



Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Lafouresse, F;Groom, JR

Title:

Friends help make lasting memories

Date:

2018-04-01

Citation:

Lafouresse, F. & Groom, J. R. (2018). Friends help make lasting memories. *Immunology and Cell Biology*, 96 (4), pp.344-346. <https://doi.org/10.1111/imcb.12036>.

Persistent Link:

<https://hdl.handle.net/11343/283872>

DR JOANNA R GROOM (Orcid ID : 0000-0001-5251-7835)

Article type : News and Commentary

ICB NEWS AND COMMENTARY

Title:

Friends help make lasting memories

Strapline

Inflammatory monocytes promote resident memory T cell persistence following lung infection

Fanny Lafouresse^{1,2} and Joanna R. Groom^{1,2*}

¹Walter and Eliza Hall Institute of Medical Research, 1G Royal Parade, Parkville, VIC 3052, Australia

²Department of Medical Biology, University of Melbourne, Parkville, VIC 3010, Australia
Australia

* Corresponding author. Email: groom@wehi.edu.au

Word Count

Text – 1170

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1111/IMCB.12036](https://doi.org/10.1111/IMCB.12036)

This article is protected by copyright. All rights reserved

CD8⁺ Resident memory T cells (Trm) “sound the alarm” to initiate the recruitment of multiple innate and adaptive cells following local, secondary challenge¹. A new study just published in *Immunology & Cell Biology* now demonstrates that Trm may also get a little help in order to be maintained in the periphery².

Partitioning of T cell memory

Following infection, the acquisition and maintenance of T cell memory is essential to confer protection. Pathogen-specific CD8⁺ T cells can differentiate into diverse memory cell subsets¹. These broadly segregate into three distinct populations; effector memory cells (Tem) which circulate throughout the body; central memory cells (Tcm) that primarily locate in lymph nodes, and tissue-resident memory cells (Trm) which are defined by their location in peripheral tissues¹. Given their strategic positioning at sites of pathogen entry, Trm represent the first T cell contact during recall responses and act to coordinate local secondary responses³⁻⁵. Understanding the lineage relationships between the different CD8⁺ T memory cells and factors that govern memory CD8⁺ T cell fate are crucial to develop better therapies against cancer and viral infection as well as to design more efficient vaccination strategies.

In this issue of *Immunology & Cell Biology*, Desai *et al.* use CCR2-deficient mice, to investigate the role of inflammatory monocytes (IMs) on CD8⁺ T cell effector and memory responses in the lung following vaccinia virus infection². IMs were shown to be expanded in the spleen and lungs following infection, and this was dramatically reduced in CCR2-deficient animals. This loss of IMs did not affect the initial differentiation of T cell effectors or the establishment of either circulating or Trm memory cells. However, 50 days following infection, CCR2^{-/-} mice had a specific loss in the CX3CR1⁻CXCR3⁺ memory population. Desai *et al.* used intravenous labeling of CD45 to determine that this compartment comprised of both circulating and resident memory, and that both of these were decreased in CCR2^{-/-} hosts. This analysis also permitted discrimination of other Tem and Trm populations within the lung. CXCR1^{int} memory CD8⁺ T cells have the highest self-renewal capacity and are the predominant subset in peripheral tissues^{4, 6}. Desai *et al.* show that this population also consists of both patrolling and resident T cell memory, however these were unaffected by loss of CCR2. How unique populations of Trm individually contribute to secondary

N&C Friends help make lasting memories

responses is not well understood. This current study provides a new model to dissect individual roles of Trm populations and how they contribute or compensate to immune protection.

The helpers of Trm differentiation and maintenance

Recently, cDC1 dendritic cells, which specifically express Clec9A (DNDR-1) were shown to be required for Trm priming and differentiation⁷. Clec9A-mediated cross presentation is specifically required for Trm priming within draining LNs. Interestingly, these interactions were not essential for formation of circulating T cell memory⁷. During T cell priming, CD4⁺ T cell help also assists Trm differentiation⁸. In the absence of CD4⁺ T cells help, CD8⁺ T cells express high levels of T-bet, which hinders Trm differentiation. Further, the lower T-bet levels expressed in the presence of help, in turn induce expression of CD103 on Trm, facilitating their correct localization in the lung^{8,9}. Finally, once formed, neutrophils provide a trail of CXCL12 which leads memory CD8 cells to the lung¹⁰. This current study adds IMs to the team of cellular players that assist the differentiation, trafficking, positioning, and maintenance of Trm (Figure 1).

How might IMs influence Trm persistence?

The expression of CXCR3 on the CX3CR1⁻ Trm population is of interest for their maintenance as memory CD8⁺ T cells are recruited into nasal-associated lymphoid tissue (NALT) through CXCR3 signaling¹¹. It is tempting to speculate that without specific cell contacts or cytokines keeping them sequestered in the lung, CX3CR1⁻CXCR3⁺ Trm may migrate to into alternative locations, such as the NALT.

The precise mechanism by which IMs regulate the persistence of Trm in the lung warrants further investigation. Desai *et al.* show that IM cell numbers decrease in the lung by day 20 following infection. This timing is prior to Trm contraction in CCR2^{-/-} mice, suggesting direct interaction between IM and Trm is not required. However, as the CX3CR1⁻ compartment was comprised of both circulating and resident memory, IMs may co-localize with Trm at other sites which then replenish lung Trm. Indeed, others have shown co-localization between IMs and intestinal Trm influences their survival and persistence¹². Further, adoptive transfer experiments of CCR2⁺Ly6C⁺ monocytes show that this population can differentiate into DCs or macrophages in the inflamed lung¹³. Therefore, it may be either of these cell types, which were not identified by Desai *et al.* that promote Trm persistence. The visualization of the distinct memory populations within their niche would greatly inform

memory CD8⁺ T cell subsets biology and clarify the requirement for cell contact and the cellular players that drive retention. As Desai *et al.* study suggests, the helpers of Trm are likely different for each Trm subtype and different again at unique peripheral sites.

IMs may also influence the local inflammatory cues that are required for Trm maintenance. Several studies have shown the cytokines IL-7, IL-15, IL-18 and TGFβ are important for Trm long-term persistence^{1,9,12,14}. Of these, IM can rapidly produce bioactive IL-18 and IL-15 following infection, although how much is produced at steady state or following pathogen clearance is unknown¹⁵. Further, as IL-15 is required for initial Trm priming, if IMs were the cellular source at this time, the phenotype of CCR2-deficient mice would be more pronounced early in infection^{7,9}.

While further studies are needed to elucidate the mechanisms by which IMs influence Trm maintenance, this intriguing study by Desai *et al.* highlights the importance of cell collaboration for long-term persistence of memory.

REFERENCES

1. Mueller SN, Mackay LK. Tissue-resident memory T cells: local specialists in immune defence. *Nat Rev Immunol* 2016; **16**: 79-89.
2. Desai P, Tahiliani V, Stanfield J, *et al.* Inflammatory monocytes contribute to the persistence of CXCR3^{hi}CX3CR1^{lo} circulating and lung-resident memory CD8⁺ T cells following respiratory virus infection. *Immunol Cell Biol* 2018; **96**: doi: 10.1111/imcb.12006 XXX-XXX ADD PAGE NOS AT ISSUE PAGING STAGE
3. Beura LK, Mitchell JS, Thompson EA, *et al.* Intravital mucosal imaging of CD8⁺ resident memory T cells shows tissue-autonomous recall responses that amplify secondary memory. *Nat Immunol* 2018; **19**: 173-182.
4. Gilchuk P, Hill TM, Guy C, *et al.* A distinct lung-interstitium-resident memory CD8⁺ T cell subset confers enhanced protection to lower respiratory tract infection. *Cell Rep* 2016; **16**: 1800-1809.

5. Schenkel JM, Fraser KA, Beura LK, *et al.* T cell memory. Resident memory CD8 T cells trigger protective innate and adaptive immune responses. *Science* 2014; **346**(6205): 98-101.
6. Gerlach C, Moseman EA, Loughhead SM, *et al.* The chemokine receptor CX3CR1 defines three antigen-experienced CD8 T cell subsets with distinct roles in immune surveillance and homeostasis. *Immunity* 2016; **45**(6): 1270-1284.
7. Iborra S, Martinez-Lopez M, Khouili SC, *et al.* Optimal generation of tissue-resident but not circulating memory T cells during viral infection requires crosspriming by DNCR-1⁺ dendritic cells. *Immunity* 2016; **45**: 847-860.
8. Laidlaw BJ, Zhang N, Marshall HD, *et al.* CD4⁺ T cell help guides formation of CD103⁺ lung-resident memory CD8⁺ T cells during influenza viral infection. *Immunity* 2014; **41**: 633-645.
9. Mackay LK, Wynne-Jones E, Freestone D, *et al.* T-box transcription factors combine with the cytokines TGF-beta and IL-15 to control tissue-resident memory T cell fate. *Immunity* 2015; **43**: 1101-1111.
10. Lim K, Hyun YM, Lambert-Emo K, *et al.* Neutrophil trails guide influenza-specific CD8⁺ T cells in the airways. *Science* 2015; **349**(6252): aaa4352.
11. Pizzolla A, Wang Z, Groom JR, *et al.* Nasal-associated lymphoid tissues (NALTs) support the recall but not priming of influenza virus-specific cytotoxic T cells. *Proc Natl Acad Sci USA* 2017; **114**: 5225-5230.
12. Bergsbaken T, Bevan MJ, Fink PJ. Local inflammatory cues regulate differentiation and persistence of CD8⁺ tissue-resident memory T cells. *Cell Rep* 2017; **19**: 114-124.
13. Landsman L, Varol C, Jung S. Distinct differentiation potential of blood monocyte subsets in the lung. *J Immunol* 2007; **178**: 2000-2007.

14. Adachi T, Kobayashi T, Sugihara E, *et al.* Hair follicle-derived IL-7 and IL-15 mediate skin-resident memory T cell homeostasis and lymphoma. *Nat Med* 2015; **21**: 1272-1279.
15. Soudja SM, Ruiz AL, Marie JC, *et al.* Inflammatory monocytes activate memory CD8⁺ T and innate NK lymphocytes independent of cognate antigen during microbial pathogen invasion. *Immunity* 2012; **37**: 549-562.

Figure 1. The friends of Trm priming, recruitment and maintenance. (a) In draining LNs, antigens are cross presented to Trm via Clec9a by cDC1s in the presence of CD4⁺ T cells. (b) This induces expression of CD103 allowing recruitment to peripheral tissues. Neutrophils lay a trail of CXCL12 to promote T cell recruitment into the lung. (c) Within the lung inflammatory monocytes (IM) promote the retention of CX3CR1- Trm potentially through direct contact or cytokine secretion. Thus, multiple cells work together to establish and maintain Trm cells.

