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Author/s:

Leong, TSH;Martin, GJO;Ashokkumar, M

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# 1 **Ultrasonic Encapsulation – A Review**

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3 Thomas S. H. Leong<sup>1,2</sup>, Gregory J. O. Martin<sup>2</sup>, Muthupandian Ashokkumar<sup>1\*</sup>

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5 <sup>1</sup> School of Chemistry, The University of Melbourne, Parkville, Victoria 3010, Australia

6 <sup>2</sup> Department of Chemical & Biomolecular Engineering, The University of Melbourne,  
7 Parkville, Victoria 3010, Australia

## 8 **Highlights**

- 9       • Ultrasonic preparation of encapsulated functional materials is reviewed.
- 10       • Mechanisms responsible for expression of functional properties are described.
- 11       • Promising applications for ultrasonically synthesized materials are identified.

12 **Keywords:** ultrasound, encapsulation, functional foods, bioproducts, emulsions,  
13 sonoprocessing, responsive polymers

14  
15 \*corresponding author

## 18 **Abstract**

19 Encapsulation of materials in particles dispersed in water has many applications in nutritional  
20 foods, imaging, energy production and therapeutic/diagnostic medicine. Ultrasonic technology  
21 has been proven effective at creating encapsulating particles and droplets with specific physical  
22 and functional properties. Examples include highly stable emulsions, functional polymeric  
23 particles with environmental sensitivity, and microspheres for encapsulating drugs for targeted  
24 delivery. This article provides an overview of the primary mechanisms arising from ultrasonics  
25 responsible for the formation of these materials, highlighting examples that show promise  
26 particularly in the development of food and bioproducts.

## 27 **1. Introduction**

28 Ultrasonics is a versatile technology with proven effectiveness to create a range of catalytic  
29 and functional materials that have applications across a multitude of fields including food [1],  
30 imaging [2], energy production [3] and therapeutic/diagnostic medicine [4]. The primary  
31 mechanism responsible for the creation of these materials is known as acoustic cavitation,  
32 which is the formation and collapse of bubbles influenced by ultrasound [5]. Ultrasound's  
33 versatility is owed in part to its broad active frequency region that can be tuned and applied  
34 specifically to control the intensity and number of cavitation events. These can be used to  
35 control aspects of materials such as particle size, surface roughness and structure.

36 Ultrasound can be used to promote the internalization of materials through a process known as  
37 encapsulation. The motivation for encapsulation is to protect, prolong or stabilize the  
38 internalised material from environmental deterioration and enables pharmaceuticals and/or  
39 nutrients to be delivered with enhanced efficacy in biological systems. These delivery systems  
40 take a number of different forms. A simple example found in foods is an emulsion [6]. The  
41 dispersion of an oil within water enables effective loading of oil soluble nutrients into aqueous  
42 food media, and is a useful strategy for preparing functional food products. Another example  
43 is the use of environmentally sensitive polymers to create core-shell structures that can be used  
44 to entrap materials such as drugs [7]. These polymer delivery agents respond to changes in pH,  
45 temperature or other external stimuli such that they release entrapped drug material only under  
46 specific conditions, thereby prolonging drug efficacy. These polymers can be synthetic or  
47 natural, such as proteins. Microspheres and microcapsules made from proteins have the  
48 advantage of being bio-compatible and bio-degradable, and have been extensively studied for  
49 pharmaceutical applications [8].

50 Whilst there are a number of reviews [4, 6, 7] covering the formation of different types of  
51 encapsulated materials, there has yet to be a review that brings together these different systems  
52 with details on how to effectively create them using ultrasonics. This review will provide a  
53 guide towards the application of ultrasound to promote encapsulation of materials, focusing on  
54 examples of relevance to the food, biomaterial and pharmaceutical industries.

## 55 **2. Theory of applied ultrasound**

56 This section provides an overview of the principles of ultrasound. A focus is made on the key  
57 physical and chemical effects of ultrasound in aqueous systems to provide background for the

58 subsequent discussion on the application of ultrasound-promoted encapsulation in aqueous  
59 systems.

## 60 *2.1 Characteristics of ultrasound*

61 Ultrasound is generally defined as sound at frequencies above 16 kHz. It is (generally) not  
62 audible when transmitted through the air. When sustained through a liquid medium (e.g. water),  
63 the ultrasonic pressure oscillations may cause in-phase expansion and contraction of the  
64 dissolved gas bubbles i.e. the bubble expands during the negative pressure cycle and contracts  
65 during the positive pressure cycle. This bubble oscillation is accompanied by diffusion of  
66 gas/vapour in and out of the bubble during the expansion and contraction respectively. The  
67 diffusion of gas in and out of the bubble is not equal [9] and under certain conditions, i.e.  
68 oscillation driven above a certain threshold pressure, the diffusion process can result in net  
69 accumulation of mass within the bubble over time. This results in net bubble growth and is  
70 known as rectified diffusion, a process unique to bubbles oscillating within a sound field. In a  
71 field containing multiple bubbles, the interaction of bubbles by collisions combining to form a  
72 larger bubble can also result in what is known as coalescence, and is another source of net  
73 bubble growth in an acoustic sound field [10].

74 Both rectified diffusion and coalescence cause bubble growth. Bubbles within a sound field  
75 will grow in size until they reach what is known as the bubble resonance size range, at which  
76 point they collapse. This formation, growth and collapse of a bubble due to the influence of  
77 ultrasound, is known as acoustic cavitation [5].

78 Of interest to material synthesis, the collapsing bubbles produce localised regions of extreme  
79 temperature and intense physical shearing. Bubbles driven at low ultrasonic frequency (~ 20-  
80 100 kHz), may collapse extremely violently, releasing sufficient energy to produce  
81 temperatures up to 10,000 K within the bubble core and pressures of several hundred  
82 atmospheres within a few hundred micron of the bubble collapse point [11]. This can lead to  
83 the formation of highly reactive radicals which can be used to promote chemical reactions. In  
84 water for example, hydrogen and hydroxyl radicals can be formed due to the splitting of the  
85 water molecule by pyrolysis. Both the physical shear and radical formation can be beneficially  
86 exploited to create materials with a range of desired functionality.

87 The ultrasound frequency regime ranges from 16 kHz to 500 MHz, although the frequency  
 88 range most suitable for processing fluids is typically between 16-3000 kHz. When ultrasound  
 89 is applied to fluids the cavitation effects are highly dependent on the frequency. The intensity  
 90 of bubble collapse (i.e. amount of energy released) and the maximum bubble size prior to  
 91 collapse (resonance size) are correlated and approximately inversely proportional to the applied  
 92 frequency [12] (see Figure 1).

93 A simple relationship that can relate the resonance size of the bubble with the frequency is  
 94 given by:

$$95 \quad F \times R \approx 3 \quad (1)$$

96 where  $F$  is the frequency in Hz and  $R$  is the bubble radius in m. Note that this equation gives  
 97 only a very approximate theoretical resonance size and that there are other factors which may  
 98 control the resonance size of the bubble [5, 13].

99 A more accurate version of (1) is the linear resonance radius which can be calculated using  
 100 the following equation [13]:

101

$$102 \quad R_r = \sqrt{\frac{3\gamma p_\infty}{\rho_L \omega^2}} \quad (2)$$

103 where  $\gamma$  is the specific heat ratio of the gas inside the bubble,  $p_\infty$  is the ambient liquid  
 104 pressure,  $\rho_L$  is the liquid density and  $\omega$  is the angular frequency of ultrasound (all in SI  
 105 units). In practice, the size for an active bubble is usually smaller than this radius due to the  
 106 nonlinear nature of the bubble pulsation [14].

107 Ultrasound can be categorized into several different regions along the frequency spectrum. The  
 108 *power ultrasound* region [12] spans the low frequency range between 16 – 100 kHz. It is  
 109 characterized by large bubble resonance sizes followed by intense bubble collapse, often  
 110 resulting in extremely strong physical effects including localized shear and high temperatures.  
 111 This category of ultrasound delivers high energy density in the order of 10-1000 W/cm<sup>2</sup>. Power  
 112 ultrasound is often selected for material processing and to some extent in synthesis, owing to  
 113 its strong physical shear and intense local temperature effects. Examples of processes in this

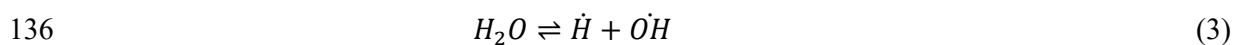
114 region include emulsification [15], homogenization [16], cell disruption [17] and  
115 polymerization [18].

116 The region between 100 - 1000 kHz is usually labelled *intermediate ultrasound*. This region  
117 results in only moderately intense bubble collapse, but importantly produces the most  
118 'sonochemically active' bubble population that results in highly efficient radical production.  
119 Koda et al. [19] and Mason et al. [20] have shown that peak radical production occurs  
120 somewhere between 400 - 800 kHz, although it also depends on the power applied and the  
121 physical and chemical properties of the fluid system. This intermediate ultrasound region is  
122 selected when chemical modification is the primary goal.

123 Above 1000 kHz, the physical effects of bubble collapse become relatively benign due to small  
124 bubble resonance size prior to collapse and a reduced proportion of bubbles undergoing  
125 cavitation due to an increased cavitation threshold. The cavitation threshold refers to the  
126 pressure (or size) above which bubble nucleation, a necessary precursor to cavitation, occurs.  
127 However, despite a reduction in cavitation, radical formation is still possible at frequencies  
128 around 1000 kHz [21] provided that sufficient energy intensity is employed. This regime,  
129 typically labelled the *diagnostic* or *megasonic* region, is used if only gentle physical effects are  
130 desired. It is particularly suitable for applications such as selective particle separation [22],  
131 where the aim is to also preserve the natural integrity of the separated product.

## 132 **2.2 Radical formation and sonochemistry**

133 The concentrated energy released during bubble collapse can split solvent/solute molecules  
134 that have diffused into the bubbles, to form radical species. For water, the following reaction  
135 may take place [23]:



137 That is, water molecules can be split into highly reactive hydrogen and hydroxyl radicals.  
138 Direct splitting of water as described above requires high temperatures resultant from high  
139 intensity bubble collapse typically seen only in the power ultrasound region. Alternate reaction  
140 pathways in water have been described by Yasui et al. [24] that allow for hydroxyl radical  
141 production even at relatively low bubble temperatures that are typical of intermediate  
142 ultrasound.

143 These radical species can be used to induce a whole range of redox reactions. As such,  
144 ultrasound can be used in organic synthesis reactions in which radicals are used to initiate and  
145 increase the reaction rates. Ultrasound has also been successfully used to increase yield by  
146 inducing modified reaction pathways that favor the formation of specific reaction products [4].

147 The radicals formed through cavitation can be used to initiate and accelerate the rates of free-  
148 radical polymerization and copolymerization [18], meaning that in some cases the use of  
149 ultrasound can obviate the need for a chemical initiator. Methyl methacrylate for example, can  
150 be polymerized by ultrasound without the use of an initiator [25], creating polymers with a  
151 molecular weight of up to 400,000 Da. Ultrasound-induced polymerization can however be  
152 complicated by the fact that the resulting polymers can also be simultaneously broken down  
153 by the intense shear forces resulting from the collapsing bubbles. These fragmented polymers  
154 may subsequently react to form side-products that may be quite different to conventional  
155 polymerization in the absence of acoustic cavitation.

156 Proteins are biological polymers that can be cross-linked to form larger networks and structures  
157 by ultrasound. These networks can be used to coat bubbles, droplets or other templates, forming  
158 rigid spheres that can be used for encapsulating materials for applications such as drug delivery  
159 [8]. The formation of protein microspheres by application of ultrasound results from a  
160 combination of shear-induced emulsification and radical formation [26, 27]. The high shear  
161 and temperature can partially unfold the proteins [28] which can then accumulate at the air or  
162 organic phase boundary and undergo cross-linking [27]. The protein cross-linking can be  
163 reversible, for instance through hydrophobic interactions and hydrogen bonding, or irreversible  
164 if covalent links are produced, for example disulphide bonds. In the latter case, this can  
165 potentially be facilitated by free radicals generated through ultrasonic cavitation [29].

166 Ultrasound can also be used to lower the temperature or pressure of some reactions, or reduce  
167 the requirement for solvents, which are expensive and often toxic. As such, ultrasonic synthesis  
168 techniques are often considered as 'green chemistry' alternatives for many applications [25].

### 169 ***2.3 Physical effects of ultrasound***

170 Physical modification of materials by ultrasound can arise from the shear forces generated  
171 during bubble collapse that are associated with pressure shockwaves, liquid microjets and  
172 acoustic streaming. Acoustic streaming is the propagation of disturbances in the fluid caused

173 by the ultrasonically induced oscillation of gas bubbles. This results in localized shear forces  
174 in the immediate vicinity of the bubbles. Liquid microjets result from the asymmetric and  
175 extremely rapid collapse of bubbles during cavitation, leading to unidirectional expulsion of  
176 high velocity jets into the surrounding fluid. Naude and Ellis [30] first hypothesized that the  
177 observed pitting of solid surfaces and particle size reduction of colloids on exposure to  
178 ultrasound was in fact due to the formation of microjets during asymmetric bubble collapse. It  
179 has since been shown that microjets with velocities in the order of 100 m/s can be formed [31],  
180 and that these can create pitting and erosion of surfaces [25]. Microjetting is also the primary  
181 cause of ultrasound-induced bulk mixing. Symmetric bubble collapse generates shockwaves  
182 that propagate radially outwards from the collapse point into the surrounding fluid. These  
183 shockwaves can be used to increase the rates of mass transfer across interfacial boundaries,  
184 enhancing the efficiency of multi-phase reactions [25]. All of these physical effects are  
185 commonly utilized in laboratory ultrasonic baths to facilitate cleaning of surfaces and  
186 dissolution of solids.

187 Although collapsing bubbles may reach temperatures of many thousands of degrees Celsius,  
188 these extreme temperatures are confined to small areas at the core of the collapsing bubble and  
189 near the bubble surface [32]. In the bulk solution, the increase in temperature resultant from a  
190 single bubble collapse is small. Nevertheless, the dissipation of heat from many cavitation  
191 bubbles, vibrating transducer surfaces, shockwave propagation and acoustic streaming can all  
192 contribute to incremental increases to the surrounding bulk temperature. If required,  
193 temperature control (e.g. a cooling jacketed reactor) can be used to prevent undesired  
194 temperature effects, e.g. denaturation of proteins in foods.

195 The physical forces resultant from cavitation can increase particle interactions in suspensions  
196 [25]. Solid powders suspended in fluids, may experience an increase to their momentum in the  
197 vicinity of a cavitation bubble, which can cause them to collide together with greater force than  
198 under quiescent conditions. Inorganic solids can be fractured and disrupted upon collision,  
199 leading to a reduction to their average particle size [25]. The minimum size achievable is  
200 dependent on characteristics of the solid, solvent and cavitation intensity. The lower limit is  
201 reached when the momentum of the particles become too small to create further impacts to  
202 cause particle fragmentation.

203 Ultrasound can be used to form encapsulating particles over a broad range of sizes, from around  
204 100 nm to 20  $\mu\text{m}$  in diameter. The particle size can be controlled to a large extent by selecting

205 appropriate sonication conditions such as power intensity, frequency and reactor configuration.  
206 The duration of processing [33], the type of reaction vessel [15] and sonifier used [34] are also  
207 variables that control the size of the particles that are formed. Importantly, in addition to being  
208 able to target a desired average particle size, ultrasound is able to produce particles with a  
209 narrow size distribution. For example, it has been shown that the use of a flow-through horn  
210 system could generate lysozyme coated nanospheres of very narrow size distribution ranging  
211 between 550-650 nm, compared with a larger 3 mm ultrasound horn that resulted in formation  
212 of particles with a broader range between 850-1200 nm [34]. Zhou et al. [35] also used high  
213 frequency ultrasound as a post-sonication technique to further narrow down the size  
214 distribution of ultrasonically-generated microspheres. By using 213 kHz ultrasound, lysozyme  
215 microspheres with a distribution of initially 0.5-4  $\mu\text{m}$  were narrowed to 0.5-2  $\mu\text{m}$  due to  
216 selective breakage of the larger microspheres by the ultrasound.

### 217 **3. Applications of ultrasonic encapsulation**

218 Ultrasonics can be used to promote specific functionality in different materials. For example,  
219 in foods comprising emulsions, the shelf-stability and physical appearance are dependent on  
220 the droplet size of the dispersed phase. Ultrasonics can be used to disperse different organic/oil  
221 phases into various aqueous phases in a controlled manner, to create emulsified products that  
222 are very shelf-stable and attractive in appearance [6]. Ultrasonics can also be used to promote  
223 the formation of polymer systems that are responsive to specific environmental conditions,  
224 such as pH and temperature. These polymer systems are useful for controlling drug release in  
225 biological systems. A combination of emulsification and polymerization can be promoted by  
226 ultrasound to form protein cross linkages, resulting in the formation of protein microspheres.  
227 These entities can be made biocompatible and biodegradable to enable their use as drug-  
228 delivery vehicles. This section will describe the effect of ultrasound on important functional  
229 properties of food emulsions (3.1), polymer particles for controlled drug release (3.2), and  
230 protein microspheres (3.3).

#### 231 **3.1 Functional food emulsions**

232 Droplet size and polydispersity are key attributes that govern the functionality and stability of  
233 emulsions. The intense shear forces generated during ultrasonic cavitation can be used to create  
234 emulsions with very small and relatively uniformly sized droplets [36]. Two mechanisms are  
235 responsible for the emulsification effect of ultrasound. First, the application of the sound field

236 produces interfacial waves, which become unstable resulting in the dispersion of the oil phase  
237 into the continuous water phase as mid- to large-sized droplets. Secondly, the physical effects  
238 resultant from cavitation break up these initially formed droplets of dispersed oil into droplets  
239 of sub-micron size [25].

240 Ultrasonics is particularly useful for the production of, for example, food emulsions. In food  
241 emulsions, the size of the emulsified droplets influences its visual appearance, mouth-feel and  
242 shelf-life stability among other things [6]. Whereas large sized emulsion droplets are  
243 characterized by a ‘milky’ opaque appearance, emulsions with emulsion droplet size (EDS)  
244 smaller than  $\sim 100$  nm, can appear translucent and almost clear [15] due to the reduction in  
245 light scattering by the smaller droplets.

246 While emulsions are inherently thermodynamically unstable, when the droplets are smaller  
247 than  $\sim 100$  nm they become kinetically stable [37]. At these sizes, the Brownian motion of the  
248 droplets overcomes the natural buoyancy force of the droplets to rise and cream. The instability  
249 is dependent instead on colloidal forces such as Ostwald Ripening [38] and droplet-droplet  
250 collisions that leads to coalescence and eventual phase separation. These are usually very slow  
251 processes, such that nano-sized emulsions are shelf-stable for many months [39].

252 The formation of nano-sized droplets requires the application of strong shear forces to break  
253 apart the liquid droplets. This is typically achieved using low frequency ultrasound in the  
254 *power ultrasound* region (20-100 kHz) delivered at high energy intensity  $> 10$  W/cm<sup>2</sup>. The  
255 emulsified droplets then need to be stabilized by a surfactant in the system, to prevent  
256 spontaneous phase separation by coalescence [15]. Midsonic and megasonic frequencies  $> 400$   
257 kHz are usually ineffective at forming emulsions, due to insufficient physical shear forces  
258 arising from the collapse of smaller resonance size bubbles at these frequencies.

259 Higher frequency ultrasound (midsonic to megasonic) has however been reported useful at  
260 forming nano-size emulsion droplets when applied following low frequency ultrasound  
261 through a process known as *tandem acoustic emulsification* [39, 40]. Oleic acid/water nano-  
262 emulsions were prepared by Kamogawa et al. [40] using this technique, while Nakabayashi et  
263 al. [39] also reported the production of transparent emulsions of ethylenedioxythiophene  
264 (EDOT) monomer formed by sequential emulsification at 20 kHz, 1.6 MHz and 2.4 MHz  
265 ultrasound. The nano-emulsions formed by Nakabayashi et al. were stable even in the absence  
266 of additional surfactant for 1 to 2 years. These nanoemulsions are not only stable and attractive

267 in appearance, but can be used to conduct direct electropolymerization (see Section 3.2) in the  
268 absence of additional surfactant [41].

269 It was proposed that the small droplets achieved in tandem acoustic emulsification upon  
270 application of higher frequency ultrasound was not due to destructive shear forces such as  
271 microjets and shockwaves prominent at low frequency ultrasound. Instead, it is due to the  
272 enhanced acceleration of solvent and the emulsion droplets caused by acoustic radiation forces  
273 and acoustic streaming [40] such that they collide together and break apart into smaller  
274 droplets. These acceleration forces become stronger with increased frequency, and the  
275 sequence in which the different frequencies of ultrasound are applied is noted to be important,  
276 with reversal of the order (i.e. high frequency followed by low frequency) resulting in  
277 ineffective emulsification.

278 Oil-in-water and water-in-oil emulsions can both be produced successfully using the tandem  
279 emulsification technique. Although the application of tandem acoustic emulsification has yet  
280 to be reported for food applications, it would be an attractive avenue for creating nano-sized  
281 surfactant-free emulsions.

282 For foods, the EDS plays a significant role in the sensory characteristics [42]. In general,  
283 emulsions containing smaller droplets have a higher viscosity [42] which are purported to  
284 provide improved sensory properties such as ‘creamier’ mouthfeel [6] in a range of products  
285 such as cheese [43] and creams [44].

286 Emulsions can be used to load hydrophobic or amphiphilic materials with biological  
287 functionality or nutritional benefit into an aqueous fluid. In the case of amphiphilic materials,  
288 it may be desirable to maximize the surface area of the droplets by reducing the emulsion  
289 droplet size. Smaller droplets are also better at retaining a larger amount of volatile material  
290 within the oil phase of an emulsion during spray drying for the production of encapsulated  
291 microparticles [45]. This is because the smaller emulsion droplets are less likely to be broken  
292 apart by the atomizer within the spray dryer [45]. Spray dried encapsulating microparticles can  
293 be used to create products that are able to mitigate the release of undesirable odors or smells  
294 e.g. fish oil powders.

295 Another way to encapsulate materials in emulsions is to create what is known as a double  
296 emulsion. Double emulsions are emulsions entrapped within emulsions. Their capacity to  
297 encapsulate aqueous components within oil droplets makes them promising delivery vehicles

298 for bioactives, for flavour masking and for fat reduction in foods [46]. The entrapped inner  
299 phase is protected from degradation by environmental factors in the external phase, and release  
300 of inner material can be delayed until it enters the digestive system, thereby masking potentially  
301 undesirable flavours. Fat reduction can be achieved without compromising the sensory properties  
302 of the fat phase by displacing fat without reducing the apparent volume fraction of the fat  
303 droplets.

304 There is significant commercial interest, with a large number of examples having been  
305 developed for the production of flavour-enhanced and reduced-fat salad dressings [47], and  
306 also reduced-fat cheese [48-50]. Instability is a potential issue for using double emulsions in  
307 food applications. Rapid phase separation can arise due to the relatively large droplets  
308 (typically greater than 20  $\mu\text{m}$ ) [51] that are formed at the low shear rates which are required to  
309 avoid release of the entrapped material. This issue may be resolved to an extent by use of  
310 ultrasonication as reported by Tang et al. [52, 53]. Ultrasonication was successfully used to  
311 form double emulsions of sub-micron size range for the purpose of aspirin encapsulation,  
312 achieving both high stability (1 month prolonged storage) and entrapment yield (up to 99%  
313 encapsulation) [53].

314 The use of ultrasonics has been compared with most conventional and state of the art  
315 emulsification techniques. Some of the more common methods applied in industrial  
316 emulsification are rotor-stator systems [54] and high pressure homogenization [55]. In addition  
317 to conventional high pressure homogenization, a modified technology known as the  
318 Microfluidizer<sup>TM</sup> involves impinging two pressurized streams against each other. The  
319 Microfluidizer<sup>TM</sup> (MF) has been shown to be highly effective at nano-emulsion preparation  
320 [56-58]. MF has relatively high energy efficiency for producing emulsions with very small and  
321 narrowly distributed EDS [57] and is commonly used in the pharmaceutical industry to make  
322 nano-emulsions. Madhi Jafari et al. [57] have compared emulsion preparation using US at  
323 matched specific energies with MF, and found comparable performance. It was found that  
324 when using matched 20 kJ/kg energy input, particle size reduction by MF achieved mean  
325 volume-weighted particle size of 0.83  $\mu\text{m}$  compared with 1.02  $\mu\text{m}$  for ultrasonication at 20  
326 kHz.

327 Emulsification by ultrasonication and MF occurs via common causal mechanisms including  
328 cavitation and shear [58]. Although MF has been noted to be superior in size reduction and  
329 generating emulsions with more narrow size distributions, ultrasonication is deemed to be

330 significantly easier to operate, clean and maintain [57]. With extended duration of processing,  
331 ultrasonication has been shown to be able to achieve comparably small emulsion droplets to  
332 MF [58]. Leong et al. [15]. demonstrated the capability of ultrasound to produce emulsions  
333 with comparable particle size to microfluidization, provided that the energy density and  
334 surfactant system was optimized.

335 Typically, the formation of emulsions requires a large amount of surfactant to cover the newly  
336 formed surfaces and hence stabilize the dispersed droplets. The tandem-emulsification  
337 technique reported by Nakabayashi et al. [39] and Kamogawa et al. [40] is exceptional in that  
338 ultrasound can produce nano-sized emulsions in the absence of surfactant, although the  
339 technique is yet to be proven for a wide range of oils. Recently, Shanmugam et al. [59] have  
340 shown that ultrasonic emulsification can be used to create stable food-based emulsions of flax  
341 seed oil directly in skim-milk without the requirement for additional surfactants. The native  
342 milk proteins are partially denatured (less than 1%) by the ultrasound and allowing them to  
343 effectively coat the formed oil droplets, stabilizing the emulsion for at least 7 days.  
344 Emulsification could not be achieved in the absence of ultrasound even when using matched  
345 applied energies in a rotor-stator system, suggesting the importance of acoustic cavitation to  
346 the stabilization process.

### 347 **3.2 Formation of polymeric particles for controlled drug release**

348 In the treatment of certain diseases, drugs must be delivered at rates corresponding to the  
349 physiological needs of the patient. In conventional drug delivery, the concentration of the drug  
350 within the patient's blood stream rises, peaks then declines. Each drug has a different  
351 concentration above which it becomes toxic and below which it is rendered ineffective.  
352 Controlled drug release is desirable in treating certain illnesses, as it enables maintenance of a  
353 drug within a desired therapeutic range with a single dose that is responsive to the needs of the  
354 patient.

355 Polymer-based materials that are sensitive to environmental factors such as temperature, pH  
356 and ionic strength, have potential to be used as responsive drug delivery vehicles [7]. If the  
357 structure of the polymer can be externally regulated (e.g. by magnetic, ultrasonic, thermal and  
358 electric stimulation) or self-regulated (i.e. by changing environmental conditions), it is possible  
359 to release the entrapped drug in a controlled manner. Ultrasonics has been investigated as a  
360 tool to assist in synthesizing such polymers with a range of functionality. For a more detailed

361 review of ultrasonically enhanced synthesis of polymers, readers are invited to read the review  
362 by Price [18].

363 As described in Section 2, acoustic cavitation leads to both chemical and physical phenomena  
364 that can be controlled to create polymers with improved rates of reaction and more defined  
365 characteristics such as molecular weight. Radical polymerization is one of the most studied  
366 [18] sonochemically-enhanced polymerization processes. The radicals formed during acoustic  
367 cavitation can be used to initiate the polymerization process in place of conventional initiators  
368 [60, 61]. A particular system in which this has been successful is in vinyl monomers such as  
369 methyl methacrylate [62]. Another notable advantage of generating radicals using ultrasound  
370 is the ability to perform the reaction at reduced temperatures (i.e. between -10 to 60 °C  
371 compared to between 50 to 100 °C for more conventional radical polymerization reactions of  
372 PMMA).

373 The physical effects of ultrasound can be used to control various properties of the resulting  
374 polymer. The intensity of the ultrasound applied, which influences the strength and number of  
375 acoustic cavitation bubbles, is one variable that can be modulated to control the yield of  
376 polymer produced as well as the final molecular weight of the resulting polymer [63]. A larger  
377 number of collapsing bubbles creates more radicals, which can increase the frequency of  
378 polymer initiation events. Simultaneously, the shearing forces resultant from the collapse of  
379 bubbles can break apart some of the long polymer chains that are formed, effectively reducing  
380 the molecular weight of the final polymer. The intensity of collapse, and the duration over  
381 which sonication is applied, can control the molecular weight of the polymers [63]. The ability  
382 to influence the polymerization process by manipulating ultrasound variables provides scope  
383 to transform conventional polymerization processes to form new materials. It should be noted  
384 though that sonication has little to no effect on the actual propagation reaction of the polymer  
385 formation. The main effects are largely confined to the initiation process and subsequent chain  
386 breakage of the formed polymer chains.

387 Ultrasound can promote emulsion polymerization to form latex particles of approximately  
388 equal size to the emulsion droplets [64]. The latex particles formed by ultrasound are typically  
389 smaller than those formed by conventional emulsion polymerization, resulting in an increased  
390 surface area [65]. Further, the use of ultrasound removes the need for chemical initiators or co-  
391 stabilizers, reduces the required reaction temperature, increases the rate of polymerization and  
392 results in higher monomer conversion and potentially higher molecular weights. As a number

393 of these advantages reduce chemical and energy consumption, ultrasonic emulsion  
394 polymerisation can be considered a 'green' alternative to conventional polymerization  
395 reactions. A proposed mechanism for ultrasonically-promoted emulsion polymerization is  
396 presented in Figure 2.

397 Ultrasonic emulsion polymerization has also been used to create latex coated magnetic  
398 nanoparticles using a simple, one-step method [66]. The particles exhibited colloidal stability  
399 for up to 12 months with no observed deterioration, and strong magnetic properties. The  
400 suspensions behaved as conventional magnetic fluids in their response to a magnetic field.

401 Temperature responsive polymers of poly(N-isopropylacrylamide) and poly(N-  
402 vinylcaprolactam) have been prepared using ultrasound, and their swelling behavior in the  
403 presence of different concentrations of surfactant (SDS) studied [67]. The potential for these  
404 types of polymers to be used as drug delivery vehicles was demonstrated by their ability to  
405 entrap rhodamine B dye. The structure of these polymers is temperature dependent. At 20 °C,  
406 the polymer chain is an expanded coil that is soluble in water resulting in clear aqueous  
407 solutions. When heated to 32 °C, the chains collapse to a globular structure which decreases  
408 the solvation properties causing the polymer solution to become turbid. The release kinetics of  
409 the entrapped rhodamine B was consequently found to be dependent on the temperature, with  
410 higher release rates occurring at 40 °C compared with 20 °C [67]. The release of dye was found  
411 to follow Fickian diffusion kinetics, with the diffusion coefficients being  $4 \times 10^{-12}$  and  $3.6 \times$   
412  $10^{-11}$  m<sup>2</sup>/s at 20 °C and 40 °C respectively i.e. an order of magnitude increase in release rate.  
413 There was an apparent limit to the maximum amount of dye release (approximately 62 %),  
414 which was attributed to the concentration gradient of the dye within the polymer and bulk  
415 solution approaching zero.

### 416 **3.3 Formation of protein-coated microspheres**

417 Ultrasonics have been used to prepare protein microspheres, which have a wide range of  
418 potential biomedical applications including acting as echo contrast agents for sonography and  
419 magnetic resonance imaging, and as vehicles for drug delivery [4]. These microspheres  
420 (typically several  $\mu\text{m}$  in diameter) consist of a protein shell surrounding a core which can be  
421 either a gas or a liquid. The solid shell is a barrier to permeation between the interior phase and  
422 the aqueous exterior, conferring long term storage stability to the protein microspheres.

423 Protein microspheres are formed by a combination of two acoustic phenomena: emulsification  
424 and cavitation. In the emulsification stage bubbles or liquid droplets are created, which acts as  
425 a 'template' for the protein shell to form around. The radicals produced by acoustic cavitation  
426 lead to the formation of superoxide species which promote the formation of intermolecular  
427 disulphide crosslinks between the proteins covering the interface. It has been shown that  
428 emulsification alone (via vortex mixing) was not sufficient to produce long-lived  
429 microbubbles, indicating that chemical cross-linking arising from ultrasonic cavitation was  
430 required to produce stable protein microbubbles [26].

431 One of the first commercially available protein microspheres were albumin-coated  
432 microbubbles, marketed under the name Albunex® and Optison™ [8]. These microspheres  
433 have been used primarily as contrast agents for ultrasound imaging, with the air-filled core  
434 providing enhanced signal response.

435 Protein microspheres can be used as drug delivery vehicles with therapeutic agents either  
436 loaded on the surface of air-filled protein microspheres, or if liquid filled, entrapped within the  
437 liquid phase of the protein microsphere. Of importance for drug delivery, they are inherently  
438 biodegradable and likely to be more biocompatible than microsphere made from synthetic  
439 polymers. They can also be functionalized with ligands (e.g. antibodies, peptides or vitamins)  
440 to target specific entities within the body.

441 To achieve further functionality, proteins may be used that infer useful biological properties to  
442 the formed microspheres. Cavalieri et al. [27] first reported the formation of lysozyme protein  
443 microspheres which were stable for several months using a sonochemical approach. Lysozyme,  
444 derived from hen egg white, has natural anti-microbial properties and the microspheres formed  
445 from lysozyme were found to retain some of the enzymatic functionality and anti-microbial  
446 activity of the native protein. This work by Cavalieri et al. [27] confirmed the need to release  
447 free thiol groups via partial protein denaturation in order to initiate crosslinking required to  
448 stabilize protein microspheres (Figure 3). Alternatively, the microbubbles can be used as a  
449 carrier for antibiotics. Avivi et al. [68] encapsulated tetracycline into bovine serum albumin-  
450 coated microspheres using a sonochemical approach, and found that up to 65% encapsulation  
451 efficiency could be achieved. Importantly, it was found that the majority of the encapsulated  
452 tetracycline, approximately 97%, was loaded within the core of the bubble and not simply  
453 adsorbed to the surface of the protein microspheres. Avivi et al. [68] confirmed that the

454 antimicrobial activity of the entrapped tetracycline when released by gentle heating was  
455 identical to equivalent amounts of free tetracycline when applied to different strains of bacteria.

456 Zhou et al. [69] used the same approach to create liquid-encapsulating lysozyme microspheres  
457 loaded with various oils (sunflower oil, tetradecane, dodecane and perfluorohexane). Liquid-  
458 filled microspheres can theoretically be loaded with significantly larger quantities of oil-soluble  
459 drugs, than air-filled bubbles where the active drug component needs to be functionalized on  
460 the surface of the bubble. The type of liquid encapsulated in the microspheres was found to  
461 influence the physical properties (i.e. size, polydispersity, and shell wall strength) of the formed  
462 microspheres.

463 The approach used to synthesize lysozyme microspheres can be extended to the synthesis of  
464 protein-mimicking polymer-coated microspheres. Cavalieri et al. [29] first reported a one-step  
465 sonochemical process to synthesise microspheres made from synthetic thiolated polymers of  
466 polymethacrylic acid. Important physical properties of the formed microspheres could be  
467 controlled by adjusting the thiol content in the macromolecules. The size, surface roughness,  
468 and shell thickness were all found to increase with increasing number of thiol groups in the  
469 monomer backbone. Recent work has further demonstrated the versatility of this method with  
470 the fabrication of new types of microspheres, including chitosan/titanium dioxide hybrids [70].  
471 These hybrid microspheres have composite properties including high mechanical strength and  
472 antibacterial activity.

473 The size of the active sonochemical region delivering the ultrasound has been shown to effect  
474 the size distribution of formed microspheres, offering a means of controlling size [34]. In this  
475 study, a novel flow-through sonication horn with a very small active sonochemical region  
476 created smaller and more monodisperse microspheres than larger diameter horns with larger  
477 sonochemical regions.

478 In addition to promoting the formation of protein microspheres, ultrasound is a potential tool  
479 for targeted drug release. Ultrasound has been shown to break apart chitosan/titanium oxide  
480 hybrid nanospheres, releasing the entrapped contents [70]. This ability could potentially be  
481 used to induce rupture of protein microspheres to increase the localized delivery of a drug to  
482 specific parts of the body.

## 483 **4 Industry application**

484 Ultrasonics has been successfully used to generate a range of functional food and biomaterials  
485 in the laboratory. Industrial uptake of ultrasonics is not currently widespread, but is gaining  
486 considerable traction. A number of potential issues identified in early studies are gradually  
487 being debunked or resolved. Some of these issues will be discussed in the following section.

### 488 **4.1 Generation of particulate metal contaminants**

489 The strong physical phenomena generated by cavitation are capable of affecting not just the  
490 product, but also the transducer and reactor surfaces. The potential for release of metallic  
491 particles into the product that may be too small to remove has raised some health concerns.  
492 Recently however, Mawson et al. [71] assessed the production of metal particulates from  
493 ultrasonic transducers and found no evidence for the formation of harmful nanoparticles (<80  
494 nm). In their study, no nano-particulate material was observed even after prolonged exposure  
495 (up to 7.5 hours) to high intensity ultrasound (20 kHz and 174 J/mL). However, most food or  
496 drug related applications involving ultrasound only require a few second of ultrasonic  
497 processing [72], meaning the risk of contamination of sensitive products such as foods and  
498 pharmaceuticals is minimal. Contamination-free reactors [73] have also been developed, and  
499 these can be used for the production of high-valued products that require the utmost purity.

### 500 **4.2 Degradation of functional properties**

501 The formation of radicals by ultrasonic cavitation can be beneficial in promoting and enhancing  
502 sonochemical reactions, but they can also potentially degrade redox sensitive components. This  
503 is a particular concern for foods and bioproducts, where the flavor and nutritional properties  
504 must be taken into account. Vitamins, fats, and other lipids are particularly susceptible to  
505 reactions induced by oxidative radicals. Fatty acid oxidation and lipolysis can significantly  
506 modify the flavor profile of the food [74] and produce off-flavors associated with ‘burnt  
507 rubber’, ‘grass’ or ‘rancid fruit’. Destruction of vitamins and anti-oxidants in foods may reduce  
508 the nutritional value or induce undesirable color changes.

509 However, these problems can be mitigated by selecting operation at lower frequency ultrasound  
510 and employing shorter exposure times [72]. As can be observed in Figure 4, radical formation  
511 at 20 kHz has been shown to be minimal relative to mid and high frequency ultrasound, and at  
512 short sonication times essentially negligible and unlikely to result in any significant change to

513 the functionality of biomaterials. For instance, for a solution of 5 % protein, the radical  
514 concentration resultant from sonication at 20 kHz for 5 min would typically be approximately  
515 less than 10 ppm (i.e. 10 moles of radical per million moles of protein). A recent study  
516 performed by Juliano et al. [75] showed that by limiting the duration of sonication (i.e. low  
517 specific energy), off-flavor volatiles from oxidation of milk fat could be kept below detectable  
518 sensory thresholds, even when operating with mid (400 kHz) or high (1 MHz) frequency  
519 ultrasound. The reason is because many natural food products such as milk contain natural anti-  
520 oxidants, which mitigate detrimental changes to the product.

#### 521 ***4.3 Augmentation or replacement of current industrial techniques***

522 Ultrasound can be used to replace or complement conventional techniques such as  
523 emulsification and polymerization that are used to create encapsulating particles in aqueous  
524 systems. Acoustic cavitation can provide efficient high-shear processing and radical  
525 polymerization as described in sections 3.1 and 3.2. However, ultrasound cannot replace  
526 industrial techniques currently used to create dried encapsulating particles, for example, spray  
527 drying, freeze drying, extrusion coating, fluidized bed coating, and coacervation [76]. Although  
528 studies have employed ultrasonic atomization as a technique to generate microcapsules at lab  
529 scale [77], spray drying is the most feasible method on an industrial scale, particularly for foods  
530 [76, 78]. Spray drying is highly energy efficient and effective technique by which internalized  
531 materials can be stabilized during storage. Emulsions can be formed as a precursor to  
532 encapsulate lipids and oil-soluble material during spray drying. Ultrasound can play a  
533 complementary role in aiding the formation of stable small-sized emulsions which may have  
534 beneficial outcomes within the spray dried product [45].

535 Hydrodynamic cavitation [79, 80], imparts similar cavitation-borne mechanisms to materials  
536 as ultrasonic cavitation, with the advantage of higher throughput due to its more conventional  
537 unit design. In some situations, hydrodynamic cavitation can be more efficient due to its ability  
538 to generate cavitation over a large volume/region [79].

#### 539 ***4.4 Towards scale-up***

540 Ultrasonic technology is yet to be widely implemented at an industrial scale, not due to  
541 uncertainty of its efficacy, but to challenges in scaling up. For a comprehensive review of the  
542 design considerations for efficient scale up of sonochemical reactors readers are directed to

543 Gogate et al. [81]. A key issue for scale-up of sonoprocessing that is worth highlighting is the  
544 strong attenuation in effectiveness with distance from the ultrasound source. This complicates  
545 scale-up as the effective volume is confined to the active sonochemical regions close to the  
546 transducers which, in some cases, can be quite small and narrow. One strategy is to use flow-  
547 through cells, where liquid passes through a narrow region close to the transducer to ensure all  
548 elements of fluid are subjected to ultrasound. Alternatively, flow chambers incorporating  
549 multiple transducer horns can be effective at providing uniform delivery of ultrasound to large  
550 volumes of material. Many commercial flow-through sonication products are now available,  
551 and can be tailored for a range of applications.

552 The current cost of the technology, although not prohibitive, is still often higher than  
553 conventional alternatives. However, the ability of ultrasound to produce unique, high-value  
554 products with improved functionality while reducing chemical and energy consumption in  
555 some applications, may compensate for the extra cost. It is envisioned that the continual  
556 development of the technology will lead to gradual industrial uptake of ultrasonics and  
557 eventually its mainstream use for the production of valuable functional materials.

## 558 **5. Conclusion**

559 The ultrasonic synthesis of functional food and bio-materials has a bright future with many yet-  
560 to-be realized commercial opportunities. Many of the issues identified in early studies are being  
561 overcome, paving the way for ultrasonic synthesis of the next generation of drug delivery  
562 agents, functional biomaterials and food products.

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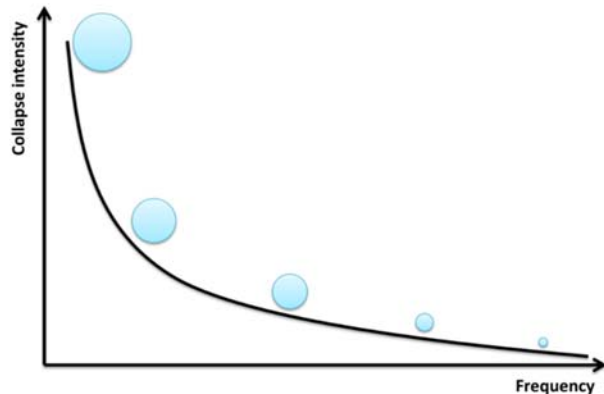
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768 **Figures**

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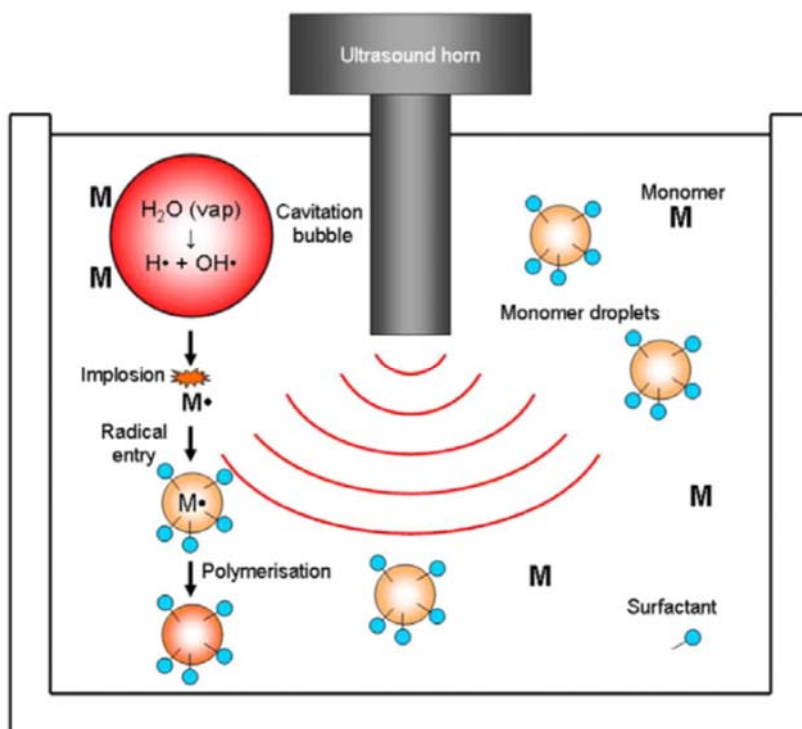


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771 Figure 1: Schematic representation of the relationship between ultrasonic frequency applied  
772 and the relative intensity and size of the collapsing bubbles. Not drawn to scale.

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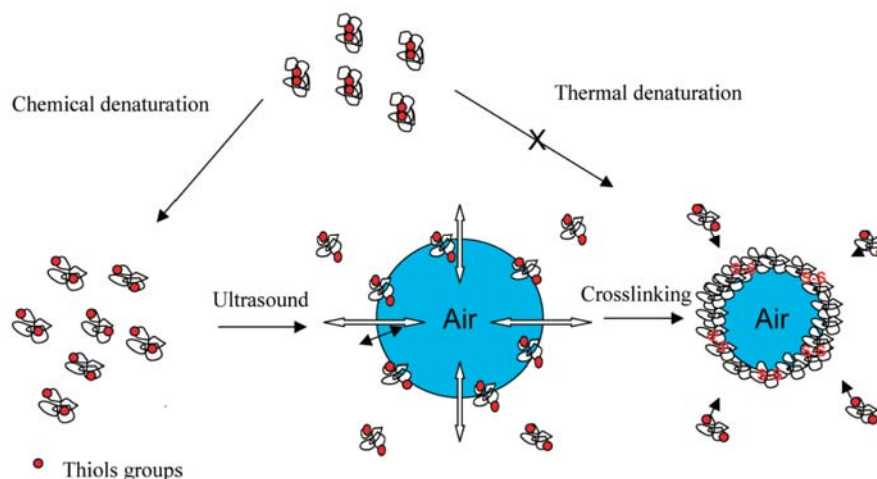


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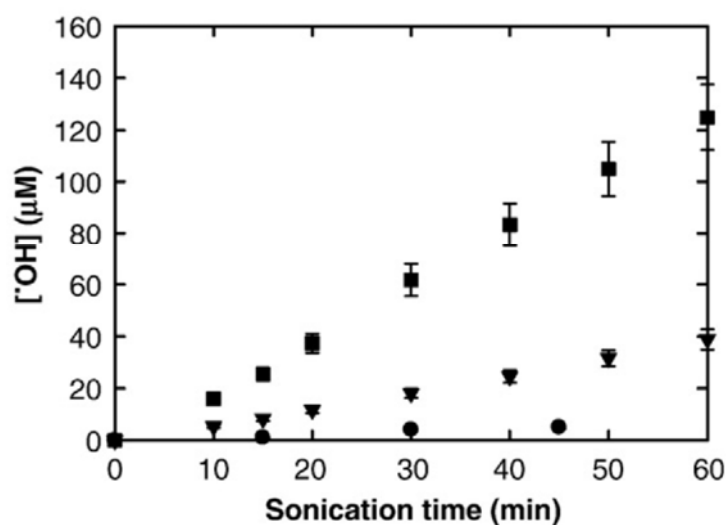
775 Figure 2: A schematic diagram of a proposed emulsion polymerization process. Reprinted  
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Figure 3: Mechanism proposed for the formation of lysozyme protein microspheres. Reprinted with permission from Cavalieri et al [27]. Copyright 2008 American Chemical Society.



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Figure 4: OH radical yield generated in water upon sonication at different acoustic frequencies with matched power  $0.9 \text{ W cm}^{-2}$ . Adapted from Ashokkumar et al. [72], Copyright 2008, with permission from Elsevier.