        |           |             | <b>54</b>         |           |             |
| <b>OCP (units)</b>  | Not available | 20-<50 | <b>9.0-43.2</b>  | 4.8-14.1  | 37.3-47.6   | <b>18.0-51.9</b>  | 14.1-21.9 | 47.4-55.9   |
| <b>OHP (units)</b>  |               |        | <b>1.7-11.2</b>  | 1.3-2.1   | 8.6-14.2    | <b>3.1-13.0</b>   | 2.8-3.6   | 11.4-14.8   |
| <b>OFP (%)</b>  |               |        | <b>75.3-92.3</b> | 72.7-77.9 | 90.0-94.7   | <b>73.6-90.1</b>  | 71.8-75.5 | 88.3-91.8   |
| <b>Number of volunteers</b>   |               |        | <b>18</b>        |           |             | <b>39</b>         |           |             |
| <b>OCP (units)</b>  | Not available | 50->80 | <b>20.2-55.7</b> | 14.4-25.5 | 49.7-60.2   | <b>21.2-61.2</b>  | 16.8-27.0 | 56.4-65.3   |
| <b>OHP (units)</b>  |               |        | <b>3.5-13.9</b>  | 2.9-4.4   | 10.6-16.5   | <b>4.0-20.9</b>   | 3.3-5.2   | 17.4-24.8   |
| <b>OFP (%)</b>  |               |        | <b>73.1-88.2</b> | 70.6-75.6 | 85.3-90.3   | <b>64.1-92.3</b>  | 59.9-68.9 | 87.5-96.5   |
|   |               |        |                  |           |             |                   |           |             |
| * Asterisk indicates common reference interval for both male and female; CI = confidence interval; APTT = activated partial thromboplastin time; PT = prothrombin time; ETP = endogenous thrombin potential; OCP = overall coagulation potential; OHP = overall haemostatic potential; OFP = overall fibrinolytic potential |               |        |                  |           |             |                   |           |             |





