



Minerva Access is the Institutional Repository of The University of Melbourne

Author/s:

Wilkinson, DP;Golding, N;Guillera-Arroita, G;Tingley, R;McCarthy, MA

Title:

Defining and evaluating predictions of joint species distribution models

Date:

2021-03-01

Citation:

Wilkinson, D. P., Golding, N., Guillera-Arroita, G., Tingley, R. & McCarthy, M. A. (2021). Defining and evaluating predictions of joint species distribution models. *Methods in Ecology and Evolution*, 12 (3), pp.394-404. <https://doi.org/10.1111/2041-210X.13518>.

Persistent Link:

<https://hdl.handle.net/11343/337967>

1

2 MR DAVID PETER WILKINSON (Orcid ID : 0000-0002-9560-6499)

3 DR NICK GOLDING (Orcid ID : 0000-0001-8916-5570)

4 DR GURUTZETA GUILLERA-ARROITA (Orcid ID : 0000-0002-8387-5739)

5

6

7 Article type : Research Article

8

9

10 Handling editor: Professor Robert Freckleton

11

12 **Running title:** Predictions for joint species distribution models13 **Title:** Defining and evaluating predictions of joint species distribution models14 David P. Wilkinson^{1*}, Nick Golding^{1,2,3}, Gurutzeta Guillera-Arroita¹, Reid Tingley⁴, Michael
15 A. McCarthy¹

16

17 1. School of BioSciences, University of Melbourne, Parkville, 3010, Victoria, Australia

18 2. Telethon Kids Institute, Perth Children's Hospital, 15 Hospital Ave, Nedlands, 6009,
19 Western Australia, Australia

20 3. Curtin University, Kent St, Bentley, 6102, Western Australia, Australia

21 4. School of Biological Sciences, Monash University, Clayton, 3800, Victoria, Australia

22 *Corresponding author: dwilkinson@unimelb.edu.au

This is the author manuscript accepted for publication and has undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record](#). Please cite this article as [doi: 10.1111/2041-210X.13518](https://doi.org/10.1111/2041-210X.13518)

This article is protected by copyright. All rights reserved

23

24 Number of words: 6744

25 Number of tables: 1

26 Number of figures: 4

27 Number of references: 45

28

29

30 **Abstract:**

- 31 1. Joint species distribution models (JSDMs) simultaneously model the distributions of
32 multiple species, while accounting for residual co-occurrence patterns. Despite
33 increasing adoption of JSDMs in the literature, the question of how to define and
34 evaluate JSDM predictions has only begun to be explored.
- 35 2. We define four different JSDM prediction types that correspond to different aspects of
36 species distribution and community assemblage processes. *Marginal* predictions are
37 environment-only predictions akin to predictions from single-species models; *joint*
38 predictions simultaneously predict entire community assemblages; and *conditional*
39 *marginal* and *conditional joint* predictions are made at the species- or assemblage-
40 level, conditional on the known occurrence state of one or more species at a site. We
41 define five different classes of metrics that can be used to evaluate these types of
42 predictions: threshold-dependent, threshold-independent, community dissimilarity,
43 species richness, and likelihood metrics.
- 44 3. We illustrate different prediction types and evaluation metrics using a case study in
45 which we fit a JSDM to a frog occurrence dataset collected in Melbourne, Australia.
- 46 4. JSDMs present opportunities to investigate facets of species distribution and
47 community assemblage processes that are not possible to explore with single-species
48 models. We show that there are a variety of different metrics available to evaluate
49 JSDM predictions, and that choice of prediction type and evaluation metric should
50 closely match the questions being investigated.

51 **Keywords:** joint species distribution models, prediction, evaluation metrics, biotic
52 interactions, community assemblage, species richness

54 1. Introduction

55 Species distribution models (SDMs) are commonly used to predict into un-sampled areas or
56 to new environmental conditions (Elith & Leathwick 2009). Joint species distribution models
57 (JSDMs) are an extension of standard correlative SDMs that allow multiple species to be
58 modelled simultaneously while accounting for species correlations not explained by available
59 environmental predictors, e.g. due to species interactions or important missing covariates
60 (Kissling *et al.* 2012; Pollock *et al.* 2014; Golding, Nunn & Purse 2015; Ovaskainen *et al.*
61 2016b; Clark *et al.* 2017). Despite increasing adoption of JSDMs in the literature, it remains
62 unclear how predictions of JSDMs differ from those of standard SDMs, and how specifically
63 JSDM predictions can be used to address different questions in ecology and conservation
64 (Ovaskainen *et al.* 2016a; Zhang *et al.* 2018; but see: Norberg *et al.* 2019).

65 Modelling distributions of single species with correlative SDMs ignores the impacts of
66 species interactions, which potentially biases estimated coefficients and resultant predictions
67 (Kissling *et al.* 2012; Wisz *et al.* 2013). There have been attempts to account for biotic
68 interactions by using other species' occurrence states as predictor variables alongside abiotic
69 variables (Leathwick & Austin 2001; Araújo & Luoto 2007; Meier *et al.* 2010; Pellissier *et al.*
70 2010), or by constraining predicted distributions to observed or predicted distributions of
71 species on which the target depends (Schweiger *et al.* 2012), but these approaches are
72 restricted to unidirectional interactions (Kissling *et al.* 2012) and require that distributions of
73 the non-target species are known *a priori* or estimated using analogous single-species SDMs.
74 Stacked species distribution models (SSDMs) combine, or stack, multiple single-species
75 SDMs to estimate community structure and species richness (Gelfand *et al.* 2005; Parviainen
76 *et al.* 2009; Mateo *et al.* 2012). Yet, it has been suggested that, because they do not account
77 for species interactions, SSDMs tend to over-predict species richness (Pineda & Lobo 2009;
78 Guisan & Rahbek 2011; Calabrese *et al.* 2014; Thuiller *et al.* 2015). By accounting for
79 interactions between multiple species, the expectation is that JSDMs might allow for more
80 accurate predictions. The captured interactions might be true biotic interactions, or reflect the
81 effect of relevant missing predictors.

82 An important decision when aiming to obtain predictions from a JSDM is defining what to
83 predict, based on the JSDM's multivariate output. In a single-species SDM, the focus of
84 prediction is clear: some feature of the target species (presence/absence, abundance, etc).

85 Multivariate JSDMs, however, have multiple response variables (one per species), all of
86 which are potentially correlated. Prediction can therefore be approached in several ways. For
87 instance, we may aim to predict community composition at sites where we have no
88 knowledge of distributions, or, we may be interested in exploring how having data on the
89 distributions of some species changes our prediction for a focal species.

90 Once we have a prediction, we need to evaluate it. Even ill-fitting models can generate
91 predictions, and poor predictions can hamper species management. More broadly, by
92 evaluating predictions we can use predictive performance to assess which combinations of
93 environmental variables best predict species distributions, compare performance between
94 different modelling approaches, and assess the reliability of the predictions we generate
95 (Guisan & Zimmermann 2000; Lawson *et al.* 2014). How do we approach this for JSDMs?
96 The substantial literature on the evaluation of SDM predictions (Fielding & Bell 1997; Liu,
97 White & Newell 2009; Lawson *et al.* 2014) covers a wide variety of metrics, but are they
98 appropriate in a multi-species context? Are the most common metrics used for single-species
99 SDMs (such as AUC) still the most relevant? JSDMs can predict community assemblages,
100 which opens up a suite of potential evaluation metrics in the form of the dissimilarity indices
101 widely used in community ecology (Legendre & De Cáceres 2013). What insight can be
102 gained from these additional metrics?

103 Our article outlines different types of prediction from JSDMs. Marginal predictions are
104 environment-only predictions that average over the occurrence and co-occurrence patterns of
105 other species, and would be the most familiar to users of single-species SDMs. Joint
106 predictions simultaneously predict the occurrence of multiple species while accounting for
107 environmental responses and species correlations. Both prediction types (marginal and joint)
108 can be calculated conditional on the known occurrence state(s) of one or more species in the
109 community. We outline which evaluation methods are appropriate for different prediction
110 types. Finally, we fit a standard JSDM to a frog occurrence dataset from Melbourne,
111 Australia. Our case study is not intended to authoritatively compare JSDM performance,
112 which would require multiple datasets and modelling methods, but rather to provide practical
113 examples of the different types of JSDM predictions and evaluation metrics.

114 **2. Materials and Methods:**

115 *2.1. Joint, marginal and conditional probabilities*

116 Probability theory defines the relationships between the joint, conditional and marginal
117 probabilities. Considering the presence/absence of two species at a single site, let A denote
118 presence of species A and B denote presence of species B (and conversely A' and B' would
119 denote the absence of species A and B respectively). The joint probability of both species
120 being present is:

$$\begin{aligned}\Pr(A,B) &= \Pr(A|B) \cdot \Pr(B) \\ &= \Pr(B|A) \cdot \Pr(A)\end{aligned}\tag{1}$$

121 where $\Pr(A|B)$ is the (conditional) probability that species A is present given species B is
122 present, and $\Pr(A)$ is the marginal probability of species A being present.

123 The two-species scenario can be extended to more species to consider the relationship
124 between the joint probability of occurrence and the probability of presences (or absences) of a
125 subset of the species conditional on the presence (or absence) of the complement of species.
126 In a four-species scenario:

$$\begin{aligned}\Pr(A,B,C,D) &= \Pr(A,B|C,D) \cdot \Pr(C,D) \\ &= \Pr(A|B,C,D) \cdot \Pr(B,C,D)\end{aligned}\tag{2}$$

127 Here we define $\Pr(A,B|C,D)$ as a joint conditional probability (the probability of the joint
128 occurrence of species A and B at the site, conditional on the presence of both species C and
129 D). $\Pr(A|B,C,D)$ is a conditional probability (the probability of the occurrence of species A at
130 the site conditional on the presence of species B, C and D).

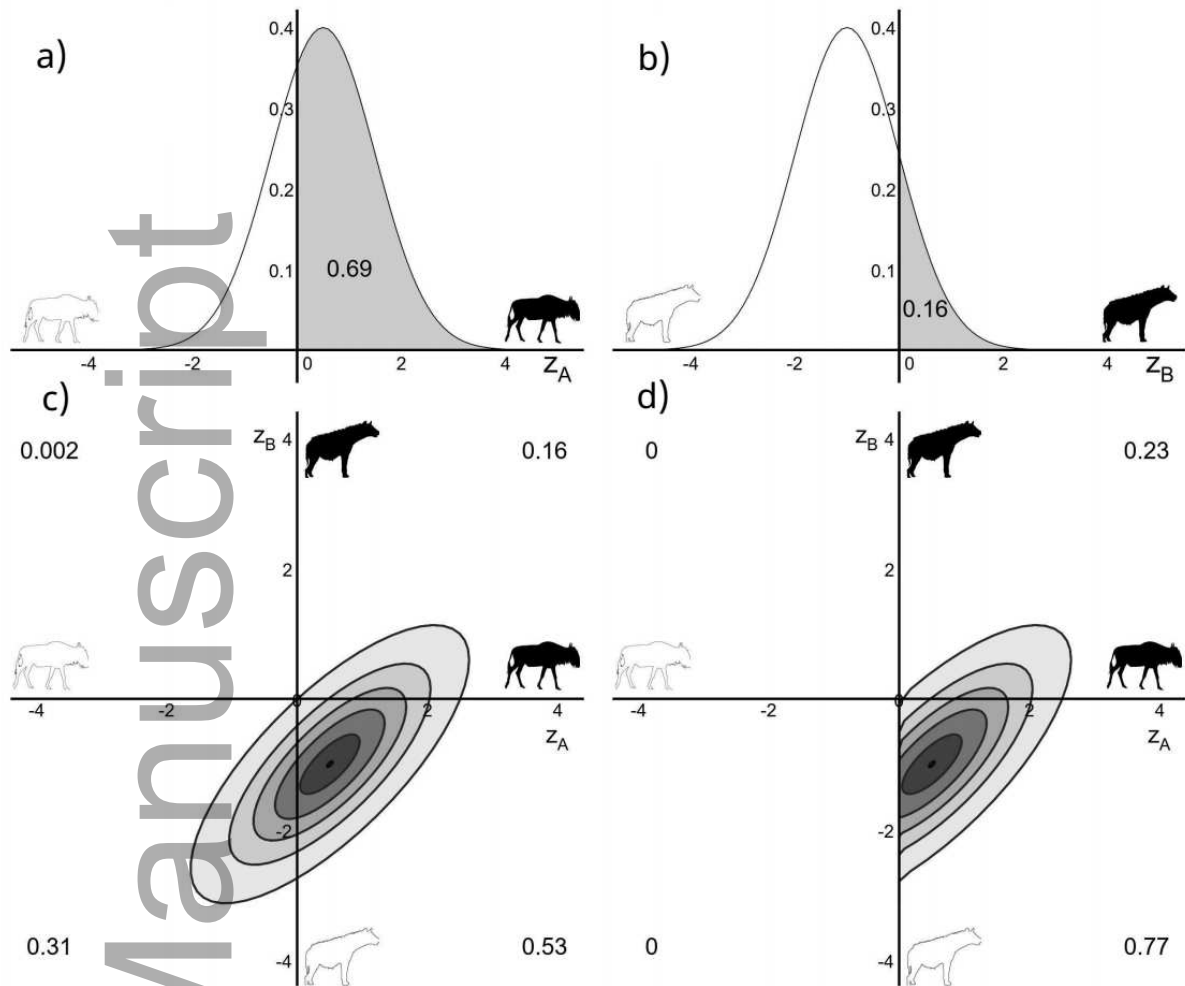
131 2.2. Joint SDMs

132 Most occurrence-based JSDMs are built on the foundation of the Chib and Greenberg (1998)
133 multivariate probit regression model. In what follows, we focus on the multivariate probit
134 model formulation, though the same principles can be applied to the multivariate logistic and
135 latent factor models that have also been used as JSDMs. For this model, the occurrence state
136 (present or absent) of species j , for $j = 1, \dots, J$, at a site i , for $i = 1, \dots, n$, is y_{ij} and modelled via
137 a normally-distributed latent variable, z_{ij} , with y_{ij} equal to 1 when $z_{ij} > 0$, and 0 otherwise
138 (see Figure 1a for a visual representation). This latent variable is not to be confused with the
139 latent variable/latent factor concept of latent factor models (Warton *et al.* 2015; Wilkinson *et*
140 *al.* 2019). The model is as follows:

141

$$\begin{aligned}
 y_{ij} &= 1(z_{ij} > 0) \\
 z_{ij} &= \mu_{ij} + e_{ij} \\
 \mu_{ij} &= \mathbf{X}_{i,\cdot} \boldsymbol{\beta}_{\cdot,j} \\
 \mathbf{e}_i &\sim MVN(\mathbf{0}, \mathbf{R})
 \end{aligned}
 \tag{3}$$

143 where the latent variable, z_{ij} , is the sum of the linear predictor, μ_{ij} , and the correlated residual
 144 error, e_{ij} . The linear predictor is the product of the measured environmental variables $\mathbf{X}_{i,\cdot}$, and
 145 their corresponding regression coefficients $\boldsymbol{\beta}_{\cdot,j}$, as in standard generalised linear models.
 146 Correlations in the residual error \mathbf{e}_i are captured in \mathbf{R} , a symmetric and positive-definite
 147 matrix; its diagonal elements are 1 and its off-diagonal elements – the residual correlations
 148 between species – are restricted between -1 and 1. The elements of \mathbf{R} reflect species co-
 149 occurrence patterns not described by the environmental predictors (i.e. species interactions, or
 150 missing predictors). Standard deviations, and in turn variances, are constrained to equal 1 in
 151 probit regression, thus covariance and correlation matrices are equivalent. Because the
 152 variance of the latent variable distribution remains constant, the probability of presence for a
 153 single species is controlled only by the mean value of the distribution, μ (compare Figures 1a
 154 and 1b).



155

156 Figure 1: Visualisations of different normally-distributed latent variables representing species
 157 probabilities of presence for two species (A and B). a) The normally distributed latent
 158 variable, z_A , with mean, μ_A , of 0.5 for species A. $\Pr(A)$ is equal to the area under the curve
 159 where $z_A > 0$, shown here in grey. b) The normally distributed latent variable, z_B , with mean,
 160 μ_B , of -1 for species B. $\Pr(B)$ is equal to the area under the curve where $z_B > 0$, shown here
 161 in grey. c) The multivariate, normally-distributed latent variable, z_{ij} , for a two species
 162 scenario. The mean of the distribution, μ_{ij} , on each species' respective axis is the same as
 163 their independent distributions in a) and b) and there is positive correlation of 0.75 between
 164 them. The contours of the probability distribution, the grey ellipses, indicate probability
 165 density values of 0.1, 0.3, 0.5, 0.7, and 0.9. The numbers in the four corners are the
 166 probabilities of the multivariate latent variable integrated in that quadrant, e.g. in the upper
 167 right quadrant; there is a probability of 0.16 that both species will occur at the site. d) The
 168 multivariate, normally-distributed latent variable from c) truncated on the known occurrence
 169 state of species A.

170 The correlation in e_i makes the space of the latent variables multivariate. Therefore, the
 171 whole species community at a site is represented by a multi-dimensional latent normal
 172 random variable with as many dimensions as species. Integration over the relevant portion of
 173 the multivariate latent variable space yields the *joint* probability of observing a given
 174 configuration of presence/absence of all species at a site. For instance, in the two-species
 175 scenario (Figure 1c), the joint probability $\Pr(A,B)$ that both species A and B are present is:

$$\Pr(z_A > 0, z_B > 0) = \int_0^{\infty} \int_0^{\infty} f(z_A, z_B) dz_A dz_B \quad (4)$$

176 Where $f(z_A, z_B)$ is the joint probability distribution for the bivariate normal. The double
 177 integral calculates the probability in the region where both z_A and z_B are greater than zero
 178 (i.e. both species are present). In this two-species case, this probability can be visualised as
 179 the volume under the corresponding region of a three-dimensional surface (as in Figure 1c).

180 Similar to Equation 1, a joint probability distribution can be written as a function of
 181 conditional and marginal density functions; therefore, the joint probability in Equation 4 can
 182 be rewritten as follows:

$$\begin{aligned} \Pr(A,B) &= \int_0^{\infty} \int_0^{\infty} f(z_A|z_B) \cdot f(z_B) dz_A dz_B \\ &= \int_0^{\infty} \int_0^{\infty} f(z_B|z_A) \cdot f(z_A) dz_B dz_A \end{aligned} \quad (5)$$

184 Here the conditional probability distribution $f(z_A|z_B)$ reflects how likely different values of
 185 z_A are (and therefore $\Pr(A)$ given a fixed value of z_B). The marginal probability distribution f
 186 (z_B) reflects how likely different values of z_B are (and therefore $\Pr(B)$) *independent* of the
 187 occurrence of species A). The order of items in the joint probability is inconsequential, so
 188 species can be marginalised out in any order (lines 1 and 2 in Equation 5 are equivalent).

189 The marginal probability is obtained by summing the probabilities for all community
 190 assemblages in which the species is present, regardless of the presence of the other species; in
 191 our two-species scenario $\Pr(A) = \Pr(A|B) + \Pr(A|B')$. Computing the *conditional*
 192 probability of presence of a species given the presence (or absence) of another species
 193 involves restricting the probability space to that indicating presence (or absence) of the other

194 species. These probability distributions can be combined in different ways to define a number
 195 of prediction types for JSDMs. We next consider four types of JSDM prediction, mapping
 196 onto different ecological questions. These are illustrated in Figure 2.

197

198 *2.3. Prediction types*

	A	B	C	Prediction Type	Notation
i	?			Marginal	$\Pr(A)$
ii	?	?	?	Joint	$\Pr(A, B, C)$
iii	?	1	0	Conditional	$\Pr(A B, C')$
iv	?	?	0	Conditional joint	$\Pr(A, B C')$
v	?	1		Conditional marginal	$\Pr(A B)$

199

200 Figure 2: Five different prediction types possible with JSDMs. The boxes show the
 201 occurrence states of species *A*, *B*, and *C*. Question marks denote the species being predicted.
 202 Empty boxes indicate that the occurrence state for those species is not informing the
 203 prediction, while 1/0 denote a known presence/absence state of species which is used to
 204 inform prediction. *Prediction Type* is the name of the prediction methodology, and *Notation*
 205 shows the corresponding probabilistic notation, following the definitions in section 2.1.

206

207 **2.3.1. Marginal prediction**

208 Similar to single-species SDM predictions, marginal JSDM predictions are based solely on
 209 environmental attributes (they do not consider the presence or absence of the other species,
 210 see Figure 2i). This corresponds to predicting, for example, the occurrence of plant species *A*
 211 based solely on its response to environmental variables, such as soil nutrient and water

212 availability, without accounting for co-occurrence patterns with species B-E in the
213 community (e.g., Ovaskainen et al 2016a, Zhang et al. 2018). Marginal JSDM predictions can
214 be calculated using the estimated regression coefficients and the corresponding covariate
215 values at each site, as in a standard generalised linear model. The sole difference between
216 marginal predictions for JSDMs and those of single-species SDMs is that inter-species
217 correlations are accounted for in the estimation of JSDM regression coefficients.

218 2.3.2. Joint prediction

219 We can predict species community composition that accounts for both the environmental
220 covariates *and* species co-occurrence, by using the joint probability distribution. This
221 corresponds to predicting the assemblage of plant species A-E in a community
222 simultaneously, while accounting for their individual responses to environmental conditions
223 (e.g. soil nutrients) and co-occurrence patterns of the species (e.g., Ovaskainen *et al* 2016a,
224 Norberg *et al* 2019).

225 Whereas the marginal prediction of a community at a given site can be represented by a
226 single vector of probabilities (one for each species), the joint prediction instead yields a
227 probability value for each possible realisation of the community composition (each
228 realisation is a vector of 1s and 0s indicating presence/absence of each species; Figure 2ii).
229 With J species, the number of possible community assemblages is 2^J . This number increases
230 very quickly with J . Whilst with three species there are eight possible assemblages, with ten
231 species there are 1024, and with twenty species over a million. It is therefore generally
232 infeasible to compute and store the probabilities of all possible assemblages when evaluating
233 the predictions. One alternative is to use the model to simulate community assemblages by
234 taking random draws from the joint probability distribution. The frequencies of the simulated
235 assemblages reflect the probability of plausible assemblages. Also, in some cases we are only
236 interested in a subset of the probabilities. For instance, for model evaluation purposes, we
237 only need to compute joint predictions for the species assemblages observed in held-out data.

238

239 2.3.3. Conditional joint prediction

240 In some cases, we might be interested in estimating a species' occurrence probability given
241 the known occurrence state of other species. If we know the correlation between species
242 (estimated by the JSDM) and the occurrence state of some of those species, we can make a

243 more informed prediction of the unknown occurrence state of the remaining species. This
244 corresponds to predicting plant species A-D in the community simultaneously when we know
245 the occurrence state of species E, and leveraging that knowledge to constrain the possible
246 predicted assemblages (e.g., Taylor-Rodriguez *et al* (2017)).

247 For each species with a known occurrence state, we can truncate the multivariate normal
248 distribution over the latent variable in the dimension that represents that species, either to be
249 positive if the species is present, or negative if it is absent (see Figure 1d for a visual
250 representation). Because probability distributions integrate to one, truncating this distribution
251 alters the probabilities of the remaining community assemblage possibilities. For instance, in
252 our example, the marginal probability of species B, $\Pr(B)$, is 0.16 (Figure 1b), but if we
253 know that species A is present, then the conditional probability of species B is $\Pr(B | A)$
254 = 0.23 (Figure 1d).

255 2.3.4. Conditional marginal prediction

256 Conditional marginal predictions, which are simultaneously conditional on and marginal to
257 the occurrence states of the other species (Figure 2v), are another type of prediction that can
258 be made with JSDMs. As per conditional joint predictions, we can make more informed
259 predictions by using the known occurrence state of other species, however, this method
260 makes use of the updated marginal distribution of the remaining species. This corresponds to
261 predicting plant species A based on its response to environmental variables (e.g., soil
262 nutrients), leveraging information on the known occurrence state of species E, and still being
263 independent of the remaining species in the community B-D. This prediction type is also
264 simple to represent; like marginal predictions, it can be stored as a vector of probabilities for
265 each species.

266 2.4. Evaluation metrics

267 To date, the choice of evaluation metric in the multi-species context of JSDMs has not been
268 thoroughly explored. In a single-species context, choice of metric depends largely on factors
269 such as data type (e.g., presence-absence vs. presence-only) and prediction format (binary or
270 probabilistic) (Lawson *et al.* 2014). Here we consider traditional single-species metrics for
271 JSDMs and new metrics that may be suitable. We broadly classify metrics for evaluating
272

273 JSDM predictions into five groups, in terms of the aspects of performance on which they
274 focus (see Table 1).

275 **Threshold-independent metrics** evaluate continuous predicted probabilities against
276 observed presence-absence data. A widely used threshold-independent metric used in single-
277 species SDMs is the Area Under the Receiver Operating Characteristic Curve (AUC), but
278 other metrics include root mean square error (RMSE), the coefficient of determination (R^2),
279 and the Pearson's or point-biserial correlation coefficient.

280 **Threshold-dependent metrics** compare binary predictions against observed presence-
281 absence data. This requires simplifying predicted values (which may be probabilities or some
282 other metric) to binary outcomes, considering them presences if they exceed a set threshold
283 value, or absences otherwise. Then, a confusion matrix contrasts observed and predicted
284 occurrence states, and metrics derived from it. Examples include precision, sensitivity, and
285 true/false positive/negative rates. The value of thresholding continuous predictions has been
286 debated in the SDM literature (Liu *et al.* 2005; Freeman & Moisen 2008; Guillera-Aroita *et*
287 *al.* 2015), for prediction and evaluation (Lawson *et al.* 2014). How to determine the threshold
288 value when binary conversion is wanted is also debated. It is common to set the threshold at
289 an arbitrary value of 0.5 (Freeman & Moisen 2008), which provides an obvious decision
290 threshold (i.e. the species is more likely to be present than absent) when predictions are
291 calibrated estimates of probability of presence. Another frequent suggestion is to set the
292 threshold to the observed prevalence of the species in question (Hanberry & He 2013). In the
293 multi-species context of JSDBMs, a logical extension of this debate is whether to define
294 community-wide or species-specific thresholds. However, Lawson *et al.* (2014) showed that
295 by using a probabilistic confusion matrix we can calculate threshold-dependent types of
296 metrics without the need to threshold probabilistic predictions. To avoid any issues with the
297 choice of threshold impacting our analysis, we followed this idea and used the probabilistic
298 confusion matrix approach when calculating our threshold-dependent metrics.

299 **Community dissimilarity indices** are widely used in community ecology to quantify the
300 dissimilarity between two realisations of species assemblages. Examples of common metrics
301 are Bray-Curtis dissimilarity and Jaccard distance (Chao *et al.* 2004). These metrics compare
302 predicted assemblages with observed assemblages and thus probabilistic predictions require
303 thresholding or samples from a binomial distribution before these metrics can be evaluated on
304 them.

305 **Species richness metrics** consider the ability of models to predict a single (but widely
 306 studied) aspect of community composition – the number of species present. We have
 307 evaluated species richness difference, defined as predicted minus observed richness, as it
 308 directly compares predictions with observed richness values.

309 **Likelihood metrics** assess model fit by computing the probability of observing a given
 310 community assemblage, assuming the model is ‘correct’ in its structure and parameter
 311 estimates. For reasons of numerical stability, it is common to work with the log of the
 312 likelihood. We use the term independent log-likelihood to represent the typical log-likelihood
 313 metric used in SSDMs. This metric assesses each species individually across all sites–
 314 computing the probability of observing that species’ presence/absence observations– and then
 315 combines these into a single metric, assuming the species’ distributions to be independent
 316 (i.e. the log-likelihoods are summed up). We can also define a joint log-likelihood that
 317 assesses all species simultaneously as an assemblage at each site, accounting for the
 318 correlation structure encoded in the JSDM formulation.

319

320 Table 1: Summary of evaluation metrics for JSDM predictions.

Name	Applicable to Binary predictions	Applicable to Probabilistic predictions	Threshold-dependent metric	Threshold-independent metric	Community dissimilarity metric	Species richness metric	Likelihood metric
Accuracy / True Skill Statistic	X	X	X				
Area under the Receiver Operating Characteristic curve (AUC)		X		X			
Bray-Curtis dissimilarity	X	X			X		
Canberra Index	X	X			X		
Cohen’s Kappa	X		X				
Diagnostic odds ratio	X	X	X				
F ₁ score	X	X	X				
False discovery ratio	X	X	X				

False negative rate	X	X	X			
False omission rate	X	X	X			
False positive rate	X	X	X			
Gower Index	X	X			X	
Gower Index (alternative)	X	X			X	
Jaccard distance	X	X			X	
Kendall rank correlation coefficient		X		X		
Kulczynski Index	X	X			X	
Log Likelihood – Independent		X				X
Log Likelihood – Joint		X				X
Mountford Index	X	X			X	
Mean error (bias)		X		X		
Mean square error		X		X		
Negative likelihood ratio	X	X	X			
Negative predictive performance	X	X	X			
Pearson correlation coefficient		X		X		
Positive likelihood ratio	X	X	X			
Positive predictive performance / Precision	X	X	X			
R ² / Coefficient of determination	X	X		X		
Raup-Crick dissimilarity	X	X	X			
Root mean square error		X		X		
Spearman rank correlation coefficient		X		X		
Species richness difference	X	X			X	
Sum of squared errors		X		X		
True negative rate / Specificity	X	X	X			
True positive rate / Sensitivity	X	X	X			
Youden's J statistic	X	X	X			

321 2.5. Case study

322 We illustrate the application of different JSDM prediction types and the appropriate
323 evaluation metric classes with a case study in which we fit a JSDM to a presence-absence
324 dataset of frog species in the Greater Melbourne area of Victoria, Australia (Parris 2006). The
325 dataset contains 9 species, 104 waterbodies (sites), and 3 measured covariates (area, road
326 density, presence of vertical wall), and was previously analysed in Pollock *et al* (2014) and
327 Wilkinson *et al* (2019). Species prevalence ranged from 0.02-0.52, and were mainly
328 positively correlated with each other except for a single species negatively correlated with the
329 rest. In our analysis, we standardised the two continuous variables and used five-fold random
330 cross validation for model evaluation.

331 We chose to use a standard multivariate probit regression JSDM (Chib& Greenberg 1998)
332 implemented with *BayesComm* v0.1-2 (Golding & Harris 2015) in R v3.5.2 (R Core Team
333 2018). The JSDM applies independent normal priors, $\beta_k \sim N(0, 100)$, on the regression
334 coefficients and an inverse Wishart prior, with $n + 2J$ degrees of freedom and scale matrix **I**,
335 on the correlation coefficients. The model was fit by MCMC using a Gibbs sampler
336 implemented in R and C++. We used a single MCMC chain of 11,000 samples, discarding
337 the first 1,000 as burn-in, to sample the posterior distribution. Model convergence was
338 examined via visual assessment of trace plots for the posterior distributions. Model fitting and
339 predictions were undertaken on The University of Melbourne's Spartan HPC infrastructure
340 (Meade *et al.* 2017).

341 Once fit to the training data, the JSDM was used to predict to the held-out test data from five-
342 fold cross validation. We calculated four prediction types: marginal, conditional marginal,
343 joint, and conditional joint. The two conditional prediction types were undertaken under the
344 assumption that we knew the occurrence state of one randomly selected "high prevalence"
345 species in the community (*Litoria ewingii*). Conditional marginal prediction returned
346 probabilistic predictions for the remaining eight species and conditional joint prediction
347 generated plausible assemblages of the remaining eight species.

348 While we have defined 35 metrics that can be calculated to assess different aspects of JSDM
349 predictions, we present only a subset in this analysis for illustrative purposes. More detail on
350 the metrics, including how they are calculated, which prediction types they are appropriate
351 for, and how to interpret them can be found in Appendix S1. Threshold-dependent and
352 threshold-independent metrics are calculated on a per-species basis, whereas community
353 dissimilarity and species richness metrics are calculated per-site. Most metrics can
354 theoretically be calculated for either species or sites (as they are just comparing two binary
355 vectors: observations and predictions) so this split is based on how they are historically used
356 in ecological literature. Metrics were evaluated once for each of the 1000 posterior samples
357 drawn.

358

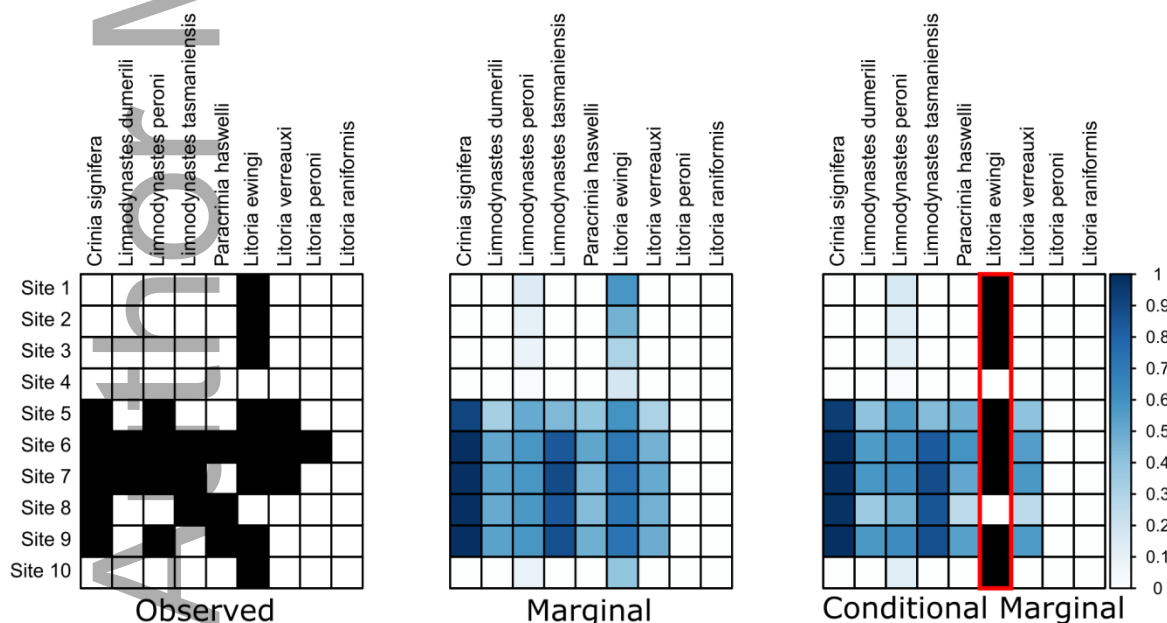
359 **3. Results**

360 The patterns of predicted probabilities of presence for each species obtained from the
361 marginal and conditional marginal predictions both broadly matched the community

362 assemblage observed in the held-out testing data (Figure 3). The AUC for most species using
 363 the marginal predictions was typically high (>0.88) with a standard deviation of <0.05 across
 364 all posterior samples. Exceptions were *L. ewingii* and *Litoria peroni*, which were predicted
 365 approximately randomly with an AUC= ~0.5, and *Litoria raniformis*, which had no recorded
 366 presences in the held-out data and was thus incompatible with AUC calculations. Marginal
 367 predictions conditioned on the known occurrence state of *L. ewingii* returned higher AUC
 368 values by a mean of 0.03 (Fig. 3). The largest AUC gain was 0.08 for *Litoria verreauxi*. For
 369 most other species-level evaluation metrics, we found the conditional marginal prediction to
 370 outperform the marginal prediction, but in most cases the differences were relatively minor.
 371 We did observe some exceptions to this, such as an increased R² value for *L. verreauxi* of
 372 0.26.

373 The conditional marginal predictions outperformed the marginal predictions for the majority
 374 of community-level metrics. For most community dissimilarity metrics, the conditional
 375 marginal predictions returned dissimilarity values between 0.05-0.1 lower than the marginal
 376 predictions which indicates a better estimate of community assemblages. Jaccard distance and
 377 Gower Index improved by 0.09, and Bray-Curtis dissimilarity improved by 0.06.

378



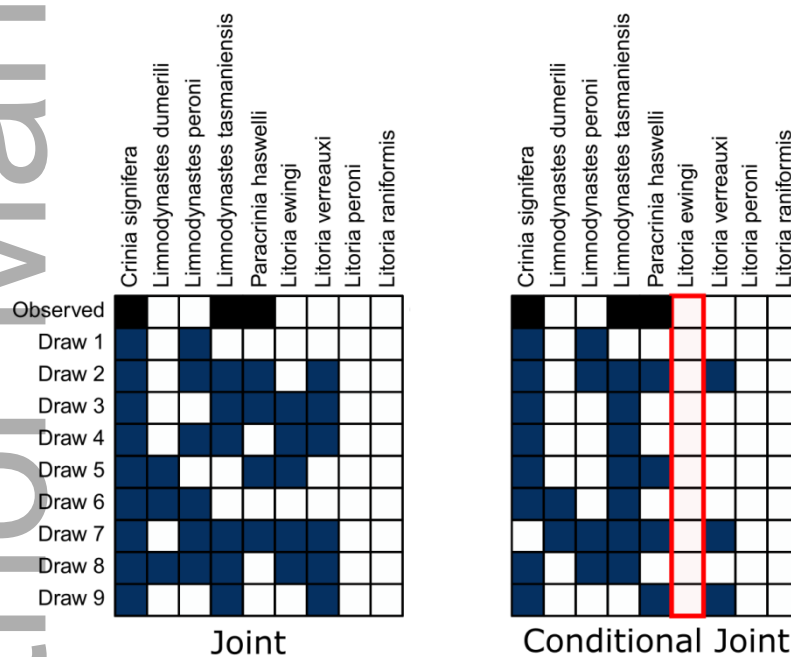
379

380 Figure 3: Species-level predictions for the frog community at a subset of 10 sites in the
 381 testing dataset. The left-hand plot depicts the known occurrence state of the species in the
 382 testing dataset (black = present, white = absent). The middle plot depicts the median

383 probability of presence from the posterior distributions of the marginal predictions. The right-
 384 hand plot depicts the median probability of presence from the posterior distributions of the
 385 conditional marginal predictions. These predictions are conditional on the known occurrence
 386 state of *L. ewingii* which are highlighted with the red border.

387 The community-level approach of joint predictions yields probabilities of specific
 388 assemblages occurring at a site or, as we present here, one can take random draws from the
 389 posterior distribution to generate plausible community assemblages at a site. In Figure 4 we
 390 show random draws from the joint and conditional joint prediction types for a single site in
 391 our held-out data. While joint predictions generally overpredicted the number of species
 392 found in the assemblage overall, conditioning on the known occurrence state (absence) of
 393 *L.ewingii* led to draws that were closer to the observed assemblage.

394



395

396 Figure 4: Binary community-level predictions of the frog community at a single site in the
 397 testing dataset. Community-level predictions here are random draws from the multivariate
 398 normal distribution representing plausible community assemblages under the environmental
 399 conditions at the site (blue = present, white = absent). The left-hand plot depicts draws of
 400 plausible community assemblages from joint predictions. The right-hand plot depicts draws
 401 of plausible community assemblages from joint predictions, conditional on the known
 402 occurrence state of *L. ewingii* at the site (highlighted with the red border). The top row of

403 both plots shows the known community assemblage in the testing dataset. The site considered
404 in this figure corresponds to site 8 in Figure 3.

405 Community dissimilarity metrics were generally between 0.02-0.05 higher for conditional
406 joint predictions relative to standard joint predictions. Raup-Crick dissimilarity was the
407 exception. Conditional joint predictions had values that were, on average, 0.25 lower;
408 however, with a standard deviation of 0.37 across all predictions, this metric is possible
409 struggling to calculate correctly across all draws. For the site considered in Figure 4, we
410 observed improvements of between 0.08-0.1 for the Bray-Curtis dissimilarity, Jaccard
411 distance, Gower index, and Canberra index.

412 As shown in Figure 4, in our dataset, both the joint and conditional joint prediction types
413 overestimated species richness. The joint prediction type had a mean species richness
414 difference estimate of 0.95 species in the held-out data, while the conditional joint prediction
415 type had a mean estimate of 0.7 species per site. By conditioning the community-level
416 prediction on the known occurrence state of a species, we are able to obtain a prediction
417 closer to the observed data.

418 **4. Discussion**

419 We have reviewed and clarified the ways in which predictions of species distributions can be
420 approached with JSDMs. Our review highlights the additional functionality that JSDMs
421 enable relative to simple stacking of single-species models. JSDMs have two main
422 advantages: an ability to partition the effect of measured variables and residual correlations
423 between species, and predictions of community assemblages that account for these
424 correlations. By partitioning the effect of measured variables from residual correlations,
425 which may include species interactions, JSDMs potentially enable more accurate estimates of
426 environmental drivers of species' distributions. In addition, by leveraging the information in
427 the residual correlations, JSDMs can potentially better predict community assemblages.

428 The different prediction types defined here for JSDMs correspond to different ecological
429 aims. Marginal predictions correspond to the traditional single-species predictions; joint
430 predictions predict entire assemblages; while the conditional and conditional marginal
431 predictions let us inform these predictions with additional information such as easy-to-detect
432 indicator species. Different aims also imply different evaluation metrics, so each practitioner

433 should determine an appropriate prediction type and evaluation metric, or metrics, for their
434 ecological question.

435 We have shown that conditioning on the known occurrence status of species can improve
436 both species- and community-level predictions by exploiting the correlations estimated in
437 JSDMs (Harris 2015). The extent of this improvement is likely dependant on the strength of
438 the correlation between the known species and other species in the community. We can
439 expect that a species that is highly correlated with the known species (positively or
440 negatively) would benefit more than a species that occurs more or less independent of the
441 known species. Therefore, benefits of JSDM predictions may be observed for only some
442 species in the community, or to differing extents between species.

443 The JSDM tended to overpredict species richness for all prediction types that account for
444 correlations between species. For our case study of nine frog species, the JSDM predicted
445 approximately one extra species per site. Zurell *et al* (2019) similarly found that a JSDM
446 overpredicted species richness compared to a SSDM. As the JSDM only overpredicted
447 species richness for prediction types that account for residual correlations, the estimated
448 correlations could potentially explain why the JSDM tended to overpredict. A largely positive
449 correlation matrix between most species combined with high marginal occurrence
450 probabilities, as seen here, could cause the JSDM to predict likely assemblages in excess of
451 restrictions, such as site carrying capacities and/or dispersal limitations.

452 We have presented JSDM prediction methods using only a single dataset for illustrative
453 purposes. A more in-depth study using a wider array of datasets is required, as dataset
454 properties likely influence the relative merits of the different prediction types. For datasets in
455 which all species are reasonably prevalent, the expected performance difference between
456 prediction types would be smaller than in scenarios of relatively sparse data, in which the
457 additional information in joint or conditional prediction types could outperform the other
458 prediction types. The value of additional information in the form of species co-occurrence
459 will also be greater when correlations between species are stronger; the known occurrence of
460 a species will provide minimal benefit if it exhibits no correlation with the other species in a
461 dataset. Future studies incorporating multiple case studies could also usefully evaluate
462 different prediction types when making vs extrapolative predictions, as our case study only
463 focused on the former type of evaluation and the latter has been shown to be more difficult
464 (Norberg *et al.* 2019).

465 A small body of literature focuses on large-scale comparisons of JSDMs and single-species
466 models. These studies have included many species and model types (Zurell *et al.* 2019;
467 Norberg *et al.* 2019) but have not addressed all of the prediction types available to JSDMs.
468 Norberg *et al.* (2019) used joint predictions but differently than presented here. First, they
469 generated binary assemblage predictions but took the average of a large number of random
470 draws to generate species-specific probabilities, which is an approximation of the marginal
471 distribution. Second, they used joint predictions to generate species richness metrics
472 equivalent to ours. Third, they used joint predictions and community dissimilarity metrics to
473 test predictions of species turnover between sites rather than to test community composition
474 at a site. To our knowledge, no JSDM studies have yet included conditional or conditional
475 marginal predictions, although they have been identified as desirable avenues of research
476 (Zurell *et al.* 2019; Norberg *et al.* 2019; Blanchet, Cazelles & Gravel 2020). Evaluations with
477 a broader array of JSDM implementations, prediction types, and datasets are warranted. Both
478 Norberg *et al.* (2019) and Zurell *et al.* (2019) included latent factor JSDMs in their
479 comparisons, whereas we use a multivariate probit model. Norberg *et al.* (2019) found that the
480 HMSC JSDM (Ovaskainen *et al.* 2016b) outperformed both other JSDMs and SSDMs, while
481 Zurell *et al.* (2019) found that the *boral* JSDM (Hui 2016) had similar results to those
482 described here. Zurell *et al.* (2019) suggested that the poor performance of *boral* was a result
483 of how the latent factor model is used when extrapolating predictions. Prediction using latent
484 factor JSDMs can be performed in two ways: (1) marginalizing over the latent factors by
485 assigning the mean value of modelled sites to prediction sites when performing regression-
486 style predictions, and (2) defining the latent factor models in the same way as multivariate
487 probit models (see Wilkinson *et al.* (2019) for notation), which lets one use the prediction
488 types defined in this paper. This second method may be a better alternative to latent factor
489 model prediction than marginalizing over the latent factors.

490 5. Conclusion

491 JSDMs enable a variety of different ways to predict species distributions and community
492 assemblages. Here we have defined environment-only marginal predictions, joint predictions
493 for whole community assemblages, and conditional marginal and conditional joint
494 predictions that can also leverage additional information on known species' occurrences.
495 Previous studies have either not considered prediction with JSDMs, focussed on marginal
496 predictions, or considered only limited aspects of joint prediction. We have also shown that
497 there are several classes of evaluation metrics that can be applied to subsets of these

498 predictions. Ecologists seeking to use these prediction methods and evaluation metrics should
499 consider which method and metric are most closely linked to the ecological question they are
500 investigating. A larger comparison of prediction types that considers different JSDM
501 implementations and multiple datasets is required to evaluate general performance of these
502 models and prediction types.

504 **Acknowledgements**

505 We thank Peter Vesk, Brendan Wintle, and Jian Yen for insightful discussions, and Els Van
506 Burm and Kristina Chyn for manuscript reviews. DW is funded by an Australian Government
507 Research Training Program Scholarship. NG, GGA and RT are supported by Australian
508 Research Council (ARC) Discovery Early Career Researcher Awards (DE180100635,
509 DE160100904 and DE170100601).

511 **Conflicts of Interest**

512 No conflicts of interest are declared.

514 **Authors' Contributions**

515 All authors conceived the ideas and methodology. DW implemented the analysis. DW led
516 writing the manuscript but all authors contributed significantly throughout and gave final
517 approval before submission.

519 **Data Accessibility**

- 520 • Script files to replicate the model running, data extraction, analysis, and plot
521 generation are available in an online repository (Wilkinson 2019).
- 522 • An anonymised version of the frog dataset is available in our online repository
523 (Wilkinson 2019). This dataset will allow you to replicate our results without the
524 ability to identify the particulars of the dataset.

525 **Reference List**

This article is protected by copyright. All rights reserved

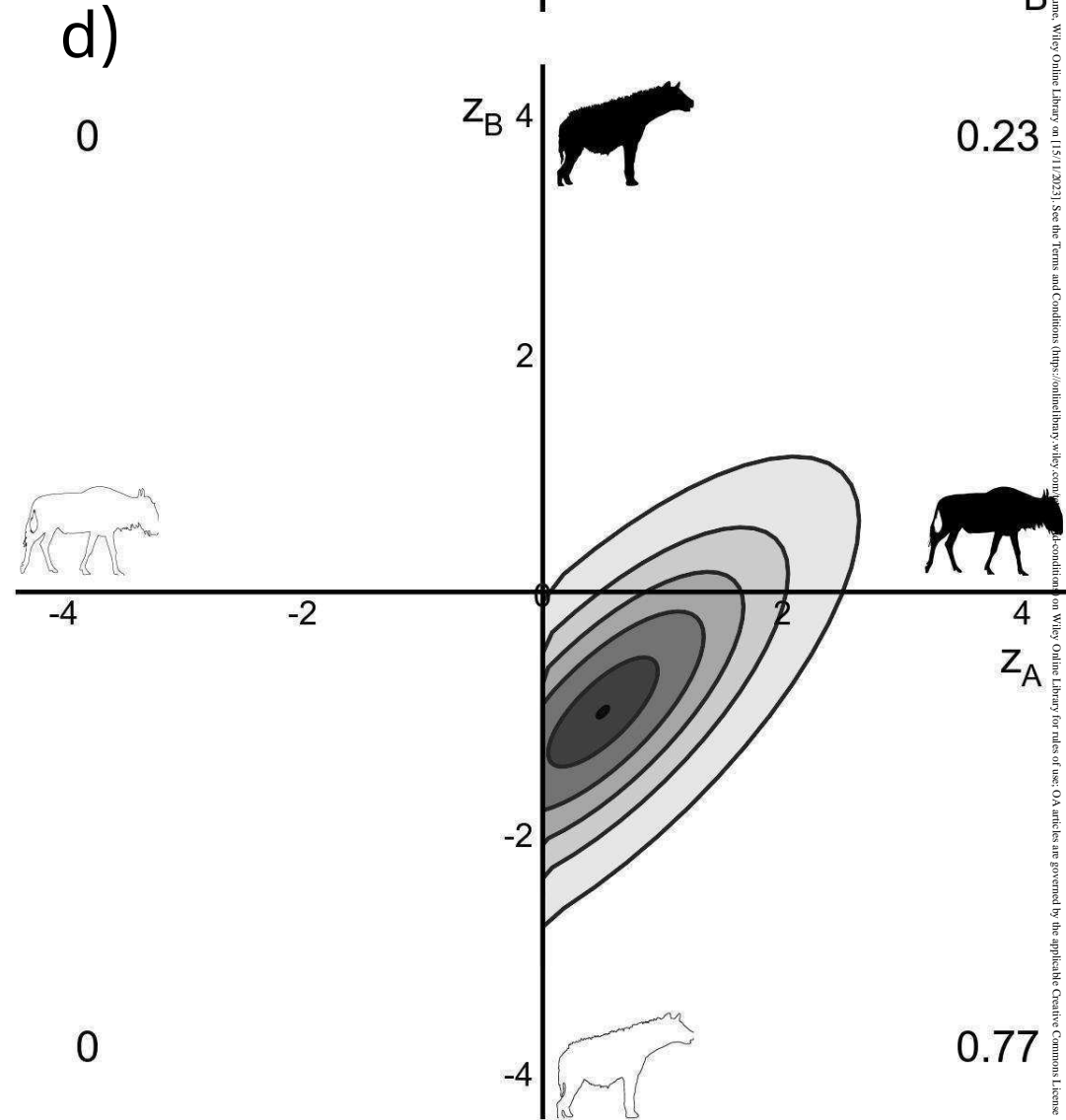
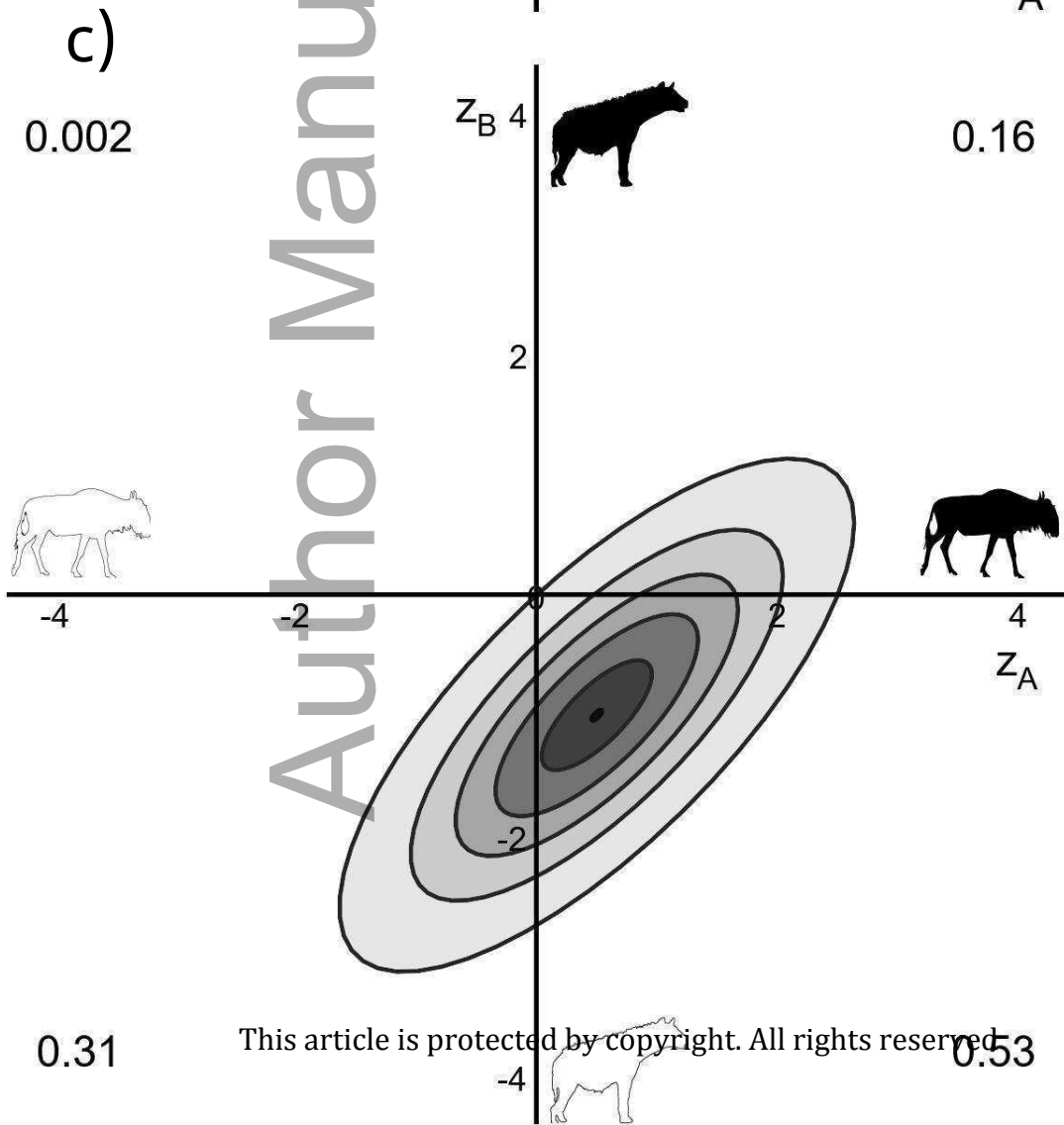
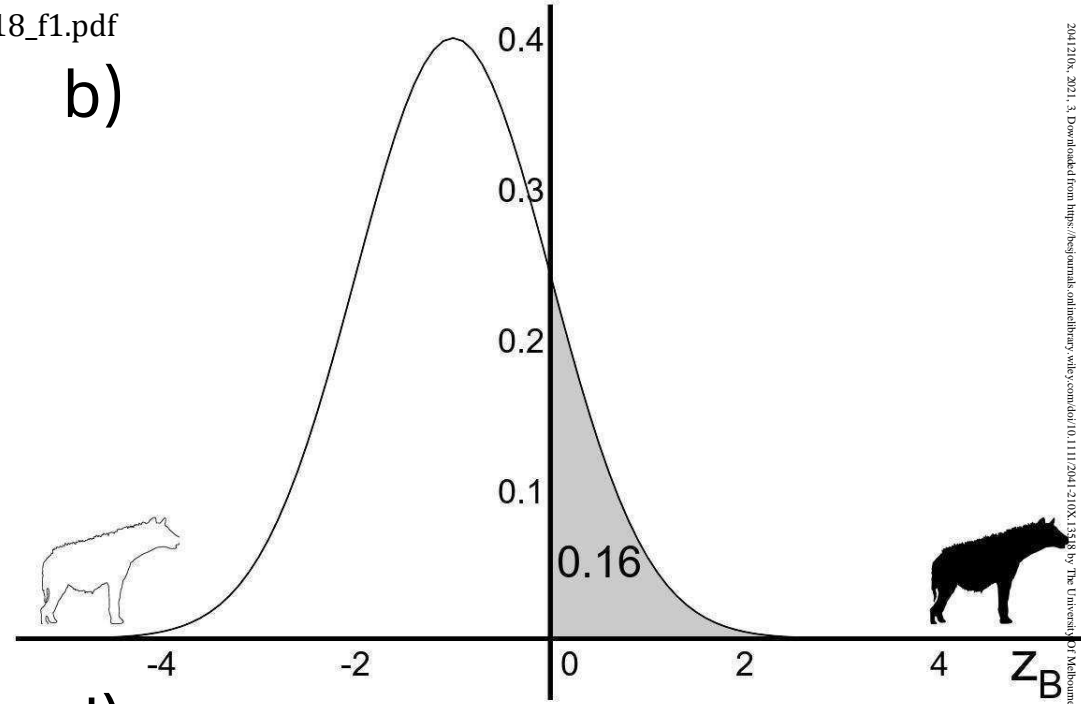
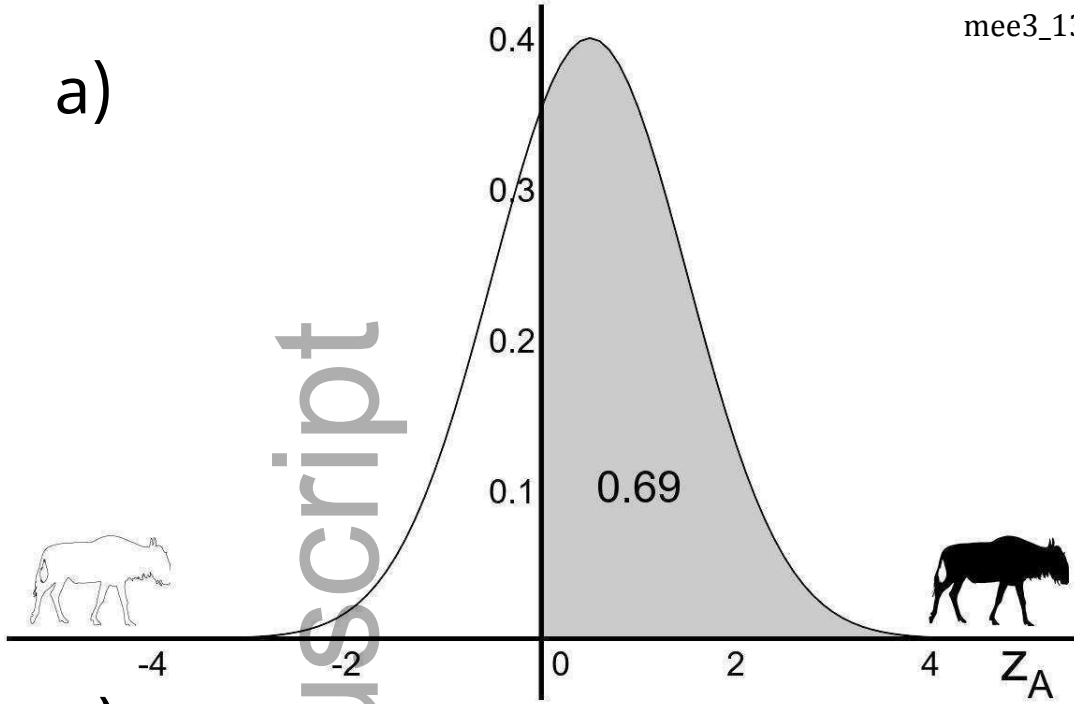
- 526 Araújo, M.B. & Luoto, M. (2007). The importance of biotic interactions for modelling
527 species distributions under climate change. *Global Ecology and Biogeography*, **16**, 743–
528 753.
- 529 Blanchet, F.G., Cazelles, K. & Gravel, D. (2020). Co-occurrence is not evidence of
530 ecological interactions. *Ecology Letters*, **23**, 1050–1063.
- 531 Calabrese, J.M., Certain, G., Kraan, C. & Dormann, C.F. (2014). Stacking species
532 distribution models and adjusting bias by linking them to macroecological models.
533 *Global Ecology and Biogeography*, **23**, 99–112.
- 534 Chao, A., Chazdon, R.L., Colwell, R.K. & Shen, T.-J. (2004). A new statistical approach for
535 assessing similarity of species composition with incidence and abundance data. *Ecology*
536 *Letters*, **8**, 148–159.
- 537 Chib, S. & Greenberg, E. (1998). Analysis of multivariate probit models. *Biometrika*, **85**,
538 347–361.
- 539 Clark, J.S., Nemergut, D., Seyednasrollah, B., Turner, P.J. & Zhang, S. (2017). Generalized
540 joint attribute modeling for biodiversity analysis: median-zero, multivariate, multifarious
541 data. *Ecological Monographs*, **87**, 34–56.
- 542 Elith, J. & Leathwick, J.J.R. (2009). Species Distribution Models: Ecological Explanation
543 and Prediction Across Space and Time. *Annual Review of Ecology, Evolution, ...*, **40**,
544 677–697.
- 545 Fielding, A.H. & Bell, J.F. (1997). A review of methods for the assessment of prediction
546 errors in conservation presence/absence models. *Environmental Conservation*, **24**, 38–
547 49.
- 548 Freeman, E.A. & Moisen, G.G. (2008). A comparison of the performance of threshold criteria
549 for binary classification in terms of predicted prevalence and kappa. *Ecological*
550 *Modelling*, **217**, 48–58.
- 551 Gelfand, A.E., Schmidt, A.M., Wu, S., Silander, J.A., Latimer, A. & Rebelo, A.G. (2005).
552 Modelling species diversity through species level hierarchical modelling. *Journal of the*
553 *Royal Statistical Society: Series C (Applied Statistics)*, **54**, 1–20.
- 554 Golding, N. & Harris, D.J. (2015). BayesComm: Bayesian Community Ecology Analysis.

- 555 Golding, N., Nunn, M.A. & Purse, B. V. (2015). Identifying biotic interactions which drive
556 the spatial distribution of a mosquito community. *Parasites & Vectors*, **8**, 367.
- 557 Guillera-Aroita, G., Lahoz-Monfort, J.J., Elith, J., Gordon, A., Kujala, H., Lentini, P.E.,
558 Mccarthy, M.A., Tingley, R. & Wintle, B.A. (2015). Is my species distribution model fit
559 for purpose? Matching data and models to applications. *Global Ecology and*
560 *Biogeography*, **24**, 276–292.
- 561 Guisan, A. & Rahbek, C. (2011). SESAM - a new framework integrating macroecological
562 and species distribution models for predicting spatio-temporal patterns of species
563 assemblages. *Journal of Biogeography*, **38**, 1433–1444.
- 564 Guisan, A. & Zimmermann, N.E. (2000). Predictive habitat distribution models in ecology.
565 *Ecological Modelling*, **135**, 147–186.
- 566 Hanberry, B.B. & He, H.S. (2013). Prevalence, statistical thresholds, and accuracy
567 assessment for species distribution models. *Web Ecol*, **13**, 13–19.
- 568 Harris, D.J. (2015). Generating realistic assemblages with a joint species distribution model
569 (D. Warton, Ed.). *Methods in Ecology and Evolution*, **6**, 465–473.
- 570 Hui, F.K.C. (2016). boral - Bayesian Ordination and Regression Analysis of Multivariate
571 Abundance Data in r. *Methods in Ecology and Evolution*, **7**, 744–750.
- 572 Kissling, W.D., Dormann, C.F., Groeneveld, J., Hickler, T., Kühn, I., McNerny, G.J.,
573 Montoya, J.M., Römermann, C., Schiffers, K., Schurr, F.M., Singer, A., Svenning, J.-C.,
574 Zimmermann, N.E. & O'Hara, R.B. (2012). Towards novel approaches to modelling
575 biotic interactions in multispecies assemblages at large spatial extents. *Journal of*
576 *Biogeography*, **39**, 2163–2178.
- 577 Lawson, C.R., Hodgson, J.A., Wilson, R.J. & Richards, S.A. (2014). Prevalence, thresholds
578 and the performance of presence-absence models. *Methods in Ecology and Evolution*, **5**,
579 54–64.
- 580 Leathwick, J.R. & Austin, M.P. (2001). Competitive interactions between tree species in New
581 Zealand old-growth indigenous forests. *Ecology*, **82**, 2560–2573.
- 582 Legendre, P. & De Cáceres, M. (2013). Beta diversity as the variance of community data:
583 Dissimilarity coefficients and partitioning. *Ecology Letters*, **16**, 951–963.

- 584 Liu, C., Berry, P.M., Dawson, T.P. & Pearson, R.G. (2005). Selecting thresholds of
585 occurrence in the prediction of species distributions. *Ecography*, **28**, 385–393.
- 586 Liu, C., White, M. & Newell, G. (2009). Measuring the accuracy of species distribution
587 models: A review. *18th World IMACS Congress and MODSIM09 International*
588 *Congress on Modelling and Simulation: Interfacing Modelling and Simulation with*
589 *Mathematical and Computational Sciences, Proceedings*, 4241–4247.
- 590 Mateo, R.G., Felicísimo, Á.M., Pottier, J., Guisan, A. & Muñoz, J. (2012). Do stacked
591 species distribution models reflect altitudinal diversity patterns? *PLoS ONE*, **7**.
- 592 Meade, B., Lafayette, L., Sauter, G. & Tosello, D. (2017). Spartan HPC-Cloud Hybrid:
593 Delivering Performance and Flexibility.
- 594 Meier, E.S., Kienast, F., Pearman, P.B., Svenning, J.-C., Thuiller, W., Araújo, M.B., Guisan,
595 A. & Zimmermann, N.E. (2010). Biotic and abiotic variables show little redundancy in
596 explaining tree species distributions. *Ecography*, **33**, 1038–1048.
- 597 Norberg, A., Abrego, N., Blanchet, F.G., Adler, F.R., Anderson, B.J., Anttila, J., Araújo,
598 M.B., Dallas, T., Dunson, D., Elith, J., Foster, S.D., Fox, R., Franklin, J., Godsoe, W.,
599 Guisan, A., O'Hara, B., Hill, N.A., Holt, R.D., Hui, F.K.C., Husby, M., Kålås, J.A.,
600 Lehtinen, A., Luoto, M., Mod, H.K., Newell, G., Renner, I., Roslin, T., Soinen, J.,
601 Thuiller, W., Vanhatalo, J., Warton, D., White, M., Zimmermann, N.E., Gravel, D. &
602 Ovaskainen, O. (2019). A comprehensive evaluation of predictive performance of 33
603 species distribution models at species and community levels. *Ecological Monographs*,
604 **89**, 1–24.
- 605 Ovaskainen, O., Abrego, N., Halme, P. & Dunson, D. (2016a). Using latent variable models
606 to identify large networks of species-to-species associations at different spatial scales.
607 *Methods in Ecology and Evolution*, **7**, 549–555.
- 608 Ovaskainen, O., Roy, D.B., Fox, R. & Anderson, B.J. (2016b). Uncovering hidden spatial
609 structure in species communities with spatially explicit joint species distribution models.
610 *Methods in Ecology and Evolution*, **7**, 428–436.
- 611 Parris, K.M. (2006). Urban amphibian assemblages as metacommunities. *Journal of Animal*
612 *Ecology*, **75**, 757–764.

- 613 Parviainen, M., Marmion, M., Luoto, M., Thuiller, W. & Heikkinen, R.K. (2009). Using
614 summed individual species models and state-of-the-art modelling techniques to identify
615 threatened plant species hotspots. *Biological Conservation*, **142**, 2501–2509.
- 616 Pellissier, L., Anne Bråthen, K., Pottier, J., Randin, C.F., Vittoz, P., Dubuis, A., Yoccoz,
617 N.G., Alm, T., Zimmermann, N.E. & Guisan, A. (2010). Species distribution models
618 reveal apparent competitive and facilitative effects of a dominant species on the
619 distribution of tundra plants. *Ecography*, **33**, 1004–1014.
- 620 Pineda, E. & Lobo, J.M. (2009). Assessing the accuracy of species distribution models to
621 predict amphibian species richness patterns. *Journal of Animal Ecology*, **78**, 182–190.
- 622 Pollock, L.J., Tingley, R., Morris, W.K., Golding, N., O’Hara, R.B., Parris, K.M., Vesk, P.A.
623 & McCarthy, M.A. (2014). Understanding co-occurrence by modelling species
624 simultaneously with a Joint Species Distribution Model (JSDM) (J. McPherson, Ed.).
625 *Methods in Ecology and Evolution*, **5**, 397–406.
- 626 R Core Team. (2018). R: A Language and Environment for Statistical Computing.
- 627 Schweiger, O., Heikkinen, R.K., Harpke, A., Hickler, T., Klotz, S., Kudrna, O., Kühn, I.,
628 Pöyry, J. & Settele, J. (2012). Increasing range mismatching of interacting species under
629 global change is related to their ecological characteristics. *Global Ecology and*
630 *Biogeography*, **21**, 88–99.
- 631 Taylor-Rodríguez, D., Kaufeld, K., Schliep, E.M., Clark, J.S. & Gelfand, A.E. (2017). Joint
632 Species Distribution Modeling: Dimension Reduction Using Dirichlet Processes.
633 *Bayesian Analysis*, **12**, 939–967.
- 634 Thuiller, W., Pollock, L.J., Gueguen, M. & Münkemüller, T. (2015). From species
635 distributions to meta-communities. *Ecology Letters*, **18**, 1321–1328.
- 636 Warton, D.I., Blanchet, F.G., O’Hara, R.B., Ovaskainen, O., Taskinen, S., Walker, S.C. &
637 Hui, F.K.C. (2015). So Many Variables: Joint Modeling in Community Ecology. *Trends*
638 *in Ecology and Evolution*, **30**, 766–779.
- 639 Wilkinson, D.P. (2019). JSDM_Prediction v0.1.0. Zenodo,
640 <https://dx.doi.org/10.5281/zenodo.3514766>.
- 641 Wilkinson, D.P., Golding, N., Guillera-Arroita, G., Tingley, R. & McCarthy, M.A. (2019). A

- 642 comparison of joint species distribution models for presence-absence data. *Methods in*
643 *Ecology and Evolution*, **10**, 198–211.
- 644 Wisz, M.S., Pottier, J., Kissling, W.D., Pellissier, L., Lenoir, J., Damgaard, C.F., Dormann,
645 C.F., Forchhammer, M.C., Grytnes, J.A., Guisan, A., Heikkinen, R.K., Høye, T.T.,
646 Kühn, I., Luoto, M., Maiorano, L., Nilsson, M.C., Normand, S., Öckinger, E., Schmidt,
647 N.M., Termansen, M., Timmermann, A., Wardle, D.A., Aastrup, P. & Svenning, J.C.
648 (2013). The role of biotic interactions in shaping distributions and realised assemblages
649 of species: Implications for species distribution modelling. *Biological Reviews*, **88**, 15–
650 30.
- 651 Zhang, C., Chen, Y., Xu, B., Xue, Y. & Ren, Y. (2018). Comparing the prediction of joint
652 species distribution models with respect to characteristics of sampling data. *Ecography*,
653 **41**, 1876–1887.
- 654 Zurell, D., Zimmermann, N.E., Gross, H., Baltensweiler, A., Sattler, T. & Wüest, R.O.
655 (2019). Testing species assemblage predictions from stacked and joint species
656 distribution models. *Journal of Biogeography*, **00**, 1–13.
- 657

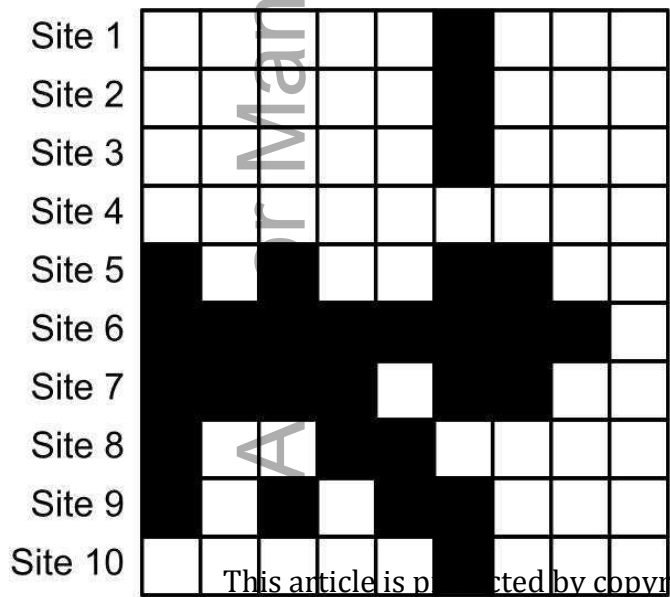


Prediction TypeNotation

	A	B	C		
i	?			Marginal	$\Pr(A)$
ii	?	?	?	Joint	$\Pr(A, B, C)$
iii	?	1	0	Conditional	$\Pr(A \mid B, C')$
iv	?	?	0	Conditional joint	$\Pr(A, B \mid C')$
v	?	1		Conditional marginal	$\Pr(A \mid B)$

This article is protected by copyright. All rights reserved

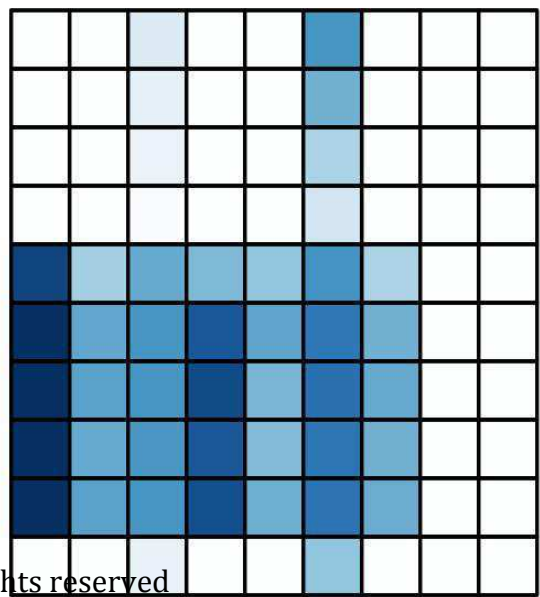
Crinia signifera
 Limnodynastes dumerilii
 Limnodynastes peroni
 Limnodynastes tasmaniensis
 Paracrinia haswelli
 Litoria ewingi
 Litoria verreauxi
 Litoria peroni
 Litoria raniformis



This article is protected by copyright. All rights reserved

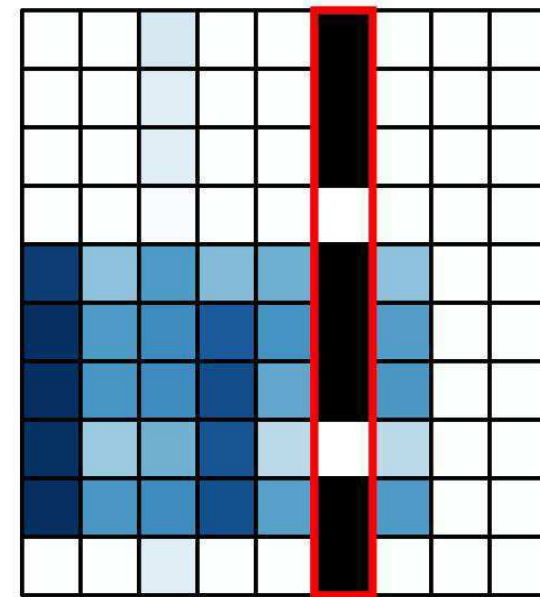
Observed

Crinia signifera
 Limnodynastes dumerilii
 Limnodynastes peroni
 Limnodynastes tasmaniensis
 Paracrinia haswelli
 Litoria ewingi
 Litoria verreauxi
 Litoria peroni
 Litoria raniformis

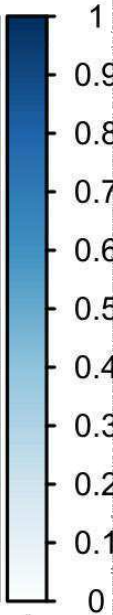


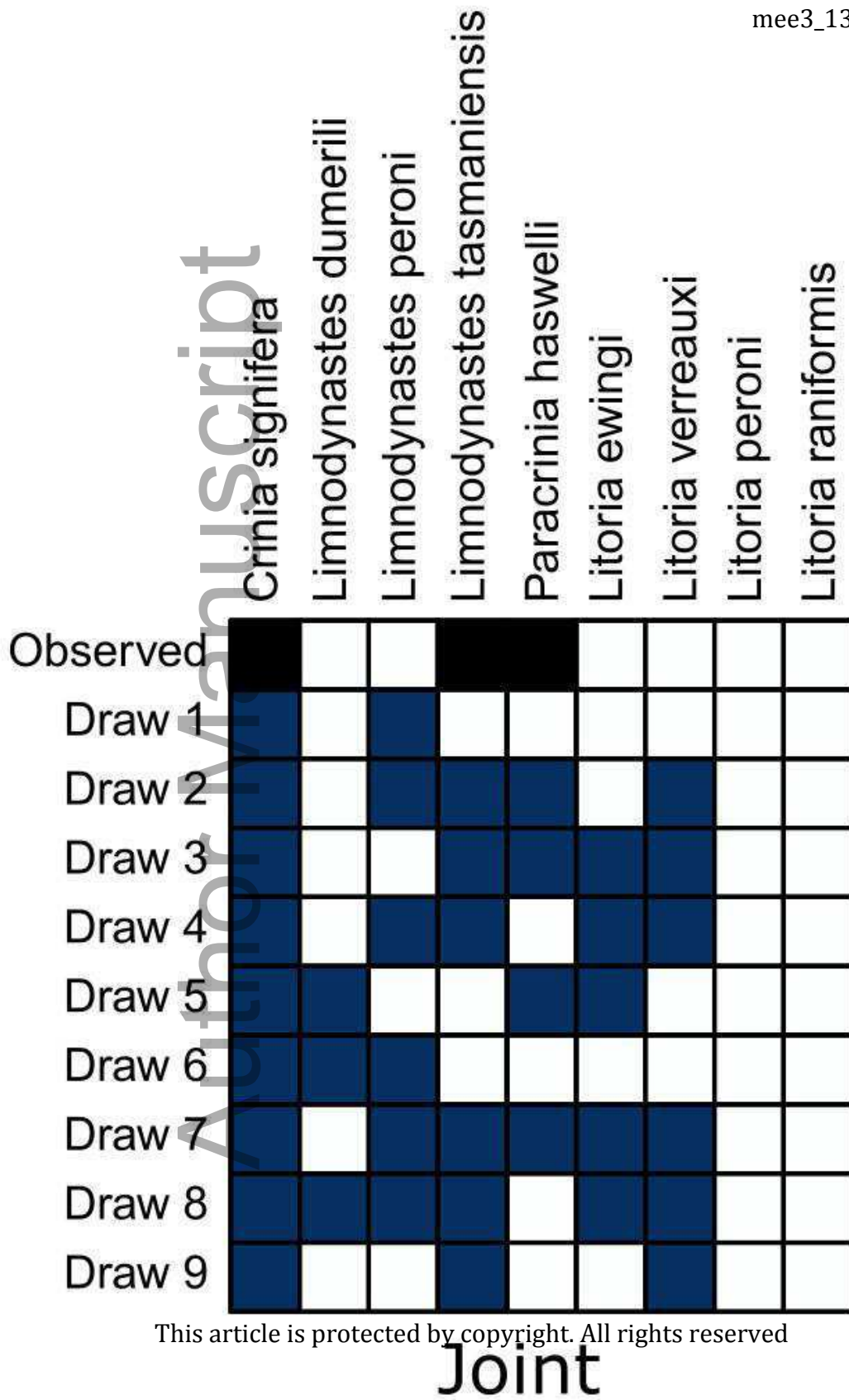
Marginal

Crinia signifera
 Limnodynastes dumerilii
 Limnodynastes peroni
 Limnodynastes tasmaniensis
 Paracrinia haswelli
 Litoria ewingi
 Litoria verreauxi
 Litoria peroni
 Litoria raniformis



Conditional Marginal





This article is protected by copyright. All rights reserved

